

RESEARCH ARTICLE

# Norovirus and Sapovirus Epidemiology and Strain Characteristics among Navajo and Apache Infants

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## Abstract

Norovirus and sapovirus are important causes of acute gastroenteritis (AGE) among American Indian infants. We investigated the prevalence and molecular epidemiology of norovirus and sapovirus in American Indian infants who have historically experienced a high burden of AGE compared to other US populations. Stool samples were collected from 241 children with AGE (cases) and from 343 infants without AGE (controls)  $\leq 9$  months of age from 2002–2004. Cases experienced forceful vomiting and/or 3 or more watery or looser-than-normal stools in 24 hours. Stools were tested by real-time RT-PCR for norovirus GI, GII and GIV and sapovirus GI, GII, GIV and GV. Positive samples were genotyped after sequencing conventional RT-PCR products. Norovirus was identified in 76 (31.5%) of the cases and 70 (20.4%) of the controls ( $p < 0.001$ ). GII.3 and GII.4 Farmington Hills were the most frequently identified genotypes in 14.5% and 30.3% of cases and 17.1% and 27.1% of controls, respectively. Sapovirus GI and GII genotypes were identified in 8 (3.3%) of cases and 8 (2.3%) of controls and a single GIV virus was detected in a control. The same norovirus and sapovirus genotypes were circulating in the general U.S. population in the same time period. The high detection rate of norovirus in healthy controls suggests significant asymptomatic transmission in young infants in these communities.

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## Introduction

Norovirus and sapovirus are genetically diverse genera in the family *Caliciviridae*. The major structural capsid protein (VP1) sequence is used to classify both norovirus and sapovirus into genogroups and genotypes. Norovirus is organized into at least seven genogroups (GI–GVII) of which GI, GII and GIV viruses are detected in humans [1]. Of the eight reported sapovirus genogroups (GI–GVIII), viruses of four (GI, GII, GIV and GV) have been detected in humans [2].

In the United States (US), norovirus causes an estimated 19–21 million cases of acute gastroenteritis (AGE) resulting in 1.7–1.9 million outpatient and 400,000 emergency department visits annually [3]. In addition, norovirus causes 56,000–71,000 hospitalizations and 570–800

deaths per year in the US, the majority occurring among young children and the elderly [3]. With the application of real-time RT-PCR diagnostics, sapovirus has been increasingly implicated as the cause of outbreaks [4, 5] and sporadic AGE [6–8]. In pediatric AGE, sapovirus detection ranges from 5–17% depending upon the country [6–10].

Norovirus and sapovirus are important causes of AGE among American Indian infants [11] who have historically experienced a higher burden of AGE compared to children of the general US population [12]. The aim of this analysis is to establish the prevalence and molecular characteristics of the norovirus and sapovirus genotypes circulating among American Indian infants who were selected for a case-control study of AGE etiology [11]. The infants selected for the case-control study were participants of the placebo arm of a rotavirus vaccine efficacy trial that was conducted in two American Indian communities in the Southwest US from 2002–2004 [13].

## Materials and Methods

### The study

Stool samples from American Indian infants  $\leq 9$  months of age that had been stored at  $-80^{\circ}\text{C}$  since the efficacy trial were selected for the case-control study and tested for adenovirus group F, astrovirus types 1–8, norovirus GI, GII and GIV, group A rotavirus and sapovirus GI, GII, GIV and GV [11]. Case and control inclusion criteria were described previously [11].

### Detection and genotyping of norovirus and sapovirus by real-time RT-PCR

Detection of noroviruses and sapoviruses by real-time RT-PCR has been described in detail elsewhere [11]. Positive specimens were genotyped by conventional RT-PCR [14, 15] and PCR products of appropriate size (norovirus GI: 330 base pairs, norovirus GII: 344 base pairs, sapovirus: 434 base pairs) were gel-purified using the QIAquick Gel Extraction Kit (Qiagen). Sanger sequencing was performed using the Big Dye Cycle Sequencing Kit (Applied Biosystems) and sequences (available upon request) were genotyped by comparison to norovirus and sapovirus reference strains in the database of the National Calicivirus laboratory at CDC. Phylogenetic analyses were performed using MEGA (version 6.0) and statistical analyses were performed in Stata (version 13).

The Institutional Review Boards of the Navajo Nation, Phoenix Area Indian Health Service, Centers for Disease Control and Prevention and Johns Hopkins Bloomberg School of Public Health approved this research, as did the Navajo and White Mountain Apache tribes. Parents provided written consented for participation of their infants in the original rotavirus vaccine trial.

## Results

Norovirus or sapovirus was detected in 84 (35%) of 241 cases and in 79 (23%) of 343 controls. Co-infections of norovirus or sapovirus with another virus were common (Table 1).

Norovirus GII viruses were detected in 71 (29.5%) of cases and in 62 (18.1%) of controls ( $p < 0.001$ ; Table 2). Between 2–3% of cases and controls were positive for norovirus GI.

Norovirus genotypes GII.3 and GII.4 predominated among cases (GII.3:  $N = 11$  [14.5%], GII.4:  $N = 23$  [30.3%]) as well as among controls (GII.3:  $N = 12$  [17.1%], GII.4:  $N = 19$  [27.1%]). Among GI viruses, GI.3b and GI.7 were most prevalent (Table 3).

Most sapovirus strains belonged to GI (53%) or GII (41%) whereas one GIV virus was detected in a control and no GV viruses were detected (Table 4).

**Table 1. Percent of enteric virus co-infections with norovirus or sapovirus.**

Co-infecting virus	Norovirus, N (%)		Sapovirus, N (%)	
	Case (N = 76)	Control* (N = 70)	Case (N = 8)	Control* (N = 9)
Astrovirus	7 (9.2)	7 (10.0)	0 (0)	2 (22.2)
Adenovirus	3 (3.9)	3 (4.3)	2 (25.0)	0 (0)
Rotavirus	8 (10.5)	9 (12.9)	3 (37.5)	2 (22.2)

\* There was one control co-infected with norovirus and sapovirus.

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## Discussion

We report the prevalence and genotype distribution of norovirus and sapovirus in American Indian infants of the US Southwest from 2002–2004. The prevalence of norovirus in cases (31.5%) was approximately twice as high compared to the estimated global prevalence of 18% [16]. The high prevalence of norovirus found in healthy controls (20.4%) is within the range (4–27%) observed in other pediatric AGE studies [17, 18–21]. Prolonged viral shedding or an altered gut microbiota have been suggested as possible explanations of high detection rates among control subjects [22, 23].

Norovirus GII.3 and GII.4 were the primary genotypes identified among American Indian infants. These genotypes have been reported globally but also vary geographically [17, 24–26]. Several studies from Africa suggested a higher prevalence of GII.3 viruses in children [24–26], but not in another study in young children the U.S. [17]. All GII.4 viruses detected in the American Indian children typed as GII.4 Farmington Hills, a variant that emerged in 2002 [27] and became the predominant strain globally [28].

This study had several limitations. The stools from this study were collected 11–13 years ago in the context of a clinical trial for a rotavirus vaccine candidate. Therefore, the characterized genotypes may not reflect the current distribution of norovirus and sapovirus circulating among American Indian infants in the US Southwest. The high prevalence of norovirus and sapovirus in healthy controls creates difficulty in the attribution of AGE etiology and dampens the association between virus detection and disease and future studies should focus on whether prolonged shedding could explain the high percentage of positives in healthy control children.

Norovirus GII.3 and GII.4 genotypes predominated among Navajo and White Mountain Apache infants in the early 2000's as well as in norovirus outbreaks and among other populations in the U.S. around that same time [27, 29]. A similar distribution of norovirus genotypes among cases and controls suggest significant transmission within the population accompanied by asymptomatic carriage and shedding of the virus. Only a few different sapovirus genotypes were detected among American Indian infants. The frequent number of co-infections in

**Table 2. Norovirus and sapovirus detected in cases and controls.**

Virus	Cases, N = 241		Controls, N = 343	
	N (%)*	N not mixed (%)	N (%)*	N not mixed (%)
Norovirus	76 (31.5)**	59 (24.5)**	70 (20.4)	50 (14.6)
GI	6 (2.5)	6 (2.5)	10 (2.9)	7 (2.0)
GII	71 (29.5)**	54 (22.4)**	62 (18.1)	44 (12.8)
Sapovirus	8 (3.3)	4 (1.7)	9 (2.6)	4 (1.2)

\* Includes some cases or controls with mixed viral detection of Group F adenovirus, astrovirus or rotavirus.

\*\* Proportion of cases positive statistically higher than controls (p<0.001).

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**Table 3. Norovirus genotype distribution among cases and healthy controls.**

Genotypes	Frequency (%)	
	Cases, N = 76	Controls, N = 70
GI genotypes		
GI.3b	3 (4.0)	1 (1.4)
GI.7	3 (4.0)	6 (8.6)
Not typed	--	3 (4.3)
GII genotypes		
GII.2	1 (1.3)	1 (1.4)
GII.3	11 (14.5)	12 (17.1)
GII.4 Farmington Hills	23 (30.3)	19 (27.1)
GII.5	8 (10.5)	9 (12.9)
GII.6	7 (9.2)	1 (1.4)
GII.7	5 (6.6)	5 (7.1)
GII.12	1 (1.3)	1 (1.4)
GII.14	--	1 (1.4)
GII.17	1 (1.3)	--
Not typed	14 (18.4)	13 (18.6)

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**Table 4. Sapovirus genotype distribution among cases and healthy controls.**

Genotypes	Frequency (%)	
	Cases, N = 8	Controls, N = 9
GI genotypes		
GI.1*	6 (75)	2 (22.2)
GI.2	--	1 (11.1)
GII genotypes		
GII.1*	2 (25)	2 (22.2)
GII.2	2 (25)	1 (11.1)
GIV	--	1 (11.1)
Not typed	1 (12.5)	4 (44.4)

\* Both GI.1 and GII.1 viruses were detected in samples from two cases and two controls

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children highlights that continued monitoring of the causes of AGE should include multi-enteric diagnostic platforms [30] to advance our understanding of the individual AGE pathogen in high-risk populations. It would be beneficial to conduct another study in this population to determine if norovirus trends and strain characteristics have changed over time.

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## Author Contributions

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**Investigation:** LG.

**Methodology:** JV.

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**Resources:** RW RR NG MS.

**Supervision:** MS UP.

**Writing – original draft:** LG JV KO UP MS.

**Writing – review & editing:** LG KO RW RR NG MS UP JV.

## References

1. Vinjé J. Advances in laboratory methods for detection and typing of norovirus. *J Clin Microbiol.* 2015; 53: 373–81. doi: [10.1128/JCM.01535-14](https://doi.org/10.1128/JCM.01535-14) PMID: [24989606](https://pubmed.ncbi.nlm.nih.gov/24989606/)
2. Oka T, Wang Q, Katayama K, Saif LJ. Comprehensive review of human sapoviruses. *Clin Microbiol Rev.* 2015; 28:32–53. doi: [10.1128/CMR.00011-14](https://doi.org/10.1128/CMR.00011-14) PMID: [25567221](https://pubmed.ncbi.nlm.nih.gov/25567221/)
3. Hall AJ, Lopman BA, Payne DC, Patel MM, Gastañaduy PA, Vinjé J, et al. Norovirus disease in the United States. *Emerg Infect Dis.* 2013; 19:1198–205. doi: [10.3201/eid1908.130465](https://doi.org/10.3201/eid1908.130465) PMID: [23876403](https://pubmed.ncbi.nlm.nih.gov/23876403/)
4. Svraka S, Vennema H, van der Veer B, Hedlund KO, Thorhagen M, Siebenga J, et al. Epidemiology and genotype analysis of emerging sapovirus-associated infections across Europe. *J Clin Microbiol.* 2010; 48: 2191–2198. doi: [10.1128/JCM.02427-09](https://doi.org/10.1128/JCM.02427-09) PMID: [20392905](https://pubmed.ncbi.nlm.nih.gov/20392905/)
5. Lee LE, Cebelinski EA, Fuller C, Keene WE, Smith K, Vinje J, et al. Sapovirus outbreaks in long-term care facilities, Oregon and Minnesota, USA, 2002–2009. *Emerg Infect Dis.* 2012; 18: 873–876. doi: [10.3201/eid1805.111843](https://doi.org/10.3201/eid1805.111843) PMID: [22516204](https://pubmed.ncbi.nlm.nih.gov/22516204/)
6. Bucardo F, Reyes Y, Svensson L, Nordgren J. Predominance of norovirus and sapovirus in Nicaragua after implementation of universal rotavirus vaccination. *PLoS One.* 2014; 9: e98201. doi: [10.1371/journal.pone.0098201](https://doi.org/10.1371/journal.pone.0098201) PMID: [24849288](https://pubmed.ncbi.nlm.nih.gov/24849288/)
7. Becker-Dreps S, Bucardo F, Vilchez S, Zambrana LE, Liu L, Weber DJ, et al. Etiology of childhood diarrhea after rotavirus vaccine introduction: a prospective, population-based study in Nicaragua. *Pediatr Infect Dis J.* 2014 Nov; 33:1156–63. doi: [10.1097/INF.0000000000000427](https://doi.org/10.1097/INF.0000000000000427) PMID: [24879131](https://pubmed.ncbi.nlm.nih.gov/24879131/)
8. Chhabra P, Payne DC, Szilagyi PG, Edwards KM, Staat MA, Shirley SH, et al. Etiology of viral gastroenteritis in children <5 years of age in the United States, 2008–2009. *J Infect Dis.* 2013; 208:790–800. doi: [10.1093/infdis/jit254](https://doi.org/10.1093/infdis/jit254) PMID: [23757337](https://pubmed.ncbi.nlm.nih.gov/23757337/)
9. Johnsen CK, Midgley S, and Bottiger B. Genetic diversity of sapovirus infections in Danish children 2005–2007. *J Clin Virol.* 2009; 46: 265–269.
10. Kirkwood C, Clark R, Bogdanovic-Sakran N, Bishop RF. A 5-year study of the prevalence and genetic diversity of human caliciviruses associated with sporadic cases of acute gastroenteritis in young children admitted to hospital in Melbourne, Australia (1998–2002). *J Med Virol.* 2005; 77: 96–101. doi: [10.1002/jmv.20419](https://doi.org/10.1002/jmv.20419) PMID: [16032716](https://pubmed.ncbi.nlm.nih.gov/16032716/)
11. Grant L, Vinjé J, Parashar U, Watt J, Reid R, Weatherholtz R, et al. Epidemiologic and clinical features of other enteric viruses associated with acute gastroenteritis in American Indian infants. *J Pediatr.* 2012; 161: 110–5.e1. doi: [10.1016/j.jpeds.2011.12.046](https://doi.org/10.1016/j.jpeds.2011.12.046) PMID: [22336577](https://pubmed.ncbi.nlm.nih.gov/22336577/)
12. Holman RC, Parashar UD, Clarke MJ, Kaufman SF, Glass RI. Trends in diarrhea-associated hospitalizations among American Indian and Alaska native children, 1980–1995. *Pediatrics.* 1999; 103:E11. PMID: [9917491](https://pubmed.ncbi.nlm.nih.gov/9917491/)
13. Grant LR, Watt JP, Weatherholtz RC, Moulton LH, Reid R, Santosham M, et al. Efficacy of a pentavalent human-bovine reassortant rotavirus vaccine against rotavirus gastroenteritis among American

- Indian children. *Pediatr Infect Dis J*. 2012; 31:184–8. doi: [10.1097/INF.0b013e3182435afe](https://doi.org/10.1097/INF.0b013e3182435afe) PMID: [22252206](https://pubmed.ncbi.nlm.nih.gov/22252206/)
14. Kojima S, Kageyama T, Fukushi S, Hoshino FB, Shinohara M, Uchida K, et al. Genogroup-specific PCR primers for detection of Norwalk-like viruses. *J Virol Methods*. 2002; 100: 107–14. PMID: [11742657](https://pubmed.ncbi.nlm.nih.gov/11742657/)
  15. Yan H, Yagyu F, Okitsu S, Nishio O, Ushijima H. Detection of norovirus (GI, GII), Sapovirus and astrovirus in fecal samples using reverse transcription single-round multiplex PCR. *J Virol Methods*. 2003; 114: 37–44. PMID: [14599677](https://pubmed.ncbi.nlm.nih.gov/14599677/)
  16. Ahmed SM, Hall AJ, Robinson AE, Verhoef L, Premkumar P, Parashar UD, et al. Global prevalence of norovirus in cases of gastroenteritis: a systematic review and meta-analysis. *Lancet Infect Dis*. 2014; 14:725–30. doi: [10.1016/S1473-3099\(14\)70767-4](https://doi.org/10.1016/S1473-3099(14)70767-4) PMID: [24981041](https://pubmed.ncbi.nlm.nih.gov/24981041/)
  17. Payne DC, Vinjé J, Szilagyi PG, Edwards KM, Staat MA, Weinberg GA, et al. Norovirus and medically attended gastroenteritis in U.S. children. *N Engl J Med*. 2013; 368: 1121–30. doi: [10.1056/NEJMsa1206589](https://doi.org/10.1056/NEJMsa1206589) PMID: [23514289](https://pubmed.ncbi.nlm.nih.gov/23514289/)
  18. Bucardo F, Nordgren J, Carlsson B, Kindberg E, Paniagua M, Möllby R, et al. Asymptomatic norovirus infections in Nicaraguan children and its association with viral properties and histo-blood group antigens. *Pediatr Infect Dis J*. 2010; 29:934–9. doi: [10.1097/INF.0b013e3181ed9f2f](https://doi.org/10.1097/INF.0b013e3181ed9f2f) PMID: [20657344](https://pubmed.ncbi.nlm.nih.gov/20657344/)
  19. Lopman BA, Trivedi T, Vicuña Y, Costantini V, Collins N, Gregoricus N, et al. Norovirus Infection and Disease in an Ecuadorian Birth Cohort: Association of Certain Norovirus Genotypes With Host FUT2 Secretor Status. *J Infect Dis*. 2015; 211:1813–21. doi: [10.1093/infdis/jiu672](https://doi.org/10.1093/infdis/jiu672) PMID: [25505295](https://pubmed.ncbi.nlm.nih.gov/25505295/)
  20. Zhang S, Chen TH, Wang J, Dong C, Pan J, Moe C, et al. Symptomatic and asymptomatic infections of rotavirus, norovirus, and adenovirus among hospitalized children in Xi'an, China. *J Med Virol*. 2011; 83:1476–84. doi: [10.1002/jmv.22108](https://doi.org/10.1002/jmv.22108) PMID: [21618552](https://pubmed.ncbi.nlm.nih.gov/21618552/)
  21. Platts-Mills JA, Babji S, Bodhidatta L, Gratz J, Haque R, Havt A, et al.; MAL-ED Network Investigators. Pathogen-specific burdens of community diarrhoea in developing countries: a multisite birth cohort study (MAL-ED). *Lancet Glob Health*. 2015 Jul 17. pii: S2214-109X(15)00151-5.
  22. Atmar RL, Opekun AR, Gilger MA, Estes MK, Crawford SE, Neill FH, et al. Norwalk virus shedding after experimental human infection. *Emerg Infect Dis*. 2008; 14:1553–7. doi: [10.3201/eid1410.080117](https://doi.org/10.3201/eid1410.080117) PMID: [18826818](https://pubmed.ncbi.nlm.nih.gov/18826818/)
  23. Baldridge MT, Nice TJ, McCune BT, Yokoyama CC, Kambal A, Wheadon M, et al. Commensal microbes and interferon-λ determine persistence of enteric murine norovirus infection. *Science*. 2015; 347:266–9. doi: [10.1126/science.1258025](https://doi.org/10.1126/science.1258025) PMID: [25431490](https://pubmed.ncbi.nlm.nih.gov/25431490/)
  24. Yassin MA, Kirby A, Mengistu AA, Arbide I, Dove W, Beyer M, et al. Unusual norovirus and rotavirus genotypes in Ethiopia. *Paediatr Int Child Health*. 2012; 32:51–5. doi: [10.1179/1465328111Y.0000000047](https://doi.org/10.1179/1465328111Y.0000000047) PMID: [22525449](https://pubmed.ncbi.nlm.nih.gov/22525449/)
  25. Hassine-Zaafraane M, Sdiri-Loulizi K, Kaplon J, Salem IB, Pothier P, Aouni M, et al. Prevalence and genetic diversity of norovirus infection in Tunisian children (2007–2010). *J Med Virol*. 2013; 85:1100–10. doi: [10.1002/jmv.23552](https://doi.org/10.1002/jmv.23552) PMID: [23532785](https://pubmed.ncbi.nlm.nih.gov/23532785/)
  26. Dove W, Cunliffe NA, Gondwe JS, Broadhead RL, Molyneux ME, Nakagomi O, et al. Detection and characterization of human caliciviruses in hospitalized children with acute gastroenteritis in Blantyre, Malawi. *J Med Virol*. 2005; 77:522–7. doi: [10.1002/jmv.20488](https://doi.org/10.1002/jmv.20488) PMID: [16254959](https://pubmed.ncbi.nlm.nih.gov/16254959/)
  27. Widdowson MA, Cramer EH, Hadley L, Bresee JS, Beard RS, Bulens SN, et al. Outbreaks of acute gastroenteritis on cruise ships and on land: identification of a predominant circulating strain of norovirus—United States, 2002. *J Infect Dis*. 2004; 190:27–36. doi: [10.1086/420888](https://doi.org/10.1086/420888) PMID: [15195240](https://pubmed.ncbi.nlm.nih.gov/15195240/)
  28. Hoa Tran TN, Trainor E, Nakagomi T, Cunliffe NA, Nakagomi O. Molecular epidemiology of noroviruses associated with acute sporadic gastroenteritis in children: global distribution of genogroups, genotypes and GI.4 variants. *J Clin Virol*. 2013; 56:185–93. doi: [10.1016/j.jcv.2012.11.011](https://doi.org/10.1016/j.jcv.2012.11.011) PMID: [23218993](https://pubmed.ncbi.nlm.nih.gov/23218993/)
  29. Blanton LH, Adams SM, Beard RS, et al. Molecular and epidemiologic trends of caliciviruses associated with outbreaks of acute gastroenteritis in the United States, 2000–2004. *J Infect Dis*. 2006; 193:413–21. doi: [10.1086/499315](https://doi.org/10.1086/499315) PMID: [16388489](https://pubmed.ncbi.nlm.nih.gov/16388489/)
  30. Gonzalez MD, Langley LC, Buchan BW, et al. Multicenter Evaluation of the Xpert Norovirus Assay for Detection of Norovirus Genogroups I and II in Fecal Specimens. *J Clin Microbiol*. 2016; 54:142–7. doi: [10.1128/JCM.02361-15](https://doi.org/10.1128/JCM.02361-15) PMID: [26560532](https://pubmed.ncbi.nlm.nih.gov/26560532/)