

# Measurement of Total Phosphorus and Polyphosphate in *Chlamydomonas reinhardtii*

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## Abstract

Phosphorus is an essential nutrient for plants. Green algae usually store excess P as polyphosphate (polyP) in the vacuoles. PolyP, a linear chain of three to hundreds of phosphate residues linked by phosphoanhydride bonds, is important for cell growth. Based on the previous method of polyP purification with silica gel columns (Werner et al., 2005; Canadell et al., 2016) in yeast cells, we developed a protocol to purify and determine the total P and polyP in *Chlamydomonas reinhardtii* by a quick, simplified, and quantitative method. We use hydrochloric acid or nitric acid to digest polyP or total P in dried cells and analyze P content using the malachite green colorimetric method. This method may be applied to other microalgae.

**Keywords:** Green algae, *Chlamydomonas*, Polyphosphate, Total phosphorus, Vacuolar phosphate, Plant nutrient

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## Background

As an essential nutrient, phosphorus is required for the constitution of cellular components such as nucleic acids, phospholipids, and ATP. P is also required for the regulation of protein phosphorylation in plants. P is stored as inorganic phosphate (Pi) in the vacuoles of land plants and as inorganic polyphosphate (polyP) in chlorophyte algae. PolyP is a linear chain composed of several phosphate residues that are linked together by phosphoanhydride bonds. PolyP is an important metabolite and a signaling molecule acting as an energy source, a regulator of gene expression, a channel-forming component, and a storage form of Pi found in green plants (Wang et al., 2021).

Since the polyP is a mixture with different chain lengths, few analytical methods are currently available for polyP quantitation. In order to determine the content of polyP, it usually has to be degraded into Pi first, and then its content can be estimated by measuring the amount of Pi released. Two experimental methods are widely used for quantitatively estimating the amount of polyP in vitro (Aschar-Sobbi et al., 2008). The first one is based on the hydrolysis of polyP by hydrochloric acid and subsequent measurement of the Pi released. The second one is based on the activity of two enzymes, exopolyphosphatase and polyphosphate kinase, which can digest and convert the polyP into Pi for measurement. However, the expression and purification of these enzymes are complicated and time consuming (Christ et al., 2020).

Here, we describe an updated protocol for quantitative measurement of the polyP content in algae. We use silica gel columns to purify polyP and then convert it into Pi through digestion by hydrochloric acid. Although this method is not very accurate, because some short-chain-length polyP could not be completely extracted by the silica gel columns, it is economical and convenient. This protocol is relatively simple and time saving, leading to a fast determination of the polyP in *Chlamydomonas*, and should be considered suitable for polyP determination in other microalgae.

## Materials and reagents

1. Centrifuge tubes (1.5, 2, 15, and 50 mL) (Sangon Biotech)
2. Sterile culture flasks (50 and 250 mL) (Sangon Biotech)
3. Pipettes (10, 20 and 200  $\mu$ L, 1 and 5 mL) (Eppendorf)
4. 96-well ELISA plate (Sangon Biotech, catalog number: F605031)
5. Petri dishes (Sangon Biotech, catalog number: F611003)
6. 47 mm filter paper sheets (Whatman, catalog number: 1822047)
7. Spin Columns CA2 from TIANGel Midi Purification kit (TIANGEN, catalog number: DP209)
8. Hydrochloric acid (HCl)
9. Nitric acid (HNO<sub>3</sub>)
10. 98% sulfuric acid (H<sub>2</sub>SO<sub>4</sub>)
11. Tris (C<sub>4</sub>H<sub>11</sub>NO<sub>3</sub>) (Sangon Biotech, catalog number: A100826)
12. Glacial acetic acid (CH<sub>3</sub>COOH) (Sangon Biotech, catalog number: A501931)
13. Ammonium chloride (NH<sub>4</sub>Cl) (Sangon Biotech, catalog number: A501569)
14. Magnesium sulfate heptahydrate (MgSO<sub>4</sub>·7H<sub>2</sub>O) (Sangon Biotech, catalog number: A610329)
15. Calcium chloride dihydrate (CaCl<sub>2</sub>·2H<sub>2</sub>O) (Sangon Biotech, catalog number: A610050)
16. Dipotassium hydrogen phosphate (K<sub>2</sub>HPO<sub>4</sub>) (Sangon Biotech, catalog number: A610447)
17. Potassium phosphate monobasic (KH<sub>2</sub>PO<sub>4</sub>) (Sangon Biotech, catalog number: A600445)
18. EDTA disodium salt (C<sub>10</sub>H<sub>14</sub>N<sub>2</sub>O<sub>8</sub>Na<sub>2</sub>·2H<sub>2</sub>O) (Sangon Biotech, catalog number: A100105)
19. Zinc sulfate heptahydrate (ZnSO<sub>4</sub>·7H<sub>2</sub>O) (Sangon Biotech, catalog number: A602906)
20. Iron(II) sulfate heptahydrate (FeSO<sub>4</sub>·7H<sub>2</sub>O) (Sangon Biotech, catalog number: A600461)
21. Boric acid (H<sub>3</sub>BO<sub>3</sub>) (Sangon Biotech, catalog number: A100588)
22. Manganese (II) chloride tetrahydrate (MnCl<sub>2</sub>·4H<sub>2</sub>O) (Sangon Biotech, catalog number: A500331)
23. Cobalt chloride hexahydrate (CoCl<sub>2</sub>·6H<sub>2</sub>O) (Sigma- Aldrich: catalog number: 255599)
24. Copper(II) sulfate pentahydrate (CuSO<sub>4</sub>·5H<sub>2</sub>O) (Sangon Biotech, catalog number: A600063)

25. Ethanol absolute (Sangon Biotech, catalog number: A500737)
26. Neutral red (Sangon Biotech, catalog number: A600895)
27. 1 M Tris-HCl (pH 7.5) (Sangon Biotech, catalog number: A610283)
28. Sodium iodide (NaI) (Sangon Biotech, catalog number: A610283)
29. Ammonium heptamolybdate tetrahydrate  $[(\text{NH}_4)_6\text{Mo}_7\text{O}_{24}\cdot 4\text{H}_2\text{O}]$  (Sangon Biotech, catalog number: A600067)
30. Malachite green oxalate salt (Sangon Biotech, catalog number: A620330)
31. TAP medium (see Recipes)
32. 40× TAP salt mixture (see Recipes)
33. Phosphate solution (50  $\mu\text{g}/\text{mL}$ ) (see Recipes)
34. Hutner's trace elements (see Recipes)
35. TAP solid medium (see Recipes)
36. 1 M sulfuric acid ( $\text{H}_2\text{SO}_4$ ) (see Recipes)
37. 2 M NaOH (see Recipes)
38. 70% ethanol (see Recipes)
39. 0.1% neutral red solution (w/v) (see Recipes)
40. 1 M Tris-HCl (pH 7.5) supplemented to 6% (v/v) with 0.1% neutral red solution (w/v) (see Recipes)
41. 6 M NaI (see Recipes)
42. Wash buffer (see Recipes)
43. 2 M hydrochloric acid (HCl) (see Recipes)
44. Phosphate (Pi) solution (10  $\mu\text{g}/\text{mL}$ ) (see Recipes)
45. 28 mM ammonium heptamolybdate in 2.1 M  $\text{H}_2\text{SO}_4$  (see Recipes)
46. 0.76 mM malachite green in 0.35% polyvinyl alcohol (see Recipes)

## Equipment

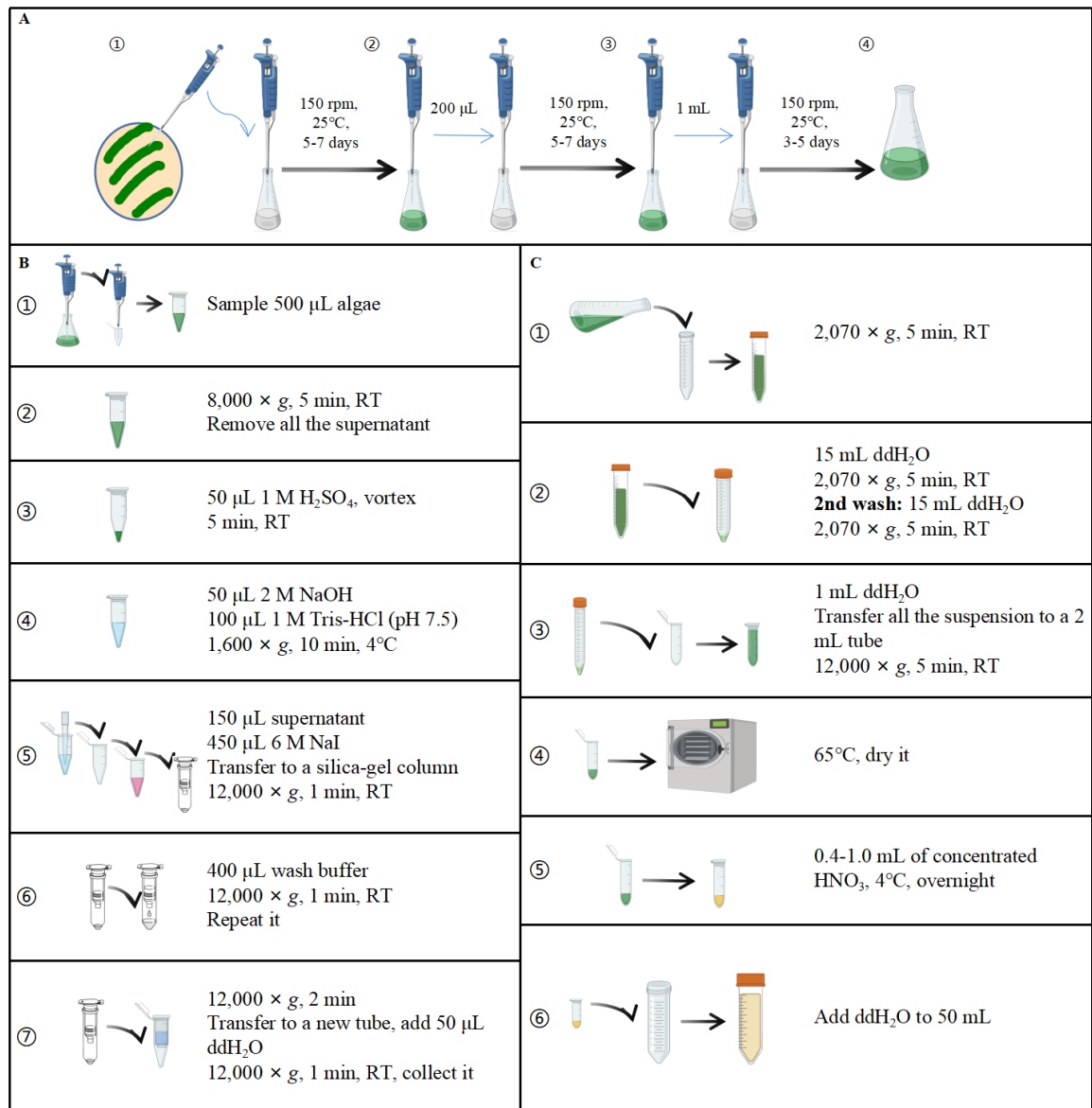
1. Spectrophotometer (Shimadzu, model: UVmini-1240)
2. Centrifuge (Thermo Scientific, catalog number: 75002411)
3. Hitachi Himac CT15RE tabletop high-speed microcentrifuge
4. Hitachi CR21N high-speed refrigerated centrifuge
5. Vortex (IKA, model: IKA MS 3 basic)
6. Thermal cycler (Thermo Scientific, catalog number: 4484073)
7. Microplate reader (Tecan, model: Infinite® F50 microplate reader)
8. Vacuum pump (JinTeng, model: GM-0.5B)
9. Analytical balance (Sangon Biotech, catalog number: G001142)
10. Air dry oven (Sangon Biotech, catalog number: G003409)
11. Clean bench (AIRTECH, catalog number: SW-CJ-2F)
12. Fume hood (ALPHA, VAV controller)
13. Nikon Digital Camera D7200

## Software

1. Microsoft Excel (Microsoft, Inc)
2. Microsoft PowerPoint (Microsoft, Inc)
3. Prism GraphPad (Dotmatic, Inc)

## Procedure

In this section, we describe the procedure to measure polyP and total P in *C. reinhardtii*. The first step is to culture algae cells. Next, collect an appropriate number of algae cells to extract P (polyP or total P) and then digest polyP or total P into Pi by hydrochloric acid or nitric acid from dried cells. To quantify the Pi released, the malachite green colorimetric method is used.



**Figure 1. Basic workflow of cell culture and P measurement.** A. Procedure for algae cell culture. B and C show the procedures for polyP and total P measurement, respectively. Drawn and composed using Microsoft PowerPoint.

### A. Culture methods

Protocols adapted from Wang et al. (2021).

Cell culture is grown using 20 mL of TAP medium in a 50 mL flask. Stationary-phase cell culture with a volume of 200 µL is transferred into 20 mL of fresh medium every week. The culture growth conditions are adjusted to a speed of 150 rpm shaking at 25 °C using a rocking incubator, and the culture itself is illuminated

continuously with white fluorescent light ( $150 \mu\text{mol photons m}^{-2} \text{s}^{-1}$ ). A cell density of approximately  $1 \times 10^7$  cells (3–5 days after transfer, late linear phase) is used for the experiment.

The following provides specific steps:

1. Pick wildtype (CC-4533) and mutant (*Crptc1*) or other desired strain(s) from a fresh plate with a sterile toothpick or pipette tip and inoculate with 20 mL of liquid medium (TAP medium or appropriate selection medium) using a 50 mL flask. Rotate at 150 rpm and 25 °C for 5–7 days under continuous white light. This primary culture is grown to a stationary phase.
2. Inoculate cultures into a 50 mL flask with 20 mL of TAP medium and adjust inoculum size appropriately to make the  $\text{OD}_{750}$  of inoculated solution approximately 0.2–0.3. Cultures will reach the stable period ( $\sim 1 \times 10^7$  cells) again after 5–7 days. The  $\text{OD}_{750}$  should be measured every 12 h to count the number of algae cells.
3. Transfer 1 mL of the secondary algae cells into a 250 mL flask containing 100 mL of TAP medium and incubate for 3–5 days.

After three times of uniformization, different algae strains reach the same or similar growth status.

Optional:

4. Determine the  $\text{OD}_{750}$  every 12 h, calculate the regression equation, and draw the growth curve.

*Note: The growth phase can be determined by measuring cell density and drawing the growth curve.*

## B. Extraction of P

This section contains two parts. The first one is the purification and degradation of polyP and the second one is the digestion of total P in algae with nitric acid. After purification and degradation, polyP and total phosphorus content is converted into Pi released and can be measured using the malachite green colorimetric method.

1. Purification and degradation of polyP

Protocol adapted from Werner et al. (2005), Canadell et al. (2016), and Wang et al. (2021).

- a. Determine the  $\text{OD}_{750}$  of the algae using a spectrophotometer and record it.
- b. Add 500  $\mu\text{L}$  of algae (record the exact weight) into a 1.5 mL centrifuge tube. After centrifugation at  $8,000 \times g$  for 5 min at room temperature, remove all the supernatant by pipetting and harvest the algae cells.

*Note: The sample can be directly purified or stored at  $-20 \text{ }^\circ\text{C}$ . For more accurate measurements of dry weight, 5 mL of algae is filtered on 47 mm filter paper sheets using a vacuum pump, and then put in a drying oven for several hours (Figure 1). The weight of the 47 mm filter paper sheet should be measured using an analytical balance before and after sample loading.*

- c. Add 50  $\mu\text{L}$  of 1 M  $\text{H}_2\text{SO}_4$  (see Recipes) into the algae, mix by pipetting or vortex to homogenate, and then place the tube at room temperature for 5 min.
- d. Neutralize the suspension with 50  $\mu\text{L}$  of 2 M NaOH (see Recipes) and 100  $\mu\text{L}$  of 1 M Tris-HCl (pH 7.5) supplemented to 6% (v/v) with 0.1% neutral red solution (see Recipes). Gently mix the tubes upside down four times.

*Note: The pH indicator helps to assure that the pH is around 7 (the sample color becomes orange red), which is a critical step for the reproducibility of polyP extraction. Samples too acidic (pink) or too basic (yellow) have to be corrected to the appropriate pH (orange red) by the addition of 2 M NaOH or 1 M  $\text{H}_2\text{SO}_4$ , respectively.*

- e. Centrifuge at  $1,600 \times g$  for 10 min at 4 °C.
- f. Drain 150  $\mu\text{L}$  of supernatant with a pipette into a new 1.5 mL centrifuge tube containing 450  $\mu\text{L}$  of 6 M NaI (freshly prepared; see Recipes).
- g. Mix by pipetting 3–5 times and then transfer to a silica gel column (Spin Columns CA2 from TIANgel Midi Purification kit). Centrifuge the column at  $12,000 \times g$  for 1 min at room temperature.
- h. Discard the flowthrough and place the column back into the collection tube. Add 400  $\mu\text{L}$  of wash buffer (see Recipes) into the column and centrifuge the column at  $12,000 \times g$  for 1 min at room temperature.
- i. Repeat step B1h.

- j. Discard the flowthrough, place the column back into the collection tube, and centrifuge at  $12,000 \times g$  for 2 min at room temperature to remove residual wash buffer.
  - k. Transfer the column into a clean 1.5 mL centrifuge tube and place the column with the cap open for 2–5 min to dry the membrane.
  - l. Add 50  $\mu\text{L}$  of ddH<sub>2</sub>O to the center of the membrane, incubate at room temperature for 2 min, and then centrifuge at  $12,000 \times g$  for 2 min at room temperature.  
*Note: The eluted polyP can be directly degraded; however, leaving it at room temperature for a longer time should be avoided. For long-term storage, it can be kept at  $-20\text{ }^\circ\text{C}$ .*
  - m. Pipette 10  $\mu\text{L}$  of eluted polyP and add 10  $\mu\text{L}$  of 2 M HCl (see Recipes), mix thoroughly, and centrifuge for several seconds.
  - n. Incubate at  $95\text{ }^\circ\text{C}$  for 30 min in the thermal cycler and then transfer onto the ice.
  - o. The degraded polyP sample can be directly measured with the malachite green colorimetric method or stored at  $-20\text{ }^\circ\text{C}$ .
2. Determination of total P
    - a. Mark a 2 mL centrifuge tube and weigh it.
    - b. Collect 10–15 mL of cells into a 15 mL centrifuge tube and centrifuge at  $2,070 \times g$  for 5 min at room temperature.
    - c. Discard the supernatant, wash the algae cells with ddH<sub>2</sub>O, and centrifuge at  $2,070 \times g$  for 5 min at room temperature.
    - d. Repeat step B2c.
    - e. Discard the supernatant, suspend the algae cells with 1 mL of ddH<sub>2</sub>O, and transfer the entire suspension to the 2 mL centrifuge tube.
    - f. Centrifuge at  $12,000 \times g$  for 5 min at room temperature and pipette all the supernatant carefully in case of loss of algae samples.
    - g. Dry and weigh the tube with algae samples.
    - h. Add 0.4–1.0 mL of concentrated HNO<sub>3</sub> and incubate overnight at  $4\text{ }^\circ\text{C}$  until the solution is clean.
    - i. Transfer the solution into a 50 mL centrifuge tube, wash the inside of the 2 mL centrifuge tube with ddH<sub>2</sub>O, transfer all the wash water into the 50 mL centrifuge tube, and dilute with ddH<sub>2</sub>O to 50 mL.
    - j. This sample can then be analyzed for P concentration determination using the malachite green colorimetric method.

### C. Quantification of P with the malachite green colorimetric method

1. Drawing of the standard curve (Table 1)

**Table 1. Standard curve**

	1	2	3	4	5	6	7
Concentration $\mu\text{g/mL}$	0	0.25	0.5	0.75	1	1.5	2
Pi solution (10 $\mu\text{g/mL}$ ) $\mu\text{L}$	0	2.5	5	7.5	10	15	20
ddH <sub>2</sub> O $\mu\text{L}$	100	97.5	95	92.5	90	85	80

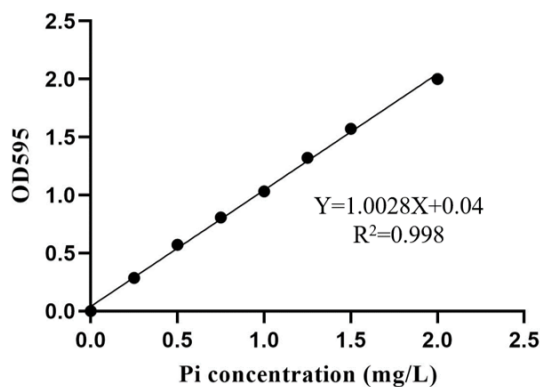
According to the Pi concentration gradient in the table, add these substances in order:

- a. Add Pi solution (10  $\mu\text{g/mL}$ ) (see Recipes) and the corresponding volume of ddH<sub>2</sub>O.
- b. Add 86  $\mu\text{L}$  of 28 mM ammonium heptamolybdate in 2.1 M H<sub>2</sub>SO<sub>4</sub> (see Recipes).
- c. Add 64  $\mu\text{L}$  of 0.76 mM malachite green in 0.35% polyvinyl alcohol (see Recipes).
- d. Leave the 96-well ELISA plate at room temperature for 30 min and measure the OD<sub>595</sub> with a microplate reader.

*Note: Do not let the reaction proceed for more than 1 h as it can cause the appearance of small precipitates in highly concentrated Pi samples and polyP degradation in background samples, thus interfering with the correct Pi measure.*

- e. Draw the standard curve by taking the volume of Pi solution (10  $\mu\text{g/mL}$ ) as the x-axis and the OD<sub>595</sub> as the y-axis. Then, the P concentration of the samples can be calculated according to the standard

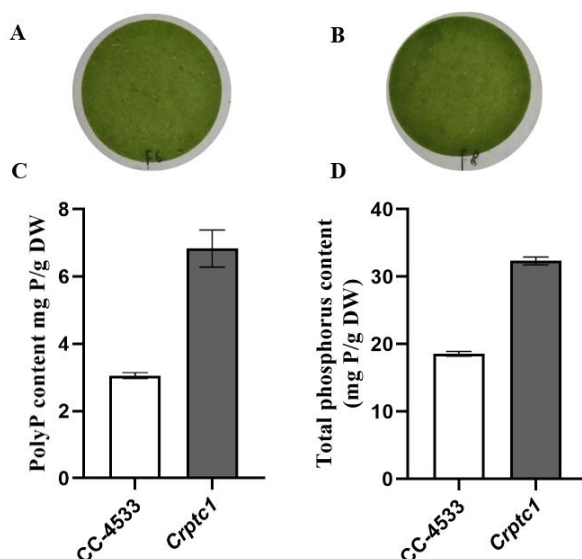
curve.



**Figure 2. Standard curve.** Pi concentration as x-axis and the OD<sub>595</sub> as the y-axis. Drawn using Microsoft Excel and Prism GraphPad.

2. Determination of P in samples
  - a. Use a 96-well plate for the reaction and measurement of polyP. To quantify released Pi, add to each well 10 μL of polyP after degradation, 90 μL of ddH<sub>2</sub>O, 86 μL of 28 mM ammonium heptamolybdate in 2.1 M H<sub>2</sub>SO<sub>4</sub> (see Recipes), and 64 μL of 0.76 mM malachite green in 0.35% polyvinyl alcohol (see Recipes), in this order. Mix the well by pipetting gently.
  - b. The blank control consists of 100 μL of ddH<sub>2</sub>O, 86 μL of 28 mM ammonium heptamolybdate in 2.1 M H<sub>2</sub>SO<sub>4</sub>, and 64 μL of 0.76 mM malachite green in 0.35% polyvinyl alcohol.
  - c. Leave the 96-well plate at room temperature for 30 min and measure the OD<sub>595</sub> with a microplate reader.

## Results



**Figure 3. The content of PolyP and total P in wildtype strain CC-4533 and the *Crptc1* mutant.** (A) and (B) show filter paper sheets of 5 mL cells through a vacuum pump. The weight of algae cells was calculated using the



dry weight difference of filter paper before and after filtration. PolyP content (C) and total P content (D) of the wildtype strain CC-4533 and the *Crptc1* mutant. CrPTC1 is a vacuole efflux phosphate transporter in algae and catalyzes Pi transport out of acidocalcisomes. The *Crptc1* mutant over-accumulates total P and polyP (Wang et al., 2021). Data is presented as the mean  $\pm$  SD from five independent experiments. Pictures were taken using a Nikon Digital Camera D7200. C and D were drawn using Microsoft Excel and Prism GraphPad.

## Recipes

### 1. TAP medium

Tris 2.42 g  
 40 $\times$  TAP salt mixture 25 mL (Recipe 2)  
 Phosphate solution (50  $\mu$ g/mL) 0.4 mL (Recipe 3)  
 Hutner's trace elements 1 mL (Recipe 4)  
 Glacial acetic acid 1 mL  
 Adjust to pH 7.0 with HCl  
 Add ddH<sub>2</sub>O to 1 L and autoclave at 121  $^{\circ}$ C for 20 min

### 2. 40 $\times$ TAP salt mixture

NH<sub>4</sub>Cl 15 g  
 MgSO<sub>4</sub>·7H<sub>2</sub>O 4 g  
 CaCl<sub>2</sub>·2H<sub>2</sub>O 2 g  
 Add ddH<sub>2</sub>O to 1 L

### 3. Phosphate solution (50 $\mu$ g/mL)

K<sub>2</sub>HPO<sub>4</sub> 28.8 g  
 KH<sub>2</sub>PO<sub>4</sub> 14.4 g  
 Add ddH<sub>2</sub>O to 100 mL

### 4. Hutner's trace elements

EDTA disodium salt 50 g  
 ZnSO<sub>4</sub>·7H<sub>2</sub>O 22 g  
 FeSO<sub>4</sub>·7H<sub>2</sub>O 4.99 g  
 H<sub>3</sub>BO<sub>3</sub> 11.4 g  
 MnCl<sub>2</sub>·4H<sub>2</sub>O 5.06 g  
 CoCl<sub>2</sub>·6H<sub>2</sub>O 1.61 g  
 CuSO<sub>4</sub>·5H<sub>2</sub>O 1.57 g  
 (NH<sub>4</sub>)<sub>6</sub>Mo<sub>7</sub>O<sub>24</sub>·4H<sub>2</sub>O 1.10 g  
 Add ddH<sub>2</sub>O to 1 L

### 5. TAP solid medium

TAP medium (Recipe 1) with 1% (w/v) agar  
 Autoclave at 121  $^{\circ}$ C for 20 min

### 6. 1 M sulfuric acid (H<sub>2</sub>SO<sub>4</sub>)

Mix 5.45 mL of 98% sulfuric acid with 94.55 mL of ddH<sub>2</sub>O and store at room temperature

### 7. 2 M NaOH

Mix 8 g of NaOH with 80 mL of ddH<sub>2</sub>O  
 Add ddH<sub>2</sub>O to 100 mL and store at room temperature



**8. 70% ethanol**

Mix 70 mL of ethanol absolute with 30 mL of ddH<sub>2</sub>O

**9. 0.1% neutral red solution (w/v)**

Mix 0.1 g of neutral red with 100 mL of 70% ethanol (Recipe 8) and store at room temperature

**10. 1 M Tris-HCl (pH 7.5) supplemented to 6% (v/v) with 0.1% neutral red solution (w/v)**

Mix 12.1 g of Tris with 70 mL of ddH<sub>2</sub>O

Add 6 mL of 0.1% neutral red solution (w/v) (Recipe 9)

Adjust pH to 7.5 with HCl

Add ddH<sub>2</sub>O to 100 mL and store at room temperature

**11. 6 M NaI**

Mix 8.9934 g of NaI with ddH<sub>2</sub>O up to 10 mL

The solution has to be freshly prepared each time and kept in darkness

**12. Wash buffer**

10 mM Tris-HCl buffer (pH 7.5), 50% ethanol, 1 mM EDTA, and 100 mM NaCl

Mix 5 mL of 1 M Tris-HCl (pH 7.5), 250 mL of 100% ethanol, 1 mL of 0.5 M EDTA, 10 mL of 5 M NaCl, and 200 mL of ddH<sub>2</sub>O

Adjust pH to 7.5 with HCl

Add ddH<sub>2</sub>O to 500 mL and store at room temperature

**13. 2 M HCl**

Mix 16.67 mL of 37% HCl with 83.33 mL of ddH<sub>2</sub>O and store at room temperature

**14. Phosphate (Pi) solution (10 µg/mL)**

Mix 50 µL of Phosphate solution (50 µg/mL; Recipe 3) and 200 µL of ddH<sub>2</sub>O

**15. 28 mM ammonium heptamolybdate in 2.1 M H<sub>2</sub>SO<sub>4</sub>**

Mix 3.46 g of ammonium heptamolybdate tetrahydrate with 80 mL of ddH<sub>2</sub>O

Add 11.2 mL of 98% H<sub>2</sub>SO<sub>4</sub>

Add ddH<sub>2</sub>O to 100 mL and store at room temperature

**16. 0.76 mM malachite green in 0.35% polyvinyl alcohol**

Mix 350 mg of polyvinyl alcohol in 100 mL of ddH<sub>2</sub>O at 80 °C and stir vigorously on a magnetic stirrer until all polyvinyl alcohol dissolves completely

Add 35 mg of malachite green oxalate salt

Add ddH<sub>2</sub>O to 100 mL and store at room temperature

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## Competing interests

No conflict of interest or competing interests declared.

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