#### Check for updates

#### OPEN ACCESS

EDITED BY Malik Hassan Mehmood, Government College University Faisalabad, Pakistan

REVIEWED BY Jiguang Wang, Shanghai Jiao Tong University, China Eman Gohar, Vanderbilt University Medical Center, United States Tahir Hussain, University of Houston, United States

\*CORRESPONDENCE Omar Z. Ameer, oaladhami@alfaisal.edu

#### SPECIALTY SECTION

This article was submitted to Cardiovascular and Smooth Muscle Pharmacology, a section of the journal Frontiers in Pharmacology

RECEIVED 20 May 2022 ACCEPTED 26 September 2022 PUBLISHED 11 October 2022

#### CITATION

Ameer OZ (2022), Hypertension in chronic kidney disease: What lies behind the scene. *Front. Pharmacol.* 13:949260. doi: 10.3389/fphar.2022.949260

#### COPYRIGHT

© 2022 Ameer. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

# Hypertension in chronic kidney disease: What lies behind the scene

### Omar Z. Ameer<sup>1,2</sup>\*

<sup>1</sup>Department of Pharmaceutical Sciences, College of Pharmacy, Alfaisal University, Riyadh, Saudi Arabia, <sup>2</sup>Department of Biomedical Sciences, Faculty of Medicine, Macquarie University, Sydney, NSW, Australia

Hypertension is a frequent condition encountered during kidney disease development and a leading cause in its progression. Hallmark factors contributing to hypertension constitute a complexity of events that progress chronic kidney disease (CKD) into end-stage renal disease (ESRD). Multiple crosstalk mechanisms are involved in sustaining the inevitable high blood pressure (BP) state in CKD, and these play an important role in the pathogenesis of increased cardiovascular (CV) events associated with CKD. The present review discusses relevant contributory mechanisms underpinning the promotion of hypertension and their consequent eventuation to renal damage and CV disease. In particular, salt and volume expansion, system (SNS) hyperactivity, sympathetic nervous upregulated renin-angiotensin-aldosterone system (RAAS), oxidative stress, vascular remodeling, endothelial dysfunction, and a range of mediators and signaling molecules which are thought to play a role in this concert of events are emphasized. As the control of high BP via therapeutic interventions can represent the key strategy to not only reduce BP but also the CV burden in kidney disease, evidence for major strategic pathways that can alleviate the progression of hypertensive kidney disease are highlighted. This review provides a particular focus on the impact of RAAS antagonists, renal nerve denervation, baroreflex stimulation, and other modalities affecting BP in the context of CKD, to provide interesting perspectives on the management of hypertensive nephropathy and associated CV comorbidities.

#### KEYWORDS

sympathetic hyperactivity, renin-angiotensin-aldosterone system, vascular remodeling, endothelial dysfunction, chronic kidney disease

## **1** Introduction

Chronic kidney disease (CKD) is the most common cause of secondary hypertension (Campese et al., 2006; Masuda and Nagata, 2020), in which hypertension is considered an important co-morbid factor associated with CKD (Baumeister et al., 2009; Middleton and Pun, 2010), and is the leading cause of death globally (Unger et al., 2020; Guan et al., 2022). Hypertension presents in approximately 80–85% of patients with CKD (Wong et al., 2006; Collins et al., 2014; Freedman and Cohen, 2016), with the more severe

glomerular diseases having a higher incidence of hypertension (Blythe, 1985; Sanchez et al., 2018). For any given cause of CKD including hypertension itself, the elevation in systemic blood pressure (BP) accentuates the rate at which glomerular filtration rate (GFR) declines (Perry et al., 1995; Ku et al., 2019), which makes hypertension an independent risk factor for end-stage renal disease (ESRD) (Tozawa et al., 2003; Salman, 2015). Previous reports have noted that hypertension increases the risk for new or recurrent cardiovascular (CV) events in individuals with stage 2-3 CKD (Muntner et al., 2005). This is also evident for genetically driven causes of CKD, with virtually all patients with polycystic kidney disease (PKD) for example, developing hypertension by the time ESRD is present (Chapman et al., 2010). Reports suggest that the high BP state is considered a strong independent risk factor for ESRD and interventions to prevent the disease need to emphasize the prevention and control of both high-normal and high BP (Klag et al., 1996; Ku et al., 2019). Hypertension is therefore associated with accelerated progression of kidney disease as well as the development and worsening of cardiovascular disease (CVD).

The kidney has an intrinsic role in the development of hypertension. There is both clinical and experimental evidence to suggest that the "BP goes with the kidney" (Adamczak et al., 2002). Indeed, it has been shown that normotensive recipients of kidneys from hypertensive donors become hypertensive in humans (Guidi et al., 1996) and in rats (Rettig et al., 1990); and conversely hypertensive recipients of normotensive kidneys may become normotensive again in both humans (Curtis et al., 1983) and rats (Patschan et al., 1997). In addition, risk factors associated with BP elevation and its development are similar to those in the general population after kidney transplantation (Sanchez et al., 2018). Presently, it remains somewhat vague as to what is the best target BP in the treatment of hypertensive patients with CV risks. As such, this is creating a grey area in the selection between reducing the risk of CV incidence and mortality, or risking having poor prognostic or damaging effects on the kidneys themselves. In the Systolic BP Intervention Trial (SPRINT) study (Group, 2015), aggressive BP reduction with antihypertensives to target a systolic BP of <120 mmHg (relative to the standard target of systolic BP < 140 mmHg) has been shown to reduce the rate of CV risks (myocardial infarction, acute coronary syndrome, heart failure, stroke, or CV deaths) or all-cause mortality. An unfortunate association noted with this outcome, however, was a higher risk of renal injury in the intensive BP lowering cohorts (Obi et al., 2018; Rocco et al., 2018), or failure to produce a benefit for kidney GFR decline or ESRD risk (Schrier et al., 2014; Cheung et al., 2017). Regardless of CKD etiology, severe and uncontrolled hypertension accelerates the loss of nephrons, and hence GFR (Ku et al., 2019). However, whether intensive BP reduction slows GFR decline is evidently still unclear. Furthermore, the disagreement on the cut-off value for hypertension treatment guidelines in adults between

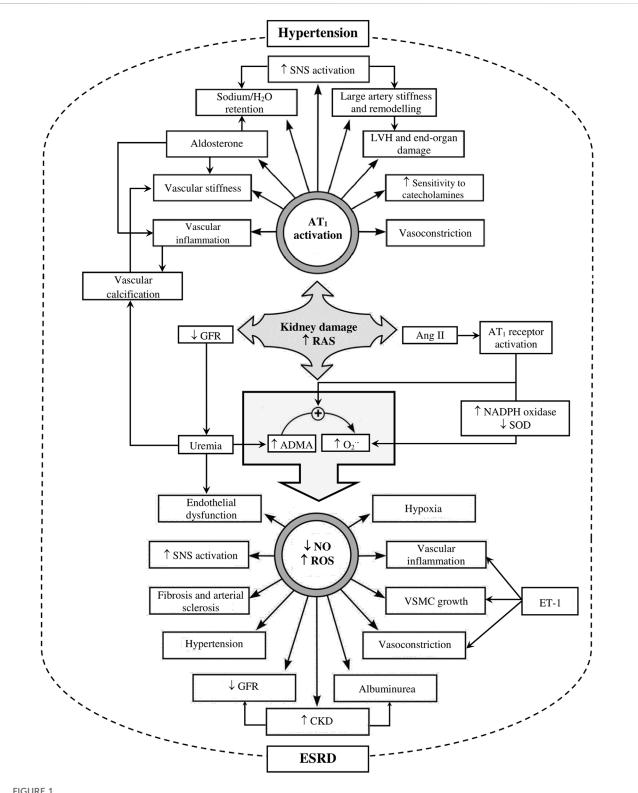
Americans (target above 130/80 mmHg) and Europeans (target above 140/90 mmHg or above 130/80 mmHg with a high CV risk) (Antza et al., 2021) creates a lot of confusion for clinicians, suggesting that a large number of patients remain without BP treatment. The clinical scenario around elevated BP with continual monitoring should be taken into careful consideration, and a decision around lowering BP treatment should always be carried out while placing the utmost priority on patient's outcomes. It is important to critically decide whether long-term treatment goals should have CV or renal protection benefits, as achieving a compatible target BP in order to balance the cardio-renal protection axis has proved challenging. Of importance to note is that the kidneys may play a key "cause and effect" role: patients with CKD have an elevated CV risk, and hypertension is one of the leading risk factors for CV events and stroke (Townsend and Taler, 2015), and is known to worsen renal impairment.

## 2 Pathogenesis of hypertension in chronic kidney disease

The pathogenesis of hypertension in patients with CKD is complex and multifactorial, and often resistant to treatment due to its complexity (Campese et al., 2006; Townsend and Taler, 2015; Hamrahian, 2022). Several putative mechanisms contribute to elevated BP in this disease, including neural and hormonal changes that often act in concert to disrupt appropriate BP regulation. The majority of these factors that contribute to the development of hypertension in CKD are described here (Figure 1).

## 2.1 Salt retention and volume expansion

Sodium retention is of primary importance, as under the influence of Ang II and aldosterone (RAS section in Figure 1), which both promote sodium reabsorption in the proximal and collecting tubules respectively (Rodriguez-Iturbe et al., 2007), there can be a total loss of GFR in renal disease. This impairs sodium excretion, and therefore leads to further sodium retention and extracellular expansion (Blaustein et al., 2006; Ku et al., 2019). Even though the degree of extracellular volume expansion may be insufficient to induce edema, hypertension can still be evoked (Vertes et al., 1969; Preston et al., 1996). Sodium retention causes hypertension through volume-dependent and volume-independent mechanisms (Sica and Carl, 2005). An excess extracellular volume (volumedependent) leads to increased perfusion of peripheral tissues, which stimulates vasoconstriction, heightening peripheral vascular resistance, and consequently rising BP (Dal Canton et al., 1996; Sica and Carl, 2005). This latter effect is ultimately due to the direct volume expansion, and indirect



#### FIGURE 1

Schematic representation of factors involved in kidney damage, hypertension and end-stage renal disease (ESRD). Figure shows complications arise upon activation of the renin-angiotensin system (RAS) secondary to kidney damage. Increased RAS activity subsequently activates angiotensin-1 receptor (AT<sub>1</sub> receptors) via increasing angiotensin II (Ang II) and decreases glomerular filtration rate (GFR). Angiotensin 1 receptor activation results in a myriad of effects such as vascular remodeling and inflammation, aldosterone release, sodium/H<sub>2</sub>O retention, disturbs the balance between antioxidant (superoxide dismutase; SOD) and oxidant (nicotine amide dinucleotide phosphate; NADPH) mechanisms, and increase sympathetic (Continued)

#### FIGURE 1 (Continued)

nervous system (SNS) activation. A reduction in GFR causes uremia, which is associated with increased asymmetric dimethylarginine (ADMA) and oxidative stress, all of which predispose to endothelial dysfunction and loss of nitric oxide (NO) functional signaling. Impairment of the NO pathway and increased reactive oxygen species (ROS) can subsequently predispose to the aggravation of SNS activation, vascular remodeling, and inflammation, further elevating systemic hypertension. Endothelin via endothelin-1 receptors (ET-1), on the other hand, can directly contribute to vasoconstriction and vascular inflammation and hypertrophy. In addition, CKD can also trigger vascular calcification, which is a hallmark of CKD complications and progression to ESRD. LVH, left ventricular hypertrophy;  $O_2^{\bullet}$ , superoxide free radical; VSMC; vascular smooth muscle cell. Figure adapted and modified from: (Palm et al., 2007; Grassi et al., 2010; Nauli et al., 2011; Briet and Schiffrin, 2013; Raina et al., 2020).

production of endogenous cardiotonic steroids or ouabain-like steroids (Schoner, 2002), which may further favor an increase in sodium load, BP and vascular remodeling (Blaustein et al., 2009). On the other hand, volume-independent mechanisms include increased vascular stiffness and increased central SNS outflow as a result of excessive sodium retention (Saxena et al., 2018) (SNS overactivity section in Figure 1). In addition to the aforementioned, excess sodium chloride load within the circulation also contributes to the autoregulation phenomena (Salem, 2002) (Figure 1). Physiologically, this autoregulation mechanism shields the capillary tuft of the glomeruli within nephrons from the elevated systemic BP. Thus, the afferent arteriole myogenic diameter changes in response to systemic BP reflexively, and sodium chloride delivery to the macula densa exerts a tubulo-glomerular feedback to maintain the intraglomerular pressure, and hence maintain a normal GFR. However, under the pathological state of chronic systemic BP elevation, the high pulsatility remodels and/or stiffens the afferent arteriole and reduces its ability to constrict and dilate. This subsequently leads to glomerular hypertension, nephrosclerosis, and progressive deterioration of kidney function. It is noteworthy to mention here with respect to certain medications like non-steroidal anti-inflammatory drugs (NSAIDs) such as indomethacin or ibuprofen, perturbs the afferent arteriole blood flow to the nephron via inhibition of the vasodilating PGI2 cyclooxygenase (COX) product (Baker and Perazella, 2020), and hence patients taking these drugs chronically can inadvertently accelerate kidney damage and impair the glomerulus shielding effect.

Alterations in vasoactive substances or abnormal ion channel function may play a role in the development of hypertension in ESRD (Martinez-Maldonado, 1998). For example, we know that sodium chloride intake potentiates the pressor action of endogenous catecholamines, such as norepinephrine (NE) (Ito et al., 1989) (Figure 1). An overt sodium intake results in stiffer blood vessels, vascular inflammatory changes, and loss in nitric oxide (NO) functionality (Figure 1); processes that are known to contribute to BP elevation (Hovater and Sanders, 2012). In the face of fluid and salt overload, in accordance with research strictly reducing sodium intake to < 1.5 g/d (Ku et al., 2019) is recommended, and that this should be the earliest nonpharmacological intervention to initiate (Ku et al., 2019; Masuda and Nagata, 2020). Such management strategies can be more robust at the start of a confirmed BP elevation, because with time, some patients can become resistant to sodiummediated changing maneuvers, by showing no discernible response in their BP when challenged by low and high sodium diets. The ability to maintain a normal BP state in the face of a sodium challenge may be hampered with progressive loss in kidney function, and a decline in GFR is aligned with a greater sensitivity of BP to salt (Koomans et al., 1982).

In view of the above, it can be concluded that salt retention can compromise CV regulation and negatively impact disease progression in CKD via both a direct and indirect actions, whereby salt expands plasma volume and alter regulation of vasoactive and hormonal substances key to BP control, respectively.

## 2.2 Sympathetic nervous system activity

Autonomic neuroregulation of CV function which governs periodic fluctuations in heart rate and BP, depends on a balance between the sympathetic and parasympathetic activity to the heart and vasculature (Salman, 2015). In CKD, a state of functional disharmony between sympathetic and parasympathetic arms of the autonomic nervous system, with altered tonic and reflex control of autonomic outflows, is seem, culminating in autonomic dysfunction and SNS hyperactivity (Salman, 2015). Overactivity of the SNS augments the high BP state via an increase in cardiac output and total peripheral resistance. This may result from direct actions on cardiac and vascular receptors, or by the renin-angiotensin system's (RAS) influence on the release of renin and sodium retention by the kidney (Martinez-Maldonado, 1998). This latter mechanism may thus contribute to target organ damage and adverse clinical outcomes, independent of BP (Kelm et al., 1996; Mancia et al., 1999; Greenwood et al., 2001) (Figure 1).

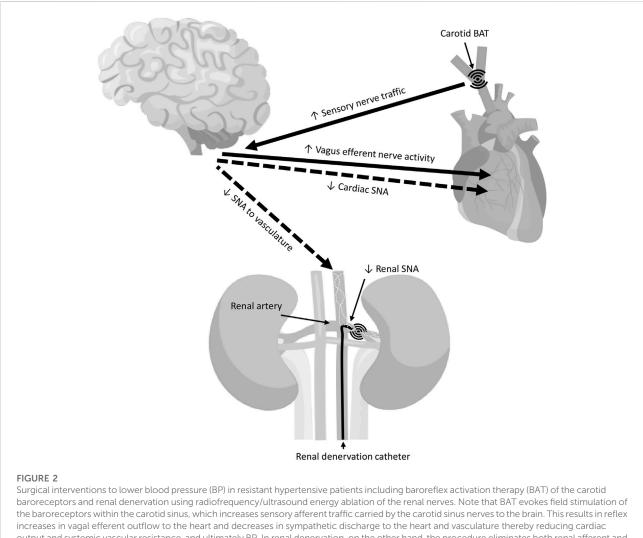
The enhanced SNS activity has been demonstrated in patients with CKD and ESRD (Campese and Krol, 2002; Neumann et al., 2004; Blankestijn, 2007; Vink et al., 2013; Sata et al., 2018). In addition to showing increased muscle sympathetic nerve activity (SNA) (Converse et al., 1992; Johansson et al., 1999; Neumann et al., 2004), renal failure patients also demonstrate increased circulating plasma catecholamines, as measured by plasma levels or total body NE spillover (Cerasola et al., 1998; Johansson et al., 1999; Zoccali et al., 2002; Klein et al., 2003a). Supporting this SNS overactivity is the evidence that administration of debrisoquin, a post-ganglionic sympathetic blocker, significantly reduces BP in hypertensive dialysis patients, but not in controls or normotensive dialysis patients (Schohn et al., 1985). The kidneys have a dual role in the pathogenic rise of sympathetic activity as they are both a recipient of efferent sympathetic signals as well as a generator of renal afferent activity (Vink et al., 2013; Sata et al., 2018). Stimulation of the renal efferent nerves results in a cascade of actions in the kidneys, and because of renal vasoconstriction, renal blood flow and GFR decrease, salt retention increases, and Ang II is produced. Here, the stimulation of Ang II is of particular importance, because Ang II directly causes vasoconstriction, has trophic effects, and as mentioned earlier, plays an important role in water and sodium reabsorption in the proximal tubule (Reid, 1992; Rodriguez-Iturbe et al., 2007) (Figure 1). The afferent signal in failing kidneys is also thought to contribute to an increase in BP.

reabsorption in the proximal tubule (Reid, 1992; Rodriguez-Iturbe et al., 2007) (Figure 1). The afferent signal in failing kidneys is also thought to contribute to an increase in BP. This is evidenced by correction of both BP and muscle SNA in patients who have undergone bilateral nephrectomy (Converse et al., 1992; Adamczak et al., 2002; Hausberg et al., 2002), and attenuation of elevated BP in animal models of renal failure, where dorsal rhizotomy (a procedure that selectively destroys the renal afferents), or renal denervation was performed (Ye et al., 1998; Campese and Krol, 2002; Ciriello and de Oliveira, 2002). Some might argue that the greater SNA in CKD might be a renal-specific response or due to a generalized stimulation of SNS activity. A study performed recently via electro-lesioning 5/6th of the cortex (glomeruli layer) of one kidney on New Zealand rabbits (Sata et al., 2020) compared renal sympathetic activity and NE spillover to those of sham control. Interestingly, their results suggested that nerve overactivity and global SNS activity originate from the kidneys, because circulating NE measurements were not the driver of increased BP in this model (Sata et al., 2020). These aforementioned results suggest that perhaps renal denervation may present a more effective treatment approach than the use of sympatholytic drugs in the context of systemic hypertension in CKD.

The relationship between increased SNS activity relative to vasculature can also be complex. As in the presence of SNS overactivity, the vasculature can become hypertrophied (Bevan, 1984), which in itself or as mediated by a complex interaction with the RAS can sensitize the vasculature to sympathetic vasomotor input (Adams et al., 1990; Smid et al., 1995); a phenomenon termed as the "vascular amplifier effect". We have previously addressed this issue in our rat model of chronic polycystic kidney disease (PKD), the Lewis polycystic kidney disease (LPK), which has an autosomal recessive variant of PKD (McCooke et al., 2012). In this study, we evaluated the extent of the relative contribution of the SNS versus the vasculature in the maintenance of hypertension in the LPK, its normotensive control (Lewis rat) and the spontaneously hypertensive rat (SHR) as a high BP rodent model (Ameer et al., 2014a). Our data revealed that despite the presence of exaggerated depressor effects in BP to direct vasodilators (response amplification) in the CKD model, the SNS overactivity as assessed by ganglionic blockade with hexamethonium still accounted for a larger depressor effect relative to that of direct vasodilation. This was further confirmed by our BP spectral analysis of Mayer's waves (low frequency power: range 0.2-0.6 and 0.4 Hz in rats), reflecting SNS vasomotor tone (Stauss, 2007). Indeed, relative to both the normotensive and hypertensive controls, the LPK demonstrated the steepest relationship when BP responses to ganglionic blockade were correlated with the low frequency oscillation in systolic BP (Ameer et al., 2014a). When directly recorded under both anesthetized (Salman et al., 2014; Salman et al., 2015a) and conscious (Salman et al., 2015b) conditions, baseline renal SNA was consistently higher in the LPK model of CKD compared to controls. Together, our data provide both direct and indirect experimental evidence of heightened sympathetic drive in CKD, and its potential contribution to the temporal decline in renal and hemodynamic function observed in this condition.

In experimental animals it has been demonstrated that ischemic metabolites or uremic toxins can evoke reflex increases in SNS activity and BP (Katholi, 1985), and that chronic stimulation of afferent nerves may lead to SNS overactivity and therefore an increase in BP too. Supporting this finding, an earlier study showed that uremic toxins have a stimulatory effect on SNS activity (Recordati et al., 1981). However, in contrast, other reports have shown ongoing increased SNA in patients with renal grafts, despite correction of uremia (Hausberg et al., 2002). These observations suggest that increased SNA appears to be driven by signals arising in the native kidneys that are perhaps independent of circulating uremia-related toxins.

In addition to the above, denervation of renal nerves around the renal artery have become an important strategy in the treatment of resistant or refractory hypertension among CKD and non-CKD patients (Burnier, 2022). The procedure itself (Figure 2) involves the use of catheter-based devices that ablate the renal afferent and efferent (sympathetic) nerves surrounding the renal arteries using either radiofrequency, ultrasound energy, or trans-arterial injection of caustic substances (Gajulapalli et al., 2020). Several trials have been conducted (Krum et al., 2009; Investigators, 2010; Esler et al., 2012; Kandzari et al., 2012; Bakris et al., 2014; Bhatt et al., 2014; Liu et al., 2017; Sievert et al., 2017; Böhm et al., 2020), however to date, there is no global census or legislative approval of renal denervation as a clinical procedure for the treatment of non-pharmacologically. hypertension Although renal denervation has played an ameliorating role in reducing elevated BP and preserving renal function in diseased kidneys in humans (Hering et al., 2012; Ott et al., 2015) and rats (Yao et al., 2017) with CKD, other factors contributing to BP elevation remain, that are unalterable by renal denervation. For example, in our studies, we have shown that despite a reduction in BP and



output and systemic vascular resistance, and ultimately BP. In renal denervation, on the other hand, the procedure eliminates both renal afferent and efferent activity including renal sympathetic nerve activity (SNA), the excessive activation of which is thought to be the driver of increased BP in hypertensive individuals. Adapted and modified from (Bhatt et al., 2019; Gajulapalli et al., 2020).

urinary protein of our CKD rat model, renal denervation did not fix the problem of increased arterial stiffness as assessed by pulse wave velocity (PWV), a hallmark of BP derived risk factor for morbidity (Yao et al., 2017). To date, the renal denervation field is challenged by: 1) a lack of trained personnel and professionals performing the procedure; 2) an unavailability of strict requirements for operation specifications; 3) nil available methods for measuring the effectiveness of denervation procedures; 4) a lack of definition with regards to which patients are optimal candidates for the denervation; 5) reported controversy in clinical outcomes, especially in relation to office or ambulatory BP readings, and GFR protection versus worsening; 6) the extent of the denervation process required; 7) nerve anatomical variations among patients; 8) the issue of re-denervation or renal nerves convergence (Yao et al., 2017) after a period of time following the procedure; 9)

variation in timing and follow-up during the trials (for example, 2–36 months); (10) the presence of good representative shamcontrols; and 11) the complications that arise following the procedures, which may range from minor adverse effects, to hematoma, renal artery narrowing or stenosis or dissection, acute renal failure, or even death (Sata et al., 2018; Ku et al., 2019; Liang et al., 2020). In view of the above, evidently a lot of factors need to be considered in renal denervation procedures, involving not just comparisons between clinical outcomes, but also cost effectiveness considerations between the denervation procedures and currently approved pharmacological therapy.

The participation of the SNS became more complex with the discovery of renalase oxidase, an amine oxidase enzyme (flavoprotein oxidase) involved in the regulation of cardiac function, BP and kidney injury (Xu et al., 2005; Wang et al., 2014; Czubilińska-Łada et al., 2020). It metabolizes

catecholamines and is mainly expressed in the glomeruli and proximal tubules of the kidney, cardiomyocytes and other tissues (Xu et al., 2005; Wang et al., 2014). Renalase is secreted into the plasma and urine of healthy persons, yet is undetectable in uremic individuals. Reports have shown that the renalase knockout mouse is hypertensive and exquisitely sensitive to cardiac ischemia (Wu et al., 2010), and develops renal tubular damage that can be ameliorated with recombinant renalase treatment (Lee et al., 2013). The administration of recombinant renalase produces a powerful and rapid hypotensive effect on rats (Schiffrin et al., 2007; Wang et al., 2017) and reduces plasma NE, left ventricular hypertrophy (LVH), and cardiac fibrosis in 5/6 nephrectomy hypertensive CKD rats (Baraka and El Ghotny, 2012). Blood renalase levels are inversely correlated with GFR and are markedly reduced in ESRD patients (Desir, 2009). Thus, SNS overactivity could be partly associated with abnormalities in the renalase pathway in CKD, as renalase deficiency may contribute to the heightened sympathetic tone observed in ESRD patients (Li et al., 2008; Desir and Peixoto, 2014). Indeed, extended research into the various pathways and mechanisms involved in this crucial enzyme activity would no doubt result in influential outcomes in tackling conditions such as high BP and kidney disease, especially when it comes to the use of this oxidase or its analogues as a therapeutic or diagnostic tool.

As previously reviewed (Salman, 2015), a major mechanism long thought to underlie altered elevated SNS activity and therefore BP in CKD is reduced arterial baroreflex function. In hypertension, it is hypothesized that the baroreceptor sensitivity is reset to a higher operating pressure. Possible explanations for this phenomenon are direct damage to the receptors, a change in the coupling between receptors and vascular walls, genetically determined properties of the receptors, and decreased distensibility of the vascular walls in which receptors are embedded (Salman, 2015). Heart rate baroreflex sensitivity, either spontaneous (Lacy et al., 2006; Studinger et al., 2006; Johansson et al., 2007) or evoked by pharmacologic manipulation of BP (Agarwal et al., 1991; Tinucci et al., 2001; Studinger et al., 2006), are blunted in CKD, and most importantly, deficits show a strong correlation with reductions in GFR (Lacy et al., 2006). This therefore suggests a direct association between deterioration in renal function and alterations in the cardiac baroreflex, rendering these markers useful in determining CV risk in CKD. In contrast to clinical studies investigating heart rate baroreflex, those directly assessing sympathetic baroreflex in CKD patients are conflicting, perhaps due to the limited number of reports directly assessing SNA baroreflex function in humans, alongside the more technically demanding experimental setup required to conduct those experiments, relative to heart rate baroreflex measurements. Ligtenberg and others (Ligtenberg et al., 1999) demonstrated comparable muscle SNA baroreflex sensitivity in hypertensive CKD patients and normotensive controls despite a resetting of the sympathetic baroreflex function to a higher BP range in the CKD group. In contrast, Tinucci and colleagues (Tinucci et al., 2001) demonstrated markedly attenuated muscle SNA baroreflex sensitivity in CKD patients relative to controls; therefore suggesting a pressing need for more clinical studies to elucidate the role of altered sympathetic baroreflex in mediating sympathetic hyperactivity in CKD. As far as animal studies are concerned, a huge body of evidence, as shown by our laboratory work (Harrison et al., 2010; Hildreth et al., 2013; Salman et al., 2014; Salman et al., 2015a; Yao et al., 2015) and others (Chen et al., 2016; Freitas et al., 2016), has consistently supported the presence of impaired cardiac and sympathetic baroreflex function in CKD - results which align more with the clinical data previously put forward by Tinucci and colleagues (Tinucci et al., 2001). Interestingly, over the last 2 decades, an overwhelming amount of attention has been directed toward correction of SNS overactivity, baroreflex dysfunction and high BP in the resistant hypertensive population. Such investigations have involved using pacing of the carotid baroreceptors (Figure 2) to increase their activity (also known as baroreflex activation therapy; BAT) (Bhatt et al., 2019) and hence reflexively reduce SNS central discharge (Illig et al., 2006; Heusser et al., 2010; Bisognano et al., 2011). This can be practically performed baroreflex amplification therapy (electrically by via barostimulator or mechanically by stent) and carotid body modulation (Groenland and Spiering, 2020). Several preclinical studies have been conducted on animals for proof of concept (Lohmeier and Iliescu, 2011), some with remarkable BP lowering effects. For example, Lohmeier and colleagues have shown that following a week of continuous baroreflex activation in dogs, a significant and sustained reduction in BP, heart rate, and NE levels, without a compensatory increase in plasma renin activity was observed (Lohmeier et al., 2004). A few clinical studies demonstrated some clinical benefits for BAT in the CKD population, including significant reductions in BP and improvements in kidney function, which could potentially lower CV risks in those patients (Wallbach et al., 2014; Beige et al., 2015; Wallbach et al., 2018). While baroreflex stimulation appears quite promising in treating resistant hypertension in CKD, longitudinal data are still inadequate and further research involving randomized, sham-controlled trials is required to define the long-term efficacy of this treatment approach and determine its ability to lower CV morbidity and mortality in CKD.

Taken together, SNS overactivity is a classical feature of CKD, and its role in the maintenance and progression of vascular disease in CKD is quite complex, given the well-established SNS interaction with vascular and hormonal mechanisms responsible for BP control. Despite a variety of interventions targeting SNS hyperactivity including non-pharmacological, pharmacological, and surgical interventions many patients remain undertreated, suggesting a pressing need for more innovative treatment approaches for sympathetic overactivity in the CKD population.

## 2.3 Renin angiotensin aldosterone system

In physiological terms, RAS has the capacity to regulate arterial pressure and sodium balance (Figure 1). When BP or perfusion falls, or SNA increases, the juxtaglomerular cells release renin which cleaves angiotensinogen, leading to an increase in Ang II levels (Morgado and Neves, 2012). The octapeptide Ang II has powerful direct vasoconstrictor effects, and stimulates the production of aldosterone, a mineralocorticoid which in turn increases renal sodium reabsorption, thus closing the regulatory feedback loop. If blood volume is normal, the increased RAS activity produces an abnormal rise in BP (Morgado and Neves, 2012). And, in patients with mild to moderate renal insufficiency, elevated plasma renin activity and Ang II concentrations correlate well with BP (Sinclair et al., 1987).

In CKD secondary to vascular disease, diabetes or PKD, reports have indicated that in areas of renal injury or ischemia induced by scarring, there is a greater production of local and intra-renal Ang II, which then exacerbates systemic hypertension (Acosta, 1982; Rosenberg et al., 1994; Fonseca et al., 2014). This increase in the RAS component is often responsible for at least part of the hypertension that persists after the restoration of normovolemia (Vertes et al., 1969; Preston et al., 1996). The hyperactivation of the RAS leads to elevation of peripheral vascular resistance, resulting in a vicious cycle and decline in cardiac performance, progressing to cardiac remodeling (Makaritsis et al., 2006). This is because Ang II itself is a proinflammatory mediator that can activate nuclear factor k-lightchain-enhancer of activated B cells (NF-kB) (Ruiz-Ortega et al., 2000) and vascular adhesion molecule-1 (VCAM-1) (Rodríguez-Vita et al., 2005). At the kidney, Ang II can promote renal fibroblasts, increase expression of transforming growth factor  $\beta 1$ (TGF-  $\beta$ 1), tissue growth factor, and fibronectin and collagen type I, which eventually increases renal fibrosis (Wolf, 1998; Lin et al., 2005). Furthermore, Ang II upregulates plasminogen activator inhibitor-1 and tissue inhibitor of matrix metalloproteinases-1, causing accumulation of extracellular matrix (Abrahamsen et al., 2002). In support of this, it has been shown that the AT<sub>1</sub> receptor antagonist, losartan, can prevent the development of CKD by maintaining renal blood flow, decreasing inflammation and tissue hypoxia following acute renal injury (Rodríguez-Romo et al., 2016).

The role of RAS is most apparent when considering the effectiveness of angiotensin-converting enzyme inhibitors (ACEi) or  $AT_1$  receptor antagonists/blockers (ARB) in controlling BP in renal insufficiency (Martinez-Maldonado, 1998), reducing muscle SNA (Ligtenberg et al., 1999; Klein et al., 2003b; Neumann et al., 2007), and providing renoprotection beyond BP control (Adamczak et al., 2002). Numerous studies have documented a key role for RAS effects on both the activity of the SNS and vascular remodeling. Development of vascular hypertrophy, remodeling and/or stiffness causes structurally based vascular amplification,

which produces an increased sensitivity of mean arterial pressure (MAP) to vasoactive agents, including an augmented response to removal of sympathetic vasoconstriction inputs, which may otherwise appear as an increase in sympathetic activity (Moretti et al., 2009).

Ang II is also known to mediate central sympathoexcitation (Veerasingham and Raizada, 2003; Fink, 2009; Sun et al., 2009). The mechanism by which Ang II is believed to alter sympathetic outflow to organs like the heart and vasculature involves activation of nicotinamide adenine dinucleotide (NADPH) oxidase and increased levels of reactive oxygen species (ROS) in neuroanatomical regions within the brain central to sympathetic stimulation (Gao et al., 2005; Gao et al., 2008; Schlaich et al., 2009). In experimental animal models, renallymediated local release of Ang II appears to mediate central activation of the SNS activity, as ARBs abrogate SNS activation caused by renal injury in rat experiments (Ye et al., 2002). A study conducted on our CKD rat as alluded to earlier, the LPK model, revealed that acute blockade of AT<sub>1</sub> receptor by IV losartan ameliorated not only BP but also improved baroreflex control of splanchnic SNA and further reduced the splanchnic, renal and lumbar SNA chemoreflex outflows in this CKD model (Yao et al., 2015); findings which thus support the role of Ang II in perpetuating the peripheral SNS reflexes during CKD progression.

With respect to the effect of Ang II on vascular remodeling in kidney disease, we have conducted a series of different experiments on our CKD rat model that evidently highlights the significance of RAS antagonism in CKD-presented vascular pathology. As an earlier study from our group has demonstrated, chronic administration of the ACEi, perindopril to the LPK rat causes a reduction in the PWV index of arterial stiffness and enhances central artery compliance (Ng et al., 2011). These results were not achieved by renal denervation of our animals (Yao et al., 2017), and whether this is the case in different CKD animal models or humans with kidney diseases has not been fully investigated. Hence, further research in this area is required in order to establish a clear ground on the clinical outcome of RAS antagonism versus that of renal denervation.

Regarding RAS pharmacological antagonism in CKD, the examination of the effect of chronic AT<sub>1</sub> blockade with valsartan on the function and structure of pathological vascular remodeling were assessed further in the LPK model of CKD in two different studies. In an earlier study, the aorta was used to investigate large artery dysfunction derived by CKD (Ameer et al., 2016). This study revealed that valsartan ameliorated the NE sympathetic mediator constricting effect on the aorta, normalized endothelium-dependent and -independent relaxations, improved NO functionality, and reduced large artery hypertrophy, wall elastin component fragmentation, calcification, vessel cystic degeneration and aortic wall matrix metalloproteinase-9 levels (Ameer et al., 2016). This myriad of effects suggests that AT<sub>1</sub> antagonism can extend to have effects

that are beneficial for CV health. In a later complementary study, the mesenteric artery was utilized to investigate small resistance artery remodeling in response to valsartan (Quek et al., 2018). Here, the chronic AT<sub>1</sub> antagonism had a detrimental effect on small artery geometry as it increased internal and external diameters and reduced the thickness of mesenteric walls alongside artery wall-lumen ratios. These effects would result in a significant impact on peripheral vascular resistance and vascular amplification (Adams et al., 1990; Smid et al., 1995; Ameer et al., 2014a), of which constitutes a hallmark of peripheral vascular resistance and a high BP that is often difficult to lower. The functional improvements in the large conduit artery seen in the earlier aorta study were also carried out in a relatively similar manner in smaller resistant mesenteric artery, especially with respect to improvement of endothelium dysfunction and NO contribution to relaxation. Furthermore, elevation of endothelium-derived hyperpolarization factor (EDHF) and the downplaying of prostanoids roles in resistant mesenteric artery of this CKD model were found to be a significant component of vascular-modulating autacoids. Interestingly, mesenteric artery biomechanical properties were further improved with valsartan treatment, as resistant artery compliance increased while intrinsic stiffness reduced; such effects could be reflected by changes in artery wall structural configuration of elastin and collagen components (Quek et al., 2018). No doubt, results from these investigations highlight the role of Ang II in driving vascular manifestations in CKD (Quek et al., 2018). Such findings also justify why early treatment with specific pharmacotherapy such as Ang II antagonism can potentially pan effects that not only induce BP reduction in CKD, but could limit pathological changes in arteries and improve the CV health beyond BP control; results of which may not be fully seen with other pharmacological interventions (Jeewandara et al., 2015; Quek et al., 2020). This likely explains why the ACEi and ARB are evidently taking the lead in treatments of hypertensive kidney disease (Unger et al., 2020).

As a result of RAS activation, the hormone aldosterone is produced. In addition to its renal sodium reabsorption effects, aldosterone is involved in various biological processes at the level of the vasculature. For instance, it can induce endothelial dysfunction through various mechanisms including reduction of NO bioavailability, increased inflammation, oxidative stress, and direct effects on glucose-6-dehydrogenase and epidermal growth factor, resulting in increased endothelial cell volume, stiffness and inability to release NO (Briet and Schiffrin, 2013). Aldosterone can also act on endothelial progenitor cells reducing vascular homing, migration, differentiation, and proliferation; the effect of which could cause reductions in circulatory colonyforming units of the endothelial cells and impairment of their migratory function (Briet and Schiffrin, 2013). Studies demonstrating the deleterious aldosterone effects suggest the involvement of this mineralocorticoid in tissue remodeling and fibrosis, as verified in various animal models of hypertension and kidney failure (Greene et al., 1996; Blasi et al., 2003; Nishiyama et al., 2004). Interestingly, increased oxidative stress and vascular inflammation are thought to be the first damaging effects of aldosterone on the vasculature, as it has been shown that in experimental uni-nephrectomized rats receiving aldosterone, a vascular increase in oxidative stress and inflammatory mediators preceded arterial fibrosis (Sun et al., 2002). Our research team has also investigated the effects of chronic aldosterone antagonism with spironolactone on the CV and renal function of the LPK model of CKD (Jeewandara et al., 2015). Unexpectedly, we found that spironolactone reduced BP in female rats only, and not in male rats, exhibiting sexual dimorphism for being resistant to BP reduction in male CKD rats. Furthermore, cardiac perivascular fibrosis, α-cardiac actin marker, and kidney function were evidently reduced with this treatment; however, spironolactone did not rectify the vascular remodeling (Jeewandara et al., 2015) and oversensitivity to constriction was seen in this model; changes that were evidently ameliorated with RAS antagonism as discussed earlier (Ameer et al., 2016; Quek et al., 2018). Thus, we know that aldosterone can exert damaging effects on the kidney and heart, that can have pathophysiological consequences (Hian et al., 2016).

Spironolactone has also been shown to help prevent cardiac and vascular fibrosis independently from reductions in BP (Benetos et al., 1997; Lacolley et al., 2001). For example, administration of spironolactone was found to prevent cardiac hypertrophic remodeling and increase expression of NADPH oxidase components in uremic rats (Michea et al., 2008), which further suggests that oxidative stress, proinflammatory and profibrotic actions of aldosterone are highly relevant pathophysiological conditions by which this mineralocorticoid is implicated in mediating CV injury (Briet and Schiffrin, 2010). A combination of RAAS axis antagonism (i.e. ACEi/or ARB plus aldosterone antagonist) is often used and can have beneficial effects in lowering proteinuria, and resistant BP in CKD patients (Unger et al., 2020); however, the establishment of ESRD progression with this combination still lacks characterization (Bolignano et al., 2014; Chung et al., 2020). Furthermore, this combination increases the risk of hyperkalemia and gynecomastia secondary to aldosterone antagonism, especially with old mineralocorticoid antagonists, and patients with such pharmacological interventions should be carefully monitored. Nevertheless, the development of new non-steroidal mineralocorticoid aldosterone receptor antagonists like finerenone has proven to have a superior safety profile to the old spironolactone and slightly newer eplerenone in heart and kidney disease patients (Capelli et al., 2020; Volpe and Patrono, 2020). The important clinical outcomes of these newly approved drugs on mortality rate, CV events, proteinuria and renal death reduction are yet to be evaluated.

As shown above, it is evident that most of our knowledge on the role of Ang II in CV control is heavily weighed on the

understanding of its modulatory action on AT<sub>1</sub> receptors. Despite decades long of physiological evidence (Johnson and Malvin, 1977; van der Mark and Kline, 1994) describing the counterregulatory role for AT<sub>2</sub> receptors, which often oppose the AT<sub>1</sub> action mediated by Ang II, their pharmacological targeting has been far less emphasized in the literature, perhaps due to lower AT<sub>2</sub> receptor expression levels in adults relative to that of AT<sub>1</sub> receptors (Ozono et al., 1997). Indeed, animal studies, using models of hypertension, heart and kidney disease and obesity, have shown that AT<sub>2</sub> receptor activation promotes diuresis and natriuresis, lowers BP, inhibits vascular calcification, suppresses chronic inflammation, mitigates cardiac damage, and improves renal structural and functional abnormalities (Johnson and Malvin, 1977; van der Mark and Kline, 1994; Gelosa et al., 2009; Kukida et al., 2019; Wang et al., 2020; Gavini et al., 2021). Further relating to this is the additive beneficial action for AT<sub>2</sub> receptor stimulation when AT<sub>1</sub> receptors are blocked, facilitating preferential stimulation of the unblocked AT<sub>2</sub> receptors (Kemp et al., 2012). It is therefore due to these beneficial outcomes that this pathway is now commonly referred to as the "protective arm" of RAS. A number of AT<sub>2</sub> receptor agonists have emerged from animal studies including Compound 21 (Kaschina et al., 2008), MOR107 (Wagenaar et al., 2013) and NP-6A4 (Gavini et al., 2021); however, to date, therapeutic effects of these compounds in ameliorating CV and renal diseases have yet to be thoroughly tested in phase II/III human trials. If proven effective, pharmacological targeting of AT<sub>2</sub> receptors may serve as a potential strategy to treat CV deficits associated with hypertension and CKD.

Collectively, one can acknowledge that RAAS plays a significant role in CKD development and progression, given its multifaceted contribution to the modulation of neuronal, vascular, and inflammatory pathways impacting CV and renovascular control. The efficacy ACEi and  $AT_1$  receptor blockers and their use as first-line agents to treat CKD patients is not surprising owing to their beneficial actions that extend beyond BP control. Moreover, the emergence of more research evidence regarding the protective role of  $AT_2$  receptors and their ability to augment the action of ARBs may represent a novel therapeutic approach to treating CV complications associated with CKD.

# 2.4 Role of the vasculature in hypertension in chronic kidney disease

One issue contributing to the high BP state in CKD is the presence of vascular changes. The vessels of CKD patients are continuously exposed to numerous hormones, factors and uremic toxins that can be responsible for the observed vascular changes and dysfunction, which in turn causes a rise in BP that is often difficult and resistant to treatment with drug

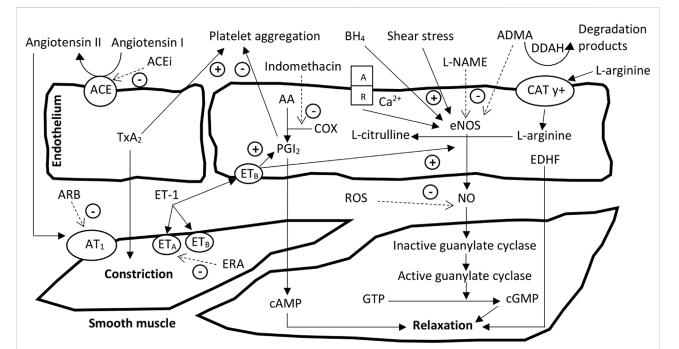
therapy. Vascular dysfunction can take place early in the progression of CKD and although it appears to be independent of hypertension development, it is considered a definite part in the hypertension sequalae (Schiffrin et al., 2007; Ameer et al., 2015; Six et al., 2020). As mentioned earlier with our findings from a CKD animal model, vascular changes may be structural, as characterized by vessel remodeling that favors increased contractility and a loss in elasticity causing vascular resistance (Ameer et al., 2014b; Ameer et al., 2016; Quek et al., 2016); or they may be associated with functional changes such as the regularly encountered defect in acetylcholine (ACh) relaxation (Figure 3) (Morris et al., 2000; Ameer et al., 2015; Roumeliotis et al., 2020; Six et al., 2020). Such changes are often associated with an overexpression of the intracellular adhesion molecule-1 (ICAM-1) and VCAM-1 (Safar et al., 2003; Rodríguez-Vita et al., 2005; Six et al., 2020). We know thus far these vascular alterations have been associated with the development of cardiac pathologies, further raising BP and worsening during CKD progression (Maizel et al., 2009). Of note is that the consistent observation of endothelium dysfunction in CKD patients can be the result of an increase in endothelial injury and a reduction in endothelial repair (Goligorsky et al., 2010), and this dysfunction has been associated with numerous systemic BP and hemodynamic alterations.

## 2.4.1 Endothelial dysfunction

Endothelial dysfunction can be described as a systemic pathological state of the endothelium (the inner most lining of blood vessels) that arises from an imbalance between vasodilating and vasoconstricting substances produced by (or acting on) the endothelium (Deanfield et al., 2005). This is because the endothelium itself acts as a dynamic and functionally complex organ that is involved in the regulation of many crucial biological mechanisms, including maintenance of vascular tone and permeability, inflammatory responses, immunity, and angiogenesis (Roumeliotis et al., 2020).

In patients with CKD, endothelial dysfunction may have a dual role, for not only is it a crucial step in the development of CVD, but dysfunction and disintegration of endothelial cells in glomerular capillaries, and particularly those capillaries in renal medulla pave the way for CKD progression (Fliser et al., 2011; Harlacher et al., 2022). Furthermore, the endothelial dysfunction may serve as an early basis of atherosclerosis during the early stages of CKD, and the prevalence of this dysfunction is progressively high toward later ESRD (Yilmaz et al., 2006). Importantly, the loss of endothelium integrity is highly paralleled by increased mortality rates from CVD incidence (Roumeliotis et al., 2020).

There are many endothelium-derived relaxing factors (EDRFs) and endothelium-derived constricting factors (EDCFs) that have been speculated to play a role in hypertension associated with kidney disease (Shultz, 1992;



#### FIGURE 3

Major signaling pathways involved in endothelial and vascular smooth muscle function. Cationic amino acid transporter-y + regulates uptake of L-arginine substrate into the endothelial cell. Agent A, acting on receptor (R) of the endothelial cell activates  $Ca^{2+}$  influx: for example, acetylcholine (ACh) acts on the endothelial muscarinic (M<sub>3</sub>) receptor causing  $Ca^{2+}$  influx. The increase in intracellular  $Ca^{2+}$  leads to activation of endothelial nitric oxide synthase (eNOS). Shear stress or substrate supplementation of tetrahydrobiopterin (BH<sub>4</sub>) can increase activation of eNOS while the latter is inhibited by asymmetric dimethylarginine (ADMA). Nitric oxide (NO) endothelial derived relaxing factor (EDRF) then diffuses to the smooth muscle cells where it activates guanylate cyclase, which results in an increase in cyclic guanosine monophosphate (cGMP), and thus initiates processes leading to relaxation. N<sup>m</sup>-nitro-L-arginine methyl ester (L-NAME) is a NOS inhibitor, and reactive oxygen species (ROS) act as NO scavengers. Endothelin is an endothelial ET<sub>B</sub>. Arachidonic acid (AA) is a membrane phospholipid that gives rise to prostanoids via cyclooxygenase (COX). Prostacyclin I<sub>2</sub> (PGI<sub>2</sub>) and thromboxane A<sub>2</sub> (TxA<sub>2</sub>) are prostanoids. The former causes vascular relaxation and inhibits platelet aggregation, while the latter produces the opposite of these effects. COX can be inhibited by nonsteroidal anti-inflammatory drugs like indomethacin and thus inhibit prostanoid formation. Angiotensin converting enzyme (ACE) produces angiotensin II (Ang II); Ang II can cause smooth muscle contraction via angiotensin receptor (AL) activation. ACE inhibitor (ACE) plocks Ang II formation. Angiotensin receptor s. Adapted and modified from (Cohen and Vanhoutte, 1995; Félétou and Vanhoutte, 2006; Wong and Vanhoutte, 2010; Raina et al., 2020; Roumeliotis et al., 2020).

Martinez-Maldonado, 1998; Zhao et al., 2021). These and pharmacological agents that are commonly used to target or influence them, are shown in Figure 3.

Endothelium dysfunction can have an impact on the function and structure of the vasculature. Functionally, impaired endothelium-dependent vasodilation has been observed in CKD and ESRD patients (Passauer et al., 2000; Thambyrajah et al., 2000; Wang et al., 2000; Amabile et al., 2005), and has been described as a pervasive phenomenon in uremic individuals, which may later influence morbidity and mortality from large vessel atherosclerotic disease and LVH (Zoccali et al., 2003). Microalbuminuria, a marker of glomerular hyperfiltration, has been correlated with and may be a manifestation of impaired endothelial function (Stehouwer and Smulders, 2006), and is suggested to participate in the mechanisms that lead to progression of renal disease (Fujihara et al., 1995). Indeed, it has been postulated that glomerular endothelial dysfunction is an early feature of essential hypertension that may precede BP elevation (Schiffrin et al., 2007). Structurally, endothelium dysfunction is implicated in arterial remodeling by several mechanisms, including inhibition of platelet aggregation and monocyte adhesion to the endothelial cell, abrogation of lowdensity lipoprotein (LDL) cholesterol oxidation, and atherosclerosis (Pannier et al., 2000; Van Varik et al., 2012; Roumeliotis et al., 2020). Furthermore, endothelium influences vascular wall homeostasis via the endothelial cell cytokines and growth factors (Csiszar et al., 2009; Urschel et al., 2012; Van Varik et al., 2012). Endothelial cells can also affect the state of vascular hypertrophy via the production of TGF-β and bone morphogenetic proteins (BMPs) which stimulate vascular smooth muscles and pericytes to remodel and deposit extracellular matrix (Boström et al., 1993; Simionescu et al., 2005; Van Varik et al., 2012). Therefore, the preservation of the functional and structural integrity of the vascular endothelial cells is fundamental for not only vasculature hemodynamics, but also the global CV health state.

#### 2.4.2 Nitric oxide signaling

Vascular endothelial cells cause vasodilatation as a normal response to increases in flow, shear stress and agonists through the release of several mediators, which include NO, prostaglandin  $I_2$  (PGI<sub>2</sub>) and EDHF (Figure 3) (Campese et al., 2006; Roumeliotis et al., 2020).

Nitric oxide is a gaseous molecule with vasorelaxant, antiinflammatory, and antithrombotic properties (Vanhoutte et al., 2016), and is probably one of the most important molecules produced by the endothelium. Reduced NO bioavailability (Figure 1) appears to be one of the main factors involved in CKD-associated endothelial dysfunction and its hallmark (Wever et al., 1999; Hasdan et al., 2002; Passauer et al., 2005; Roumeliotis et al., 2020).

Nitric oxide is synthesized by the enzyme nitric oxide synthase (NOS) from the precursor amino acid L-arginine (L-Arg) (Figure 3). Nitric oxide synthase is present in three isoforms named after the different tissues they were initially discovered in (Lüscher and Vanhoutte, 1990; Bruno and Taddei, 2011), though each distinct isoform has since been discovered to also be expressed in other tissues. Neuronal NOS (nNOS) is constitutively expressed in central and peripheral nervous system neurons, where NO acts as an important neurotransmitter and plays a key role in the autonomic regulation of CV function (Patel et al., 2001; Hirooka et al., 2011), and is also expressed in skeletal muscle and the epithelial layers of the airways. Neuronal NOS is also present in macrophages and endothelial cells, where it seems to play a role in the regulation of basal vascular tone (Seddon et al., 2008), and has been implicated in the rise of central SNS activity in CKD patients (Park et al., 2015). Inducible NOS (iNOS) is expressed in a number of different cellular types and its activity is minimal under baseline conditions, but it is progressively induced by inflammatory stimuli (Lüscher and Vanhoutte, 1990) and was originally discovered in macrophages. Endothelial NOS (eNOS) is constitutively expressed not only in the vascular endothelial cells, but in various cell types including neuronal cells (Bruno and Taddei, 2011), myocardial cells, and skeletal muscle. Nitric oxide released from the endothelium is triggered by receptor-mediated mechanisms including ACh, bradykinin, serotonin, substance P, adenosine diphosphate and triphosphate, histamine and endothelin-1 via endothelin B receptors, which are all Ca2+dependent (Figure 3) (Roumeliotis et al., 2020); and by nonspecific mechanical stimuli such as shear stress by blood flow or hormonal growth factors like vascular endothelial growth factor, insulin and adiponectin - all of which are Ca2+-

independent mechanisms (H Heiss and M Dirsch, 2014; Roumeliotis et al., 2020). Remarkably, the shear stress on the vascular walls is the most powerful mechanism of stimulated NO release (Bruno and Taddei, 2011). Endothelial-derived NO has the capacity to maintain vascular tone and to produce vasodilatation through the activation of guanylate cyclase and cyclic guanosine monophosphate (cGMP) (Figure 3) (Bruno and Taddei, 2011; Roumeliotis et al., 2020).

It has been reported that various factors can impair the NO pathway during CKD and consequently cause further elevation in BP (Figure 1). The oxidative stress state is one example, whereby an imbalance between antioxidants and oxidants leads to potential damage (Sies, 1997; Vaziri et al., 1998; Daenen et al., 2019). Thus, reductions in antioxidant mechanisms such as catalase, superoxide dismutase (SOD) and glutathione dismutase (Vaziri et al., 2002); and elevation in oxidation mechanisms such as superoxide, hydrogen peroxide (Vaziri et al., 2002), activation of the reduced form of NADPH oxidase, xanthine oxidase, uncoupled eNOS, myeloperoxidase, and mitochondrial oxidases (Vaziri et al., 2002; Daenen et al., 2019) could be potential pathways that lead to an oxidative stress state, which would result in increased ROS in uremia. Uncoupling of eNOS under pathological conditions results in ROS production, caused by electron transfer within eNOS active sites to uncouple it from L-Arg and oxygen reduction to superoxide free radical  $(O_2^{\bullet})$  (Alp and Channon, 2004).

In addition, it has been demonstrated that vascular smooth muscle NADPH oxidase is activated by Ang II and subsequently increases vascular ROS levels (Griendling et al., 1994), with the NADPH p47 (phox) subunit being probably one of the most important oxidation sources in the vasculature that is also upregulated by Ang II (Touyz et al., 2005); a finding which has been evident with the increased aortic expression of p47 phox toward ESRD in a CKD rat model (Ameer et al., 2015). The oxidant free radical rapidly reacts with NO to form the highly reactive intermediate peroxynitrite (ONOO-) (Daenen et al., 2019). The formation of these nitroso-compounds has multiple negative effects, reducing NO availability, exerting direct vasoconstriction, and impairing the activity of prostacyclin synthase and eNOS (Munzel et al., 2010), thus contributing to an increase in BP in CKD (Hu et al., 1998). Furthermore, it has been established that lipid oxidation products such as oxidized LDL promote the displacement of eNOS from its surface membrane caveolae to the cell cytoplasm, where its activity is consequently directly inhibited (Blair et al., 1999; Gharavi et al., 2006; Roumeliotis et al., 2020). This occurrence can be translated to the inverse relationship between oxidized-LDL with vascular distensibility, such as the carotid artery in CKD patients (Nawrot et al., 2010).

Renal failure also plays a role as a trigger for inflammatory responses, which influence NO bioavailability (Schiffrin et al., 2007). Hypoxia, tumor necrosis factor- $\alpha$  and inflammatory cytokines are main stimuli that can negatively influence eNOS

expression (Lüscher and Vanhoutte, 1990). Moreover, accumulation of inflammatory markers such as C-reactive proteins, uremic toxins, advanced glycation end products (AGE) (Vaziri et al., 1998; Jofré et al., 2006) and Ang IImediated amplification of inflammatory mechanisms (Pecoits-Filho et al., 2003) can also impact NO bioavailability negatively. It has been demonstrated that cell transfusion of healthy endothelial cells into 5/6 nephrectomized rats improves NOmediated endothelial dysfunction in the renal failure state (Pacurari et al., 2013). Therefore, it is likely that the concomitant uremia state associated with renal impairment can be a trigger for diminished NO bioavailability and progressive endothelial dysfunction; a vascular calamity that can further deteriorate renal function in CKD (Schiffrin et al., 2007; Roumeliotis et al., 2020).

The L-Arg and pteridine cofactor tetrahydrobiopterin (BH<sub>4</sub>) are essential precursors for NO production (Figure 3) (Daenen et al., 2019), and their reduction has been associated with limiting NO bioavailability and decreased endotheliumdependent relaxation (Moreau et al., 2012; Zhang et al., 2012). Supporting the former precursor involvement, we found that acute addition of L-Arg in vitro vascular preparations modestly improves the endothelium-dependent relaxation of the aorta in a polycystic CKD rat model (Ameer et al., 2015; Ameer et al., 2016). Although transiently demonstrated, this effect would roll out the absence of defects in L-Arg uptake through the endothelial cationic transporter (CAT y+) of L-Arg (Figure 3) (Schiffrin et al., 2007; Roumeliotis et al., 2020), at least in the early stages of renal impairment of this CKD model. Despite that there has been no absolute cutoff regarding the benefits or harm of the acute or chronic L-Arg supplementation in CKD (Peters et al., 1999; Cross et al., 2001; Düsing et al., 2021), L-Arg deficiency is becoming a leading factor for eNOS deficits, and its substitution can show some positive CV effects in renal insufficiency (Amador-Martínez et al., 2019; Düsing et al., 2021; Hsu and Tain, 2021). Note however that others have reported that chronic L-Arg intake might predispose to endothelial cells aging (Scalera et al., 2009), and its acute intake does not improve the endothelial dysfunction encountered in kidney disease (Cross et al., 2001). With respect to the effects of BH4 on CV health, several studies have demonstrated a role for this NOS re-coupler in various conditions, including but not limited to: improving endothelial function and arterial stiffness in post-menopausal women (Moreau et al., 2012), oxidative stress in arterial diseases (Maier et al., 2000; Cosentino et al., 2008), hypertension (Porkert et al., 2008), NO-mediated vascular relaxation in type II diabetes (Heitzer et al., 2000), protection against atherosclerosis and vascular inflammation in a mice model (Schmidt et al., 2010), reduced elevated BP, improved kidney function in 5/6 nephrectomy rat model (Podjarny et al., 2004), reduced arterial wall thickness and collagen remodeling in a chronic ischemia rat model (Akaihata et al., 2020), and reversal

or alleviation of vascular pathologies associated with CKD (Ameer et al., 2014b; Ameer et al., 2015; Quek et al., 2016). Despite the aforementioned, it is still critical to know that certain underlying etiologies like oxidative stress may still hamper the beneficial effects of BH<sub>4</sub> treatment, especially in kidney disease (Channon, 2020). It is therefore crucial to revisit the effect of these bio-precursors and reveal their key role in NOS-mediated activities, especially in the treatment of the high BP state in CKD animal models and humans, and research and optimization of NO re-coupler and precursors are of importance within this area of NO-linked vascular pathology.

Another feature associated with impaired NO signaling is the accumulation of asymmetric dimethylarginine (ADMA) (Figures 1, 3) (Channon, 2020), and its less researched inactive enantiomer symmetric dimethylarginine (SDMA) (El-Khoury et al., 2016; Six et al., 2020). Asymmetric dimethylarginine, a naturally occurring amino acid, is an endogenous competitive inhibitor of NOS (Vallance and Leiper, 2004), which can cause endothelial dysfunction and/or reduced NO production particularly in renal failure (Leone et al., 1992; Zoccali et al., 2001), and is associated with increased CV risk (Bruno and Taddei, 2011). It is also reported to be elevated in CKD (Leone et al., 1992; Fleck et al., 2001) and ESRD patients on dialysis (Anderstam et al., 1997). Interestingly, ADMA levels have been found to correlate with the severity of atherosclerosis (Kielstein et al., 1999), CV events, and mortality in CKD patients (Zoccali et al., 2001), and even with renal disease state progression (Fliser et al., 2005). Asymmetric dimethylarginine levels can be influenced by Ang II through the activation of AT<sub>1</sub> receptors; thus, Ang II can indirectly predispose to further NOS inhibition (Palm et al., 2007), perpetuating to eNOS uncoupling with free radicals generation (Vallance and Leiper, 2004). Asymmetric dimethylarginine is mainly metabolized by dimethylarginine dimethylaminohydrolase (DDAH) and cleared by the kidney (Figure 3) (Schiffrin et al., 2007; Roumeliotis et al., 2020).

The synthesis of ADMA requires the enzyme protein arginine methyltransferase, which methylates arginine residues (Schiffrin et al., 2007; Channon, 2020). Asymmetric dimethylarginine enters vascular endothelial cells via the same CAT y + transporter of L-Arg (Figure 3). The activity of this transporter colocalizes with caveolin-bound NOS, which suggests that CAT y + transporter activity may be a determinant of the local concentrations of ADMA. The ADMA then competes with L-Arg substrates for transport into the cell. Thus, ADMA may block entry of L-Arg, resulting in a decrease in NO synthesis (Schiffrin et al., 2007; Channon, 2020). Adding to these complexities, DDAH-mediated catabolism of ADMA is severely hampered by the presence of circulating inflammatory cytokines, ROS, hyperlipidemia, hyperglycemia and their related consequence including AGEs (Roumeliotis et al., 2020); many of which are associated with the uremic state in CKD (Schiffrin et al., 2007; Roumeliotis et al., 2020).

#### 2.4.3 Prostanoids signaling

The prostanoids are eicosanoids derived from autocoids from the membrane of the cell, and are part of the EDRFs and EDCFs (Zhao et al., 2021). The prostanoids are a family of lipid mediators generated by the action of cyclooxygenase (COX) on a 20-carbon unsaturated fatty acid, arachidonic acid. Two major isozymes encoded by different genes, COX<sub>1</sub> and COX<sub>2</sub>, mediate this production process (Smith et al., 2000) and are differentially effective, mainly as vasodilators and vasoconstrictors regulating vascular tone (Figure 3) (Qi et al., 2002; Zewde and Mattson, 2004), while the newly discovered variant COX<sub>3</sub> enzyme is still under investigation with particular importance in brain tumors (Zhao et al., 2021). Prostanoids encompass prostaglandins (PGI<sub>2</sub>, PGD<sub>2</sub>, PGE<sub>2</sub> and PGF<sub>2</sub> $\alpha$ ) and thromboxane (TxA<sub>2</sub>), which are generated in a paracrine or autocrine manner, and are all implicated in CVD (Smyth et al., 2009; Wu et al., 2021), especially in conditions associated with a state of downregulation of EDRFs and upregulation of EDCFs, such as hypertension (Liang et al., 2019). Prostanoid analogues are becoming a good therapeutic key in the treatment of pulmonary hypertension (Pan et al., 2020) while their synthesis inhibitors like the NSAIDs are often associated with CV and renal deleterious effects, in particular in polypharmacytreated patients (Cabassi et al., 2020). Prostaglandin I2 is most abundant in the aorta followed by TxA2, PGE2, and PGF2a (Qi et al., 2006). Because of their short half-life, prostanoids neither circulate nor impact directly on systemic vascular tone. However, they may modulate local vascular tone at the site of their formation and affect systemic BP (Smyth et al., 2009; Wu et al., 2021). Prostanoids are involved in maintaining normal homeostasis and kidney function, thus PGI2 deficiency can induce renal fibrosis and vascular injury, including arteriosclerosis and wall thickening of the aorta. Prostaglandin I<sub>2</sub> signaling is essential to oppose vasoconstrictor pathways for maintenance of renal homeostasis, and is an important vasodilatory renal prostanoid, which can potentially influence numerous pathological mechanisms implicated in various aspects of renal disease, including renal hemodynamic changes, changes in GFR, oxidative stress, inflammatory processes, and platelet aggregation in renovascular disease complicated by hypertension (Nasrallah and Hébert, 2005; Zhao et al., 2021). While, the downstream of COX<sub>2</sub> derived prostanoids like TxA<sub>2</sub> is a key contributing factor in mediating vasoconstriction (Figure 3) and is implicated in endothelial dysfunction and raising BP (Zhao et al., 2021). Furthermore, it has been suggested that impairment of prostanoids-mediated relaxation may contribute to endothelial dysfunction in the aorta of hypertensive rats (Tang et al., 2008). Experimental evidence revealed that upon chronic Ang II inhibition by ACEi, suppression of vasoconstrictor prostanoids production and improvement of flow-mediated dilation in arteries from congestive heart failure rats were achieved (Varin et al., 2000). It is noteworthy to mention that oxidative stress causes an

upregulation of COXs in hypertension (Paravicini and Touyz, 2008) and CKD, accelerating CVD incidence (Mathew et al., 2017), and the high uremic solute indole-3 acetic acid triggers COX<sub>2</sub> expression elevation through endothelial inflammation and oxidation (Dou et al., 2015). Thus, the existence of positive feedback between ROS and COX, and vasoconstricting prostanoids altogether exacerbate BP elevation. Furthermore, a crosstalk relationship between Ang II and the COX system exists, whereby COX activity has been shown to modulate the pressor effect of Ang II on BP (Qi et al., 2002), and Ang II itself acts as a COX<sub>2</sub> inducer (Zhao et al., 2021). With current advances in research, there is mounting evidence suggesting that the therapeutic potential of drug discoveries is partially dependent on their ability to target prostanoid biosynthesis and their receptors in the treatment of various CVDs including hypertension and nephropathy. Thus far, research efforts are still required to investigate and pinpoint the exact contribution of targeting COX or particular prostanoid receptors, and to clarify the misconception arising from animal research and human studies relating to prostanoids as a therapeutic target.

# 2.4.4 Endothelium-derived hyperpolarizing factor

Although EDHF's exact identity remains unknown, EDHF is evidently an important NO, PGI2-independent agonist induced relaxation pathway (Figure 3) (Luksha et al., 2009), and TxA2 receptor antagonist (Zhao et al., 2021). It acts via relaxing the vascular smooth muscle cell (VSMC) through putative potassium channels and desensitizes the cell to vasoconstrictor stimuli (Chen and Suzuki, 1990). Notably, reserved EDHF protection may serve as a compensatory vasodilatory mechanism in the disease state (Katz and Krum, 2001; Quek et al., 2016; Lobov and Ivanova, 2020). Reports have demonstrated the presence of extensive interaction between NO and EDHF, and that EDHF activity might be enhanced or depressed depending on the state of hypertension, vessel studied, and species (Luksha et al., 2009; Lobov and Ivanova, 2020). For example, EDHF responses were shown to be preserved in mesenteric arteries from the LPK (Quek et al., 2016) and one kidney-one clip Sprague-Dawley rat model of renovascular hypertension (Christensen et al., 2007), yet diminished in the mesenteric arteries from Wistar rats of the same renal failure model (Vettoretti et al., 2006). In contrast, others have reported upregulation of EDHF responses in carotid arteries from two kidney-one clip Wistar rats (Sendao Oliveira and Bendhack, 2004), and an impairment of EDHF mediated relaxation in response to bradykinin but not ACh in subcutaneous resistance arteries from uremic patients (Luksha et al., 2012). Despite this mixed evidence, EDHF as a part of endothelium mechanisms are indeed an important NO deficiency backup component, and are implicated in aggravation of the hypertensive state and end-organ damage (Luksha et al., 2009).

Endothelial dysfunction may result in abnormalities that are not manifested as reduced endothelium-dependent vasodilation (Deng et al., 1995). For example, a dysfunctional endothelium can be associated with changes of endothelial cell phenotypes with increased proliferation, anoikis, altered morphology, production of C-reactive protein, and other inflammatory and thrombogenic mediators, including monocyte chemotactic protein-1 and plasminogen activator inhibitor-1, upregulated adhesion molecules, and enhanced thrombogenicity and adhesiveness for circulating cells (Safar et al., 2003). In a similar fashion reviewed here with respect to oxidative stress affecting NO and prostanoids, ROS and, most importantly superoxide anion, decreases EDHF-induced vasorelaxation; a process which involves decreasing the activity of calciumsensitive potassium channels and acting on cellular junctions, which thus alters the spread of hyperpolarization of the endothelial cells at the VSMCs (Griffith et al., 2005; Kusama et al., 2005).

#### 2.4.5 Endothelin signaling

Endothelin is a family of four 21 amino acid peptides, and endothelin-1 (ET-1), the main isoform, is produced by endothelial cells. Although the other potent vasoconstrictors and pro-fibrotic growth factors isoforms ET-2 and ET-3 are known (Yanagisawa et al., 1988), ET-1, is the predominant vascular isoform with the greatest regulatory effect on vascular tone, and is integral in the regulation of renal and CV pathophysiology (Raina et al., 2020). Endothelin can mediate vasoconstriction when it binds to its A and B receptors (ET\_A and ET\_B) in VSMCs, vasodilation  $\mathrm{ET}_{\mathrm{B}}\text{-}$ mediated via prostacyclin, NO release from the endothelial cells (Figure 3) (de Nucci et al., 1988), and natriuresis in the kidneys (Schneider and Mann, 2014). Nevertheless, endothelin vasoconstricting effect outweighs the vasorelaxing effect, as the latter remains very small and is almost negligible (Channick and Rubin, 2002). It has been found that ET-1 contributes to the pathogenesis and maintenance of hypertension, arterial stiffness and enhancement of CV risk factors such as oxidative stress, inflammation, and endothelial dysfunction (Dhaun et al., 2006; Dhaun et al., 2011; Raina et al., 2020). Research evidence also supported a role for endothelin in regulating autonomic function, demonstrating a modulatory action for endothelin on carotid bodies, superior cervical ganglion and nodose ganglion to influence baroreflex and chemoreflex regulation. Further, a role for endothelin in promoting catecholamine release from the sympathetic nerve terminals and adrenal glands also has been shown (Mortensen, 1999; Salman, 2015). In the kidney, ET-1 via ET<sub>A</sub> facilitates fibrotic pathways to cause nephropathy progression in conditions such as hypertension, diabetes, dyslipidemia, glomerular sclerosis, and autosomal dominant PKD (Wesson, 2006; Dhaun et al., 2011; Kohan and Barton, 2014; Raina et al., 2020). Furthermore, ET-1 expression may be pathologically

enhanced in kidney disease via uremic toxins of AGEs (Six et al., 2020).

A significant crosstalk exists between the RAAS and ET system (Figure 1), as Ang II induces aldosterone release from the adrenal cortex, which in turn increases renal ET-1 expression (Mennuni et al., 2014). Concurrently, renal ET-1 expression is also increased secondary to a rise in vascular NADPH oxidase, NF-kB and ROS due to Ang II actions (Mennuni et al., 2014; Raina et al., 2020). The resultant increased ET-1 activation causes severe renal cortex and medullary vasculature constriction, which eventually reduces renal blood flow and initiates renal fibrosis, a process already paved by the profibrotic effects of ROS and NO level elevations and reductions, respectively (Mennuni et al., 2014).

Endothelin receptor antagonists (ERAs) (Figure 3) have been implicated both in human and animal research studies, especially when needed to tackle hypertension and kidney disease (Goddard et al., 2004; Kohan and Barton, 2014; Breyer and Susztak, 2016; Trachtman et al., 2018; Raina et al., 2020); the ultimate etiologies for ESRD (Mennuni et al., 2014). For example, in the experimental rat model of hypertension and NO dysfunction with deoxycorticosterone acetate (DOCA)-salt and nitric oxide synthase inhibitor Nw-nitro-L-arginine (NOARG), a prototype ET<sub>A</sub> antagonist treatment produced a major protection against hypertensive nephron damage evidently mediated via NF-kB and ET-1/ET<sub>A</sub> pathways (Kimura et al., 2012). Interestingly, it has also been demonstrated that the  $ET_A$ antagonist avosentan, in combination with the ARB antagonist valsartan, gave rise to albuminuria reduction and improved mortality rate in a transgenic hypertensive rat model, relative to the ARB monotherapy (Baltatu et al., 2014); hence supporting the existence of a reno-protective effect for ERA and crosstalk effects of ET with RAS. Such effects have given rise to the clinical study DUET on the new and first in its' class sparsentan, which acts as a dual ET<sub>A</sub> and AT<sub>1</sub> antagonist (Trachtman et al., 2018). In this study, the dual blockade by sparsentan was compared to the conventional ARB irbesartan in focal segmental glomerulosclerosis kidney disease patients, and the results showed that sparsentan was tolerable for up to 8 weeks of treatment with a better reno-protection and proteinuria reduction against that of irbesartan (Trachtman et al., 2018).

Despite the promising research pointing to ERAs as a possible therapeutic option in hypertensive nephropathy, and due to the widespread physiological activity of endothelin across multiple organ systems, ERAs use has unfortunately been accompanied with significant adverse effects. Such effects have ranged from the dangerous and common fluid retention, to exacerbating congestive heart failure, and also included less common hepato-and testicular toxicities and teratogenicity (Kohan and Barton, 2014; Raina et al., 2020). Thus, these side effects severely restrict patients' eligibility for treatment and limits its clinical usefulness. Current standard available therapy involves ACEi/ARB and may also include the

10.3389/fphar.2022.949260

increasingly popular use of sodium-glucose cotransporter-2 inhibitors (SGLT<sub>2</sub>i); hence ERAs are used less often, and when or if considered, involve very patient-specific cases. Certainly, more preclinical and clinical studies are needed to be conducted on certain underlying conditions of hypertension and kidney disease.

## 2.5 Miscellaneous

#### 2.5.1 Parathyroid hormone

Parathyroid hormone (PTH) rises early in CKD and is a culprit contributor to its uremia toxicity. A combination of a decline in kidney function, impairment in phosphate excretion, and failure to synthesize calcitriol the bioactive form of vitamin D, all collectively lead to this maladaptive process of rising PTH level and parathyroid hyperplasia (Cunningham et al., 2011). It has been established that the phosphatonin fibroblast growth factor-23 (FGF-23) has a central role in the regulation of phosphate-vitamin D homeostasis, as this FGF-23 concentration increases secondary to hyperphosphatemia in CKD (Slatopolsky et al., 1999; Six et al., 2020). The progression to secondary hyperparathyroidism causes elevation in the intracellular calcium and plasma phosphate concentrations, and these play their part in increased vascular constriction and raising BP (Raine et al., 1993; Six et al., 2020), where the PTH itself also directly affects vascular tone (Rubin et al., 2005).

Hyperparathyroidism is highly associated with vascular calcification in dialysis patients (Moldovan et al., 2010) and should be considered in risk stratification for kidney-linked mortality in CKD patients (De Nicola et al., 2014). Parathyroidectomy may result in BP improvement in CKD (Morgado and Neves, 2012), and lowering PTH production by the chronic administration of an active vitamin D can reduce both intracellular calcium and systemic BP (Raine et al., 1993). Furthermore, administration of sevelamer as a calcium-free phosphate binder to lower phosphate has also proven to be good for control of serum PTH and FGF-23 levels (Six et al., 2020), and hence would slow the progression of vascular calcification in CKD. Adding to its ability to control phosphate, sevelamer also has pleiotropic effects that can be useful in correcting lipid abnormalities, reducing oxidative stress and inflammatory markers (Nikolov et al., 2006).

### 2.5.2 Erythropoietin

Erythropoietin, implicated in the treatment of anemia associated with CKD, is overlooked as a causative agent for BP elevation (Krapf and Hulter, 2009; Agarwal, 2018). Most hypertension cases take place with erythropoietin in conditions of severe renal impairment and with dialysis (Krapf and Hulter, 2009), while others of animals or human studies reporting the administration of erythropoietin with normal kidneys showed less advanced BP effects (Agarwal, 2018). Nevertheless, advanced increases in BP associated with erythropoietin use can be triggered by numerous mechanisms, such as: the degree of elevation in the hematocrit or blood viscosity (Raine, 1988; Eschbach et al., 1989), enhanced pressor responsiveness to NE and Ang II, increased cytosolic calcium in VSMCs, increased blood serotonin or endothelin levels (Vaziri, 1999), and increased vasoconstricting (PGF<sub>2a</sub> and TxA<sub>2</sub>) and decreased vasorelaxing PGI<sub>2</sub> prostanoid release (Agarwal, 2018). Endothelial dysfunction and the consequent augmented hypertension (Vaziri and Zhou, 2009; Agarwal, 2018) can result in vascular smooth muscle thickening and remodeling, inhibition of endothelial NO through increased production of ADMA (Scalera et al., 2005; Vaziri and Zhou, 2009), increased oxidative stress (Rancourt et al., 2010; Vaziri, 2010), or signaling through ET<sub>A</sub> receptor (Briet et al., 2013).

### 2.5.3 Ion channels and signaling

Abnormal signaling pathways through ion channels on the vascular wall may predispose to elevated BP in CKD (Martinez-Maldonado, 1998; Li and Fung, 2019). For example, transient receptor potential cation channels linked to ion transport are also reported to play a role in the pathogenesis of arterial systemic hypertension in CKD (Mene et al., 2013), through regulation of vascular smooth muscle contraction, renal perfusion or hemodynamics, as well as the total body balance of divalent cations. Experimental evidence suggests that expression and/or activity of the depolarizing voltage-gated calcium channels is enhanced during hypertension, whereas the function of the hyperpolarizing potassium channels may be declined (Martinez-Maldonado, 1998). A direct link exists between the density of epithelial sodium channels (ENaC) and vascular functionality. Indeed, overexpression of ENaC promotes arterial stiffness, increased myogenic tone, and endothelium morphology dysregulation and remodeling (Li and Fung, 2019), potentially contributing to heightened BP. Increased activity of protein kinase and increased cytosolic calcium levels may further inhibit the activation of potassium channels, thus increasing the likelihood of arterial smooth muscle depolarization, raising vascular tone, and ultimately arterial pressure (Sobey, 2001).

Interestingly, the effect of endogenous digitalis-like substances or more accurately its close relative ouabains on sodium or volume-dependent hypertension was reported as an associated factor in driving the high BP state in CKD (Morgado and Neves, 2012; Blaustein and Hamlyn, 2020). These include quite a few substances produced in the adrenal gland or in the hypothalamus that additionally cause inhibition of Na-K-ATPase in the cell membranes, and are natriuretic and vasoconstrictors (Takahashi et al., 2011). The elevated levels of these digitalis-like substances in CKD (Hamlyn et al., 1996; Komiyama et al., 2005; Blaustein and Hamlyn, 2020), secondary to an increased sodium content and hypervolemia, may also contribute to the hypertension commonly seen in CKD patients.

Lastly, it is progressively important to know the beneficial effects of SGLT<sub>2</sub>-inhibition with drugs like canagliflozin or empagliflozin is widely focused on CV volume and glycemic control via the induction of natriuresis and osmotic diuresis (Lytvyn et al., 2017). However, the success of SGLT<sub>2</sub>i in slowing CKD progression with and without diabetes (Wanner et al., 2016; Heerspink et al., 2020), may be related to their bonus effects on reducing oxidative stress, vascular alterations and providing significant renoprotection (Fernandez-Fernandez et al., 2020).

## 2.5.4 Uremic state

In the past 2 decades, investigations using uremic serum from CKD patients have been unravelling the complex composition of uremic toxins and their deleterious effect on BP and kidney disease progression. Those uremic toxins can affect endothelial cell function and viability, via inducing endothelial cell apoptosis, and mitigating its proliferation and migration. For example, following cell activation or apoptosis, the cellular plasma membrane shed what is known as the endothelial microparticles/fragments, which are remarkably increased in CKD (Al-Massarani et al., 2008). These microparticles act to express externalized phosphatidylserine group and endothelium specific antigens, hence culminating into vascular dysfunction (Amabile et al., 2005), impairment of NO signaling pathway, and hypertension in the long run. Other uremic toxins like inorganic phosphate, p-cresyl sulfate, and indoxyl sulfate are known toxins to increase endothelial microparticles shedding and apoptosis, causing augmentation of vascular contractility, impairment of vascular relaxation, NO reduction, and enhancement of ROS production (Figure 1). The gut microflora derived-p-cresyl sulfate, and its role in vascular disease in CKD, opened the gate to investigate the effect of intestinal bacteria in deriving these uremic toxins. Thus, modification of microflora in kidney disease patients could have a lead potential in reducing the effect of these circulating uremic toxin and their evoked renal fibrosis and CV calamities (Gao and Liu, 2017; Castillo-Rodriguez et al., 2018; Six et al., 2020). This has been the target of some promising preclinical data (Plata et al., 2019; Sato et al., 2020) that is yet to be thoroughly studied, evaluated and applied systematically (Rayego-Mateos and Valdivielso, 2020). Lastly, the aforementioned FGF-23, and its co-receptor klotho have been the focus of investigations of many studies in CKD humans and animals, as those signaling entities play an important role in heart and blood vessel functions and structures (Gutiérrez et al., 2009; Mirza et al., 2009). With worsening CKD, klotho deficiency is becoming a status quo condition, as circulating klotho levels decline, arterial stiffness and hypertension ensue (Gao et al., 2016). Furthermore, a klotho/FGF-23 deregulation state observed in CKD is increasingly becoming linked to resulting CV pathologies such as LVH, vascular calcification, and

endothelial dysfunction (Olauson and Larsson, 2013; Erben, 2016).

## 2.5.5 Effect of sex

Sex differences in CKD appear to play a major but ill-defined role in the incidence, prevalence, and progression of renal disease. Furthermore, our knowledge of the molecular mechanisms governing CV control and renal function in both genders remains limited, which is perhaps, in part, due to the fact that most research studies are primarily conducted in the male gender then findings are generalized to both males and females. Another possible reason for this scarcity of information is that most women with CKD are postmenopausal, which could explain why the influence of female hormones tends to be overlooked. This section therefore provides compelling evidence regarding the importance of sexual dimorphism in investigating pathophysiologic mechanisms relating to CVD in the CKD population.

Clinical studies on CVD in CKD patients suggest that women carry a relatively lower risk of developing CVD and related complications compared with men (Shajahan et al., 2021); however, it remains not entirely understood if these effects, as shown in hypertension, are driven by a differential influence of sex on CV regulatory control mechanisms in CKD. In relation to sex differences in vascular dysfunction, younger women (<55 years old) with CKD appear to show greater endothelium-dependent and -independent relaxation of the brachial artery than men, and that these differences disappear when women's age exceeds 55 years (Kruse et al., 2020). These data strongly support the notion that vascular dysfunction in women is not as pronounced as it is in men. With RAS, on the other hand, it has been shown that Ang II induces lower increases in BP and proteinuria in female SHRs compared with males. Although the precise mechanisms underlying these differences are incompletely understood, it is believed that estrogens in females may decrease the synthesis of ACE while testosterone in males may promote increased renin release (Quan et al., 2004). Female SHRs also seem to demonstrate greater expression levels of the "protective" AT<sub>2</sub> receptors (Hilliard et al., 2014), and exhibit more profound vasodilatory responses to the AT<sub>2</sub> agonist Compound 21 (Hilliard et al., 2012). Together, these data, although yet to be replicated in CKD, potentially indicate that male sex is associated with a greater RAS activity compared with females.

In relation to neural control of CV function in CKD, despite many studies reporting autonomic deficits in the CKD population, which contribute to sympathetic hyperactivity and increased CV risk in CKD, there are hardly any reports that dissect study variables based on sex. Our group was the first to show that impaired cardiac and sympathetic baroreflex control in male CKD rats was likely driven by temporal deficits in the functionality of the afferent and central components of the baroreflex arc (Salman et al., 2014), whereas impaired heart rate and sympathetic baroreflex in female rats appeared to result primarily from a sole decline in the central processing of baroreceptor afferent input (Salman et al., 2015a). These results indicate that female CKD rats express fewer autonomic deficits compared with males; however, if similar sex differences in autonomic function can be identified in humans with CKD remains to be determined.

Apart from the role of the vascular-hormonal-neural triad in the development of CVD in CKD, there is now growing evidence supporting a role for inflammation in the development and maintenance of hypertension and renal injury (Caillon and Schiffrin, 2016). It is therefore possible that the pathology of CVD in CKD is also contingent upon sex differences in the expression levels of key inflammatory markers driving the hypertensive state in both sexes. For instance, hypertension is associated with increased renal T cells infiltration (Caillon and Schiffrin, 2016); and the number of T regulatory cells (Tregs), which play a pivotal role in maintaining immune homeostasis via suppressing effector T cells activation (Mahajan et al., 2006) and modulating RAS, is lower in many models of hypertension (Barhoumi et al., 2011; Amador et al., 2014; Fabbiano et al., 2015). Indeed, female SHRs express more renal Tregs, which corresponds with a lower BP and less proteinuria and renal injury (Tipton et al., 2012). However, the role of Tregs in altering the course of CKD progression in both males and females has yet to be addressed in experimental and clinical studies.

## **3** Conclusion

Hypertension, the leading cause of CV incidence derived from and sustained by CKD development and progression, is a crucial challenge for modern medicine, given the rising numbers in morbidity and mortality worldwide. While numerous potential pathophysiological mechanisms have been uncovered in recent years, many details remain poorly understood or unknown, and therefore, continued research and investigation to explore and navigate therapeutic options are imperative in hypertensive kidney disease. Here, we put forward relevant findings which demonstrate mechanisms underpinning raised BP in CKD, while taking into account the existence of multifactorial entities that can initiate or come later into play in the disease process. A deeper understanding of the molecular mechanisms and pathways will allow the identification of key targets of pharmacological or nonpharmacological therapies that are able to achieve an

appropriate control of renal damage in hypertension. Consequently, this would give the chance for novel therapeutic strategies to overcome current conventional treatments by their ability to interact with the cardio-renal system. The hormonal, neural, and vascular arms are integral in the regulation of renal and CV physiology, and they amalgamate complex mechanisms, often by interacting with each other, and inadvertently pave the way for the pathogenesis of hypertensive nephropathy. Knowing that, however, more research is required to transfer promising experimental results into routine clinical practice. Although the use of current pharmacological agents targeting systems like the RAAS, SNS, ET-1, voltage-gated Ca2+ channels, COX, and renal Na<sup>+</sup>/K<sup>+</sup>/2Cl<sup>-</sup>, ENaC and SGLT<sub>2</sub> have translated into benefit of use and reductions in CV morbidity and mortality, there remains room for improvement in controlling hypertensive kidney disease, especially when it comes to halting, reversing, or even modulating its adverse action on the CV system.

## Author contributions

OA contributed to the literature review, prepared the figures, drafted the manuscript and approved and submitted the final version of the manuscript.

## Acknowledgments

The author thanks Dr Ibrahim M. Salman, Dr Ko Jin Quek and Dr Jacqueline K. Phillips for their insights and useful comments during the production of this manuscript.

# Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

## Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

# References

Abrahamsen, C. T., Pullen, M. A., Schnackenberg, C. G., Grygielko, E. T., Edwards, R. M., Laping, N. J., et al. (2002). Effects of angiotensins II and IV on blood pressure, renal function, and PAI-1 expression in the heart and kidney of the rat. *Pharmacology* 66, 26–30. doi:10.1159/000063252

Acosta, J. H. (1982). Hypertension in chronic renal disease. Kidney Int. 22, 702-712. doi:10.1038/ki.1982.233

Adamczak, M., Zeier, M., Dikow, R., and Ritz, E. (2002). Kidney and hypertension. *Kidney Int.* 61, S62–S67. doi:10.1046/j.1523-1755.61.s80.28.x

Adams, M. A., Bobik, A., and Korner, P. I. (1990). Enalapril can prevent vascular amplifier development in spontaneously hypertensive rats. *Hypertension* 16, 252–260. doi:10.1161/01.hyp.16.3.252

Agarwal, A., Anand, I. S., Sakhuja, V., and Chugh, K. S. (1991). Effect of dialysis and renal transplantation on autonomic dysfunction in chronic renal failure. *Kidney Int.* 40, 489–495. doi:10.1038/ki.1991.236

Agarwal, R. (2018). Mechanisms and mediators of hypertension induced by erythropoietin and related molecules. *Nephrol. Dial. Transpl.* 33, 1690–1698. doi:10. 1093/ndt/gfx324

Akaihata, H., Hata, J., Tanji, R., Honda-Takinami, R., Matsuoka, K., Sato, Y., et al. (2020). Tetrahydrobiopterin prevents chronic ischemia-related lower urinary tract dysfunction through the maintenance of nitric oxide bioavailability. *Sci. Rep.* 10, 19844. doi:10.1038/s41598-020-76948-9

Alp, N. J., and Channon, K. M. (2004). Regulation of endothelial nitric oxide synthase by tetrahydrobiopterin in vascular disease. *Arterioscler. Thromb. Vasc. Biol.* 24, 413–420. doi:10.1161/01.ATV.0000110785.96039.f6

Al-Massarani, G., Vacher-Coponat, H., Paul, P., Widemann, A., Arnaud, L., Loundou, A., et al. (2008). Impact of immunosuppressive treatment on endothelial biomarkers after kidney transplantation. *Am. J. Transpl.* 8, 2360–2367. doi:10.1111/j.1600-6143.2008.02399.x

Amabile, N., Guérin, A. P., Leroyer, A., Mallat, Z., Nguyen, C., Boddaert, J., et al. (2005). Circulating endothelial microparticles are associated with vascular dysfunction in patients with end-stage renal failure. *J. Am. Soc. Nephrol.* 16, 3381–3388. doi:10.1681/ASN.2005050535

Amador, C. A., Barrientos, V., Peña, J., Herrada, A. A., González, M., Valdés, S., et al. (2014). Spironolactone decreases DOCA-salt-induced organ damage by blocking the activation of T helper 17 and the downregulation of regulatory T lymphocytes. *Hypertension* 63, 797–803. doi:10.1161/HYPERTENSIONAHA.113. 02883

Amador-Martínez, I., Pérez-Villalva, R., Uribe, N., Cortés-González, C., Bobadilla, N. A., and Barrera-Chimal, J. (2019). Reduced endothelial nitric oxide synthase activation contributes to cardiovascular injury during chronic kidney disease progression. *Am. J. Physiol. Ren. Physiol.* 317, F275–F285. doi:10. 1152/aircnal.00020.2019

Ameer, O. Z., Boyd, R., Butlin, M., Avolio, A. P., and Phillips, J. K. (2015). Abnormalities associated with progressive aortic vascular dysfunction in chronic kidney disease. *Front. Physiol.* 6, 150. doi:10.3389/fphys.2015.00150

Ameer, O. Z., Butlin, M., Kaschina, E., Sommerfeld, M., Avolio, A. P., and Phillips, J. K. (2016). Long-term angiotensin II receptor blockade limits hypertension, aortic dysfunction, and structural remodeling in a rat model of chronic kidney disease. J. Vasc. Res. 53, 216–229. doi:10.1159/000452411

Ameer, O. Z., Hildreth, C. M., and Phillips, J. K. (2014a). Sympathetic overactivity prevails over the vascular amplifier phenomena in a chronic kidney disease rat model of hypertension. *Physiol. Rep.* 2, e12205. doi:10. 14814/phy2.12205

Ameer, O. Z., Salman, I. M., Avolio, A. P., Phillips, J. K., and Butlin, M. (2014b). Opposing changes in thoracic and abdominal aortic biomechanical properties in rodent models of vascular calcification and hypertension. *Am. J. Physiol. Heart Circ. Physiol.* 307, H143–H151. doi:10.1152/ajpheart.00139.2014

Anderstam, B., Katzarski, K., and Bergström, J. (1997). Serum levels of NG, NG-dimethyl-L-arginine, a potential endogenous nitric oxide inhibitor in dialysis patients. *J. Am. Soc. Nephrol.* 8, 1437–1442. doi:10.1681/ASN. V891437

Antza, C., Doundoulakis, I., Stabouli, S., and Kotsis, V. (2021). American, European and international hypertension guidelines: Time to shake hands? *Int. J. Cardiol. Hypertens.* 8, 100075. doi:10.1016/j.ijchy.2020.100075

Baker, M., and Perazella, M. A. (2020). NSAIDs in CKD: Are they safe? Am. J. Kidney Dis. 76, 546–557. doi:10.1053/j.ajkd.2020.03.023

Bakris, G. L., Townsend, R. R., Liu, M., Cohen, S. A., D'agostino, R., Flack, J. M., et al. (2014). Impact of renal denervation on 24-hour ambulatory blood pressure: Results from SYMPLICITY HTN-3. J. Am. Coll. Cardiol. 64, 1071–1078. doi:10. 1016/j.jacc.2014.05.012 Baltatu, O. C., Zaugg, C. E., Schumacher, C., Louie, P., Campos, L. A., and Bader, M. (2014). Avosentan is protective in hypertensive nephropathy at doses not causing fluid retention. *Pharmacol. Res.* 80, 9–13. doi:10.1016/j.phrs. 2013.12.003

Baraka, A., and El Ghotny, S. (2012). Cardioprotective effect of renalase in 5/ 6 nephrectomized rats. J. Cardiovasc. Pharmacol. Ther. 17, 412–416. doi:10.1177/ 1074248412446977

Barhoumi, T., Kasal, D. A., Li, M. W., Shbat, L., Laurant, P., Neves, M. F., et al. (2011). T regulatory lymphocytes prevent angiotensin II-induced hypertension and vascular injury. *Hypertension* 57, 469–476. doi:10.1161/HYPERTENSIONAHA. 110.162941

Baumeister, S. E., Böger, C. A., Krämer, B. K., Döring, A., Eheberg, D., Fischer, B., et al. (2009). Effect of chronic kidney disease and comorbid conditions on health care costs: A 10-year observational study in a general population. *Am. J. Nephrol.* 31, 222–229. doi:10.1159/000272937

Beige, J., Koziolek, M. J., Hennig, G., Hamza, A., Wendt, R., Müller, G. A., et al. (2015). Baroreflex activation therapy in patients with end-stage renal failure: Proof of concept. *J. Hypertens.* 33, 2344–2349. doi:10.1097/HJH.0000000000000697

Benetos, A., Lacolley, P., and Safar, M. (1997). Prevention of aortic fibrosis by spironolactone in spontaneously hypertensive rats. *Arterioscler. Thromb. Vasc. Biol.* 17, 1152–1156. doi:10.1161/01.atv.17.6.1152

Bevan, R. D. (1984). Trophic effects of peripheral adrenergic nerves on vascular structure. *Hypertension* 6, III19–26. doi:10.1161/01.hyp.6.6\_pt\_2.iii19

Bhatt, D., Kristensen, A. M., Pareek, M., and Olsen, M. H. (2019). Baroreflex activation therapy for resistant hypertension and heart failure. U. S. Card. Rev. 13, 83–87.

Bhatt, D. L., Kandzari, D. E., O'neill, W. W., D'agostino, R., Flack, J. M., Katzen, B. T., et al. (2014). A controlled trial of renal denervation for resistant hypertension. *N. Engl. J. Med.* 370, 1393–1401. doi:10.1056/NEJMoa1402670

Bisognano, J. D., Bakris, G., Nadim, M. K., Sanchez, L., Kroon, A. A., Schafer, J., et al. (2011). Baroreflex activation therapy lowers blood pressure in patients with resistant hypertension: Results from the double-blind, randomized, placebocontrolled rheos pivotal trial. *J. Am. Coll. Cardiol.* 58, 765–773. doi:10.1016/j. jacc.2011.06.008

Blair, A., Shaul, P. W., Yuhanna, I. S., Conrad, P. A., and Smart, E. J. (1999). Oxidized low density lipoprotein displaces endothelial nitric-oxide synthase (eNOS) from plasmalemmal caveolae and impairs eNOS activation. *J. Biol. Chem.* 274, 32512–32519. doi:10.1074/jbc.274.45.32512

Blankestijn, P. J. (2007). Sympathetic hyperactivity—A hidden enemy in chronic kidney disease patients. *Perit. Dial. Int.* 27, S293–S297. doi:10.1177/089686080702702s50

Blasi, E. R., Rocha, R., Rudolph, A. E., Blomme, E. A., Polly, M. L., and Mcmahon, E. G. (2003). Aldosterone/salt induces renal inflammation and fibrosis in hypertensive rats. *Kidney Int.* 63, 1791–1800. doi:10.1046/j.1523-1755.2003.00929.x

Blaustein, M. P., and Hamlyn, J. M. (2020). Ouabain, endogenous ouabain and ouabain-like factors: The Na+ pump/ouabain receptor, its linkage to NCX, and its myriad functions. *Cell calcium* 86, 102159. doi:10.1016/j.ceca.2020.102159

Blaustein, M. P., Zhang, J., Chen, L., and Hamilton, B. P. (2006). How does salt retention raise blood pressure? *Am. J. Physiol. Regul. Integr. Comp. Physiol.* 290, R514–R523. doi:10.1152/ajpregu.00819.2005

Blaustein, M. P., Zhang, J., Chen, L., Song, H., Raina, H., Kinsey, S. P., et al. (2009). The pump, the exchanger, and endogenous ouabain: Signaling mechanisms that link salt retention to hypertension. *Hypertension* 53, 291–298. doi:10.1161/HYPERTENSIONAHA.108.119974

Blythe, W. B. (1985). Natural history of hypertension in renal parenchymal disease. Am. J. Kidney Dis. 5, A50-A56. doi:10.1016/s0272-6386(85)80066-4

Böhm, M., Kario, K., Kandzari, D. E., Mahfoud, F., Weber, M. A., Schmieder, R. E., et al. (2020). Efficacy of catheter-based renal denervation in the absence of antihypertensive medications (SPYRAL HTN-OFF MED pivotal): A multicentre, randomised, sham-controlled trial. *Lancet* 395, 1444–1451. doi:10.1016/S0140-6736(20)30554-7

Bolignano, D., Palmer, S. C., Navaneethan, S. D., and Strippoli, G. F. (2014). Aldosterone antagonists for preventing the progression of chronic kidney disease. *Cochrane Database Syst. Rev.* doi:10.1002/14651858.cd007004.pub3

Boström, K., Watson, K., Horn, S., Wortham, C., Herman, I., and Demer, L. (1993). Bone morphogenetic protein expression in human atherosclerotic lesions. *J. Clin. Invest.* 91, 1800–1809. doi:10.1172/JCI116391

Breyer, M. D., and Susztak, K. (2016). Developing treatments for chronic kidney disease in the 21st century. *Semin. Nephrol.* 36, 436–447. doi:10.1016/j.semnephrol. 2016.08.001()

Briet, M., Barhoumi, T., Mian, M. O., Sierra, C., Boutouyrie, P., Davidman, M., et al. (2013). Effects of recombinant human erythropoietin on resistance artery

endothelial function in stage 4 chronic kidney disease. J. Am. Heart Assoc. 2, e000128. doi:10.1161/JAHA.113.000128

Briet, M., and Schiffrin, E. L. (2010). Aldosterone: Effects on the kidney and cardiovascular system. Nat. Rev. Nephrol. 6, 261-273. doi:10.1038/nrneph.2010.30

Briet, M., and Schiffrin, E. L. (2013). Vascular actions of aldosterone. J. Vasc. Res. 50, 89–99. doi:10.1159/000345243

Bruno, R. M., and Taddei, S. (2011). "Nitric oxide," in *Encyclopedia of exercise medicine in health and disease*. Editors F. C. Mooren and J. S. Skinner (Berlin, Heidelberg, Germany: Springer-Verlag).

Burnier, M. (2022). Renal denervation for patients with chronic kidney disease and resistant hypertension: Effective and safe but still not the panacea. Oxford University Press.

Cabassi, A., Tedeschi, S., Perlini, S., Verzicco, I., Volpi, R., Gonzi, G., et al. (2020). Non-steroidal anti-inflammatory drug effects on renal and cardiovascular function: From physiology to clinical practice. *Eur. J. Prev. Cardiol.* 27, 850–867. doi:10.1177/ 2047487319848105

Caillon, A., and Schiffrin, E. L. (2016). Role of inflammation and immunity in hypertension: Recent epidemiological, laboratory, and clinical evidence. *Curr. Hypertens. Rep.* 18, 21. doi:10.1007/s11906-016-0628-7

Campese, V., Mitra, N., and Sandee, D. (2006). Hypertension in renal parenchymal disease: Why is it so resistant to treatment? *Kidney Int.* 69, 967–973. doi:10.1038/sj.ki.5000177

Campese, V. M., and Krol, E. (2002). Neurogenic factors in renal hypertension. Curr. Hypertens. Rep. 4, 256–260. doi:10.1007/s11906-002-0016-3

Capelli, I., Gasperoni, L., Ruggeri, M., Donati, G., Baraldi, O., Sorrenti, G., et al. (2020). New mineralocorticoid receptor antagonists: Update on their use in chronic kidney disease and heart failure. *J. Nephrol.* 33, 37–48. doi:10.1007/s40620-019-00600-7

Castillo-Rodriguez, E., Fernandez-Prado, R., Esteras, R., Perez-Gomez, M. V., Gracia-Iguacel, C., Fernandez-Fernandez, B., et al. (2018). Impact of altered intestinal microbiota on chronic kidney disease progression. *Toxins* 10, 300. doi:10.3390/toxins10070300

Cerasola, G., Vecchi, M., Mule, G., Cottone, S., Mangano, M. T., Andronico, G., et al. (1998). Sympathetic activity and blood pressure pattern in autosomal dominant polycystic kidney disease hypertensives. *Am. J. Nephrol.* 18, 391–398. doi:10.1159/000013382

Channick, R. N., and Rubin, L. J. (2002). Endothelin receptor antagonism: A new era in the treatment of pulmonary arterial hypertension. *Adv. Pulm. Hypertens.* 1, 13–17. doi:10.21693/1933-088x-1.1.13

Channon, K. M. (2020). Tetrahydrobiopterin and nitric oxide synthase recouplers. Handb. Exp. Pharmacol. 264, 339-352. doi:10.1007/164\_2020\_390

Chapman, A. B., Stepniakowski, K., and Rahbari-Oskoui, F. (2010). Hypertension in autosomal dominant polycystic kidney disease. *Adv. Chronic Kidney Dis.* 17, 153–163. doi:10.1053/j.ackd.2010.01.001

Chen, G., and Suzuki, H. (1990). Calcium dependency of the endotheliumdependent hyperpolarization in smooth muscle cells of the rabbit carotid artery. J. Physiol. 421, 521–534. doi:10.1113/jphysiol.1990.sp017959

Chen, H.-H., Cheng, P.-W., Ho, W.-Y., Lu, P.-J., Lai, C.-C., Tseng, Y.-M., et al. (2016). Renal denervation improves the baroreflex and GABA system in chronic kidney diseaseinduced hypertension. *Sci. Rep.* 6, 38447. doi:10.1038/srep38447

Cheung, A. K., Rahman, M., Reboussin, D. M., Craven, T. E., Greene, T., Kimmel, P. L., et al. (2017). Effects of intensive BP control in CKD. J. Am. Soc. Nephrol. 28, 2812–2823. doi:10.1681/ASN.2017020148

Christensen, F. H., Stankevicius, E., Hansen, T., Jørgensen, M. M., Valverde, V. L., Simonsen, U., et al. (2007). Flow-and acetylcholine-induced dilatation in small arteries from rats with renovascular hypertension—Effect of tempol treatment. *Eur. J. Pharmacol.* 566, 160–166. doi:10.1016/j.ejphar.2007.03.058

Chung, E. Y., Ruospo, M., Natale, P., Bolignano, D., Navaneethan, S. D., Palmer, S. C., et al. (2020). Aldosterone antagonists in addition to renin angiotensin system antagonists for preventing the progression of chronic kidney disease. *Cochrane Database Syst. Rev.* doi:10.1002/14651858.cd007004.pub4

Ciriello, J., and De Oliveira, C. V. (2002). Renal afferents and hypertension. *Curr. Hypertens. Rep.* 4, 136–142. doi:10.1007/s11906-002-0038-x

Cohen, R. A., and Vanhoutte, P. M. (1995). Endothelium-dependent hyperpolarization: Beyond nitric oxide and cyclic GMP. *Circulation* 92, 3337–3349. doi:10.1161/01.cir.92.11.3337

Collins, A. J., Foley, R. N., Chavers, B., Gilbertson, D., Herzog, C., Ishani, A., et al. (2014). US renal data system 2013 annual data report. *Am. J. Kidney Dis.* 63, A7. doi:10.1053/j.ajkd.2013.11.001

Converse, R. L., Jacobsen, T. N., Toto, R. D., Jost, C. M., Cosentino, F., Fouad-Tarazi, F., et al. (1992). Sympathetic overactivity in patients with chronic renal failure. *N. Engl. J. Med.* 327, 1912–1918. doi:10.1056/NEJM199212313272704

Cosentino, F., Hürlimann, D., Gatti, C. D., Chenevard, R., Blau, N., Alp, N. J., et al. (2008). Chronic treatment with tetrahydrobiopterin reverses endothelial dysfunction and oxidative stress in hypercholesterolaemia. *Heart* 94, 487–492. doi:10.1136/hrt.2007.122184

Cross, J. M., Donald, A. E., Kharbanda, R., Deanfield, J. E., Woolfson, R. G., and Macallister, R. J. (2001). Acute administration of L-arginine does not improve arterial endothelial function in chronic renal failure. *Kidney Int.* 60, 2318–2323. doi:10.1046/j.1523-1755.2001.00059.x

Csiszar, A., Lehoux, S., and Ungvari, Z. (2009). Hemodynamic forces, vascular oxidative stress, and regulation of BMP-2/4 expression. *Antioxid. Redox Signal.* 11, 1683–1697. doi:10.1089/ars.2008.2401

Cunningham, J., Locatelli, F., and Rodriguez, M. (2011). Secondary hyperparathyroidism: Pathogenesis, disease progression, and therapeutic options. *Clin. J. Am. Soc. Nephrol.* 6, 913–921. doi:10.2215/CJN.06040710

Curtis, J. J., Luke, R. G., Dustan, H. P., Kashgarian, M., Whelchel, J. D., Jones, P., et al. (1983). Remission of essential hypertension after renal transplantation. *N. Engl. J. Med.* 309, 1009–1015. doi:10.1056/NEJM198310273091702

Czubilińska-Łada, J., Gliwińska, A., Badeński, A., and Szczepańska, M. (2020). Associations between renalase concentration and the occurrence of selected diseases. *Endokrynol. Pol.* 71, 334–342. doi:10.5603/EP.a2020.0044

Daenen, K., Andries, A., Mekahli, D., Van Schepdael, A., Jouret, F., and Bammens, B. (2019). Oxidative stress in chronic kidney disease. *Pediatr. Nephrol.* 34, 975–991. doi:10.1007/s00467-018-4005-4

Dal Canton, A., Fornoni, A., Fasoli, G., Ranghino, A., and Semeraro, L. (1996). "Volume-dependent factors in hypertension in chronic renal failure," in *Clinical hypertension in nephrology* (Karger Publishers), 26–30.

De Nicola, L., Conte, G., Chiodini, P., D'angio, P., Donnarumma, G., and Minutolo, R. (2014). Interaction between phosphorus and parathyroid hormone in non-dialysis CKD patients under nephrology care. *J. Nephrol.* 27, 57–63. doi:10. 1007/s40620-013-0001-8

De Nucci, G., Thomas, R., D'orleans-Juste, P., Antunes, E., Walder, C., Warner, T. D., et al. (1988). Pressor effects of circulating endothelin are limited by its removal in the pulmonary circulation and by the release of prostacyclin and endothelium-derived relaxing factor. *Proc. Natl. Acad. Sci. U. S. A.* 85, 9797–9800. doi:10.1073/pnas.85.24.9797

Deanfield, J., Donald, A., Ferri, C., Giannattasio, C., Halcox, J., Halligan, S., et al. (2005). Endothelial function and dysfunction. Part I: Methodological issues for assessment in the different vascular beds: A statement by the working group on endothelin and endothelial factors of the European society of hypertension. *J. Hypertens.* 23, 7–17. doi:10.1097/00004872-200501000-00004

Deng, L.-Y., Jin-Sheng, L., and Schiffrin, E. L. (1995). Endothelium-dependent relaxation of small arteries from essential hypertensive patients: Mechanisms and comparison with normotensive subjects and with responses of vessels from spontaneously hypertensive rats. *Clin. Sci.* 88, 611–622. doi:10.1042/cs0880611

Desir, G. V., and Peixoto, A. J. (2014). Renalase in hypertension and kidney disease. Nephrol. Dial. Transpl. 29, 22-28. doi:10.1093/ndt/gft083

Desir, G. V. (2009). Regulation of blood pressure and cardiovascular function by renalase. *Kidney Int.* 76, 366–370. doi:10.1038/ki.2009.169

Dhaun, N., Goddard, J., and Webb, D. (2006). The endothelin system and its antagonism in chronic kidney disease. *J. Am. Soc. Nephrol.* 17, 943–955. doi:10. 1681/ASN.2005121256

Dhaun, N., Macintyre, I. M., Kerr, D., Melville, V., Johnston, N. R., Haughie, S., et al. (2011). Selective endothelin-A receptor antagonism reduces proteinuria, blood pressure, and arterial stiffness in chronic proteinuric kidney disease. *Hypertension* 57, 772–779. doi:10.1161/HYPERTENSIONAHA.110.167486

Dou, L., Sallée, M., Cerini, C., Poitevin, S., Gondouin, B., Jourde-Chiche, N., et al. (2015). The cardiovascular effect of the uremic solute indole-3 acetic acid. *J. Am. Soc. Nephrol.* 26, 876–887. doi:10.1681/ASN.2013121283

Düsing, P., Zietzer, A., Goody, P. R., Hosen, M. R., Kurts, C., Nickenig, G., et al. (2021). Vascular pathologies in chronic kidney disease: Pathophysiological mechanisms and novel therapeutic approaches. *J. Mol. Med.* 99, 335–348. doi:10.1007/s00109-021-02037-7

El-Khoury, J. M., Bunch, D. R., Hu, B., Payto, D., Reineks, E. Z., and Wang, S. (2016). Comparison of symmetric dimethylarginine with creatinine, cystatin C and their eGFR equations as markers of kidney function. *Clin. Biochem.* 49, 1140–1143. doi:10.1016/j.clinbiochem.2016.07.009

Erben, R. G. (2016). Update on FGF23 and klotho signaling. *Mol. Cell. Endocrinol.* 432, 56–65. doi:10.1016/j.mce.2016.05.008

Eschbach, J. W., Kelly, M. R., Haley, N. R., Abels, R. I., and Adamson, J. W. (1989). Treatment of the anemia of progressive renal failure with recombinant human erythropoietin. *N. Engl. J. Med.* 321, 158–163. doi:10.1056/NEJM198907203210305

Esler, M. D., Krum, H., Schlaich, M., Schmieder, R. E., Böhm, M., Sobotka, P. A., et al. (2012). Renal sympathetic denervation for treatment of drug-resistant

hypertension: One-year results from the symplicity HTN-2 randomized, controlled trial. *Circulation* 126, 2976–2982. doi:10.1161/CIRCULATIONAHA.112.130880

Fabbiano, S., Menacho-Márquez, M., Robles-Valero, J., Pericacho, M., Matesanz-Marín, A., García-Macías, C., et al. (2015). Immunosuppression-independent role of regulatory T cells against hypertension-driven renal dysfunctions. *Mol. Cell. Biol.* 35, 3528–3546. doi:10.1128/MCB.00518-15

Félétou, M., and Vanhoutte, P. M. (2006). Endothelial dysfunction: A multifaceted disorder (the wiggers award lecture). *Am. J. Physiol. Heart Circ. Physiol.* 291, H985–H1002. doi:10.1152/ajpheart.00292.2006

Fernandez-Fernandez, B., Sarafidis, P., Kanbay, M., Navarro-González, J. F., Soler, M. J., Górriz, J. L., et al. (2020). SGLT2 inhibitors for non-diabetic kidney disease: Drugs to treat CKD that also improve glycaemia. Oxford University Press.

Fink, G. D. (2009). Arthur C. Corcoran Memorial Lecture. Sympathetic activity, vascular capacitance, and long-term regulation of arterial pressure. *Hypertension* 53, 307–312. doi:10.1161/HYPERTENSIONAHA.108.119990

Fleck, C., Janz, A., Schweitzer, F., Karge, E., Schwertfeger, M., and Stein, G. (2001). Serum concentrations of asymmetric (ADMA) and symmetric (SDMA) dimethylarginine in renal failure patients. *Kidney Int. Suppl.* 59, S14–S18. doi:10.1046/j.1523-1755.2001.59780014.x

Fliser, D., Kronenberg, F., Kielstein, J. T., Morath, C., Bode-Böger, S. M., Haller, H., et al. (2005). Asymmetric dimethylarginine and progression of chronic kidney disease: The mild to moderate kidney disease study. *J. Am. Soc. Nephrol.* 16, 2456–2461. doi:10.1681/ASN.2005020179

Fliser, D., Wiecek, A., Suleymanlar, G., Ortiz, A., Massy, Z., Lindholm, B., et al. (2011). The dysfunctional endothelium in CKD and in cardiovascular disease: Mapping the origin (s) of cardiovascular problems in CKD and of kidney disease in cardiovascular conditions for a research agenda. *Kidney Int. Suppl.* 1, 6–9. doi:10. 1038/kisup.2011.6

Fonseca, J. M., Bastos, A. P., Amaral, A. G., Sousa, M. F., Souza, L. E., Malheiros, D. M., et al. (2014). Renal cyst growth is the main determinant for hypertension and concentrating deficit in Pkd1-deficient mice. *Kidney Int.* 85, 1137–1150. doi:10.1038/ki.2013.501

Freedman, B. I., and Cohen, A. H. (2016). Hypertension-attributed nephropathy: what's in a name? *Nat. Rev. Nephrol.* 12, 27–36. doi:10.1038/nrneph.2015.172

Freitas, F. F., Araujo, G., Porto, M. L., Freitas, F. P., Graceli, J. B., Balarini, C. M., et al. (2016). Increased blood pressure variability prior to chronic kidney disease exacerbates renal dysfunction in rats. *Front. Physiol.* 7, 428. doi:10.3389/fphys.2016. 00428

Fujihara, C. K., De Nucci, G., and Zatz, R. (1995). Chronic nitric oxide synthase inhibition aggravates glomerular injury in rats with subtotal nephrectomy. *J. Am. Soc. Nephrol.* 5, 1498–1507. doi:10.1681/ASN.V571498

Greenwood, J. P., Scott, E. M., Stoker, J. B., and Mary, D. A. (2001). Hypertensive left ventricular hypertrophy: Relation to peripheral sympathetic drive. *J. Am. Coll. Cardiol.* 38, 1711–1717. doi:10.1016/s0735-1097(01)01600-x

Gajulapalli, R. D., Chahine, J., Rader, F., and Aneja, A. (2020). Procedures and devices to treat resistant hypertension in chronic kidney disease. *Cleve. Clin. J. Med.* 87, 435–443. doi:10.3949/ccjm.87a.19099

Gao, D., Zuo, Z., Tian, J., Ali, Q., Lin, Y., Lei, H., et al. (2016). Activation of SIRT1 attenuates klotho deficiency-induced arterial stiffness and hypertension by enhancing AMP-activated protein kinase activity. *Hypertension* 68, 1191–1199. doi:10.1161/HYPERTENSIONAHA.116.07709

Gao, H., and Liu, S. (2017). Role of uremic toxin indoxyl sulfate in the progression of cardiovascular disease. *Life Sci.* 185, 23–29. doi:10.1016/j.lfs.2017.07.027

Gao, L., Wang, W.-Z., Wang, W., and Zucker, I. H. (2008). Imbalance of angiotensin type 1 receptor and angiotensin II type 2 receptor in the rostral ventrolateral medulla potential mechanism for sympathetic overactivity in heart failure. *Hypertension* 52, 708–714. doi:10.1161/HYPERTENSIONAHA. 108.116228

Gao, L., Wang, W., Li, Y.-L., Schultz, H. D., Liu, D., Cornish, K. G., et al. (2005). Sympathoexcitation by central ANG II: Roles for AT1 receptor upregulation and NAD (P) H oxidase in RVLM. *Am. J. Physiol. Heart Circ. Physiol.* 288, H2271–H2279. doi:10.1152/ajpheart.00949.2004

Gavini, M. P., Mahmood, A., Belenchia, A. M., Beauparlant, P., Kumar, S. A., Ardhanari, S., et al. (2021). Suppression of inflammatory cardiac cytokine network in rats with untreated obesity and pre-diabetes by AT2 receptor agonist NP-6A4. *Front. Pharmacol.* 12, 693167. doi:10.3389/fphar.2021.693167

Gelosa, P., Pignieri, A., Fändriks, L., De Gasparo, M., Hallberg, A., Banfi, C., et al. (2009). Stimulation of AT2 receptor exerts beneficial effects in stroke-prone rats: Focus on renal damage. *J. Hypertens.* 27, 2444–2451. doi:10.1097/HJH. 0b013e3283311ba1

Gharavi, N. M., Baker, N. A., Mouillesseaux, K. P., Yeung, W., Honda, H. M., Hsieh, X., et al. (2006). Role of endothelial nitric oxide synthase in the regulation of SREBP activation by oxidized phospholipids. *Circ. Res.* 98, 768–776. doi:10.1161/01. RES.0000215343.89308.93

Goddard, J., Johnston, N. R., Hand, M. F., Cumming, A. D., Rabelink, T. J., Rankin, A. J., et al. (2004). Endothelin-A receptor antagonism reduces blood pressure and increases renal blood flow in hypertensive patients with chronic renal failure: A comparison of selective and combined endothelin receptor blockade. *Circulation* 109, 1186–1193. doi:10.1161/01.CIR.0000118499. 69469.51

Goligorsky, M. S., Yasuda, K., and Ratliff, B. (2010). Dysfunctional endothelial progenitor cells in chronic kidney disease. *J. Am. Soc. Nephrol.* 21, 911–919. doi:10. 1681/ASN.2009111119

Grassi, G., Seravalle, G., and Quarti-Trevano, F. (2010). The 'neuroadrenergic hypothesis' in hypertension: Current evidence. *Exp. Physiol.* 95, 581–586. doi:10. 1113/expphysiol.2009.047381

Greene, E., Kren, S., and Hostetter, T. (1996). Role of aldosterone in the remnant kidney model in the rat. J. Clin. Invest. 98, 1063–1068. doi:10.1172/JCI118867

Griendling, K. K., Minieri, C. A., Ollerenshaw, J. D., and Alexander, R. W. (1994). Angiotensin II stimulates NADH and NADPH oxidase activity in cultured vascular smooth muscle cells. *Circ. Res.* 74, 1141–1148. doi:10.1161/01.res.74.6.1141

Griffith, T. M., Chaytor, A. T., Bakker, L. M., and Edwards, D. H. (2005). 5-Methyltetrahydrofolate and tetrahydrobiopterin can modulate electrotonically mediated endothelium-dependent vascular relaxation. *Proc. Natl. Acad. Sci. U. S. A.* 102, 7008–7013. doi:10.1073/pnas.0408919102

Groenland, E. H., and Spiering, W. (2020). Baroreflex amplification and carotid body modulation for the treatment of resistant hypertension. *Curr. Hypertens. Rep.* 22, 27–28. doi:10.1007/s11906-020-1024-x

Group, S. R. (2015). A randomized trial of intensive versus standard bloodpressure control. N. Engl. J. Med. Overseas. Ed. 373, 2103-2116. doi:10.1056/ nejmoa1511939

Guan, Z., Intapad, S., Palygin, O., and Sullivan, J. C. (2022). Hypertension and chronic kidney injury or failure, volume II. *Front. Physiology* 2479.

Guidi, E., Menghetti, D., Milani, S., Montagnino, G., Palazzi, P., and Bianchi, G. (1996). Hypertension may be transplanted with the kidney in humans: A long-term historical prospective follow-up of recipients grafted with kidneys coming from donors with or without hypertension in their families. J. Am. Soc. Nephrol. 7, 1131–1138. doi:10.1681/ASN.V781131

Gutiérrez, O. M., Januzzi, J. L., Isakova, T., Laliberte, K., Smith, K., Collerone, G., et al. (2009). Fibroblast growth factor-23 and left ventricular hypertrophy in chronic kidney disease. *Circulation* 119, 2545–2552. doi:10.1161/CIRCULATIONAHA.108. 844506

H Heiss, E., and M Dirsch, V. (2014). Regulation of eNOS enzyme activity by posttranslational modification. *Curr. Pharm. Des.* 20, 3503–3513. doi:10.2174/13816128113196660745

Hamlyn, J. M., Hamilton, B. P., and Manunta, P. (1996). Endogenous ouabain, sodium balance and blood pressure: A review and a hypothesis. *J. Hypertens.* 14, 151–167. doi:10.1097/00004872-199602000-00002

Hamrahian, S. M. (2022). "Hypertension and cardiovascular disease in patients with chronic kidney disease," in *Approaches to chronic kidney disease* (Springer), 281–295.

Harlacher, E., Wollenhaupt, J., Baaten, C. C., and Noels, H. (2022). Impact of uremic toxins on endothelial dysfunction in chronic kidney disease: A systematic review. *Int. J. Mol. Sci.* 23, 531. doi:10.3390/ijms23010531

Harrison, J. L., Hildreth, C. M., Callahan, S. M., Goodchild, A. K., and Phillips, J. K. (2010). Cardiovascular autonomic dysfunction in a novel rodent model of polycystic kidney disease. *Auton. Neurosci.* 152, 60–66. doi:10.1016/j.autneu.2009.09.019

Hasdan, G., Benchetrit, S., Rashid, G., Green, J., Bernheim, J., and Rathaus, M. (2002). Endothelial dysfunction and hypertension in 5/6 nephrectomized rats are mediated by vascular superoxide. *Kidney Int.* 61, 586–590. doi:10.1046/j.1523-1755. 2002.00166.x

Hausberg, M., Kosch, M., Harmelink, P., Barenbrock, M., Hohage, H., Kisters, K., et al. (2002). Sympathetic nerve activity in end-stage renal disease. *Circulation* 106, 1974–1979. doi:10.1161/01.cir.0000034043.16664.96

Heerspink, H. J., Stefánsson, B. V., Correa-Rotter, R., Chertow, G. M., Greene, T., Hou, F.-F., et al. (2020). Dapagliflozin in patients with chronic kidney disease. *N. Engl. J. Med.* 383, 1436–1446. doi:10.1056/NEJMoa2024816

Heitzer, T., Krohn, K., Albers, S., and Meinertz, T. (2000). Tetrahydrobiopterin improves endothelium-dependent vasodilation by increasing nitric oxide activity in patients with Type II diabetes mellitus. *Diabetologia* 43, 1435–1438. doi:10.1007/ s001250051551

Hering, D., Mahfoud, F., Walton, A. S., Krum, H., Lambert, G. W., Lambert, E. A., et al. (2012). Renal denervation in moderate to severe CKD. *J. Am. Soc. Nephrol.* 23, 1250–1257. doi:10.1681/ASN.2011111062

Heusser, K., Tank, J., Engeli, S., Diedrich, A., Menne, J., Eckert, S., et al. (2010). Carotid baroreceptor stimulation, sympathetic activity, baroreflex function, and blood pressure in hypertensive patients. *Hypertension* 55, 619–626. doi:10.1161/ HYPERTENSIONAHA.109.140665

Hian, C. K., Lee, C. L., and Thomas, W. (2016). Renin-angiotensin-aldosterone system antagonism and polycystic kidney disease progression. *Nephron* 134, 59–63. doi:10.1159/000448296

Hildreth, C. M., Kandukuri, D. S., Goodchild, A. K., and Phillips, J. K. (2013). Temporal development of baroreceptor dysfunction in a rodent model of chronic kidney disease. *Clin. Exp. Pharmacol. Physiol.* 40, 458–465. doi:10.1111/1440-1681. 12110

Hilliard, L. M., Chow, C. L., Mirabito, K. M., Steckelings, U. M., Unger, T., Widdop, R. E., et al. (2014). Angiotensin type 2 receptor stimulation increases renal function in female, but not male, spontaneously hypertensive rats. *Hypertension* 64, 378–383. doi:10.1161/HYPERTENSIONAHA.113.02809

Hilliard, L. M., Jones, E. S., Steckelings, U. M., Unger, T., Widdop, R. E., and Denton, K. M. (2012). Sex-specific influence of angiotensin type 2 receptor stimulation on renal function: A novel therapeutic target for hypertension. *Hypertension* 59, 409–414. doi:10.1161/HYPERTENSIONAHA.111.184986

Hirooka, Y., Kishi, T., Sakai, K., Takeshita, A., and Sunagawa, K. (2011). Imbalance of central nitric oxide and reactive oxygen species in the regulation of sympathetic activity and neural mechanisms of hypertension. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* 300, R818–R826. doi:10.1152/ajpregu.00426.2010

Hovater, M. B., and Sanders, P. W. (2012). Effect of dietary salt on regulation of TGF- $\beta$  in the kidney. *Semin. Nephrol.* 32, 269–276. doi:10.1016/j.semnephrol.2012. 04.006()

Hsu, C.-N., and Tain, Y.-L. (2021). Developmental origins of kidney disease: Why oxidative stress matters? *Antioxidants* 10, 33. doi:10.3390/antiox10010033

Hu, Q., Corda, S., Zweier, J. L., Capogrossi, M. C., and Ziegelstein, R. C. (1998). Hydrogen peroxide induces intracellular calcium oscillations in human aortic endothelial cells. *Circulation* 97, 268–275. doi:10.1161/01.cir.97.3.268

Illig, K. A., Levy, M., Sanchez, L., Trachiotis, G. D., Shanley, C., Irwin, E., et al. (2006). An implantable carotid sinus stimulator for drug-resistant hypertension: Surgical technique and short-term outcome from the multicenter phase II rheos feasibility trial. *J. Vasc. Surg.* 44, 1213–1218. doi:10.1016/j.jvs.2006.08.024

Investigators, S. H.-., Esler, M. D., Krum, H., Sobotka, P. A., Schlaich, M. P., Schmieder, R. E., et al. (2010). Renal sympathetic denervation in patients with treatment-resistant hypertension (the symplicity HTN-2 trial): A randomised controlled trial. *Lancet* 376, 1903–1909. doi:10.1016/S0140-6736(10)62039-9

Ito, Y., Noda, H., Isaka, M., Ando, K., Sato, Y., and Fujita, T. (1989). Norepinephrine responsiveness in patients with borderline hypertension under three different sodium balances. *Clin. Exp. Hypertens. A* 11, 363–370. doi:10.3109/ 10641968909045442

Jeewandara, T. M., Ameer, O. Z., Boyd, R., Wyse, B. F., Underwood, C. F., and Phillips, J. K. (2015). Protective cardiorenal effects of spironolactone in a rodent model of polycystic kidney disease. *Clin. Exp. Pharmacol. Physiol.* 42, 353–360. doi:10.1111/1440-1681.12372

Jofré, R., Rodriguez-Benitez, P., López-Gómez, J. M., and Pérez-Garcia, R. (2006). Inflammatory syndrome in patients on hemodialysis. J. Am. Soc. Nephrol. 17, S274–S280. doi:10.1681/ASN.2006080926

Johansson, M., Elam, M., Rundqvist, B., Eisenhofer, G., Herlitz, H., Lambert, G., et al. (1999). Increased sympathetic nerve activity in renovascular hypertension. *Circulation* 99, 2537–2542. doi:10.1161/01.cir.99.19.2537

Johansson, M., Gao, S. A., Friberg, P., Annerstedt, M., Carlström, J., Ivarsson, T., et al. (2007). Baroreflex effectiveness index and baroreflex sensitivity predict all-cause mortality and sudden death in hypertensive patients with chronic renal failure. *J. Hypertens.* 25, 163–168. doi:10.1097/01.hjh. 0000254377.18983.eb

Johnson, M. D., and Malvin, R. L. (1977). Stimulation of renal sodium reabsorption by angiotensin II. Am. J. Physiol. 232, F298-F306. doi:10.1152/ajprenal.1977.232.4.F298

Kandzari, D. E., Bhatt, D. L., Sobotka, P. A., O'neill, W. W., Esler, M., Flack, J. M., et al. (2012). Catheter-based renal denervation for resistant hypertension: Rationale and design of the SYMPLICITY HTN-3 trial. *Clin. Cardiol.* 35, 528–535. doi:10. 1002/cl.22008

Kaschina, E., Grzesiak, A., Li, J., Foryst-Ludwig, A., Timm, M., Rompe, F., et al. (2008). Angiotensin II type 2 receptor stimulation: A novel option of therapeutic interference with the renin-angiotensin system in myocardial infarction? *Circulation* 118, 2523–2532. doi:10.1161/CIRCULATIONAHA.108.784868

Katholi, R. (1985). Renal nerves and hypertension: An update. Fed. Proc. 44, 2846-2850.()

Katz, S. D., and Krum, H. (2001). Acetylcholine-mediated vasodilation in the forearm circulation of patients with heart failure: Indirect evidence for the role of endothelium-derived hyperpolarizing factor. *Am. J. Cardiol.* 87, 1089–1092. doi:10. 1016/s0002-9149(01)01466-7

Kelm, M., Schäfer, S., Mingers, S., Heydthausen, M., Vogt, M., Motz, W., et al. (1996). Left ventricular mass is linked to cardiac noradrenaline in normotensive and hypertensive patients. *J. Hypertens.* 14, 1357–1364. doi:10.1097/00004872-199611000-00015

Kemp, B. A., Bell, J. F., Rottkamp, D. M., Howell, N. L., Shao, W., Navar, L. G., et al. (2012). Intrarenal angiotensin III is the predominant agonist for proximal tubule angiotensin type 2 receptors. *Hypertension* 60, 387–395. doi:10.1161/HYPERTENSIONAHA.112.191403

Kielstein, J. T., Böger, R. H., Bode-Böger, S. M., Schäffer, J., Barbey, M., Koch, K. M., et al. (1999). Asymmetric dimethylarginine plasma concentrations differ in patients with end-stage renal disease Relationship to treatment method and atherosclerotic disease. J. Am. Soc. Nephrol. 10, 594–600. doi:10.1681/ASN. V103594

Kimura, K., Ohkita, M., Koyama, M., and Matsumura, Y. (2012). Reduced NO production rapidly aggravates renal function through the NF- $\kappa$ B/ET-1/ETA receptor pathway in DOCA-salt-induced hypertensive rats. *Life Sci.* 91, 644–650. doi:10.1016/j.lfs.2012.04.022

Klag, M. J., Whelton, P. K., Randall, B. L., Neaton, J. D., Brancati, F. L., Ford, C. E., et al. (1996). Blood pressure and end-stage renal disease in men. *N. Engl. J. Med.* 334, 13–18. doi:10.1056/NEJM199601043340103

Klein, I. H., Ligtenberg, G., Neumann, J., Oey, P. L., Koomans, H. A., and Blankestijn, P. J. (2003a). Sympathetic nerve activity is inappropriately increased in chronic renal disease. J. Am. Soc. Nephrol. 14, 3239–3244. doi:10.1097/01.asn.0000098687.01005.a5

Klein, I. H., Ligtenberg, G., Oey, P. L., Koomans, H. A., and Blankestijn, P. J. (2003b). Enalapril and losartan reduce sympathetic hyperactivity in patients with chronic renal failure. J. Am. Soc. Nephrol. 14, 425–430. doi:10.1097/01.asn.0000045049.72965.b7

Kohan, D. E., and Barton, M. (2014). Endothelin and endothelin antagonists in chronic kidney disease. *Kidney Int.* 86, 896–904. doi:10.1038/ki.2014.143

Komiyama, Y., Dong, X. H., Nishimura, N., Masaki, H., Yoshika, M., Masuda, M., et al. (2005). A novel endogenous digitalis, telocinobufagin, exhibits elevated plasma levels in patients with terminal renal failure. *Clin. Biochem.* 38, 36–45. doi:10.1016/j. clinbiochem.2004.08.005

Koomans, H., Roos, J., Boer, P., Geyskes, G., and Mees, E. (1982). Salt sensitivity of blood pressure in chronic renal failure. Evidence for renal control of body fluid distribution in man. *Hypertension* 4, 190–197. doi:10.1161/01.hyp.4.2.190

Krapf, R., and Hulter, H. N. (2009). Arterial hypertension induced by erythropoietin and erythropoiesis-stimulating agents (ESA). *Clin. J. Am. Soc. Nephrol.* 4, 470–480. doi:10.2215/CJN.05040908

Krum, H., Schlaich, M., Whitbourn, R., Sobotka, P. A., Sadowski, J., Bartus, K., et al. (2009). Catheter-based renal sympathetic denervation for resistant hypertension: A multicentre safety and proof-of-principle cohort study. *Lancet* 373, 1275–1281. doi:10.1016/S0140-6736(09)60566-3

Kruse, N. T., You, Z., Moreau, K., Kendrick, J., and Jalal, D. (2020). Sex differences in endothelial function in chronic kidney disease. *Am. J. Physiol. Ren. Physiol.* 319, F33–f40. doi:10.1152/ajprenal.00156.2020

Ku, E., Lee, B. J., Wei, J., and Weir, M. R. (2019). Hypertension in CKD: Core curriculum 2019. Am. J. Kidney Dis. 74, 120–131. doi:10.1053/j.ajkd.2018.12.044

Kukida, M., Mogi, M., Kan-No, H., Tsukuda, K., Bai, H. Y., Shan, B. S., et al. (2019). AT2 receptor stimulation inhibits phosphate-induced vascular calcification. *Kidney Int.* 95, 138–148. doi:10.1016/j.kint.2018.07.028

Kusama, N., Kajikuri, J., Yamamoto, T., Watanabe, Y., Suzuki, Y., Katsuya, H., et al. (2005). Reduced hyperpolarization in endothelial cells of rabbit aortic valve following chronic nitroglycerine administration. *Br. J. Pharmacol.* 146, 487–497. doi:10.1038/sj.bip.0706363

Lacolley, P., Safar, M. E., Lucet, B., Ledudal, K., Labat, C., and Benetos, A. (2001). Prevention of aortic and cardiac fibrosis by spironolactone in old normotensive rats. J. Am. Coll. Cardiol. 37, 662–667. doi:10.1016/s0735-1097(00)01129-3

Lacy, P., Carr, S. J., O'brien, D., Fentum, B., Williams, B., Paul, S. K., et al. (2006). Reduced glomerular filtration rate in pre-dialysis non-diabetic chronic kidney disease patients is associated with impaired baroreceptor sensitivity and reduced vascular compliance. *Clin. Sci.* 110, 101–108. doi:10.1042/CS20050192

Lee, H. T., Kim, J. Y., Kim, M., Wang, P., Tang, L., Baroni, S., et al. (2013). Renalase protects against ischemic AKI. *J. Am. Soc. Nephrol.* 24, 445–455. doi:10. 1681/ASN.2012090943

Leone, A., Moncada, S., Vallance, P., Calver, A., and Collier, J. (1992). Accumulation of an endogenous inhibitor of nitric oxide synthesis in chronic renal failure. *Lancet* 339, 572–575. doi:10.1016/0140-6736(92)90865-z Li, G., Xu, J., Wang, P., Velazquez, H., Li, Y., Wu, Y., et al. (2008). Catecholamines regulate the activity, secretion, and synthesis of renalase. *Circulation* 117, 1277–1282. doi:10.1161/CIRCULATIONAHA.107.732032

Li, Q., and Fung, E. (2019). Multifaceted functions of epithelial Na+ channel in modulating blood pressure. *Hypertension* 73, 273–281. doi:10.1161/HYPERTENSIONAHA.118.12330

Liang, B., Zhao, Y.-X., and Gu, N. (2020). Renal denervation for resistant hypertension: Where do we stand? *Curr. Hypertens. Rep.* 22, 83–89. doi:10. 1007/s11906-020-01094-6

Liang, C., Zhang, Y., Zhuo, D., Lo, C.-Y., Yu, L., Lau, C.-W., et al. (2019). Endothelial cell transient receptor potential channel C5 (TRPC5) is essential for endothelium-dependent contraction in mouse carotid arteries. *Biochem. Pharmacol.* 159, 11–24. doi:10.1016/j.bcp.2018.11.002

Ligtenberg, G., Blankestijn, P. J., Oey, P. L., Klein, I. H., Dijkhorst-Oei, L.-T., Boomsma, F., et al. (1999). Reduction of sympathetic hyperactivity by enalapril in patients with chronic renal failure. *N. Engl. J. Med.* 340, 1321–1328. doi:10.1056/ NEJM199904293401704

Lin, S.-L., Chen, R.-H., Chen, Y.-M., Chiang, W.-C., Lai, C.-F., Wu, K.-D., et al. (2005). Pentoxifylline attenuates tubulointerstitial fibrosis by blocking Smad3/4-activated transcription and profibrogenic effects of connective tissue growth factor. *J. Am. Soc. Nephrol.* 16, 2702–2713. doi:10.1681/ASN.2005040435

Liu, Z., Shen, L., Huang, W., Zhao, X., Fang, W., Wang, C., et al. (2017). Efficacy and safety of renal denervation for Chinese patients with resistant hypertension using a microirrigated catheter: Study design and protocol for a prospective multicentre randomised controlled trial. *BMJ open* 7, e015672. doi:10.1136/ bmjopen-2016-015672

Lobov, G., and Ivanova, G. (2020). Regulation of arterial tone in rats fed a long-term high-salt diet. *J. Evol. Biochem. Physiol.* 57, 145–155. doi:10.1134/s0022093021010142

Lohmeier, T. E., and Iliescu, R. (2011). Chronic lowering of blood pressure by carotid baroreflex activation: Mechanisms and potential for hypertension therapy. *Hypertension* 57, 880–886. doi:10.1161/HYPERTENSIONAHA.108.119859

Lohmeier, T. E., Irwin, E. D., Rossing, M. A., Serdar, D. J., and Kieval, R. S. (2004). Prolonged activation of the baroreflex produces sustained hypotension. *Hypertension* 43, 306–311. doi:10.1161/01.HYP.0000111837.73693.9b

Luksha, L., Agewall, S., and Kublickiene, K. (2009). Endothelium-derived hyperpolarizing factor in vascular physiology and cardiovascular disease. *Atherosclerosis* 202, 330–344. doi:10.1016/j.atherosclerosis.2008.06.008

Luksha, L., Stenvinkel, P., Hammarqvist, F., Carrero, J. J., Davidge, S. T., and Kublickiene, K. (2012). Mechanisms of endothelial dysfunction in resistance arteries from patients with end-stage renal disease. *PloS one* 7, e36056. doi:10. 1371/journal.pone.0036056

Lüscher, T. F., and Vanhoutte, P. M. (1990). The endothelium: Modulator of cardiovascular function. Boca Raton, FL: CRC Press.

Lytvyn, Y., Bjornstad, P., Udell, J. A., Lovshin, J. A., and Cherney, D. Z. (2017). Sodium glucose cotransporter-2 inhibition in heart failure: Potential mechanisms, clinical applications, and summary of clinical trials. *Circulation* 136, 1643–1658. doi:10.1161/CIRCULATIONAHA.117.030012

Middleton, J. P., and Pun, P. H. (2010). Hypertension, chronic kidney disease, and the development of cardiovascular risk: A joint primacy. *Kidney Int.* 77, 753–755. doi:10.1038/ki.2010.19

Mahajan, D., Wang, Y., Qin, X., Wang, Y., Zheng, G., Wang, Y. M., et al. (2006). CD4+CD25+ regulatory T cells protect against injury in an innate murine model of chronic kidney disease. *J. Am. Soc. Nephrol.* 17, 2731–2741. doi:10.1681/ASN. 2005080842

Maier, W., Cosentino, F., Lütolf, R. B., Fleisch, M., Seiler, C., Hess, O. M., et al. (2000). Tetrahydrobiopterin improves endothelial function in patients with coronary artery disease. J. Cardiovasc. Pharmacol. 35, 173–178. doi:10.1097/00005344-200002000-00001

Maizel, J., Six, I., Slama, M., Tribouilloy, C., Sevestre, H., Poirot, S., et al. (2009). Mechanisms of aortic and cardiac dysfunction in uremic mice with aortic calcification. *Circulation* 119, 306–313, doi:10.1161/CIRCULATIONAHA.108.797407

Makaritsis, K. P., Liakopoulos, V., Leivaditis, K., Eleftheriadis, T., and Stefanidis, I. (2006). Adaptation of renal function in heart failure. *Ren. Fail.* 28, 527–535. doi:10.1080/08860220600839522

Mancia, G., Grassi, G., Giannattasio, C., and Seravalle, G. (1999). Sympathetic activation in the pathogenesis of hypertension and progression of organ damage. *Hypertension* 34, 724–728. doi:10.1161/01.hyp.34.4.724

Martinez-Maldonado, M. (1998). Hypertension in end-stage renal disease. Kidney Int. Suppl. 54, S67-S72. doi:10.1046/j.1523-1755.1998.06816.x

Masuda, T., and Nagata, D. (2020). Recent advances in the management of secondary hypertension: Chronic kidney disease. *Hypertens. Res.* 43, 869–875. doi:10.1038/s41440-020-0491-4

Mathew, R. O., Bangalore, S., Lavelle, M. P., Pellikka, P. A., Sidhu, M. S., Boden, W. E., et al. (2017). Diagnosis and management of atherosclerotic cardiovascular disease in chronic kidney disease: A review. *Kidney Int.* 91, 797–807. doi:10.1016/j. kint.2016.09.049

Mccooke, J. K., Appels, R., Barrero, R. A., Ding, A., Ozimek-Kulik, J. E., Bellgard, M. I., et al. (2012). A novel mutation causing nephronophthisis in the Lewis polycystic kidney rat localises to a conserved RCC1 domain in Nek8. *BMC Genomics* 13, 393–2164. doi:10.1186/1471-2164-13-393

Mene, P., Punzo, G., and Pirozzi, N. (2013). TRP channels as therapeutic targets in kidney disease and hypertension. *Curr. Top. Med. Chem.* 13, 386–397. doi:10. 2174/1568026611313030013

Mennuni, S., Rubattu, S., Pierelli, G., Tocci, G., Fofi, C., and Volpe, M. (2014). Hypertension and kidneys: Unraveling complex molecular mechanisms underlying hypertensive renal damage. *J. Hum. Hypertens.* 28, 74–79. doi:10.1038/jhh.2013.55

Michea, L., Villagrán, A., Urzúa, A., Kuntsmann, S., Venegas, P., Carrasco, L., et al. (2008). Mineralocorticoid receptor antagonism attenuates cardiac hypertrophy and prevents oxidative stress in uremic rats. *Hypertension* 52, 295–300. doi:10.1161/HYPERTENSIONAHA.107.109645

Mirza, M. A., Hansen, T., Johansson, L., Ahlström, H., Larsson, A., Lind, L., et al. (2009). Relationship between circulating FGF23 and total body atherosclerosis in the community. *Nephrol. Dial. Transpl.* 24, 3125–3131. doi:10.1093/ndt/gfp205

Moldovan, D., Rusu, C., Patiu, I., Racasan, S., Orasan, R., Kacso, I., et al. (2010). Could the serum parathormone be a predictive marker for peripheral vascular calcifications in chronic dialysis patients? Experience of a single center in transylvania. *Acta Endocrinol. (Copenh).*, 43–55. doi:10.4183/aeb.2010.43

Moreau, K. L., Meditz, A., Deane, K. D., and Kohrt, W. M. (2012). Tetrahydrobiopterin improves endothelial function and decreases arterial stiffness in estrogen-deficient postmenopausal women. *Am. J. Physiol. Heart Circ. Physiol.* 302, H1211–H1218. doi:10.1152/ajpheart.01065.2011

Moretti, J. L., Burke, S. L., Evans, R. G., Lambert, G. W., and Head, G. A. (2009). Enhanced responses to ganglion blockade do not reflect sympathetic nervous system contribution to angiotensin II-induced hypertension. *J. Hypertens.* 27, 1838–1848. doi:10.1097/HJH.0b013e32832dd0d8

Morgado, E., and Neves, P. L. (2012). Hypertension and chronic kidney disease: Cause and consequence-therapeutic considerations. *Antihypertens. drugs* 45. doi:10.5772/36916

Morris, S. T., Mcmurray, J. J., Rodger, R. S., and Jardine, A. G. (2000). Impaired endothelium-dependent vasodilatation in uraemia. *Nephrol. Dial. Transpl.* 15, 1194–1200. doi:10.1093/ndt/15.8.1194

Mortensen, L. H. (1999). Endothelin and the central and peripheral nervous systems: A decade of endothelin research. *Clin. Exp. Pharmacol. Physiol.* 26, 980–984. doi:10.1046/j.1440-1681.1999.03175.x

Muntner, P., He, J., Astor, B. C., Folsom, A. R., and Coresh, J. (2005). Traditional and nontraditional risk factors predict coronary heart disease in chronic kidney disease: Results from the atherosclerosis risk in communities study. J. Am. Soc. Nephrol. 16, 529–538. doi:10.1681/ASN.2004080656

Munzel, T., Gori, T., Bruno, R. M., and Taddei, S. (2010). Is oxidative stress a therapeutic target in cardiovascular disease? *Eur. Heart J.* 31, 2741–2748. doi:10. 1093/eurheartj/ehq396

Nasrallah, R., and Hébert, R. L. (2005). Prostacyclin signaling in the kidney: Implications for health and disease. *Am. J. Physiol. Ren. Physiol.* 289, F235–F246. doi:10.1152/ajprenal.00454.2004

Nauli, S. M., Jin, X., and Hierck, B. P. (2011). The mechanosensory role of primary cilia in vascular hypertension. *Int. J. Vasc. Med.*, 2011. 376281. doi:10.1155/2011/376281

Nawrot, T. S., Staessen, J. A., Holvoet, P., Struijker-Boudier, H. A., Schiffers, P., Van Bortel, L. M., et al. (2010). Telomere length and its associations with oxidized-LDL, carotid artery distensibility and smoking. *Front. Biosci.* 2, 1164–1168. doi:10. 2741/e176

Neumann, J., Ligtenberg, G., Klein, I. H., Boer, P., Oey, P. L., Koomans, H. A., et al. (2007). Sympathetic hyperactivity in hypertensive chronic kidney disease patients is reduced during standard treatment. *Hypertension* 49, 506–510. doi:10. 1161/01.HYP.0000256530.39695.a3

Neumann, J., Ligtenberg, G., Klein, I. I., Koomans, H. A., and Blankestijn, P. J. (2004). Sympathetic hyperactivity in chronic kidney disease: Pathogenesis, clinical relevance, and treatment. *Kidney Int.* 65, 1568–1576. doi:10.1111/j.1523-1755.2004. 00552.x

Ng, K., Hildreth, C. M., Avolio, A. P., and Phillips, J. K. (2011). Angiotensinconverting enzyme inhibitor limits pulse-wave velocity and aortic calcification in a rat model of cystic renal disease. *Am. J. Physiol. Ren. Physiol.* 301, F959–F966. doi:10.1152/ajprenal.00393.2011 Nikolov, I., Joki, N., Maizel, J., Lacour, B., Drüeke, T., and Massy, Z. (2006). Pleiotropic effects of the non-calcium phosphate binder sevelamer. *Kidney Int.* 70, S16–S23. doi:10.1038/sj.ki.5001994

Nishiyama, A., Yao, L., Nagai, Y., Miyata, K., Yoshizumi, M., Kagami, S., et al. (2004). Possible contributions of reactive oxygen species and mitogen-activated protein kinase to renal injury in aldosterone/salt-induced hypertensive rats. *Hypertension* 43, 841–848. doi:10.1161/01.HYP.0000118519.66430.22

Obi, Y., Kalantar-Zadeh, K., Shintani, A., Kovesdy, C., and Hamano, T. (2018). Estimated glomerular filtration rate and the risk-benefit profile of intensive blood pressure control amongst nondiabetic patients: A post hoc analysis of a randomized clinical trial. *J. Intern. Med.* 283, 314–327. doi:10. 1111/joim.12701

Olauson, H., and Larsson, T. E. (2013). FGF23 and Klotho in chronic kidney disease. *Curr. Opin. Nephrol. Hypertens.* 22, 397–404. doi:10.1097/MNH. 0b013e32836213ee

Ott, C., Mahfoud, F., Schmid, A., Toennes, S. W., Ewen, S., Ditting, T., et al. (2015). Renal denervation preserves renal function in patients with chronic kidney disease and resistant hypertension. *J. Hypertens.* 33, 1261–1266. doi:10.1097/HJH. 00000000000556

Ozono, R., Wang, Z. Q., Moore, A. F., Inagami, T., Siragy, H. M., and Carey, R. M. (1997). Expression of the subtype 2 angiotensin (AT2) receptor protein in rat kidney. *Hypertension* 30, 1238–1246. doi:10.1161/01.hyp.30.5.1238

Pacurari, M., Xing, D., Hilgers, R. H., Guo, Y. Y., Yang, Z., and Hage, F. G. (2013). Endothelial cell transfusion ameliorates endothelial dysfunction in 5/ 6 nephrectomized rats. *Am. J. Physiol. Heart Circ. Physiol.* 305, H1256–H1264. doi:10.1152/ajpheart.00132.2013

Palm, F., Onozato, M. L., Luo, Z., and Wilcox, C. S. (2007). Dimethylarginine dimethylaminohydrolase (DDAH): Expression, regulation, and function in the cardiovascular and renal systems. *Am. J. Physiol. Heart Circ. Physiol.* 293, H3227–H3245. doi:10.1152/ajpheart.00998.2007

Pan, I. Z., Carey, J. R., Jacobs, J. A., Dechand, J., Sessions, J. J., Sorensen, T., et al. (2020). Transitioning between prostanoid therapies in pulmonary arterial hypertension. *Front. Med.* 7, 81. doi:10.3389/fmed.2020.00081

Pannier, B., Guerin, A. P., Marchais, S. J., Metivier, F., Safar, M. E., and London, G. M. (2000). Postischemic vasodilation, endothelial activation, and cardiovascular remodeling in end-stage renal disease. *Kidney Int.* 57, 1091–1099. doi:10.1046/j.1523-1755.2000.00936.x

Paravicini, T. M., and Touyz, R. M. (2008). NADPH oxidases, reactive oxygen species, and hypertension: Clinical implications and therapeutic possibilities. *Diabetes care* 31, S170-S180. doi:10.2337/dc08-s247

Park, J., Liao, P., Sher, S., Lyles, R. H., Deveaux, D. D., and Quyyumi, A. A. (2015). Tetrahydrobiopterin lowers muscle sympathetic nerve activity and improves augmentation index in patients with chronic kidney disease. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* 308, R208–R218. doi:10.1152/ajpregu.00409.2014

Passauer, J., Bussemaker, E., Range, U., Plug, M., and Gross, P. (2000). Evidence *in vivo* showing increase of baseline nitric oxide generation and impairment of endothelium-dependent vasodilation in normotensive patients on chronic hemodialysis. *J. Am. Soc. Nephrol.* 11, 1726–1734. doi:10.1681/ASN.V1191726

Passauer, J., Pistrosch, F., Büssemaker, E., Lässig, G., Herbrig, K., and Gross, P. (2005). Reduced agonist-induced endothelium-dependent vasodilation in uremia is attributable to an impairment of vascular nitric oxide. *J. Am. Soc. Nephrol.* 16, 959–965. doi:10.1681/ASN.2004070582

Patel, K. P., Li, Y. F., and Hirooka, Y. (2001). Role of nitric oxide in central sympathetic outflow. *Exp. Biol. Med.* 226, 814–824. doi:10.1177/153537020122600902

Patschan, O., Kuttler, B., Heemann, U., Uber, A., and Rettig, R. (1997). Kidneys from normotensive donors lower blood pressure in young transplanted spontaneously hypertensive rats. *Am. J. Physiol.* 273, R175–R180. doi:10.1152/ajpregu.1997.273.1.R175

Pecoits-Filho, R., Heimbürger, O., Bárány, P., Suliman, M., Fehrman-Ekholm, I., Lindholm, B., et al. (2003). Associations between circulating inflammatory markers and residual renal function in CRF patients. *Am. J. Kidney Dis.* 41, 1212–1218. doi:10.1016/s0272-6386(03)00353-6

Perry, H. M., Miller, J. P., Fornoff, J. R., Baty, J. D., Sambhi, M. P., Rutan, G., et al. (1995). Early predictors of 15-year end-stage renal disease in hypertensive patients. *Hypertension* 25, 587–594. doi:10.1161/01.hyp.25.4.587

Peters, H., Border, W. A., and Noble, N. A. (1999). From rats to man: A perspective on dietary L-arginine supplementation in human renal disease. *Nephrol. Dial. Transpl.* 14, 1640–1650. doi:10.1093/ndt/14.7.1640

Plata, C., Cruz, C., Cervantes, L. G., and Ramírez, V. (2019). The gut microbiota and its relationship with chronic kidney disease. *Int. Urol. Nephrol.* 51, 2209–2226. doi:10.1007/s11255-019-02291-2

Podjarny, E., Hasdan, G., Bernheim, J., Rashid, G., Green, J., Korzets, Z., et al. (2004). Effect of chronic tetrahydrobiopterin supplementation on blood pressure

and proteinuria in 5/6 nephrectomized rats. Nephrol. Dial. Transpl. 19, 2223-2227. doi:10.1093/ndt/gfh383

Porkert, M., Sher, S., Reddy, U., Cheema, F., Niessner, C., Kolm, P., et al. (2008). Tetrahydrobiopterin: A novel antihypertensive therapy. *J. Hum. Hypertens.* 22, 401–407. doi:10.1038/sj.jhh.1002329

Preston, R. A., Singer, I., and Epstein, M. (1996). Renal parenchymal hypertension: Current concepts of pathogenesis and management. *Arch. Intern. Med.* 156, 602–611. doi:10.1001/archinte.156.6.602

Qi, Z., Cai, H., Morrow, J. D., and Breyer, M. D. (2006). Differentiation of cyclooxygenase 1-and 2-derived prostanoids in mouse kidney and aorta. *Hypertension* 48, 323–328. doi:10.1161/01.HYP.0000231934.67549.b7

Qi, Z., Hao, C. M., Langenbach, R. I., Breyer, R. M., Redha, R., Morrow, J. D., et al. (2002). Opposite effects of cyclooxygenase-1 and -2 activity on the pressor response to angiotensin II. *J. Clin. Invest.* 110, 61–69. doi:10.1172/JCI14752

Quan, A., Chakravarty, S., Chen, J. K., Chen, J. C., Loleh, S., Saini, N., et al. (2004). Androgens augment proximal tubule transport. *Am. J. Physiol. Ren. Physiol.* 287, F452–F459. doi:10.1152/ajprenal.00188.2003

Quek, K. J., Ameer, O. Z., and Phillips, J. K. (2020). Amlodipine improves vessel function and remodelling in the Lewis Polycystic Kidney rat mesenteric artery. *Am. J. Hypertens.* 

Quek, K. J., Ameer, O. Z., and Phillips, J. K. (2018). AT1 receptor antagonism improves structural, functional, and biomechanical properties in resistance arteries in a rodent chronic kidney disease model. *Am. J. Hypertens.* 31, 696–705. doi:10. 1093/ajh/hpy021

Quek, K. J., Boyd, R., Ameer, O. Z., Zangerl, B., Butlin, M., Murphy, T. V., et al. (2016). Progressive vascular remodelling, endothelial dysfunction and stiffness in mesenteric resistance arteries in a rodent model of chronic kidney disease. *Vasc. Pharmacol.* 81, 42–52. doi:10.1016/j.vph.2015.12.004

Raina, R., Chauvin, A., Chakraborty, R., Nair, N., Shah, H., Krishnappa, V., et al. (2020). The role of endothelin and endothelin antagonists in chronic kidney disease. *Kidney Dis.* 6, 22–34. doi:10.1159/000504623

Raine, A. E., Bedford, L., Simpson, A. W., Ashley, C. C., Brown, R., Woodhead, J. S., et al. (1993). Hyperparathyroidism, platelet intracellular free calcium and hypertension in chronic renal failure. *Kidney Int.* 43, 700–705. doi:10.1038/ki. 1993.100

Raine, A. (1988). Hypertension, blood viscosity, and cardiovascular morbidity in renal failure: Implications of erythropoietin therapy. *Lancet* 331, 97–100. doi:10. 1016/s0140-6736(88)90293-0

Rancourt, M. E., Rodrigue, M. E., Agharazii, M., Lariviere, R., and Lebel, M. (2010). Role of oxidative stress in erythropoietin-induced hypertension in uremic rats. *Am. J. Hypertens.* 23, 314–320. doi:10.1038/ajh.2009.242

Rayego-Mateos, S., and Valdivielso, J. M. (2020). New therapeutic targets in chronic kidney disease progression and renal fibrosis. *Expert Opin. Ther. Targets* 24, 655–670. doi:10.1080/14728222.2020.1762173

Recordati, G., Moss, N. G., Genovesi, S., and Rogenes, P. (1981). Renal chemoreceptors. J. Auton. Nerv. Syst. 3, 237-251. doi:10.1016/0165-1838(81) 90066-7

Reid, I. (1992). Interactions between ANG II, sympathetic nervous system, and baroreceptor reflexes in regulation of blood pressure. *Am. J. Physiol.* 262, E763–E778. doi:10.1152/ajpendo.1992.262.6.E763

Rettig, R., Folberth, C., Stauss, H., Kopf, D., Waldherr, R., and Unger, T. (1990). Role of the kidney in primary hypertension: A renal transplantation study in rats. *Am. J. Physiol.* 258, F606–F611. doi:10.1152/ajprenal.1990.258.3.F606

Rocco, M. V., Sink, K. M., Lovato, L. C., Wolfgram, D. F., Wiegmann, T. B., Wall, B. M., et al. (2018). Effects of intensive blood pressure treatment on acute kidney injury events in the Systolic Blood Pressure Intervention Trial (SPRINT). *Am. J. Kidney Dis.* 71, 352–361. doi:10.1053/j.ajkd.2017.08.021

Rodriguez-Iturbe, B., Romero, F., and Johnson, R. J. (2007). Pathophysiological mechanisms of salt-dependent hypertension. *Am. J. Kidney Dis.* 50, 655–672. doi:10.1053/j.ajkd.2007.05.025

Rodríguez-Romo, R., Benítez, K., Barrera-Chimal, J., Pérez-Villalva, R., Gómez, A., Aguilar-León, D., et al. (2016). AT1 receptor antagonism before ischemia prevents the transition of acute kidney injury to chronic kidney disease. *Kidney Int.* 89, 363–373. doi:10.1038/ki.2015.320

Rodríguez-Vita, J., Sánchez-López, E., Esteban, V., Rupérez, M., Egido, J., and Ruiz-Ortega, M. (2005). Angiotensin II activates the Smad pathway in vascular smooth muscle cells by a transforming growth factor- $\beta$ -independent mechanism. *Circulation* 111, 2509–2517. doi:10.1161/01.CIR.0000165133.84978.E2

Rosenberg, M. E., Smith, L. J., Correa-Rotter, R., and Hostetter, T. H. (1994). The paradox of the renin-angiotensin system in chronic renal disease. *Kidney Int.* 45, 403–410. doi:10.1038/ki.1994.52

Roumeliotis, S., Mallamaci, F., and Zoccali, C. (2020). Endothelial dysfunction in chronic kidney disease, from biology to clinical outcomes: A 2020 update. *J. Clin. Med.* 9, 2359. doi:10.3390/jcm9082359

Rubin, M. R., Maurer, M. S., Mcmahon, D. J., Bilezikian, J. P., and Silverberg, S. J. (2005). Arterial stiffness in mild primary hyperparathyroidism. *J. Clin. Endocrinol. Metab.* 90, 3326–3330. doi:10.1210/jc.2004-1400

Ruiz-Ortega, M., Lorenzo, O., RupéRez, M. N., KöNig, S., Wittig, B., and Egido, J. S. (2000). Angiotensin II activates nuclear transcription factor kappaB through AT(1) and AT(2) in vascular smooth muscle cells: Molecular mechanisms. *Circ. Res.* 86, 1266–1272. doi:10.1161/01.res.86.12.1266

Safar, M. E., Levy, B. I., and Struijker-Boudier, H. (2003). Current perspectives on arterial stiffness and pulse pressure in hypertension and cardiovascular diseases. *Circulation* 107, 2864–2869. doi:10.1161/01.CIR.0000069826.36125.B4

Salem, M. M. (2002). Pathophysiology of hypertension in renal failure. Semin. Nephrol. 22, 17-26. doi:10.1053/snep.2002.28640()

Salman, I. M. (2015). Cardiovascular autonomic dysfunction in chronic kidney disease: A comprehensive review. *Curr. Hypertens. Rep.* 17, 59. doi:10.1007/s11906-015-0571-z

Salman, I. M., Hildreth, C. M., Ameer, O. Z., and Phillips, J. K. (2014). Differential contribution of afferent and central pathways to the development of baroreflex dysfunction in chronic kidney disease. *Hypertension* 63, 804–810. doi:10.1161/ HYPERTENSIONAHA.113.02110

Salman, I. M., Phillips, J. K., Ameer, O. Z., and Hildreth, C. M. (2015a). Abnormal central control underlies impaired baroreflex control of heart rate and sympathetic nerve activity in female Lewis polycystic kidney rats. *J. Hypertens.* 33, 1418–1428. doi:10.1097/HJH.00000000000572

Salman, I. M., Sarma Kandukuri, D., Harrison, J. L., Hildreth, C. M., and Phillips, J. K. (2015b). Direct conscious telemetry recordings demonstrate increased renal sympathetic nerve activity in rats with chronic kidney disease. *Front. Physiol.* 6, 218. doi:10.3389/fphys.2015.00218

Sanchez, O. A., Ferrara, L. K., Rein, S., Berglund, D., Matas, A. J., and Ibrahim, H. N. (2018). Hypertension after kidney donation: Incidence, predictors, and correlates. *Am. J. Transpl.* 18, 2534–2543. doi:10.1111/ajt.14713

Sata, Y., Burke, S. L., Watson, A. M., Jha, J. C., Gueguen, C., Eikelis, N., et al. (2020). Differential sympathetic response to lesion-induced chronic kidney disease in rabbits. *Kidney Int.* 98, 906–917. doi:10.1016/j.kint.2020.03.039

Sata, Y., Head, G. A., Denton, K., May, C. N., and Schlaich, M. P. (2018). Role of the sympathetic nervous system and its modulation in renal hypertension. *Front. Med.* 5, 82. doi:10.3389/fmed.2018.00082

Sato, E., Hosomi, K., Sekimoto, A., Mishima, E., Oe, Y., Saigusa, D., et al. (2020). Effects of the oral adsorbent AST-120 on fecal p-cresol and indole levels and on the gut microbiota composition. *Biochem. Biophys. Res. Commun.* 525, 773–779. doi:10. 1016/j.bbrc.2020.02.141

Saxena, T., Ali, A. O., and Saxena, M. (2018). Pathophysiology of essential hypertension: An update. *Expert Rev. cardiovasc. Ther.* 16, 879–887. doi:10.1080/14779072.2018.1540301

Scalera, F., Closs, E. I., Flick, E., Martens-Lobenhoffer, J., Boissel, J. P., Lendeckel, U., et al. (2009). Paradoxical effect of L-arginine: Acceleration of endothelial cell senescence. *Biochem. Biophys. Res. Commun.* 386, 650–655. doi:10.1016/j.bbrc.2009.06.091

Scalera, F., Kielstein, J. T., Martens-Lobenhoffer, J., Postel, S. C., Täger, M., and Bode-Böger, S. M. (2005). Erythropoietin increases asymmetric dimethylarginine in endothelial cells: Role of dimethylarginine dimethylaminohydrolase. *J. Am. Soc. Nephrol.* 16, 892–898. doi:10.1681/ASN.2004090735

Schiffrin, E. L., Lipman, M. L., and Mann, J. F. (2007). Chronic kidney disease: Effects on the cardiovascular system. *Circulation* 116, 85–97. doi:10.1161/ CIRCULATIONAHA.106.678342

Schlaich, M. P., Socratous, F., Hennebry, S., Eikelis, N., Lambert, E. A., Straznicky, N., et al. (2009). Sympathetic activation in chronic renal failure. *J. Am. Soc. Nephrol.* 20, 933–939. doi:10.1681/ASN.2008040402

Schmidt, T. S., Mcneill, E., Douglas, G., Crabtree, M. J., Hale, A. B., Khoo, J., et al. (2010). Tetrahydrobiopterin supplementation reduces atherosclerosis and vascular inflammation in apolipoprotein E-knockout mice. *Clin. Sci.* 119, 131–142. doi:10.1042/CS20090559

Schneider, M. P., and Mann, J. F. (2014). Endothelin antagonism for patients with chronic kidney disease: Still a hope for the future. *Nephrol. Dial. Transpl.* 29, i69–i73. doi:10.1093/ndt/gft339

Schohn, D., Weidmann, P., Jahn, H., and Beretta-Piccoli, C. (1985). Norepinephrine-related mechanism in hypertension accompanying renal failure. *Kidney Int.* 28, 814–822. doi:10.1038/ki.1985.203

Schoner, W. (2002). Endogenous cardiac glycosides, a new class of steroid hormones. *Eur. J. Biochem.* 269, 2440–2448. doi:10.1046/j.1432-1033.2002.02911.x

Schrier, R. W., Abebe, K. Z., Perrone, R. D., Torres, V. E., Braun, W. E., Steinman, T. I., et al. (2014). Blood pressure in early autosomal dominant polycystic kidney disease. *N. Engl. J. Med.* 371, 2255–2266. doi:10.1056/NEJMoa1402685

Seddon, M. D., Chowienczyk, P. J., Brett, S. E., Casadei, B., and Shah, A. M. (2008). Neuronal nitric oxide synthase regulates basal microvascular tone in humans *in vivo*. *Circulation* 117, 1991–1996. doi:10.1161/CIRCULATIONAHA.107.744540

Sendao Oliveira, A., and Bendhack, L. M. (2004). Relaxation induced by acetylcholine involves endothelium-derived hyperpolarizing factor in 2-kidney 1-clip hypertensive rat carotid arteries. *Pharmacology* 72, 231–239. doi:10.1159/000080378

Shajahan, S., Amin, J., Phillips, J. K., and Hildreth, C. M. (2021). Relationship between sex and cardiovascular mortality in chronic kidney disease: A systematic review and meta-analysis. *PLoS One* 16, e0254554. doi:10.1371/journal.pone.0254554

Shultz, P. J. (1992). An emerging role for endothelin in renal disease. J. Lab. Clin. Med. 119, 448-449.

Sica, D., and Carl, D. (2005). Pathologic basis and treatment considerations in chronic kidney disease-related hypertension. *Semin. Nephrol.* 25, 246–251. doi:10. 1016/j.semnephrol.2005.03.001()

Sies, H. (1997). Oxidative stress: Oxidants and antioxidants. *Exp. Physiol.* 82, 291–295. doi:10.1113/expphysiol.1997.sp004024

Sievert, H., Schofer, J., Ormiston, J., Hoppe, U. C., Meredith, I., Walters, D. L., et al. (2017). Bipolar radiofrequency renal denervation with the vessix catheter in patients with resistant hypertension: 2-year results from the REDUCE-HTN trial. *J. Hum. Hypertens.* 31, 366–368. doi:10.1038/jhh.2016.82

Simionescu, A., Philips, K., and Vyavahare, N. (2005). Elastin-derived peptides and TGF-beta1 induce osteogenic responses in smooth muscle cells. *Biochem. Biophys. Res. Commun.* 334, 524–532. doi:10.1016/j.bbrc.2005.06.119

Sinclair, A. M., Isles, C. G., Brown, I., Cameron, H., Murray, G. D., and Robertson, J. W. (1987). Secondary hypertension in a blood pressure clinic. *Arch. Intern. Med.* 147, 1289–1293. doi:10.1001/archinte.147.7.1289

Six, I., Flissi, N., Lenglet, G., Louvet, L., Kamel, S., Gallet, M., et al. (2020). Uremic toxins and vascular dysfunction. *Toxins* 12, 404. doi:10.3390/toxins12060404

Slatopolsky, E., Brown, A., and Dusso, A. (1999). Pathogenesis of secondary hyperparathyroidism. *Kidney Int. Suppl.* 56, S14–S19. doi:10.1046/j.1523-1755. 1999.07304.x

Smid, S. D., Frewin, D. B., Wyartt, C. L., and Head, R. J. (1995). Functional tolerance to α-adrenergic receptor blockade in the spontaneously hypertensive rat highlights the multifunctional role of vascular angiotensin II in the development of hypertension. J. Vasc. Res. 32, 247–253. doi:10.1159/000159099

Smith, W. L., Dewitt, D. L., and Garavito, R. M. (2000). Cyclooxygenases: Structural, cellular, and molecular biology. *Annu. Rev. Biochem.* 69, 145–182. doi:10.1146/annurev.biochem.69.1.145

Smyth, E. M., Grosser, T., Wang, M., Yu, Y., and Fitzgerald, G. A. (2009). Prostanoids in health and disease. J. Lipid Res. 50, S423–S428. doi:10.1194/jlr.R800094-JLR200

Sobey, C. G. (2001). Potassium channel function in vascular disease. Arterioscler. Thromb. Vasc. Biol. 21, 28–38. doi:10.1161/01.atv.21.1.28

Stauss, H. M. (2007). Identification of blood pressure control mechanisms by power spectral analysis. *Clin. Exp. Pharmacol. Physiol.* 34, 362–368. doi:10.1111/j. 1440-1681.2007.04588.x

Stehouwer, C. D., and Smulders, Y. M. (2006). Microalbuminuria and risk for cardiovascular disease: Analysis of potential mechanisms. J. Am. Soc. Nephrol. 17, 2106–2111. doi:10.1681/ASN.2005121288

Studinger, P., Lenard, Z., Mersich, B., Reusz, G., and Kollai, M. (2006). Determinants of baroreflex function in juvenile end-stage renal disease. *Kidney Int.* 69, 2236–2242. doi:10.1038/sj.ki.5000307

Sun, C., Zubcevic, J., Polson, J. W., Potts, J. T., Diez-Freire, C., Zhang, Q., et al. (2009). Shift to an involvement of phosphatidylinositol 3-kinase in angiotensin II actions on nucleus tractus solitarii neurons of the spontaneously hypertensive rat. *Circ. Res.* 105, 1248–1255. doi:10.1161/CIRCRESAHA.109.208926

Sun, Y., Zhang, J., Lu, L., Chen, S. S., Quinn, M. T., and Weber, K. T. (2002). Aldosterone-induced inflammation in the rat heart: Role of oxidative stress. *Am. J. Pathol.* 161, 1773–1781. doi:10.1016/S0002-9440(10)64454-9

Takahashi, H., Yoshika, M., Komiyama, Y., and Nishimura, M. (2011). The central mechanism underlying hypertension: A review of the roles of sodium ions, epithelial sodium channels, the renin–angiotensin–aldosterone system, oxidative stress and endogenous digitalis in the brain. *Hypertens. Res.* 34, 1147–1160. doi:10.1038/hr.2011.105

Tang, E. H., Jensen, B. L., Skott, O., Leung, G. P., Feletou, M., Man, R. Y., et al. (2008). The role of prostaglandin E and thromboxane-prostanoid receptors in the response to prostaglandin E2 in the aorta of Wistar Kyoto rats and spontaneously hypertensive rats. *Cardiovasc. Res.* 78, 130–138. doi:10.1093/cvr/cvm112

Thambyrajah, J., Landray, M. J., Mcglynn, F. J., Jones, H. J., Wheeler, D. C., and Townend, J. N. (2000). Abnormalities of endothelial function in patients with predialysis renal failure. *Heart* 83, 205–209. doi:10.1136/heart.83.2.205

Tinucci, T., Abrahao, S., Santello, J., and Mion, D. (2001). Mild chronic renal insufficiency induces sympathetic overactivity. *J. Hum. Hypertens.* 15, 401–406. doi:10.1038/sj.jhh.1001149

Tipton, A. J., Baban, B., and Sullivan, J. C. (2012). Female spontaneously hypertensive rats have greater renal anti-inflammatory T lymphocyte infiltration than males. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* 303, R359–R367. doi:10. 1152/ajpregu.00246.2012

Touyz, R., Yao, G., Quinn, M., Pagano, P., and Schiffrin, E. (2005). p47phox associates with the cytoskeleton through cortactin in human vascular smooth muscle cells role in NAD (P) H oxidase regulation by angiotensin II. *Arterioscler. Thromb. Vasc. Biol.* 25, 512–518. doi:10.1161/01.ATV.0000154141.66879.98

Townsend, R. R., and Taler, S. J. (2015). Management of hypertension in chronic kidney disease. Nat. Rev. Nephrol. 11, 555–563. doi:10.1038/nrneph.2015.114

Tozawa, M., Iseki, K., Iseki, C., Kinjo, K., Ikemiya, Y., and Takishita, S. (2003). Blood pressure predicts risk of developing end-stage renal disease in men and women. *Hypertension* 41, 1341–1345. doi:10.1161/01.HYP. 0000069699.92349.8C

Trachtman, H., Nelson, P., Adler, S., Campbell, K. N., Chaudhuri, A., Derebail, V. K., et al. (2018). Duet: A phase 2 study evaluating the efficacy and safety of sparsentan in patients with FSGS. *J. Am. Soc. Nephrol.* 29, 2745–2754. doi:10.1681/ASN.2018010091

Unger, T., Borghi, C., Charchar, F., Khan, N. A., Poulter, N. R., Prabhakaran, D., et al. (2020). 2020 International Society of Hypertension global hypertension practice guidelines. *Hypertension* 75, 1334–1357. doi:10.1161/HYPERTENSIONAHA.120.15026

Urschel, K., Cicha, I., Daniel, W. G., and Garlichs, C. D. (2012). Shear stress patterns affect the secreted chemokine profile in endothelial cells. *Clin. Hemorheol. Microcirc.* 50, 143–152. doi:10.3233/CH-2011-1450

Vallance, P., and Leiper, J. (2004). Cardiovascular biology of the asymmetric dimethylarginine: Dimethylarginine dimethylaminohydrolase pathway. *Arterioscler. Thromb. Vasc. Biol.* 24, 1023–1030. doi:10.1161/01.ATV.0000128897.54893.26

Van Der Mark, J., and Kline, R. L. (1994). Altered pressure natriuresis in chronic angiotensin II hypertension in rats. *Am. J. Physiol.* 266, R739–R748. doi:10.1152/ajpregu.1994.266.3.R739

Van Varik, B., Rennenberg, R., Reutelingsperger, C., Kroon, A., De Leeuw, P., and Schurgers, L. J. (2012). Mechanisms of arterial remodeling: Lessons from genetic diseases. *Front. Genet.* 3, 290. doi:10.3389/fgene.2012.00290

Vanhoutte, P. M., Zhao, Y., Xu, A., and Leung, S. W. (2016). Thirty years of saying NO: Sources, fate, actions, and misfortunes of the endotheliumderived vasodilator mediator. *Circ. Res.* 119, 375-396. doi:10.1161/ CIRCRESAHA.116.306531

Varin, R., Mulder, P., Tamion, F., Richard, V., Henry, J.-P., Lallemand, F., et al. (2000). Improvement of endothelial function by chronic angiotensin-converting enzyme inhibition in heart failure role of nitric oxide, prostanoids, oxidant stress, and bradykinin. *Circulation* 102, 351–356. doi:10.1161/01.cir.102.3.351

Vaziri, N. D. (1999). Mechanism of erythropoietin-induced hypertension. Am. J. Kidney Dis. 33, 821-828. doi:10.1016/s0272-6386(99)70413-0

Vaziri, N. D., Ni, Z., Oveisi, F., Liang, K., and Pandian, R. (2002). Enhanced nitric oxide inactivation and protein nitration by reactive oxygen species in renal insufficiency. *Hypertension* 39, 135–141. doi:10.1161/hy0102.100540

Vaziri, N. D., Oveisi, F., and Ding, Y. (1998). Role of increased oxygen free radical activity in the pathogenesis of uremic hypertension. *Kidney Int.* 53, 1748–1754. doi:10.1046/j.1523-1755.1998.00947.x

Vaziri, N. D. (2010). Role of oxidative stress in the pathogenesis of erythropoietin-induced hypertension. *Am. J. Hypertens.* 23, 226-227. doi:10.1038/ajh.2009.259

Vaziri, N. D., and Zhou, X. J. (2009). Potential mechanisms of adverse outcomes in trials of anemia correction with erythropoietin in chronic kidney disease. *Nephrol. Dial. Transpl.* 24, 1082–1088. doi:10.1093/ndt/gfn601

Veerasingham, S. J., and Raizada, M. K. (2003). Brain renin-angiotensin system dysfunction in hypertension: Recent advances and perspectives. *Br. J. Pharmacol.* 139, 191–202. doi:10.1038/sj.bjp.0705262

Vertes, V., Cangiano, J. L., Berman, L. B., and Gould, A. (1969). Hypertension in endstage renal disease. N. Engl. J. Med. 280, 978–981. doi:10.1056/NEJM196905012801802

Vettoretti, S., Ochodnicky, P., Buikema, H., Henning, R. H., Kluppel, C. A., De Zeeuw, D., et al. (2006). Altered myogenic constriction and endothelium-derived hyperpolarizing factor-mediated relaxation in small mesenteric arteries of hypertensive subtotally nephrectomized rats. *J. Hypertens.* 24, 2215–2223. doi:10.1097/01.hjlh.0000249699.04113.36

Vink, E. E., De Jager, R. L., and Blankestijn, P. J. (2013). Sympathetic hyperactivity in chronic kidney disease: Pathophysiology and (new) treatment options. *Curr. Hypertens. Rep.* 15, 95–101. doi:10.1007/s11906-013-0328-5

Volpe, M., and Patrono, C. (2020). Aldosterone receptor antagonism in patients with diabetes and chronic kidney disease: New promises and old problems. *Eur. Heart J.* 42, 14–15. doi:10.1093/eurheartj/ehaa992

Wagenaar, G. T., Laghmani El, H., Fidder, M., Sengers, R. M., De Visser, Y. P., De Vries, L., et al. (2013). Agonists of MAS oncogene and angiotensin II type 2 receptors attenuate cardiopulmonary disease in rats with neonatal hyperoxia-induced lung injury. *Am. J. Physiol. Lung Cell. Mol. Physiol.* 305, L341–L351. doi:10.1152/ajplung.00360.2012

Wallbach, M., Lehnig, L.-Y., Schroer, C., Hasenfuss, G., Müller, G. A., Wachter, R., et al. (2014). Impact of baroreflex activation therapy on renal function-a pilot study. *Am. J. Nephrol.* 40, 371–380. doi:10.1159/000368723

Wallbach, M., Zürbig, P., Dihazi, H., Müller, G. A., Wachter, R., Beige, J., et al. (2018). Kidney protective effects of baroreflex activation therapy in patients with resistant hypertension. *J. Clin. Hypertens.* 20, 1519–1526. doi:10.1111/jch.13365

Wang, D., Iversen, J., and Strandgaard, S. (2000). Endothelium-dependent relaxation of small resistance vessels is impaired in patients with autosomal dominant polycystic kidney disease. J. Am. Soc. Nephrol. 11, 1371–1376. doi:10.1681/ASN.V1181371

Wang, L., Velazquez, H., Moeckel, G., Chang, J., Ham, A., Lee, H. T., et al. (2014). Renalase prevents AKI independent of amine oxidase activity. *J. Am. Soc. Nephrol.* 25, 1226–1235. doi:10.1681/ASN.2013060665

Wang, Y., Han, L., Shen, M., Jones, E. S., Spizzo, I., Walton, S. L., et al. (2020). Serelaxin and the AT(2) receptor agonist CGP42112 evoked a similar, nonadditive, cardiac antifibrotic effect in high salt-fed mice that were refractory to candesartan cilexetil. ACS Pharmacol. Transl. Sci. 3, 76–87. doi:10.1021/acsptsci.9b00095

Wang, Y., Safirstein, R., Velazquez, H., Guo, X. J., Hollander, L., Chang, J., et al. (2017). Extracellular renalase protects cells and organs by outside-in signalling. *J. Cell. Mol. Med.* 21, 1260–1265. doi:10.1111/jcmm.13062

Wanner, C., Inzucchi, S. E., Lachin, J. M., Fitchett, D., Von Eynatten, M., Mattheus, M., et al. (2016). Empagliflozin and progression of kidney disease in type 2 diabetes. *N. Engl. J. Med.* 375, 323–334. doi:10.1056/NEJMoa1515920

Wesson, D. E. (2006). Endothelin role in kidney acidification. Semin. Nephrol. 26, 393–398. doi:10.1016/j.semnephrol.2006.07.006()

Wever, R., Boer, P., Hijmering, M., Stroes, E., Verhaar, M., Kastelein, J., et al. (1999). Nitric oxide production is reduced in patients with chronic renal failure. *Arterioscler. Thromb. Vasc. Biol.* 19, 1168–1172. doi:10.1161/01.atv.19.5.1168

Wolf, G. (1998). Link between angiotensin II and TGF- $\beta$  in the kidney. Min. Electrolyte Metab. 24, 174–180. doi:10.1159/000057367

Wong, H., Mylrea, K., Feber, J., Drukker, A., and Filler, G. (2006). Prevalence of complications in children with chronic kidney disease according to KDOQI. *Kidney Int.* 70, 585–590. doi:10.1038/sj.ki.5001608

Wong, M. S., and Vanhoutte, P. M. (2010). COX-Mediated endotheliumdependent contractions: From the past to recent discoveries. *Acta Pharmacol. Sin.* 31, 1095–1102. doi:10.1038/aps.2010.127

Wu, J., Fang, S., Lu, K.-T., Wackman, K., Schwartzman, M. L., Dikalov, S. I., et al. (2021). EP3 (E-Prostanoid 3) receptor mediates impaired vasodilation in a mouse model of salt-sensitive hypertension. *Hypertension* 77, 1399–1411. doi:10.1161/HYPERTENSIONAHA.120.16518

Wu, Y., Xu, J., Velazquez, H., Wang, P., Li, G., Liu, D., et al. (2010). Renalase deficiency aggravates ischemic myocardial damage. *Kidney Int.* 79, 853–860. doi:10. 1038/ki.2010.488

Xu, J., Li, G., Wang, P., Velazquez, H., Yao, X., Li, Y., et al. (2005). Renalase is a novel, soluble monoamine oxidase that regulates cardiac function and blood pressure. *J. Clin. Invest.* 115, 1275–1280. doi:10.1172/JCI24066

Yanagisawa, M., Kurihara, H., Kimura, S., Tomobe, Y., Kobayashi, M., Mitsui, Y., et al. (1988). A novel potent vasoconstrictor peptide produced by vascular endothelial cells. *nature* 332, 411–415. doi:10.1038/332411a0

Yao, Y., Hildreth, C. M., Farnham, M. M., Saha, M., Sun, Q.-J., Pilowsky, P. M., et al. (2015). The effect of losartan on differential reflex control of sympathetic nerve activity in chronic kidney disease. *J. Hypertens.* 33, 1249–1260. doi:10.1097/HJH. 00000000000535

Yao, Y., Hildreth, C. M., Li, S., Boyd, R., Kouchaki, Z., Butlin, M., et al. "Increased arterial stiffness does not respond to renal denervation in an animal model of secondary hypertension," in 2017 39th Annual International Conference of the IEEE Engineering in Medicine and Biology Society (EMBC IEEE), 258–261.

Ye, S., Gamburd, M., Mozayeni, P., Koss, M., and Campese, V. M. (1998). A limited renal injury may cause a permanent form of neurogenic hypertension. *Am. J. Hypertens.* 11, 723–728. doi:10.1016/s0895-7061(98)00030-2

Ye, S., Zhong, H., Duong, V. N., and Campese, V. M. (2002). Losartan reduces central and peripheral sympathetic nerve activity in a rat model of neurogenic hypertension. *Hypertension* 39, 1101–1106. doi:10.1161/01.hyp.0000018590.26853.c7

Yilmaz, M. I., Saglam, M., Caglar, K., Cakir, E., Sonmez, A., Ozgurtas, T., et al. (2006). The determinants of endothelial dysfunction in CKD: Oxidative stress and asymmetric dimethylarginine. *Am. J. Kidney Dis.* 47, 42–50. doi:10.1053/j.ajkd. 2005.09.029

Zewde, T., and Mattson, D. L. (2004). Inhibition of cyclooxygenase-2 in the rat renal medulla leads to sodium-sensitive hypertension. *Hypertension* 44, 424–428. doi:10.1161/01.HYP.0000140924.91479.03

Zhang, Z., Wang, M., Xue, S.-J., Liu, D.-H., and Tang, Y.-B. (2012). Simvastatin ameliorates angiotensin II-induced endothelial dysfunction through restoration of Rho-BH4-eNOS-NO pathway. *Cardiovasc. Drugs Ther.* 26, 31–40. doi:10.1007/s10557-011-6351-3

Zhao, S., Cheng, C. K., Zhang, C.-L., and Huang, Y. (2021). Interplay between oxidative stress, cyclooxygenases, and prostanoids in cardiovascular diseases. *Antioxid. Redox Signal.* 34, 784–799. doi:10.1089/ars.2020.8105

Zoccali, C., Bode-Böger, S. M., Mallamaci, F., Benedetto, F. A., Tripepi, G., Malatino, L. S., et al. (2001). Plasma concentration of asymmetrical dimethylarginine and mortality in patients with end-stage renal disease: A prospective study. *Lancet* 358, 2113–2117. doi:10.1016/s0140-6736(01)07217-8

Zoccali, C., Mallamaci, F., Parlongo, S., Cutrupi, S., Benedetto, F. A., Tripepi, G., et al. (2002). Plasma norepinephrine predicts survival and incident cardiovascular events in patients with end-stage renal disease. *Circulation* 105, 1354–1359. doi:10. 1161/hc1102.105261

Zoccali, C., Mallamaci, F., and Tripepi, G. (2003). Traditional and emerging cardiovascular risk factors in end-stage renal disease. *Kidney Int.* 63, S105–S110. doi:10.1046/j.1523-1755.63.s85.25.x

# Glossary

CKD Chronic kidney disease ESRD end-stage renal disease CV cardiovascular CVD cardiovascular disease BP blood pressure SNS sympathetic nervous system RAAS renin-angiotensin-aldosterone system RAS renin-angiotensin system GFR glomerular filtration rate PKD polycystic kidney disease BAT baroreflex activation therapy mmHg millimeter mercury SPRINT Systolic BP Intervention Trial AT1 angiotensin-1 receptor AT2 angiotensin-2 receptor ET1 endothelin-1 receptors Ang II angiotensin II SOD superoxide dismutase NADPH nicotine amide dinucleotide phosphate ADMA asymmetric dimethylarginine SDMA symmetric dimethylarginine NO nitric oxide ROS reactive oxygen species LVH left ventricular hypertrophy O2-- superoxide free radical VSMC vascular smooth muscle cell NSAIDs non-steroidal anti-inflammatory drugs PGI2 prostacyclin I2 COX cyclooxygenase NE norepinephrine SNA sympathetic nerve activity LPK Lewis polycystic kidney disease SHR spontaneously hypertensive rat Hz hertz PWV pulse wave velocity NF-kB nuclear factor κ-light-chain-enhancer of activated B cells VCAM-1 vascular adhesion molecule-1

ACEi angiotensin-converting enzyme inhibitor MAP mean arterial pressure ARB AT1 receptor antagonist/blocker EDHF endothelium-derived hyperpolarization factor ICAM-1 intracellular adhesion molecule-1 EDRFs endothelium-derived relaxing factor EDCFs endothelium-derived constricting factors ACh acetylcholine M3 muscarinic 3 receptors Ca2+ calcium eNOS endothelial nitric oxide synthase cGMP cyclic guanosine monophosphate 1-NAME Nw-nitro-l-arginine methyl ester AA arachidonic acid TxA2 thromboxane A2 ACE Angiotensin converting enzyme ERA endothelin receptor antagonist LDL low-density lipoprotein **TGF-\beta1** transforming growth factor  $\beta$ 1 BMPs bone morphogenetic proteins NOS nitric oxide synthase L-Arg l-arginine nNOS neuronal nitric oxide synthase iNOS inducible nitric oxide synthase ONOO- peroxynitrite AGE advanced glycation end products BH4 tetrahydrobiopterin CAT y+ cationic transporter DDAH dimethylarginine dimethylaminohydrolase PG prostaglandin DOCA deoxycorticosterone acetate NOARG nitrogen oxide synthase inhibitor Nw-nitro-l-arginine SGLT2i sodium-glucose cotransporter-2 inhibitor PTH parathyroid hormone FGF-23 fibroblast growth factor-23 ENaC epithelial sodium channels Na-K-ATPase sodium-potassium atpase transporter Tregs T regulatory cells