

# Draft Genome Sequence of *Clostridium botulinum* B2 450 Strain from Wound Botulism in a Drug User in Italy

Silvia Fillo,<sup>a</sup> Francesco Giordani,<sup>a</sup> Anna Anselmo,<sup>a</sup> Antonella Fortunato,<sup>a</sup> Anna Maria Palozzi,<sup>a</sup> Riccardo De Santis,<sup>a</sup> Andrea Ciammaruconi,<sup>a</sup> Ferdinando Spagnolo,<sup>a</sup> Fabrizio Anniballi,<sup>b</sup> Alfonsina Fiore,<sup>b</sup> Bruna Auricchio,<sup>b</sup> Dario De Medici,<sup>b</sup> Florigio Lista<sup>a</sup>

Histology and Molecular Biology Section, Army Medical and Veterinary Research Center, Rome, Italy<sup>a</sup>; National Reference Center for Botulism, Department of Veterinary Public Health and Food Safety, Istituto Superiore di Sanità (ISS), Rome, Italy<sup>b</sup>

**Here, we report the draft genome sequence of *Clostridium botulinum* B2 450, responsible for the first reported case of wound botulism in a drug user in Italy.**

Received 20 February 2015 Accepted 23 February 2015 Published 2 April 2015

**Citation** Fillo S, Giordani F, Anselmo A, Fortunato A, Palozzi AM, De Santis R, Ciammaruconi A, Spagnolo F, Anniballi F, Fiore A, Auricchio B, De Medici D, Lista F. 2015. Draft genome sequence of *Clostridium botulinum* B2 450 strain from wound botulism in a drug user in Italy. *Genome Announc* 3(2):e00238-15. doi:10.1128/genomeA.00238-15.

**Copyright** © 2015 Fillo et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 3.0 Unported license](https://creativecommons.org/licenses/by/3.0/).

Address correspondence to Florigio Lista, romano.lista@gmail.com.

*Clostridium botulinum* is a microorganism able to produce the botulinum neurotoxin (BoNT), a powerful poison that causes botulism, a serious neuroparalytic disease. There are 3 mainly clinical manifestations of botulism: foodborne (ingestion of BoNT-contaminated foods), intestinal (BoNT is produced by intestinal colonization of *C. botulinum*), and wound (*C. botulinum* spores germinate and synthesize BoNT in contaminated wounds) (1). Based on its physiological characteristics, *C. botulinum* is divided into four different groups (I to IV), so phylogenetically different that they can be considered separate species (2, 3). Moreover, other species, such as *Clostridium baratii* and *Clostridium butyricum*, are able to produce botulinum neurotoxins type F and E, respectively. The BoNT is classified by 8 serotypes (A to H), each of which is divided into subtypes. To date, 16 full genomes and several draft assemblies of *C. botulinum* are available, prevalently isolated from food borne and infant cases. A5(B3')H04402 065 (accession no. FR773526) is the only full genome sequence originating from a wound botulism case (4, 5).

The B2 450 strain was isolated in 2009 from wound exudate of a heroin user patient, in Messina, Sicily, Italy (6). Sequencing was performed using both Roche 454 GS FLX Titanium and Illumina MiSeq platforms. Roche sequencing generated 286,392 single-end reads, with 112,174,234 total sequenced bases, ~28-fold coverage. The reads were *de novo* assembled with the GS Assembler software (Newbler package) into 160 contigs, and 2,430,881 paired-end reads were produced by Illumina sequencing with 731,610,908 total sequenced bases and ~185-fold coverage. Illumina reads were assembled with Abyss-pe producing 497 contigs. 454 and Miseq contigs were combined using Minimus2 software, and 18 contigs were obtained.

A well-known flaw of the 454 platform is the erroneous determination of homopolymer lengths (7); 1,218 homopolymeric stretches were corrected according to Illumina sequences. Nine gaps were closed using 454 and Miseq reads or Sanger sequencing.

The final draft assembly consists of 9 chromosomal contigs, for a total length of 4,070,655 bp, and one plasmidic contig, 250,014

bp long, containing the BoNT/B2 gene. The genome has a G+C content of 27.8%. The 8 remaining gaps are due to repeated sequences (rRNA operon and beta-N-acetyl-glucosamidase genes).

The 16S rRNA gene sequence of B2 450 belongs to group I showing 99.8% similarity with the A1 ATCC 3502 16S sequence (NC\_009495) (3). Comparing some gene sequences (rpoB-mdh-aroE-hsp60-aceK-oppB-recA) (8), the B2 450 strain appears phylogenetically closer to *Clostridium sporogenes*, with 99.99% similarity to ATCC 15579 (ABKW0200000), than to all other *C. botulinum* strains (96.14% to A1 ATCC 3502 and 95.79% to A3 Loch Maree; NC\_010520). *C. sporogenes* phylogenetically belongs to group I (9). The 450 strain may represent a *C. sporogenes* lineage that has recently acquired the BoNT/B gene (perhaps through the plasmid) (10).

**Nucleotide sequence accession number.** The genome sequence of *C. botulinum* B2 450 was deposited at DDBJ/EMBL/GenBank under the accession no. [JXSU000000000](https://www.ncbi.nlm.nih.gov/nuccore/JXSU000000000). The version described in this paper is the first version.

## ACKNOWLEDGMENT

This work was supported by Italian Ministry of Defense, SEGREDIFESA/DNA-5 Department of Technological Innovation (EBLN project).

## REFERENCES

- Sobel J. 2005. Botulism. *Clin Infect Dis* 41:1167–1173. <http://dx.doi.org/10.1086/444507>.
- Collins MD, East AK. 1998. Phylogeny and taxonomy of the foodborne pathogen *Clostridium botulinum* and its neurotoxins. *J Appl Microbiol* 84:5–17. <http://dx.doi.org/10.1046/j.1365-2672.1997.00313.x>.
- Hill KK, Smith TJ, Helma CH, Ticknor LO, Foley BT, Svensson RT, Brown JL, Johnson EA, Smith LA, Okinaka RT, Jackson PJ, Marks JD. 2007. Genetic diversity among botulinum neurotoxin-producing clostridial strains. *J Bacteriol* 189:818–832. <http://dx.doi.org/10.1128/JB.01180-06>.
- Peck MW, Stringer SC, Carter AT. 2011. *Clostridium botulinum* in the post-genomic era. *Food Microbiol* 28:183–191. <http://dx.doi.org/10.1016/j.fm.2010.03.005>.
- Carter AT, Pearson BM, Crossman LC, Drou N, Heavens D, Baker D,

- Febrer M, Caccamo M, Grant KA, Peck MW. 2011. Complete genome sequence of the proteolytic *Clostridium botulinum* type A5 (B3') strain H04402 065. *J Bacteriol* 193:2351–2352. <http://dx.doi.org/10.1128/JB.00072-11>.
6. Rodolico C, Barca E, Fenicia L, Anniballi F, Sinardi AU, Girlanda P. 2010. Wound botulism in drug users: a still underestimated diagnosis. *Neurol Sci* 31:825–827. <http://dx.doi.org/10.1007/s10072-010-0350-1>.
7. Balzer S, Malde K, Jonassen I. 2011. Systematic exploration of error sources in pyrosequencing flowgram data. *Bioinformatics* 27:i304–i309. <http://dx.doi.org/10.1093/bioinformatics/btr251>.
8. Jacobson MJ, Lin G, Whittam TS, Johnson EA. 2008. Phylogenetic analysis of *Clostridium botulinum* type A by multi-locus sequence typing. *Microbiology* 154:2408–2415. <http://dx.doi.org/10.1099/mic.0.2008/016915-0>.
9. Olsen JS, Scholz H, Fillo S, Ramisse V, Lista F, Trømborg AK, Aarskaug T, Thrane I, Blatny JM. 2014. Analysis of the genetic distribution among members of *Clostridium botulinum* group I using a novel multilocus sequence typing (MLST) assay. *J Microbiol Methods* 96:84–91. <http://dx.doi.org/10.1016/j.mimet.2013.11.003>.
10. Smith TJ, Hill KK, Foley BT, Detter JC, Munk AC, Bruce DC, Doggett NA, Smith LA, Marks JD, Xie G, Brettin TS. 2007. Analysis of the neurotoxin complex genes in *Clostridium botulinum* A1-A4 and B1 strains: BoNT/A3, /Ba4 and /B1 clusters are located within plasmids. *PLoS One* 2:e1271. <http://dx.doi.org/10.1371/journal.pone.0001271>.