





# Complete Genome Sequence of *Aeromonas hydrophila* Bacteriophage BUCT552

Rujia Chen,<sup>a</sup> Xuling Xu,<sup>c</sup> Pengjun Han,<sup>a</sup> Yunjia Hu,<sup>a</sup> Hongbo Qin,<sup>a</sup> Mengzhe Li,<sup>a</sup> Xiaoping An,<sup>a</sup>  Lihua Song,<sup>a</sup>  Huahao Fan,<sup>a</sup> Yigang Tong<sup>a,b</sup>

<sup>a</sup>College of Life Science and Technology, Beijing University of Chemical Technology, Beijing, China

<sup>b</sup>Beijing Advanced Innovation Center for Soft Matter Science and Engineering (BAIC-SM), Beijing University of Chemical Technology, Beijing, China

<sup>c</sup>Phagelux (Nanjing) Bio-tech Co., Ltd., Nanjing, China

**ABSTRACT** We report the complete genome sequence of *Aeromonas hydrophila* bacteriophage BUCT552 whose full length of the linear dsDNA genome is 59,685 bp and G+C content is 60.0%. It contains 74 open reading frames but no tRNA. The results of TEM showed BUCT552 is a member of the family *Siphoviridae*.

**A** *Aeromonas hydrophila* belongs to *Vibrionaceae*, a family of Gram-negative bacteria (1, 2) and can be found in different aquatic environments, foods, and food processing systems (3). It is an emerging bacterial pathogen, and not only a thorny problem in the aquaculture industry, but also a severe threat to human life and health (4). Bacteriophages, as a very potential antibiotic substitute, have shown good application prospects in the prevention and treatment of human and animal diseases (5–8).

In this research, *A. hydrophila* bacteriophage BUCT552 was isolated from the sewage of Zhongcai Aquatic Products Wholesale Market in Nanjing City, Jiangsu Province, China (118.7674° E, 32.0415° N) in August 2020. *A. hydrophila* host strains were cultivated in tryptic soy broth (TSB) at 29°C, 200 r/min for 8 h. Individual phages were isolated using the enrichment method, then purified with three rounds of plaque assays on double-layer tryptic soy agar (TSA) plates at 29°C overnight, obtaining clear single plaques. The phage morphology was observed by uranyl acetate negative-stain method (9). Phage genomic DNA was extracted using the phenol-chloroform method (10). The Illumina MiSeq sequencer was used to sequence the whole genome of the phage following the NEBNext Ultra II DNA library prep kit for Illumina (NEB, E7805) to construct paired-end 2 × 150-bp reads sequencing library. The genome sequence was assembled using SPAdes v3.13.0 (11). RAST (<http://rast.nmpdr.org/>) online annotation was used to quickly annotate the whole genome, and then the online NCBI BLASTp tool (<http://www.ncbi.nlm.nih.gov/BLAST>) was used to confirm RAST annotations (12). Then, tRNAscan-SE v2.0 was used (<http://lowelab.ucsc.edu/cgi-bin/tRNAscan-SE2.cgi>) (13) to predict tRNA in the genome. All tools were run with default parameters unless otherwise specified.

Transmission electron micrograph of BUCT552 showed that the capsid was a regular icosahedral body with a diameter of 55 ± 3 nm and a tail length of 200 ± 3 nm (Fig. 1). According to traditional morphology-based classification, BUCT552 belongs to the family *Siphoviridae*. The genome phage BUCT552 assembled as a circular contig of length 59,685 bp (repeats are typically omitted when reporting the length of circular contigs), and the GC content is 60.0%. The average coverage depth of the genome is 190.0. The results of RAST gene annotation showed a total of 74 open reading frames (ORF), and no tRNA was predicted in the genome. There are 24 predicted ORFs which encode structure, packaging, replication, modification, and lysis genes, thus the gene density is 92.5%. Its genome was compared with other known phages in BLASTn, showing that the three closest phages are *Aeromonas* phage LAH\_7, vB\_AhyS-A18P4, BUCT551, and

**Editor** Simon Roux, DOE Joint Genome Institute

**Copyright** © 2022 Chen et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

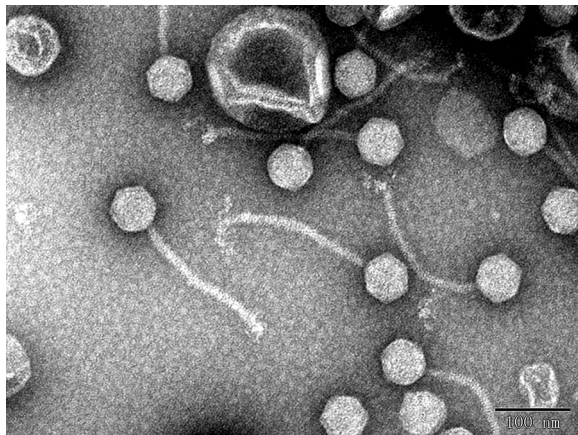
Address correspondence to Lihua Song, [songjihua@gmail.com](mailto:songjihua@gmail.com), Huahao Fan, [fanhuahao@mail.buct.edu.cn](mailto:fanhuahao@mail.buct.edu.cn), or Yigang Tong, [tong.yigang@gmail.com](mailto:tong.yigang@gmail.com).

The authors declare no conflict of interest.

**Received** 3 December 2021

**Accepted** 27 January 2022

**Published** 17 February 2022



**FIG. 1** The morphology of phage BUCT552 was examined with a transmission electron microscope (JEM-1200EX, Japan) at 80 kV. Four virions were measured in the figure.

the query cover was 3%, 8%, and 3% with the corresponding identity 75.59%, 76.30%, and 75.22%, respectively.

In this study, a lytic phage BUCT552 which can infect and lyse *A. hydrophila* was isolated from sewage. The phage displays a maximum of 8% similarity with known phages, and it enriches our understanding about the diversity of *A. hydrophila* phages.

**Data availability.** The accession number of whole genome sequence of phage BUCT552 in the GenBank database is [MW978786](https://www.ncbi.nlm.nih.gov/nuclseq/MW978786). The accession number of the raw reads in NCBI SRA is [SRR17399946](https://www.ncbi.nlm.nih.gov/sra/SRR17399946).

## ACKNOWLEDGMENTS

This research was supported by Funds for First-class Discipline Construction (No. XK1805, No. XK1803-06), National Key Research and Development Program of China (2020YFA0712100, 2020YFC0840805, 19SWAQ06, 2018YFA0903000, 2020YFC2005405), Inner Mongolia Key Research and Development Program (No. 2019ZD006), NSFC-MFST project (China-Mongolia) (No. 31961143024), and Fundamental Research Funds for Central Universities (No. BUCTRC201917, BUCTZY2022).

## REFERENCES

- Danaher PJ, Mueller WP. 2011. *Aeromonas hydrophila* septic arthritis. *Mil Med* 176:1444–1446. <https://doi.org/10.7205/milmed-d-11-00211>.
- Awan F, Dong Y, Wang N, Liu J, Ma K, Liu Y. 2018. The fight for invincibility: Environmental stress response mechanisms and *Aeromonas hydrophila*. *Microb Pathog* 116:135–145. <https://doi.org/10.1016/j.micpath.2018.01.023>.
- Stratev D, Odeyemi OA. 2016. Antimicrobial resistance of *Aeromonas hydrophila* isolated from different food sources: a mini-review. *J Infect Public Health* 9:535–544. <https://doi.org/10.1016/j.jiph.2015.10.006>.
- Janda JM, Abbott SL. 2010. The genus *Aeromonas*: taxonomy, pathogenicity, and infection. *Clin Microbiol Rev* 23:35–73. <https://doi.org/10.1128/CMR.00039-09>.
- Dedrick RM, Guerrero-Bustamante CA, Garlena RA, Russell DA, Ford K, Harris K, Gilmour KC, Soothill J, Jacobs-Sera D, Schooley RT, Hatfull GF, Spencer H. 2019. Engineered bacteriophages for treatment of a patient with a disseminated drug-resistant *Mycobacterium abscessus*. *Nat Med* 25:730–733. <https://doi.org/10.1038/s41591-019-0437-z>.
- Le TS, Nguyen TH, Vo HP, Doan VC, Nguyen HL, Tran MT, Tran TT, Southgate PC, Kurtböke Dİ. 2018. Protective effects of bacteriophages against *aeromonas hydrophila* species causing motile *aeromonas* septicemia (MAS) in striped catfish. *Antibiotics* 7:16. <https://doi.org/10.3390/antibiotics7010016>.
- Lim TH, Kim MS, Lee DH, Lee YN, Park JK, Youn HN, Lee HJ, Yang SY, Cho YW, Lee JB, Park SY, Choi IS, Song CS. 2012. Use of bacteriophage for biological control of *Salmonella* Enteritidis infection in chicken. *Res in Veterinary Science* 93:1173–1178. <https://doi.org/10.1016/j.rvsc.2012.06.004>.
- Międzybrodzki R, Borysowski J, Weber-Dąbrowska B, Fortuna W, Letkiewicz S, Szufnarowski K, Pawełczyk Z, Rogóż P, Klak M, Wojtasik E, Górski A. 2012. Clinical aspects of phage therapy. *Adv Virus Res* 83:73–121. <https://doi.org/10.1016/B978-0-12-394438-2.00003-7>.
- Furfaro LL, Payne MS, Chang BJ. 2020. Host range, morphological and genomic characterisation of bacteriophages with activity against clinical *Streptococcus agalactiae* isolates. *PLoS One* 15:e0235002. <https://doi.org/10.1371/journal.pone.0235002>.
- Lu S, Le S, Tan Y, Zhu J, Li M, Rao X, Zou L, Li S, Wang J, Jin X, Huang G, Zhang L, Zhao X, Hu F. 2013. Genomic and proteomic analyses of the terminally redundant genome of the *Pseudomonas aeruginosa* phage PaP1: establishment of genus PaP1-like phages. *PLoS One* 8:e62933. <https://doi.org/10.1371/journal.pone.0062933>.
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Pribelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J Comput Biol* 19:455–477. <https://doi.org/10.1089/cmb.2012.0021>.
- Overbeek R, Olson R, Pusch GD, Olsen GJ, Davis JJ, Disz T, Edwards RA, Gerdes S, Parrello B, Shukla M, Vonstein V, Wattam AR, Xia F, Stevens R. 2014. The SEED and the rapid annotation of microbial genomes using subsystems technology (RAST). *Nucleic Acids Res* 42:D206–214. <https://doi.org/10.1093/nar/gkt1226>.
- Lowe TM, Chan PP. 2016. tRNAscan-SE On-line: integrating search and context for analysis of transfer RNA genes. *Nucleic Acids Res* 44:W54–57. <https://doi.org/10.1093/nar/gkw413>.