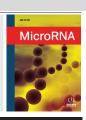
# **REVIEW ARTICLE**



Human Papillomavirus Infections, Cervical Cancer and MicroRNAs: An Overview and Implications for Public Health



Michela Lucia Sammarco<sup>1</sup>, Manuela Tamburro<sup>1</sup>, Alessandra Pulliero<sup>2</sup>, Alberto Izzotti<sup>2,3</sup> and Giancarlo Ripabelli<sup>1,\*</sup>

<sup>1</sup>Department of Medicine and Health Sciences "Vincenzo Tiberio", University of Molise, Campobasso, Italy; <sup>2</sup>Department of Health Sciences, University of Genoa, Genoa, Italy; <sup>3</sup>IRCCS Policlinico San Martino, Genoa, Italy

ARTICLE HISTORY

Received: June 14, 2019 Revised: August 21, 2019 Accepted: September 23, 2019

DOI: 10.2174/2211536608666191026115045



Abstract: Human Papillomavirus (HPV) is among the most common sexually transmitted infections in both females and males across the world that generally do not cause symptoms and are characterized by high rates of clearance. Persistent infections due at least to twelve well-recognized High-Risk (HR) or oncogenic genotypes, although less frequent, can occur, leading to diseases and malignancies, principally cervical cancer. Three vaccination strategies are currently available for preventing certain HR HPVs-associated diseases, infections due to HPV6 and HPV11 low-risk types, as well as for providing cross-protection against non-vaccine genotypes. Nevertheless, the limited vaccine coverage hampers reducing the burden of HPV-related diseases globally. For HR HPV types, especially HPV16 and HPV18, the E6 and E7 oncoproteins are needed for cancer development. As for other tumors, even in cervical cancer, non-coding microRNAs (miRNAs) are involved in posttranscriptional regulation, resulting in aberrant expression profiles. In this study, we provide a summary of the epidemiological background for HPV occurrence and available immunization programs. In addition, we present an overview of the most relevant evidence of miRNAs deregulation in cervical cancer, underlining that targeting these biomolecules could lead to wide translational perspectives, allowing better diagnosis, prognosis and therapeutics, and with valuable applications in the field of prevention. The literature on this topic is rapidly growing, but advanced investigations are required to achieve more consistent findings on the up-regulated and down-regulated miRNAs in cervical carcinogenesis. Because the expression of miRNAs is heterogeneously reported, it may be valuable to assess factors and risks related to individual susceptibility.

Keywords: E6 oncoprotein, E7 oncoprotein, immunization programs, miRNA, oncoviruses, tumor biomarkers.

# **1. INTRODUCTION**

Infectious agents are responsible for 20-25% of all cancer cases worldwide [1]. It has been estimated that nearly 15% of human cancers are associated with viral infections [2, 3], and in this context, a number of different viruses can contribute to several steps into carcinogenic processes. Among these viruses, Human Papillomavirus (HPV) has a prominent position, causing 30% of all infectious agent related cancers [4].

HPVs are extremely diverse at the level of genotype, epithelial tropism, and pathogenicity, being classified into cutaneous and mucosal groups [5]. HPVs are further divided into Low-Risk (LR) and High-Risk (HR) types, depending on Among more than 200 recognized HPV genotypes, HPV5 and HPV8 are the most commonly associated with cutaneous squamous cell carcinoma in patients with epidermodysplasia verruciformis [8]. The LR mucosal HPVs, especially HPV6 and HPV11, commonly cause genital warts, while the mucosal HR HPV16 and HPV18 are primarily involved in squamous intraepithelial lesions, which can progress to invasive squamous cell carcinoma [8, 9], even though different oncogenic types are becoming emergent [10, 11]. While HPVs have been associated with oral and numerous malignancies, including cancers of head and neck, anus, vulva, vagina, and penis, the infections are responsible for nearly all cervical cancers [7].

the lesion propensity for malignant progression [6, 7].

HPV outcomes could be avoided through the adoption of prophylactic measures, mainly with vaccination. The currently available immunization strategies have been shown to be effective in reducing genital warts, cervical disease, and

<sup>\*</sup>Address correspondence to this author at the Department of Medicine and Health Sciences "Vincenzo Tiberio", University of Molise, Campobasso, Italy; Tel: +39 0874 404961/743; Fax: +39 0874 404778; E-mail: ripab@unimol.it

anogenital dysplasia, but global vaccination rates are still under the required target level [12], including in Italy [13].

Understanding of the natural history of HPV infection and cancer progression requires a holistic epidemiological and clinical approach, taking into account the essential features related to cellular and molecular biology, transcriptional and post-transcriptional regulation. During malignancy progress, HPV genome integration into the host cell results in the primary viral E6 and E7 oncoproteins, whose overexpression contributes to cervical carcinoma development [6], inhibiting the functions of tumor suppressor p53, and promoting degradation of Retinoblastoma protein (pRb), respectively [8, 9].

In cervical cancer, as well as in many other tumors, short, non-coding single strands of RNAs, named as microRNAs (miRNAs), play a critical role, since their deregulation has been widely observed [14, 15]. Oncogenic viruses express miRNAs that may regulate their own gene expression, as well as influence the host's gene expression [16]. Contribution of miRNAs into the carcinogenic processes can be summarized as follows: miRNA genes can be located at susceptible sites in the genome/regions amplified or deleted in human cancers; since miRNAs are involved in the control of cell proliferation and apoptosis, their deregulation may contribute to proliferative diseases, including cancers; altered miRNA expression has been observed in malignant tumors and tumor cell lines compared to normal tissues [17, 18].

In this study, due to the continuous evolution of the epidemiological patterns related to these viruses, which are responsible of the most common sexually transmitted infections, and the high burden of the induced cancers, we reviewed the latest available evidences on HPV occurrence and immunization programs, tailored for the principal risk groups, and provided a description of the significant aspects concerning the miRNAs expression in cervical cancer in the current vaccination era.

## 2. HPV INFECTIONS AND CANCERS: EPIDEMIO-LOGICAL PICTURE AND TARGETED IMMUNIZA-TION STRATEGIES

HPVs, double-stranded DNA tumor viruses belonging to the *Papillomaviridae* family, represent the most frequent viral agents involved in infectious agents-associated oncogenesis [16], with peak prevalence in early adulthood [19]. Approximately, 280 papillomavirus types are described in vertebrates, and over 200 types can infect humans [20]. While infections usually do not cause symptoms, resolve spontaneously [21] and are generally cleared by the immune system within 1-2 years of onset, 10-20% can latently persist, leading to progression towards precancerous lesions, as well as to various forms of invasive cancer [22]. Furthermore, the persistence of HR HPV infections represents the main risk factor for cancer development [19].

Indeed, it is well known that all HPVs, mainly LR HPV6 and HPV11 genotypes, can induce proliferative benign lesions at the site of infection, such as papillomas or warts [23]. However, at least 12 viral types, HPV16, 18, 31, 33, 35, 39, 45, 51, 52, 56, 58, and 59, are highly related with malignant neoplasms [7, 19], and are classified as HR HPVs into Group 1 carcinogenic agents by the International Agency for Research on Cancer [5, 24]. Additional 13 HPVs were classified as either probably, or possibly, carcinogenic (Group 2A and B) based on the limited evidence and/or their close phylogenetic placement with other carcinogenic HPVs [5].

HR HPVs are not only responsible for almost all cases of cervical cancer, but also for a substantial proportion of vulvar, vaginal, penile, anal, oropharyngeal, and cutaneous squamous carcinomas [25]. Current epidemiological data confirm that HPV16 represents the most recurrent HR HPV identified in cervical cancers, but also is the most common in all HPV-related cancer sites, followed by HPV18 [7, 26, 27], which together approximately account for more than 70% of all cases of squamous cell carcinoma [28].

To date, HPV remains the most common sexually transmitted infection, and despite the availability of prevention strategies, the associated cancers represent one of the foremost causes of morbidity and mortality globally [29]. Epidemiological data regarding the global disease burden underlines that HPV is responsible of 630,000 (4.5%) new cancer cases worldwide per year: 570,000 (8.6%) cases per year in women, and 60,000 (0.8%) cases in men [30, 31]. Cervical cancer represents the second most common neoplastic pathology among women [7, 32], and accounts for 83% of HPV-attributable cancer [30], with an estimated 500,000 diagnosed cases each year, resulting in more than 250,000 deaths [33].

Since the past three decades, increased rates of anal cancer have been also reported, globally accounting for 35,000 cases per year (half occurring in men), with a high proportion associated with the HR HPV16 and HPV18 types. Furthermore, 13,000 penile and 12,000 vaginal cancer cases, 8,500 cases of vulvar carcinoma, and 38,000 head and neck cancer cases have been attributed to HPV, of which 21,000 cases are oropharyngeal cancers, occurring in most developed countries [30]. HPV-attributable head and neck squamous cell cancers are mainly represented by Oropharyngeal Cancers (OPCs), which have two distinct etiologies: the HPV infection, with a proportion of cases alarmingly increasing in some high-income countries, and the additional risk factors from tobacco/alcohol consumption [34]. HPV-positive OPC patients are epidemiologically and clinically distinct compared with their HPV-negative counterparts, being younger, and having less exposure to tobacco and alcohol, higher socioeconomic and educational status with better outcomes [35]. In the newly published 8<sup>th</sup> edition TNM (using the "tumor - T" (extent of primary tumor), "lymph node - N" (absence/presence and extent of overt regional lymph nodes), and "metastasis - M" (absence/presence of distant metastasis) attributes) [36, 37] stage system for head and neck cancer, significant changes have been introduced to HPVmediated OPC [38-40]. HPV-specific staging is necessary for several reasons: relevance to discussion with patient/family; clinical trials design since HPV-positive and HPV-negative OPCs are separately addressed; and practice guidelines may be different; separate classification for HPV-

positive and negative OPCs may be further applicable for both clinical care and cancer surveillance [38].

Prophylactic HPV vaccines based on recombinantly expressed virus-like particles, with direct and indirect effects by cross-protection against HPV types not included in the formulations, have been developed, following two main approaches: one targeting young women prior to sexual debut, and the other including young men of the same age range [12].

To date, three vaccines licensed for use by the US Food and Drug Administration (FDA) have demonstrated a robust immune response [41], and have been recommended at age 11 or 12 years [42]. The Advisory Committee on Immunization Practices also recommends vaccination for females aged 13 through 26 years and males aged 13 through 21 years not previously vaccinated, and through age 26 years for men who have sex with men and immunocompromised subjects (including those with HIV infection) whether not previously vaccinated [42]. However, a universal vaccination strategy would have a huge impact, because it could be effective for preventing up to 90% of HPV-related diseases. In particular, current vaccination is available through: bivalent HPV (2vHPV) vaccine approved since 2009 (Cervarix; Glaxo SmithKline, Rixensart, Belgium) targeting HR HPV16 and HPV18; quadrivalent HPV (4vHPV) vaccine approved since 2006 (Gardasil; Merck & Co, Inc., Whitehouse Station, NJ) protecting against LR HPV6 and 11, and HR HPV16, and HPV18; nonavalent HPV (9vHPV) vaccine introduced in 2014 (Gardasil 9, Merck & Co, Inc. Whitehouse Station, NJ) for preventing infections by LR HPV6 and 11 and HR HPV16 and 18, and five additional oncogenic types, HPV31, 33, 45, 52, 58 [43]. The nonavalent vaccine was approved by FDA for use in females and males aged 9-14 years and represents, as from 2016, the only vaccine available for use in the United States, due to broader coverage against HR HPVs, while Cervarix<sup>®</sup> and Gardasil<sup>®</sup> for males and females aged 11-12 years are still used in other countries, including Italy.

Starting with vaccines against 2vHPV and 4vHPV, which became available more than 10 years ago, over 80 countries introduced a national HPV vaccination program, and as of 2016, globally it has been estimated that 1.4% of the total population and 6.1% of women aged 10-20 years received a full vaccine course, with a heterogeneous coverage among countries [12]. It has been reported that 33.6% of females aged 10-20 years received the full course of the vaccine in highly developed countries compared to 2.7% in less developed ones [44].

In Europe, analysis of full-course vaccination by geographical region revealed that coverage among the female population was 4.3% (14 million) and that 68% of the vaccinated women were predominantly from high-income countries, as compared to 28% from upper-middle-income countries. In particular, Australia and New Zealand reported the highest age-specific coverage rates, reaching nearly 70% of females aged 15-19 years. Therefore, the vaccination compliance rate is still far from the ideal threshold, underlin-

ing the urgent need to address effective strategies and to overcome vaccination barriers.

In Italy, the latest data related to HPV vaccine coverage up to December 2017 among females aged 12-20 years revealed a lower coverage in the primary vaccination group aged 12 years as compared to other age groups [45]. In the oldest cohorts (1997-2001) to whom vaccination was offered at no cost in most of the Italian regions until the eighteenth year, coverage for at least one vaccine dose was 73-76%, and 69-72% for the complete cycle. Therefore, the national HPV vaccination coverage in girls is still below the 95% optimal threshold as established by the National Vaccine Prevention Plan 2017-2019. At the regional level, 95% coverage for all cohorts is not reached in any Region, whilst the coverage in boys is very far from the objectives by the National Vaccine Prevention Plan, which has indicated a gradual 60% threshold for 2017 up to 95% in 2019 [45]. The wide variability of vaccination coverage amongst Italian regions underlines that targeted interventions are needed in specific geographical contexts, and that, although the anti-HPV vaccination is not comprised within the mandatory vaccinations by the Italian law n. 119/2017, it is included within the so-called "Essential Level of Care" (Livelli Essenziali di Assistenza-LEA), which must be guaranteed to all citizens by every Region.

#### 3. INSIGHTS ON MIRNA AND CERVICAL CANCER

MiRNAs are short 18-25 nucleotide non-coding RNAs that bind to the target mRNA typically in the 3'-untranslated region (3'-UTR) [46], causing mRNA degradation, or ceasing their translation. There are two principal mechanisms associated with miRNA functions: first, miRNAs bind specifically to sites in the 3'-UTR of targeted mRNAs, resulting in mRNA degradation; second, miRNAs exert partial base complementarity to the targeted mRNAs and suppress the translation processes, inhibiting protein synthesis. However, each miRNA may target more than 100 mRNAs, and at the same time, each mRNA may contain multiple binding sites for different miRNAs [47].

Research showed that these small molecules are involved in the expression of at least one-third of human genes [48], and that their functions can provide various translational applications in cancer, mainly for diagnostic, prognostic, and therapeutic approaches [49, 50]. By interfering with mRNA translation, miRNAs act as post-transcriptional regulators of gene expression, playing extensive effects on cellular functions, such as development, differentiation, proliferation, inflammation, immune responses, stress responses, apoptosis, invasion, and metastatization [51], thus with high potential to be used as diseases biomarkers [49].

Aberrant (dysregulated) miRNAs expression, with silencing or over-expression (de-silencing), has been demonstrated in several tumor types, including cervical cancer cell lines, acting as either oncogenic or tumor-suppressive agents [52, 53]. Despite evidence reporting critical roles of miRNAs in the carcinogenesis, it remains unresolved whether altered miRNAs expression is the cause or the consequence of malignant transformation [54, 55]. Oncoviruses, like HPV, express miRNAs that can significantly contribute to carcinogenic processes by regulating viral gene expression or influencing host gene expression [16]. The major mechanisms involved in carcinogenesis encompass the de-regulation of oncogenes and tumor suppressor genes by viral genome integration into host genome (*i.e.*, HPV, retroviruses), and the modulation of viral oncogenes expression (*i.e.*, herpesviruses) by causing DNA damage and host cell transformation through the inactivation of major regulators of genome stability and cell cycle [16].

The HPV genome is characterized by early-region genes E1-E7 encoding for the regulatory proteins E1, E2, E4, E5, E6, and E7, followed by a late region with L1 and L2 capsid proteins, a long control region with regulatory sequences, and a viral origin of replication [56]. Amongst all, E6 and E7 represent the main oncoproteins in HPVs, whose overexpression is a pre-requisite for tumor development being necessary and sufficient for HPV-mediated oncogenesis [5]; the E5 protein also contributes to increased cancer risk [57]. In particular, the E6 protein inactivates tumor suppressor p53 function [9, 58], whereas protein E7 binds Rb retinoblastoma protein, which is involved with cell cycle progression [9]. The virus integration into the host genome is a crucial event in HPV-related carcinogenesis and influences the stage of the disease and tumor type (cervical, anal, penile, head and neck cancer, etc.) [59].

During a persistent infection correlated with cervical lesions, HR HPV E6 and E7 can promote DNA damage *via* p53 and pRb inactivation, resulting in the impaired DNA checkpoint controls, leading to the subsequent transformation into cancer cells [9]. Hence, profiling expression of miRNAs in HPV-related malignancies is pivotal for identifying novel biomarkers for pre-malignant stages and for patients who require individualized treatments [60-62].

In the last decade, numerous studies have been conducted to better understand the role of miRNAs in HPV-related tumors, particularly cervical cancer [47, 63-66], although findings are generally contradictory. In detail, there are oncomiRs that include miRNAs significantly over-expressed in tumor tissues during HPV infection and lead to the onset of aberrant cell proliferation and growth [67], and tumor suppressor miRNAs that are under-expressed in HPV-positive tumors compared with normal non-cancerous tissues [68].

Generally, a differential miRNAs expression in invasive HPV positive squamous cell carcinomas compared to control tissue has been observed [69-73], revealing up- or downregulation (Table 1) of specific miRNAs [54]. Indeed, the abnormal/altered expression of miRNAs leads to tumor genesis and modulate disease progression (Fig. 1), as well as variations in different cervical cancer stages (*i.e.*, Cervical Intraepithelial Neoplasia - CIN grade I, II, and III) [73]. Furthermore, an increasing number of dysregulated miRNAs during the progression of CIN1-3 to cervical cancer has been reported. Several mechanisms can control miRNA expression, and are associated with the combination of chromosomal defects, such as deletions, amplifications or mutations with other genetic/epigenetic events, causing up- or down-regulation of miRNAs [74].

#### 

MiRNA	References
Up-regulation	
miR-9	[60, 63, 65, 75-82]
miR-10a	[60, 64, 65, 75, 83]
miR-16	[52, 61-64, 75, 84]
miR-17	[75, 84-87]
miR-20a	[60, 66, 86, 88-93]
miR-20b	[65, 72, 75, 79, 84, 89, 94, 95]
miR-21	[52, 61, 70, 71, 75, 76, 78, 84-87, 90, 96-114]
miR-27a	[53, 62, 64, 85, 115, 116]
miR-31	[62, 71, 76, 85, 89, 96, 98, 117-120]
miR-92a	[62, 66, 75, 118, 121-125]
miR-93	[75, 84, 89, 118, 126, 127]
miR-106a	[64, 75, 84, 118, 128, 129]
miR-146a	[52, 75, 90, 103, 130, 131]
miR-155	[52, 53, 75, 78, 79, 84, 117, 118, 132-134]
miR-196a	[60, 64, 135-139]
miR-199b	[63, 79, 85, 140, 141]
miR-200a	[61, 75, 89, 105, 117, 126, 142]
miR-205	[52, 64, 85, 117, 138, 143-145]
miR-210	[66, 82, 85, 89, 146]
Down-regulation	
miR-29a	[52, 62, 64, 84, 90, 118, 147, 148]
miR-34a	[53, 73, 79, 86, 90, 114, 132, 149-154]
miR-99a	[61, 64, 73, 75, 89, 110, 118]
miR-100	[52, 62, 89, 75, 84, 86, 118, 155, 156]
miR-125b	[52, 61, 78, 75, 84, 117, 118, 151, 157]
miR-126	[52, 86, 117, 158-160]
miR-143	[52, 64, 69, 70, 85, 89, 99, 117, 148, 161-164]
miR-145	[52, 64, 69, 77, 84, 85, 89, 118, 139, 165-169]
miR-149	[61, 63, 75, 84, 85, 170]
miR-193b	[61, 65, 75, 79, 96]
miR-195	[52, 75, 76, 78, 84, 89, 110, 118, 171-175]
miR-203	[53, 61, 63-65, 75, 79, 85, 86, 90, 176]
miR-214	[89, 98, 126, 177-183]
miR-218	[69, 71, 73, 75, 76, 89, 110, 118, 147, 150, 184-191]
miR-375	[75, 84, 118, 150, 192, 193]
miR-424	[52, 78, 117, 118, 150, 194-198]
miR-497	[69, 75, 76, 84, 89, 199, 200]

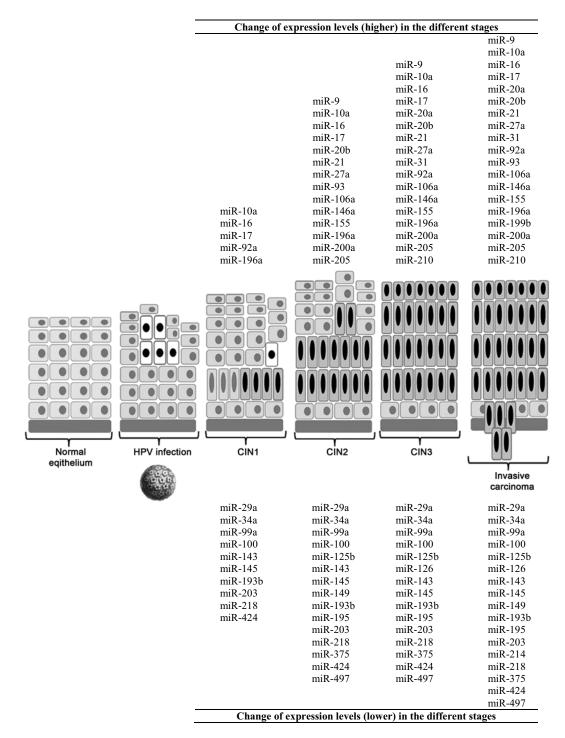


Fig. (1). Interaction between cervical epithelium, HPV infection, and the most significant miRNAs dysregulated in the cervical cancer progression (modified from Pardini *et al.* [14]).

As shown in Table 1, many studies investigated the role of miRNAs involved in the development of cervical cancer, underlining their crucial biological roles. Findings achieved from the available studies indicate that over 250 miRNAs are expressed differently in cervical cancer, being up- or downregulated.

In the present report, we described the most relevant miRNAs and their function at different stages in the cervical cancer progression (cited in at least 5 articles with similar findings; Table 1). Among the up-regulated miRNAs, miR-21 and miR-155 have been widely reported to promote cell migration and invasion (Fig. 1), as well as lymph node metastasis in cervical cancer [52, 53, 66, 70, 71, 75, 76, 78, 79, 84-87, 90, 96-114, 117, 118, 132-134]. Additional up-regulated miRNAs have been involved in the initiation and progression of cervical carcinogenesis, since the aberrant expression of miR-20b, miR-31, miR-92a, miR-146a, and miR-205 positively correlated with cell proliferation, enhancement of the migration and invasion abilities of cancer cells, and vessel invasion [52, 62, 64-66, 71, 72, 75, 76, 79, 84, 85, 89, 90, 94-96, 98, 103, 117-125, 130, 131, 138, 143-145]. Furthermore, the up-regulation of miR-196a has been linked to an enhanced proliferative ability of cervical cancer cells, advanced tumor stage, and poor overall survival in patients [60, 64, 135-139], and miR-9 expression has been found significantly over-expressed in squamous cervical cancer [60, 63, 65, 75-82].

Different miRNAs are down-regulated as well, such as: miR-34a and miR-214, both promoting cell proliferation, migration, invasion, and angiogenesis [53, 73, 79, 86, 89, 90, 98, 114, 126, 132, 149, 150-154, 177-183]; miR-143, involved in the regulation of cell migration and invasion [52, 64, 69, 70, 85, 89, 99, 117, 148, 161-164]; miR-145, whose low expression levels were significantly associated with poor cancer differentiation and lymph node metastasis [52, 64, 69, 77, 84, 85, 89, 118, 139, 165-169]; miR-195, whose reduced expression has been associated to advanced clinical stage in cancer patients and lymph node metastasis [52, 75, 76, 78, 84, 89, 110, 118, 171-175]; miR-203, linked to lymph node metastasis [53, 62-65, 75, 79, 85, 86, 90, 176]; miR-218, associated with cell migration and invasion, induction of epithelial-mesenchymal transition and invasion [69, 71, 73, 75, 76, 89, 110, 118, 147, 150, 184-191]; miR-375, involved in cell proliferation, migration and invasion, modulation of epithelial-mesenchymal transition and correlated with pelvic lymph node metastases [75, 84, 118, 150, 192, 193]; miR-424, positively correlating with poor tumor differentiation, advanced clinical stage, lymph node metastasis and other low prognostic clinical/pathological parameters [52, 78, 117, 118, 150, 194-198]. Interestingly, the expression levels of miR-34a, miR-218, miR-375, and miR-424 have been found decreased simultaneously to an increase of disease severity [14].

# CONCLUSION

miRNAs regulate diverse biological processes in the host-pathogen interactions and represent an attractive target due to the critical function in several signaling pathways. These small biomolecules are commonly encoded by viruses that undergo long-term persistent infection, including HPV, and can act as oncogenes to promote carcinogenesis, or as tumor suppressors targeting oncogene mRNAs. The miRNAs dysregulation causes abnormal cell growth and differentiation, leading to cancer and many other diseases. Indeed, miRNA expression profiles have been found to be highly altered in several cancers compared to healthy controls.

Papillomaviruses can cause several types of cancers, but only persistent infections can lead to malignant tumors. Although oncogenic genotypes are etiologically associated with cervical cancer, the HPV infection alone is not sufficient to induce the malignant transformation and to explain the development of cervical cancer, implying that molecular factors at multiple levels should be considered, including miRNAs and their altered expression profiles. Hence, a systematic and rigorous appraisal of the alterations of miRNAs during oncogenic HPVs transformation can significantly improve knowledge of cervical carcinogenesis processes, with potentially enormous public health impact in drawing successful diagnostic, prognostic, and therapeutic strategies.

Tumors related to oncoviruses, including HPV, have been extensively studied with the aim to detect markers and treatment targets. Studies focused on cervical cancer molecular epidemiology predominate, as it is the most common malignancy in women worldwide. Public health efforts should be implemented to reduce the attributable morbidity and mortality that can be achieved only with significant improvements in all levels of prevention, particularly reinforcing and supporting vaccination compliance.

Cervical cancer cells display the aberrant expression of a considerable number of miRNAs, both oncogenic and tumorsuppressive, and specific up-regulated or down-regulated miRNAs have been shown to correlate with tumors. Despite the uncertainty in the functional effects of the miRNAs dysregulation on the cervical oncogenesis, the evaluation of their expression profiles have already provided noteworthy insights, and might also be valuable to assess factors and specific risks related to individual susceptibility, as well as useful for the clinical histopathological classification. Since miRNAs expression is altered in cancer tissues, investigation of the differential profiles of certain miRNAs may lead to crucial advancements for cancer diagnosis, facilitating disease classification and monitoring its development and progression, prognosis, and treatment.

The present study provides an overview of the contribution of miRNAs into cervical cancer, focusing on the recently available evidence, which is somewhat contradictory about their influence on gene expression in the human host. Several differentially expressed miRNAs in cervical cancer tissues or cells have been reviewed, but it could not be concluded on a validated panel of viral miRNAs in cervical infected cells. Despite the extensive studies carried out, to date, no widely accepted HPV-encoded miRNAs signature can be definitely recognized, maybe due to the lack of broadly applicable methodology and of suitable/standardized laboratory models, which are required. Although additional investigations are consistently needed for a better comprehension of miRNAs pathways and expression profiles that can significantly improve the understanding of epidemiology of HPV infections and related diseases, the reviewed evidence underlines the emerging role of miRNAs in cervical carcinogenesis mechanism as key mediators of gene expression, and the usefulness of tracking their deregulation as potential predictive biomarkers in the field of infectious diseases prevention and therapeutic decisions remains to be established. Promising applications of miRNAs could also be addressed for the improvement of vaccination programs since the current HPV-targeting vaccines are effective for preventing infections and neoplastic diseases, but the established or pre-existing infections could not be cleared, being the target antigens not expressed in the infected basal epithelial cells [201-203].

In conclusion, miRNAs could represent effective markers of progression of pre-neoplastic lesions to invasive and metastatic disease, providing a dynamic prognostic factor. Therefore, since miRNAs have a vital role in all stages of cervical cancer progression, from cell invasion and migration to eventual tumor metastasis, the combined use of a selected panel of oncomir and tumor suppressor miRNAs could provide an enhanced diagnostic and prognostic approach, with increased accuracy with respect to any single miRNA marker.

### **CONSENT FOR PUBLICATION**

Not applicable.

### FUNDING

None.

### **CONFLICT OF INTEREST**

The authors declare no conflict of interest, financial or otherwise.

## **ACKNOWLEDGEMENTS**

The authors acknowledge Dr. Jim McLauchlin, Public Health England, London, UK, for the helpful comments and revisions provided to the manuscript.

## REFERENCES

 Damania B. A virological perspective on cancer. PLoS Pathog 2016; 12(2): e1005326.

http://dx.doi.org/10.1371/journal.ppat.1005326 PMID: 26866686 [2] Mesri EA, Feitelson MA, Munger K. Human viral oncogenesis: a

- [2] Mesh EA, Peterson MA, Murger K. Human vitar oncogenesis. a cancer hallmarks analysis. Cell Host Microbe 2014; 15(3): 266-82. http://dx.doi.org/10.1016/j.chom.2014.02.011 PMID: 24629334
- Zapatka M, Borozan I, Brewer DS, *et al.* The landscape of viral associations in human cancers. bioRxiv The Preprint 2018; 12: 39. [doi: https://doi.org/10.1101/465757
- [4] Bravo IG, de Sanjosé S, Gottschling M. The clinical importance of understanding the evolution of papillomaviruses. Trends Microbiol 2010; 18(10): 432-8.
- http://dx.doi.org/10.1016/j.tim.2010.07.008 PMID: 20739182
   [5] McBride AA. Oncogenic human papillomaviruses. Philos Trans R Soc Lond B Biol Sci 2017; 372(1732): 20160273. http://dx.doi.org/10.1098/rstb.2016.0273 PMID: 28893940
- [6] McLaughlin-Drubin ME, Münger K. The human papillomavirus E7 oncoprotein. Virology 2009; 384(2): 335-44.
- http://dx.doi.org/10.1016/j.virol.2008.10.006 PMID: 19007963
  [7] Tsakogiannis D, Gartzonika C, Levidiotou-Stefanou S, Markoulatos P. Molecular approaches for HPV genotyping and HPV-DNA physical status. Exp Rev Mol Med 2017; 19: e1. http://dx.doi.org/10.1017/erm.2017.2 PMID: 28162121
- [8] Haley CT, Mui UN, Vangipuram R, et al. Human oncoviruses: mucocutaneous manifestations, pathogenesis, therapeutics, and prevention (Part I: papillomaviruses and Merkel cell polyomavirus). J Am Acad Dermatol 2018; S0190-9622: 32965-7.
- [9] Viarisio D, Gissmann L, Tommasino M. Human papillomaviruses and carcinogenesis: well-established and novel models. Curr Opin Virol 2017; 26: 56-62.

http://dx.doi.org/10.1016/j.coviro.2017.07.014 PMID: 28778034

[10] Sammarco ML, Ucciferri C, Tamburro M, Falasca K, Ripabelli G, Vecchiet J. High prevalence of human papillomavirus type 58 in HIV infected men who have sex with men: a preliminary report in Central Italy. J Med Virol 2016; 88(5): 911-4. http://dx.doi.org/10.1002/jmv.24406 PMID: 26467111

- [11] Ucciferri C, Tamburro M, Falasca K, Sammarco ML, Ripabelli G, Vecchiet J. Prevalence of anal, oral, penile and urethral human papillomavirus in HIV infected and HIV uninfected men who have sex with men. J Med Virol 2018; 90(2): 358-66. http://dx.doi.org/10.1002/jmv.24943 PMID: 28906006
- [12] St Laurent J, Luckett R, Feldman S. HPV vaccination and the effects on rates of HPV-related cancers. Curr Prob Cancer 2018; 42(5): 493-506. http://dx.doi.org/10.1016/j.currproblcancer.2018.06.004 PMID: 30041818
- [13] Sheikh S, Biundo E, Courcier S, et al. A report on the status of vaccination in Europe. Vaccine 2018; 36(33): 4979-92. http://dx.doi.org/10.1016/j.vaccine.2018.06.044 PMID: 30037416
- [14] Pardini B, De Maria D, Francavilla A, Di Gaetano C, Ronco G, Naccarati A. MicroRNAs as markers of progression in cervical cancer: a systematic review. BMC Cancer 2018; 18(1): 696. http://dx.doi.org/10.1186/s12885-018-4590-4 PMID: 29945565
- [15] Araldi RP, Sant'Ana TA, Módolo DG, *et al.* The Human Papillomavirus (HPV)-related cancer biology: an overview. Biomed Pharmacother 2018; 106: 1537-56.
- http://dx.doi.org/10.1016/j.biopha.2018.06.149 PMID: 30119229
  [16] Vojtechova Z, Tachezy R. The role of miRNAs in virus-mediated
- oncogenesis. Int J Mol Sci 2018; 19(4): E1217. http://dx.doi.org/10.3390/ijms19041217 PMID: 29673190
- [17] Calin GA, Croce CM. MicroRNA-cancer connection: the beginning of a new tale. Cancer Res 2006; 66(15): 7390-4. http://dx.doi.org/10.1158/0008-5472.CAN-06-0800 PMID: 16885332
- [18] Gaur A, Jewell DA, Liang Y, et al. Characterization of microRNA expression levels and their biological correlates in human cancer cell lines. Cancer Res 2007; 67(6): 2456-68. http://dx.doi.org/10.1158/0008-5472.CAN-06-2698 PMID: 17363563
- [19] de Sanjosé S, Brotons M, Pavón MA. The natural history of human papillomavirus infection. Best Pract Res Clin Obstet Gynaecol 2018; 47: 2-13.

http://dx.doi.org/10.1016/j.bpobgyn.2017.08.015 PMID: 28964706 Araldi RP, Assaf SMR, Carvalho RF, et al. Papillomaviruses: a

- [20] Araldi RP, Assaf SMR, Carvalho RF, et al. Papillomaviruses: systematic review. Genet Mol Biol 2017; 40(1): 1-21. http://dx.doi.org/10.1590/1678-4685-gmb-2016-0128
   PMID: 28212457
- [21] Sammarco ML, Del Riccio I, Tamburro M, Grasso GM, Ripabelli G. Type-specific persistence and associated risk factors of human papillomavirus infections in women living in central Italy. Eur J Obstet Gynecol Reprod Biol 2013; 168(2): 222-6. http://dx.doi.org/10.1016/j.ejogrb.2013.01.012 PMID: 23395560
- [22] Shanmugasundaram S, You J. Targeting persistent human papillomavirus infection. Viruses 2017; 9(8): E229. http://dx.doi.org/10.3390/v9080229 PMID: 28820433
- [23] Serrano B, de Sanjosé S, Tous S, et al. Human papillomavirus genotype attribution for HPVs 6, 11, 16, 18, 31, 33, 45, 52 and 58 in female anogenital lesions. Eur J Cancer 2015; 51(13): 1732-41. http://dx.doi.org/10.1016/j.ejca.2015.06.001 PMID: 26121913
- [24] Tommasino M. The human papillomavirus family and its role in carcinogenesis. Semin Cancer Biol 2014; 26: 13-21. http://dx.doi.org/10.1016/j.semcancer.2013.11.002 PMID: 24316445
- [25] Serrano B, Brotons M, Bosch FX, Bruni L. Epidemiology and burden of HPV-related disease. Best Pract Res Clin Obstet Gynaecol 2018; 47: 14-26.

http://dx.doi.org/10.1016/j.bpobgyn.2017.08.006 PMID: 29037457

- [26] Ripabelli G, Grasso GM, Del Riccio I, Tamburro M, Sammarco ML. Prevalence and genotype identification of human papillomavirus in women undergoing voluntary cervical cancer screening in Molise, central Italy. Cancer Epidemiol 2010; 34(2): 162-7. http://dx.doi.org/10.1016/j.canep.2009.12.010 PMID: 20080070
- [27] Kaliff M, Sorbe B, Mordhorst LB, Helenius G, Karlsson MG, Lillsunde-Larsson G. Findings of multiple HPV genotypes in cervical carcinoma are associated with poor cancer-specific survival in a Swedish cohort of cervical cancer primarily treated with radiotherapy. Oncotarget 2018; 9(27): 18786-96.
- http://dx.doi.org/10.18632/oncotarget.24666 PMID: 29721161 [28] Lowy DR, Schiller JT. Reducing HPV-associated cancer globally.

http://dx.doi.org/10.1158/1940-6207.CAPR-11-0542 PMID: 22219162

- [29] Brianti P, De Flammineis E, Mercuri SR. Review of HPV-related diseases and cancers. New Microbiol 2017; 40(2): 80-5. PMID: 28368072
- [30] de Martel C, Plummer M, Vignat J, Franceschi S. Worldwide burden of cancer attributable to HPV by site, country and HPV type. Int J Cancer 2017; 141(4): 664-70. http://dx.doi.org/10.1002/ijc.30716 PMID: 28369882
- [31] World Health Organization (WHO). Human papillomavirus vaccines WHO position paper. World Health Organization, Geneva, Italy. 2017; 92: 241-68. Available from: http://apps.who.int/iris/ bitstream/10665/255353/1/WER9219.pdf?ua=1
- [32] Ferlay J, Soerjomataram I, Dikshit R, et al. Cancer incidence and mortality worldwide: sources, methods and major patterns in GLOBOCAN 2012. Int J Cancer 2015; 136(5): E359-86. http://dx.doi.org/10.1002/ijc.29210 PMID: 25220842
- [33] Plummer M, de Martel C, Vignat J, Ferlay J, Bray F, Franceschi S. Global burden of cancers attributable to infections in 2012: a synthetic analysis. Lancet Glob Health 2016; 4(9): e609-16. http://dx.doi.org/10.1016/S2214-109X(16)30143-7 PMID: 27470177
- [34] Marur S, D'Souza G, Westra WH, Forastiere AA. HPV-associated head and neck cancer: a virus-related cancer epidemic. Lancet Oncol 2010; 11(8): 781-9. http://dx.doi.org/10.1016/S1470-2045(10)70017-6 PMID: 20451455
- [35] Evans M, Newcombe R, Fiander A, *et al.* Human Papillomavirusassociated oropharyngeal cancer: an observational study of diagnosis, prevalence and prognosis in a UK population. BMC Cancer 2013; 13: 220.
- http://dx.doi.org/10.1186/1471-2407-13-220 PMID: 23634887
   [36] Amin M, Edge S, Greene F, *et al.* AJCC cancer staging manual. 8<sup>th</sup> Ed. New York: Springer June 2018; pp. 1-507. http://dx.doi.org/10.1007/978-3-319-40618-3
- [37] Brierley J, Gospodarowicz M, Wittekind C. UICC TNM classification of malignant tumours. 8<sup>th</sup> Ed. Chichester: Wiley 2017.
- [38] Huang SH, O'Sullivan B. Overview of the 8<sup>th</sup> Edition TNM classification for head and neck cancer. Curr Treat Options Oncol 2017; 18(7): 40.
- [39] O'Sullivan B. Head and neck tumours. Editors Brierley J, Gospodarowicz M, Ch W, *et al.* UICC TNM classification of malignant tumours. 8<sup>th</sup> Ed. Chichester: Wiley 2017; pp. 17-54.
- [40] O'Sullivan B, Lydiatt W, Haughey BH, et al. HPVmediated (p16+) oropharyngeal cancer. In: Amin M, Edge S, Greene F, et al, editors AJCC cancer staging manual. 8<sup>th</sup> Ed. New York: Springer; 2017; pp. 113-21.
- [41] Mollers M, Vossen JM, Scherpenisse M, van der Klis FRM, Meijer CJLM, de Melker HE. Review: current knowledge on the role of HPV antibodies after natural infection and vaccination: implications for monitoring an HPV vaccination programme. J Med Virol 2013; 85(8): 1379-85. http://dx.doi.org/10.1002/jmv.23616 PMID: 23722396
- [42] Markowitz LE, Dunne EF, Saraiya M, et al. Centers for Disease Control and Prevention (CDC). Human papillomavirus vaccination: recommendations of the Advisory Committee On Immunization Practices (ACIP). MMWR Recomm Rep 2014; 63(RR-05): 1-30. PMID: 25167164
- [43] Food and Drug Administration. December 10, 2014 Approval letter-GARDASIL 9. Silver Spring, MD: US Department of Health and Human Services, Food and Drug Administration; 2014. Available at http://www.fda.gov/BiologicsBloodVaccines/Vaccines/ ApprovedProducts/ucm426520.htmExternal
- [44] Bruni L, Diaz M, Barrionuevo-Rosas L, et al. Global estimates of human papillomavirus vaccination coverage by region and income level: a pooled analysis. Lancet Glob Health 2016; 4(7): e453-63. http://dx.doi.org/10.1016/S2214-109X(16)30099-7 PMID: 27340003
- [45] Italian Ministry of Health. General Administration of Health Prevention, office of prevention of communicable diseases and international prophylaxis. Comment on HPV vaccination coverage on 31/12/2017. Available at: http://www.salute.gov.it/imgs/C\_17\_ tavole\_27\_allegati\_itemAllegati\_1\_fileAllegati\_itemFile\_1\_file.pdf

- [46] Felekkis K, Touvana E, Stefanou Ch, Deltas C. microRNAs: a newly described class of encoded molecules that play a role in health and disease. Hippokratia 2010; 14(4): 236-40. PMID: 21311629
- [47] Laengsri V, Kerdpin U, Plabplueng C, Treeratanapiboon L, Nuchnoi P. Cervical cancer markers: epigenetics and microRNAs. Lab Med 2018; 49(2): 97-111. http://dx.doi.org/10.1093/labmed/lmx080 PMID: 29378033
- [48] Lewis BP, Burge CB, Bartel DP. Conserved seed pairing, often flanked by adenosines, indicates that thousands of human genes are microRNA targets. Cell 2005; 120(1): 15-20. http://dx.doi.org/10.1016/j.cell.2004.12.035 PMID: 15652477
- [49] Kanekura K, Nishi H, Isaka K, Kuroda M. MicroRNA and gynecologic cancers. J Obstet Gynaecol Res 2016; 42(6): 612-7. http://dx.doi.org/10.1111/jog.12995 PMID: 27098274
- [50] Kwok GT, Zhao JT, Weiss J, et al. Translational applications of microRNAs in cancer, and therapeutic implications. Noncoding RNA Res 2017; 2(3-4): 143-50. http://dx.doi.org/10.1016/j.ncrna.2017.12.002 PMID: 30159433
- [51] Mangino G, Chiantore MV, Iuliano M, Fiorucci G, Romeo G. Inflammatory microenvironment and human papillomavirusinduced carcinogenesis. Cytokine Growth Factor Rev 2016; 30: 103-11.

http://dx.doi.org/10.1016/j.cytogfr.2016.03.007 PMID: 27021827

- [52] Wang X, Tang S, Le SY, et al. Aberrant expression of oncogenic and tumor-suppressive microRNAs in cervical cancer is required for cancer cell growth. PLoS One 2008; 3(7): e2557. http://dx.doi.org/10.1371/journal.pone.0002557 PMID: 18596939
- [53] Gocze K, Gombos K, Kovacs K, Juhasz K, Gocze P, Kiss I. MicroRNA expressions in HPV-induced cervical dysplasia and cancer. Anticancer Res 2015; 35(1): 523-30. PMID: 25550598
- [54] Shishodia G, Verma G, Das BC, Bharti AC. miRNA as viral transcription tuners in HPV-mediated cervical carcinogenesis. Front Biosci (Schol Ed) 2018; 10: 21-47. http://dx.doi.org/10.2741/s499 PMID: 28930517
- Izzotti A. MicroRNA from small oligunucletoides to giant players of biological processes and diseases. MicroRNA 2019; 8(1): 2-3. http://dx.doi.org/10.2174/221153660801181024142302 PMID: 30511596
- [56] Graham SV. Human papillomavirus: gene expression, regulation and prospects for novel diagnostic methods and antiviral therapies. Future Microbiol 2010; 5(10): 1493-506. http://dx.doi.org/10.2217/fmb.10.107 PMID: 21073310
- [57] Schiffman M, Herrero R, Desalle R, *et al.* The carcinogenicity of human papillomavirus types reflects viral evolution. Virology 2005; 337(1): 76-84.

http://dx.doi.org/10.1016/j.virol.2005.04.002 PMID: 15914222

- [58] Mortensen F, Schneider D, Barbic T, et al. Role of ubiquitin and the HPV E6 oncoprotein in E6AP-mediated ubiquitination. Proc Natl Acad Sci USA 2015; 112(32): 9872-7. http://dx.doi.org/10.1073/pnas.1505923112 PMID: 26216987
- [59] Vinokurova S, Wentzensen N, Kraus I, et al. Type-dependent integration frequency of human papillomavirus genomes in cervical lesions. Cancer Res 2008; 68(1): 307-13. http://dx.doi.org/10.1158/0008-5472.CAN-07-2754 PMID: 18172324
- [60] Xin F, Liu P, Ma CF. A circulating serum miRNA panel as early detection biomarkers of cervical intraepithelial neoplasia. Eur Rev Med Pharmacol Sci 2016; 20(23): 4846-51. PMID: 27981553
- [61] Gao C, Zhou C, Zhuang J, et al. MicroRNA expression in cervical cancer: novel diagnostic and prognostic biomarkers. J Cell Biochem 2018; 119(8): 7080-90. http://dx.doi.org/10.1002/jcb.27029 PMID: 29737570
- [62] Wang X, Wang HK, Li Y, et al. microRNAs are biomarkers of oncogenic human papillomavirus infections. Proc Natl Acad Sci USA 2014; 111(11): 4262-7.
- http://dx.doi.org/10.1073/pnas.1401430111 PMID: 24591631
   [63] Lee JW, Choi CH, Choi JJ, *et al.* Altered microRNA expression in cervical carcinomas. Clin Cancer Res 2008; 14(9): 2535-42. http://dx.doi.org/10.1158/1078-0432.CCR-07-1231
   PMID: 18451214

- [64] Pereira PM, Marques JP, Soares AR, Carreto L, Santos MA. MicroRNA expression variability in human cervical tissues. PLoS One 2010; 5(7): e11780. http://dx.doi.org/10.1371/journal.pone.0011780 PMID: 20668671
- [65] Cheung TH, Man KN, Yu MY, et al. Dysregulated microRNAs in the pathogenesis and progression of cervical neoplasm. Cell Cycle 2012; 11(15): 2876-84. http://dx.doi.org/10.4161/cc.21278 PMID: 22801550
- [66] Liu SS, Chan KKL, Chu DKH, *et al.* Oncogenic microRNA signature for early diagnosis of cervical intraepithelial neoplasia and cancer. Mol Oncol 2018; 12(12): 2009-22.
- http://dx.doi.org/10.1002/1878-0261.12383 PMID: 30221475
   [67] Tomari Y, Zamore PD. MicroRNA biogenesis: drosha can't cut it without a partner. Curr Biol 2005; 15(2): R61-4.
- http://dx.doi.org/10.1016/j.cub.2004.12.057 PMID: 15668159
  [68] Baffa R, Fassan M, Volinia S, *et al.* MicroRNA expression profiling of human metastatic cancers identifies cancer gene targets. J Pathol 2009; 219(2): 214-21.
- http://dx.doi.org/10.1002/path.2586 PMID: 19593777
   [69] Martinez I, Gardiner AS, Board KF, Monzon FA, Edwards RP, Khan SA. Human papillomavirus type 16 reduces the expression of microRNA-218 in cervical carcinoma cells. Oncogene 2008; 27(18): 2575-82.
- http://dx.doi.org/10.1038/sj.onc.1210919 PMID: 17998940
  [70] Lui WO, Pourmand N, Patterson BK, Fire A. Patterns of known and novel small RNAs in human cervical cancer. Cancer Res 2007; 67(13): 6031-43. http://dx.doi.org/10.1158/0008-5472.CAN-06-0561
- PMID: 17616659
   [71] Zeng K, Mo X, Liu F, Hu X. Differential expression of microRNAs in cervical cancer and cervical precancerous lesions. Cancer Res Prev Treat 2014: 7: 024.
- [72] Kawai S, Fujii T, Kukimoto I, *et al.* Identification of miRNAs in cervical mucus as a novel diagnostic marker for cervical neoplasia. Sci Rep 2018; 8(1): 7070.

http://dx.doi.org/10.1038/s41598-018-25310-1 PMID: 29728572
 [73] Nair VB, Manasa VG, Sinto MS, Jayasree K, James FV, Kannan S. Differential expression of microRNAs in uterine cervical cancer and its implications in carcinogenesis; an integrative approach. Int J Gynecol Cancer 2018; 28(3): 553-62.

http://dx.doi.org/10.1097/IGC.000000000001203 PMID: 29466255

- Sharma G, Dua P, Agarwal SM. A comprehensive review of dysregulated miRNAs involved in cervical cancer. Curr Genomics 2014; 15(4): 310-23. http://dx.doi.org/10.2174/1389202915666140528003249 PMID: 25132800
- [75] Wilting SM, Snijders PJF, Verlaat W, et al. Altered microRNA expression associated with chromosomal changes contributes to cervical carcinogenesis. Oncogene 2013; 32(1): 106-16. http://dx.doi.org/10.1038/onc.2012.20 PMID: 22330141
- Zeng K, Zheng W, Mo X, et al. Dysregulated microRNAs involved in the progression of cervical neoplasm. Arch Gynecol Obstet 2015; 292(4): 905-13. http://dx.doi.org/10.1007/s00404-015-3702-5 PMID: 25851497
- [77] Azizmohammadi S, Safari A, Azizmohammadi S, et al. Molecular identification of miR-145 and miR-9 expression level as prognostic biomarkers for early-stage cervical cancer detection. QJM 2017; 110(1): 11-5.
- http://dx.doi.org/10.1093/qjmed/hcw101 PMID: 27345415
  [78] Park S, Eom K, Kim J, *et al.* MiR-9, miR-21, and miR-155 as potential biomarkers for HPV positive and negative cervical cancer. BMC Cancer 2017; 17(1): 658. http://dx.doi.org/10.1186/s12885-017-3642-5 PMID: 28934937
- [79] Harden ME, Prasad N, Griffiths A, Munger K. Modulation of microRNA-mRNA target pairs by human papillomavirus 16 oncoproteins. MBio 2017; 8(1): e02170-16. http://dx.doi.org/10.1128/mBio.02170-16 PMID: 28049151
- [80] Aishanjiang A, Rouzi N, Jiao Z, et al. MicroRNA-9 enhances invasion and migration of cervical carcinomas by directly targeting FOXO1. Eur Rev Med Pharmacol Sci 2018; 22(8): 2253-60. PMID: 29762826

- [81] Zhang H, Zhang Z, Wang S, Zhang S, Bi J. The mechanisms involved in miR-9 regulated apoptosis in cervical cancer by targeting FOXO3. Biomed Pharmacother 2018; 102: 626-32. http://dx.doi.org/10.1016/j.biopha.2018.03.019 PMID: 29602130
- [82] Nilsen A, Jonsson M, Aarnes EK, Kristensen GB, Lyng H. Reference microRNAs for RT-qPCR assays in cervical cancer patients and their application to studies of HPV16 and hypoxia biomarkers. Transl Oncol 2019; 12(3): 576-84. http://dx.doi.org/10.1016/j.tranon.2018.12.010 PMID: 30660934
- [83] Long MJ, Wu FX, Li P, Liu M, Li X, Tang H. MicroRNA-10a targets CHL1 and promotes cell growth, migration and invasion in human cervical cancer cells. Cancer Lett 2012; 324(2): 186-96. http://dx.doi.org/10.1016/j.canlet.2012.05.022 PMID: 22634495
- [84] Lajer CB, Garnæs E, Friis-Hansen L, et al. The role of miRNAs in Human Papilloma Virus (HPV)-associated cancers: bridging between HPV-related head and neck cancer and cervical cancer. Br J Cancer 2012; 106(9): 1526-34. http://dx.doi.org/10.1038/bjc.2012.109 PMID: 22472886
- [85] Liu L, Yu X, Guo X, et al. mR-143 is downregulated in cervical cancer and promotes apoptosis and inhibits tumor formation by targeting Bcl-2. Mol Med Rep 2012; 5(3): 753-60.
   PMID: 22160209
- [86] Chen J, Yao D, Li Y, et al. Serum microRNA expression levels can predict lymph node metastasis in patients with early-stage cervical squamous cell carcinoma. Int J Mol Med 2013; 32(3): 557-67. http://dx.doi.org/10.3892/ijmm.2013.1424 PMID: 23799609
- [87] Cai L, Wang W, Li X, et al. MicroRNA-21-5p induces the metastatic phenotype of human cervical carcinoma cells in vitro by targeting the von Hippel-Lindau tumor suppressor. Oncol Lett 2018; 15(4): 5213-9.

http://dx.doi.org/10.3892/ol.2018.7937 PMID: 29552160

[88] Kang HW, Wang F, Wei Q, et al. miR-20a promotes migration and invasion by regulating TNKS2 in human cervical cancer cells. FEBS Lett 2012; 586(6): 897-904.

http://dx.doi.org/10.1016/j.febslet.2012.02.020 PMID: 22449978
[89] Rao Q, Shen Q, Zhou H, Peng Y, Li J, Lin Z. Aberrant microRNA expression in human cervical carcinomas. Med Oncol 2012; 29(2): 1242-8.

http://dx.doi.org/10.1007/s12032-011-9830-2 PMID: 21264530

[90] Zhao S, Yao DS, Chen JY, Ding N. Aberrant expression of miR-20a and miR-203 in cervical cancer. Asian Pac J Cancer Prev 2013; 14(4): 2289-93.

http://dx.doi.org/10.7314/APJCP.2013.14.4.2289 PMID: 23725129 Zhao S, Yao D, Chen J, Ding N, Ren F. MiR-20a promotes cervical

[91] Zhao S, Yao D, Chen J, Ding N, Ren F. MiR-20a promotes cervical cancer proliferation and metastasis *in vitro* and *in vivo*. PLoS One 2015; 10(3): e0120905.

http://dx.doi.org/10.1371/journal.pone.0120905 PMID: 25803820

- [92] Liu X. Up-regulation of miR-20a by HPV16 E6 exerts growthpromoting effects by targeting PDCD6 in cervical carcinoma cells. Biomed Pharmacother 2018; 102: 996-1002. http://dx.doi.org/10.1016/j.biopha.2018.03.154 PMID: 29710555
- [93] Zhou Q, Dong J, Luo R, Zhou X, Wang J, Chen F. MicroRNA-20a regulates cell proliferation, apoptosis and autophagy by targeting thrombospondin 2 in cervical cancer. Eur J Pharmacol 2019; 844: 102-9.

http://dx.doi.org/10.1016/j.ejphar.2018.11.043 PMID: 30513279

- [94] Ma D, Zhang YY, Guo YL, Li ZJ, Geng L. Profiling of microRNA-mRNA reveals roles of microRNAs in cervical cancer. Chin Med J (Engl) 2012; 125(23): 4270-6. PMID: 23217399
- [95] Cheng Y, Geng L, Zhao L, Zuo P, Wang J. Human papillomavirus E6-regulated microRNA-20b promotes invasion in cervical cancer by targeting tissue inhibitor of metalloproteinase 2. Mol Med Rep 2017; 16(4): 5464-70.

http://dx.doi.org/10.3892/mmr.2017.7231 PMID: 28849054

- [96] Muralidhar B, Goldstein LD, Ng G, et al. Global microRNA profiles in cervical squamous cell carcinoma depend on Drosha expression levels. J Pathol 2007; 212(4): 368-77. http://dx.doi.org/10.1002/path.2179 PMID: 17471471
- [97] Yao Q, Xu H, Zhang QQ, Zhou H, Qu LH. MicroRNA-21 promotes cell proliferation and down-regulates the expression of Programmed Cell Death 4 (PDCD4) in HeLa cervical carcinoma cells. Biochem Biophys Res Commun 2009; 388(3): 539-42. http://dx.doi.org/10.1016/j.bbrc.2009.08.044 PMID: 19682430

- [98] Zhang Y, Dai Y, Huang Y, et al. Microarray profile of microribonucleic acid in tumor tissue from cervical squamous cell carcinoma without human papillomavirus. J Obstet Gynaecol Res 2009; 35(5): 842-9. http://dx.doi.org/10.1111/j.1447-0756.2009.01055.x PMID: 20149030
- [99] Deftereos G, Corrie SR, Feng Q, *et al.* Expression of mir-21 and mir-143 in cervical specimens ranging from histologically normal through to invasive cervical cancer. PLoS One 2011; 6(12): e28423.
- http://dx.doi.org/10.1371/journal.pone.0028423 PMID: 22194833
- [100] Yao T, Lin Z. MiR-21 is involved in cervical squamous cell tumorigenesis and regulates CCL20. Biochim Biophys Acta 2012; 1822(2): 248-60.
- http://dx.doi.org/10.1016/j.bbadis.2011.09.018 PMID: 22001440
  [101] Han Y, Xu GX, Lu H, *et al.* Dysregulation of miRNA-21 and their potential as biomarkers for the diagnosis of cervical cancer. Int J Clin Exp Pathol 2015; 8(6): 7131-9.
  PMID: 26261606
- [102] Li H, Sun J. Value of microRNA-21 in early diagnosis of cervical cancer. J Qiqihar Univ Med 2014; 481-2.
- [103] Liu J, Sun H, Wang X, et al. Increased exosomal microRNA-21 and microRNA-146a levels in the cervicovaginal lavage specimens of patients with cervical cancer. Int J Mol Sci 2014; 15(1): 758-73. http://dx.doi.org/10.3390/ijms15010758 PMID: 24406730
- [104] Bumrungthai S, Ekalaksananan T, Evans MF, et al. Up-regulation of miR-21 is associated with cervicitis and human papillomavirus infection in cervical tissues. PLoS One 2015; 10(5): e0127109. http://dx.doi.org/10.1371/journal.pone.0127109 PMID: 26010154
- [105] Jia W, Wu Y, Zhang Q, Gao GE, Zhang C, Xiang Y. Expression profile of circulating microRNAs as a promising fingerprint for cervical cancer diagnosis and monitoring. Mol Clin Oncol 2015; 3(4): 851-8.
- http://dx.doi.org/10.3892/mco.2015.560 PMID: 26171195 [106] Shishodia G, Verma G, Srivastava Y, Mehrotra R, Das BC, Bharti
- AC. Deregulation of microRNAs Let-7a and miR-21 mediate aberrant STAT3 signaling during human papillomavirus-induced cervical carcinogenesis: role of E6 oncoprotein. BMC Cancer 2014; 14: 996.
- http://dx.doi.org/10.1186/1471-2407-14-996 PMID: 25539644
  [107] Xu J, Zhang W, Lv Q, Zhu D. Overexpression of miR-21 promotes the proliferation and migration of cervical cancer cells *via* the inhibition of PTEN. Oncol Rep 2015; 33(6): 3108-16.
  http://dx.doi.org/10.3892/or.2015.3931 PMID: 25963606
- [108] Du G, Cao D, Meng L. miR-21 inhibitor suppresses cell proliferation and colony formation through regulating the PTEN/AKT pathway and improves paclitaxel sensitivity in cervical cancer cells. Mol Med Rep 2017; 15(5): 2713-9. http://dx.doi.org/10.3892/mmr.2017.6340 PMID: 28447761
- [109] Feng Y, Zou W, Hu C, et al. Modulation of CASC2/miR-21/PTEN pathway sensitizes cervical cancer to cisplatin. Arch Biochem Biophys 2017; 623-624: 20-30. http://dx.doi.org/10.1016/j.abb.2017.05.001 PMID: 28495512
- [110] Lin W, Feng M, Chen G, Zhou Z, Li J, Ye Y. Characterization of the microRNA profile in early-stage cervical squamous cell carcinoma by next-generation sequencing. Oncol Rep 2017; 37(3): 1477-86.
- http://dx.doi.org/10.3892/or.2017.5372 PMID: 28098890
  [111] Wei WF, Han LF, Liu D, *et al.* Orthotopic xenograft mouse model of cervical cancer for studying the role of microrna-21 in promoting lymph node metastasis. Int J Gynecol Cancer 2017; 27(8): 1587-95. http://dx.doi.org/10.1097/IGC.00000000001059
  PMID: 28945212
- [112] Xu L, Xu Q, Li X, Zhang X. MicroRNA-21 regulates the proliferation and apoptosis of cervical cancer cells *via* tumor necrosis factor-α. Mol Med Rep 2017; 16(4): 4659-63. http://dx.doi.org/10.3892/mmr.2017.7143 PMID: 28765959
- [113] Zhang Z, Wang J, Wang X, Song W, Shi Y, Zhang L. MicroRNA-21 promotes proliferation, migration, and invasion of cervical cancer through targeting TIMP3. Arch Gynecol Obstet 2018; 297(2): 433-42.
   http://dx.doi.org/10.1007/s00404.017.4508.cp.MID: 20177501

http://dx.doi.org/10.1007/s00404-017-4598-z PMID: 29177591

- [114] Zhu Y, Han Y, Tian T, et al. MiR-21-5p, miR-34a, and human telomerase RNA component as surrogate markers for cervical cancer progression. Pathol Res Pract 2018; 214(3): 374-9. http://dx.doi.org/10.1016/j.prp.2018.01.001 PMID: 29487007
- [115] Shi J, Zhang L. Clinical significance of miR-27a expression in the serum and tissue of patients with cervical squamous cell carcinomas. Chin J Clin Obstet Gynecol 2014; 49: 172-4.
- [116] Sun Y, Yang X, Liu M, Tang H. B4GALT3 up-regulation by miR-27a contributes to the oncogenic activity in human cervical cancer cells. Cancer Lett 2016; 375(2): 284-92. http://dx.doi.org/10.1016/j.canlet.2016.03.016 PMID: 26987623
- [117] Witten D, Tibshirani R, Gu SG, Fire A, Lui WO. Ultra-high throughput sequencing-based small RNA discovery and discrete statistical biomarker analysis in a collection of cervical tumours and matched controls. BMC Biol 2010; 8: 58. http://dx.doi.org/10.1186/1741-7007-8-58 PMID: 20459774
- [118] Li Y, Wang F, Xu J, et al. Progressive miRNA expression profiles in cervical carcinogenesis and identification of HPV-related target genes for miR-29. J Pathol 2011; 224(4): 484-95. http://dx.doi.org/10.1002/path.2873 PMID: 21503900
- [119] Zheng W, Liu Z, Zhang W, Hu X. miR-31 functions as an oncogene in cervical cancer. Arch Gynecol Obstet 2015; 292(5): 1083-9.

http://dx.doi.org/10.1007/s00404-015-3713-2 PMID: 25894339

- [120] Wang N, Li Y, Zhou J. miR-31 functions as an oncomir which promotes epithelial-mesenchymal transition *via* regulating BAP1 in cervical cancer. BioMed Res Int 2017; 2017: 6361420. http://dx.doi.org/10.1155/2017/6361420 PMID: 29159179
- [121] Zhou C, Shen L, Mao L, Wang B, Li Y, Yu H. miR-92a is upregulated in cervical cancer and promotes cell proliferation and invasion by targeting FBXW7. Biochem Biophys Res Commun 2015; 458(1): 63-9.

http://dx.doi.org/10.1016/j.bbrc.2015.01.066 PMID: 25623537

[122] Kong Q, Tang Z, Xiang F, et al. Diagnostic value of serum hsamir-92a in patients with cervical cancer. Clin Lab 2017; 63(2): 335-40.

http://dx.doi.org/10.7754/Clin.Lab.2016.160610 PMID: 28182350

- [123] Luo S, Li N, Yu S, Chen L, Liu C, Rong J. MicroRNA-92a promotes cell viability and invasion in cervical cancer via directly targeting Dickkopf-related protein 3. Exp Ther Med 2017; 14(2): 1227-34.
- http://dx.doi.org/10.3892/etm.2017.4586 PMID: 28810582 [124] Su Z, Yang H, Zhao M, Wang Y, Deng G, Chen R. MicroRNA-92a promotes cell proliferation in cervical cancer *via* inhibiting p21 expression and promoting cell cycle progression. Oncol Res 2017; 25(1): 137-45. http://dx.doi.org/10.3727/096504016X14732772150262 PMID: 28081742
- [125] Li ZH, Li L, Kang LP, Wang Y. MicroRNA-92a promotes tumor growth and suppresses immune function through activation of MAPK/ERK signaling pathway by inhibiting PTEN in mice bearing U14 cervical cancer. Cancer Med 2018. Epub ahead of print http://dx.doi.org/10.1002/cam4.1329 PMID: 29752775
- [126] Wang F, Liu M, Li X, Tang H. MiR-214 reduces cell survival and enhances cisplatin-induced cytotoxicity via down-regulation of Bcl2l2 in cervical cancer cells. FEBS Lett 2013; 587(5): 488-95. http://dx.doi.org/10.1016/j.febslet.2013.01.016 PMID: 23337879
- [127] Zhang X, Li F, Zhu L. Clinical significance and functions of microRNA-93/CDKN1A axis in human cervical cancer. Life Sci 2018; 209: 242-8.

http://dx.doi.org/10.1016/j.lfs.2018.08.021 PMID: 30098344

[128] Li X, Zhou Q, Tao L, Yu C. MicroRNA-106a promotes cell migration and invasion by targeting tissue inhibitor of matrix metalloproteinase 2 in cervical cancer. Oncol Rep 2017; 38(3): 1774-82.

http://dx.doi.org/10.3892/or.2017.5832 PMID: 28731196

- [129] Edatt L, Maurya AK, Raji G, Kunhiraman H, Kumar SVB. MicroRNA106a regulates matrix metalloprotease 9 in a sirtuin-1 dependent mechanism. J Cell Physiol 2018; 233(1): 238-48. http://dx.doi.org/10.1002/jcp.25870 PMID: 28233301
- [130] Wang H-W, Terinate P, Gao Y, Kalra KL. Investigation of microRNA-146a and microRNA-218 expression in cervical cancer. FASEB J 2011; 25(1): lb1-1130.6.

- [131] Hu Q, Song J, Ding B, Cui Y, Liang J, Han S. miR-146a promotes cervical cancer cell viability *via* targeting IRAK1 and TRAF6. Oncol Rep 2018; 39(6): 3015-24. http://dx.doi.org/10.3892/or.2018.6391 PMID: 29693168
- [132] Li L, Lin X, Wen W. Development and clinical primary application of SYBR Green I FQ-PCR with stem-loop RT primer to detect miR-155 [J]. Lab Med 2010; 4: 011.
- [133] Lao G, Liu P, Wu Q, et al. Mir-155 promotes cervical cancer cell proliferation through suppression of its target gene LKB1. Tumour Biol 2014; 35(12): 11933-8. http://dx.doi.org/10.1007/s13277-014-2479-7 PMID: 25155037
- [134] Zhang Y, Wang ZC, Zhang ZS, Chen F. MicroRNA-155 regulates cervical cancer via inducing Th17/Treg imbalance. Eur Rev Med Pharmacol Sci 2018; 22(12): 3719-26.
   PMID: 29949145
- [135] Zhang J, Zheng F, Yu G, Yin Y, Lu Q. miR-196a targets netrin 4 and regulates cell proliferation and migration of cervical cancer cells. Biochem Biophys Res Commun 2013; 440(4): 582-8. http://dx.doi.org/10.1016/j.bbrc.2013.09.142 PMID: 24120501
- [136] Hou T, Ou J, Zhao X, Huang X, Huang Y, Zhang Y. MicroRNA-196a promotes cervical cancer proliferation through the regulation of FOXO1 and p27Kip1. Br J Cancer 2014; 110(5): 1260-8. http://dx.doi.org/10.1038/bjc.2013.829 PMID: 24423924
- [137] Villegas-Ruiz V, Juárez-Méndez S, Pérez-González OA, et al. Heterogeneity of microRNAs expression in cervical cancer cells: over-expression of miR-196a. Int J Clin Exp Pathol 2014; 7(4): 1389-401. PMID: 24817935
- [138] Yang W, Hong L, Xu X, Wang Q, Huang J, Jiang L. LncRNA GAS5 suppresses the tumorigenesis of cervical cancer by downregulating miR-196a and miR-205. Tumour Biol 2017; 39(7): 1010428317711315. http://dx.doi.org/10.1177/1010428317711315 PMID: 28671039
- [139] Chen Z, Zhang M, Qiao Y, Yang J, Yin Q. MicroRNA-1297 contributes to the progression of human cervical carcinoma through PTEN. Artif Cells Nanomed Biotechnol 2018; 46(Suppl 2): 1120-6. http://dx.doi.org/10.1080/21691401.2018.1479711
   PMID: 29916735
- [140] Li S, Wang X, Song B, Zhou Y. Expression of miR-199b in cervical cancer tissues and its clinical significance. Chin J Cancer Prev Treat 2012; 19(17): 1335-8.
- [141] Xu LJ, Duan Y, Wang P, Yin HQ. MiR-199b-5p promotes tumor growth and metastasis in cervical cancer by down-regulating KLK10. Biochem Biophys Res Commun 2018; 503(2): 556-63. http://dx.doi.org/10.1016/j.bbrc.2018.05.165 PMID: 29807015
- [142] Zhu H, Zheng T, Yu J, Zhou L, Wang L. LncRNA XIST accelerates cervical cancer progression *via* upregulating Fus through competitively binding with miR-200a. Biomed Pharmacother 2018; 105: 789-97.
- http://dx.doi.org/10.1016/j.biopha.2018.05.053 PMID: 29909347
- [143] Xie H, Zhao Y, Caramuta S, Larsson C, Lui WO. miR-205 expression promotes cell proliferation and migration of human cervical cancer cells. PLoS One 2012; 7(10): e46990. http://dx.doi.org/10.1371/journal.pone.0046990 PMID: 23056551
- [144] Ma Q, Wan G, Wang S, Yang W, Zhang J, Yao X. Serum microRNA-205 as a novel biomarker for cervical cancer patients. Cancer Cell Int 2014; 14: 81.
- http://dx.doi.org/10.1186/s12935-014-0081-0 PMID: 25788864
   [145] Xie H, Norman I, Hjerpe A, *et al.* Evaluation of microRNA-205 expression as a potential triage marker for patients with low-grade squamous intraepithelial lesions. Oncol Lett 2017; 13(5): 3586-98. http://dx.doi.org/10.3892/ol.2017.5909 PMID: 28529583
- [146] Phuah NH, Azmi MN, Awang K, Nagoor NH. Down-regulation of microRNA-210 confers sensitivity towards 1'S-1'-Acetoxychavicol Acetate (ACA) in cervical cancer cells by targeting SMAD4. Mol Cells 2017; 40(4): 291-8.
  - http://dx.doi.org/10.14348/molcells.2017.2285 PMID: 28401751
- [147] Yamamoto N, Kinoshita T, Nohata N, et al. Tumor-suppressive microRNA-29a inhibits cancer cell migration and invasion via targeting HSP47 in cervical squamous cell carcinoma. Int J Oncol 2013; 43(6): 1855-63. http://dx.doi.org/10.3892/ijo.2013.2145 PMID: 24141696
- [148] Wu Y. Expression and significance of MiR-29a in cervical cancer tissues. Chin J Gen Practice 2013; 11: 1401-2.

- [149] Wang X, Wang HK, McCoy JP, et al. Oncogenic HPV infection interrupts the expression of tumor-suppressive miR-34a through viral oncoprotein E6. RNA 2009; 15(4): 637-47. http://dx.doi.org/10.1261/rna.1442309 PMID: 19258450
- [150] Tian Y, Zhang YZ, Chen W. MicroRNA-199a-3p and microRNA-34a regulate apoptosis in human osteosarcoma cells. Biosci Rep 2014; 34(4): e00132. http://dx.doi.org/10.1042/BSR20140084 PMID: 24957404
- [151] Ribeiro J, Marinho-Dias J, Monteiro P, et al. miR-34a and miR-125b expression in HPV infection and cervical cancer development. BioMed Res Int 2015; 2015: 304584.
- http://dx.doi.org/10.1155/2015/304584 PMID: 26180794
   [152] Chandrasekaran KS, Sathyanarayanan A, Karunagaran D. Down-regulation of HMGB1 by miR-34a is sufficient to suppress proliferation, migration and invasion of human cervical and colorectal cancer cells. Tumour Biol 2016; 37(10): 13155-66. http://dx.doi.org/10.1007/s13277-016-5261-1 PMID: 27456356
- [153] Wang JH, Zhang L, Ma YW, *et al.* microRNA-34a-upregulated retinoic acid-inducible Gene-I promotes apoptosis and delays cell cycle transition in cervical cancer cells. DNA Cell Biol 2016; 35(6): 267-79.
  - http://dx.doi.org/10.1089/dna.2015.3130 PMID: 26910120
- [154] Chen AH, Qin YE, Tang WF, Tao J, Song HM, Zuo M. MiR-34a and miR-206 act as novel prognostic and therapy biomarkers in cervical cancer. Cancer Cell Int 2017; 17: 63. http://dx.doi.org/10.1186/s12935-017-0431-9 PMID: 28615991
- [155] Li BH, Zhou JS, Ye F, et al. Reduced miR-100 expression in cervical cancer and precursors and its carcinogenic effect through targeting PLK1 protein. Eur J Cancer 2011; 47(14): 2166-74. http://dx.doi.org/10.1016/j.ejca.2011.04.037 PMID: 21636267
- [156] Cheng J, Zhao H, Yin YX. Expression of miR-101 in cervical cancer tissue and its clinical significance. Maternal Child Health Care China 2012; 7: 053.
- [157] Cui F, Li X, Zhu X, et al. MiR-125b inhibits tumor growth and promotes apoptosis of cervical cancer cells by targeting phosphoinositide 3-kinase catalytic subunit delta. Cell Physiol Biochem 2012; 30(5): 1310-8. http://dx.doi.org/10.1159/000343320 PMID: 23160634
- [158] Yu Q, Liu SL, Wang H, Shi G, Yang P, Chen XL. miR-126 Suppresses the proliferation of cervical cancer cells and alters cell sensitivity to the chemotherapeutic drug bleomycin. Asian Pac J Cancer Prev 2014; 14(11): 6569-72. http://dx.doi.org/10.7314/APJCP.2013.14.11.6569 PMID: 24377569
- [159] Huang TH, Chu TY. Repression of miR-126 and upregulation of adrenomedullin in the stromal endothelium by cancer-stromal cross talks confers angiogenesis of cervical cancer. Oncogene 2014; 33(28): 3636-47.
  - http://dx.doi.org/10.1038/onc.2013.335 PMID: 24037526
- [160] Wang C, Zhou B, Liu M, Liu Y, Gao R. miR-126-5p restoration promotes cell apoptosis in cervical cancer by targeting Bcl2l2. Oncol Res 2017; 25(4): 463-70. http://dx.doi.org/10.3727/096504016X14685034103879 PMID: 28438233
- [161] Che Y, Lu A, Liao Y. Expression of miR-143 in cervical tissue and its significance. J Med Postgra 2014; 27: 510-2.
- [162] Lin C, Huang F, Zhang YJ, Tuokan T, Kuerban G. Roles of MiR-101 and its target gene Cox-2 in early diagnosis of cervical cancer in Uygur women. Asian Pac J Cancer Prev 2014; 15(1): 45-8. http://dx.doi.org/10.7314/APJCP.2014.15.1.45 PMID: 24528073
- [163] Zhang L, Niyazi HE, Zhao HR, et al. Effects of miRNA-143 and the non-coding RNA MALAT1 on the pathogenesis and metastasis of HeLa cells. Genet Mol Res 2017; 16(1): 16. http://dx.doi.org/10.4238/gmr16019269 PMID: 28252165
- [164] Zhou M, Chen X, Wu J, He X, Ren R. MicroRNA-143 regulates cell migration and invasion by targeting GOLM1 in cervical cancer. Oncol Lett 2018; 16(5): 6393-400. http://dx.doi.org/10.3892/ol.2018.9441 PMID: 30405775
- [165] Shi M, Du L, Liu D, et al. Glucocorticoid regulation of a novel HPV-E6-p53-miR-145 pathway modulates invasion and therapy resistance of cervical cancer cells. J Pathol 2012; 228(2): 148-57. http://dx.doi.org/10.1002/path.3997 PMID: 22287315

- [166] Xing AY, Wang B, Shi DB, et al. Deregulated expression of miR-145 in manifold human cancer cells. Exp Mol Pathol 2013; 95(1): 91-7. http://dx.doi.org/10.1016/j.yexmp.2013.05.003 PMID: 23714355
- [167] Sathyanarayanan A, Chandrasekaran KS, Karunagaran D. microRNA-145 modulates epithelial-mesenchymal transition and suppresses proliferation, migration and invasion by targeting SIP1 in human cervical cancer cells. Cell Oncol (Dordr) 2017; 40(2): 119-31.

http://dx.doi.org/10.1007/s13402-016-0307-3 PMID: 27933466

- [168] Wei H, Wen-Ming C, Jun-Bo J. Plasma miR-145 as a novel biomarker for the diagnosis and radiosensitivity prediction of human cervical cancer. J Int Med Res 2017; 45(3): 1054-60. http://dx.doi.org/10.1177/0300060517709614 PMID: 28534701
- [169] Zhou X, Yue Y, Wang R, Gong B, Duan Z. MicroRNA-145 inhibits tumorigenesis and invasion of cervical cancer stem cells. Int J Oncol 2017; 50(3): 853-62. http://dx.doi.org/10.3892/ijo.2017.3857 PMID: 28112371
- [170] Qian B, Zhao L, Wang X, et al. miR-149 regulates the proliferation and apoptosis of cervical cancer cells by targeting GIT1. Biomed Pharmacother 2018; 105: 1106-16. http://dx.doi.org/10.1016/j.biopha.2018.06.075 PMID: 30021347
- [171] Du X, Lin LI, Zhang L, Jiang J. microRNA-195 inhibits the proliferation, migration and invasion of cervical cancer cells *via* the inhibition of CCND2 and MYB expression. Oncol Lett 2015; 10(4): 2639-43.
- http://dx.doi.org/10.3892/ol.2015.3541 PMID: 26622903
  [172] Wang N, Wei H, Yin D, *et al.* MicroRNA-195 inhibits proliferation of cervical cancer cells by targeting cyclin D1a. Tumour Biol 2016; 37(4): 4711-20.
- http://dx.doi.org/10.1007/s13277-015-4292-3 PMID: 26511972
  [173] Song R, Cong L, Ni G, *et al.* MicroRNA-195 inhibits the behavior of cervical cancer tumors by directly targeting HDGF. Oncol Lett 2017; 14(1): 767-75.
- http://dx.doi.org/10.3892/ol.2017.6210 PMID: 28693232
  [174] Li M, Ren CX, Zhang JM, *et al.* The Effects of miR-195-5p/MMP14 on proliferation and invasion of cervical carcinoma cells through TNF signaling pathway based on bioinformatics analysis of microarray profiling. Cell Physiol Biochem 2018; 50(4): 1398-413.
- http://dx.doi.org/10.1159/000494602 PMID: 30355924
  [175] Zhong J, Yuan H, Xu X, Kong S. MicroRNA-195 inhibits cell proliferation, migration and invasion by targeting defective in cullin neddylation 1 domain containing 1 in cervical cancer. Int J Mol Med 2018; 42(2): 779-88.
  PMID: 29750306
- [176] Zhu X, Er K, Mao C, et al. miR-203 suppresses tumor growth and angiogenesis by targeting VEGFA in cervical cancer. Cell Physiol Biochem 2013; 32(1): 64-73. http://dx.doi.org/10.1159/000350125 PMID: 23867971
- [177] Yang Z, Chen S, Luan X, et al. MicroRNA-214 is aberrantly expressed in cervical cancers and inhibits the growth of HeLa cells. IUBMB Life 2009; 61(11): 1075-82. http://dx.doi.org/10.1002/iub.252 PMID: 19859982
- [178] Qiang R, Wang F, Shi LY, et al. Plexin-B1 is a target of miR-214 in cervical cancer and promotes the growth and invasion of HeLa cells. Int J Biochem Cell Biol 2011; 43(4): 632-41. http://dx.doi.org/10.1016/j.biocel.2011.01.002 PMID: 21216304
- [179] Peng RQ, Wan HY, Li HF, Liu M, Li X, Tang H. MicroRNA-214 suppresses growth and invasiveness of cervical cancer cells by targeting UDP-N-acetyl-α-D-galactosamine:polypeptide N-acetylgalactosaminyltransferase 7. J Biol Chem 2012; 287(17): 14301-9. http://dx.doi.org/10.1074/jbc.M111.337642 PMID: 22399294
- [180] Chandrasekaran KS, Sathyanarayanan A, Karunagaran D. miR-214 activates TP53 but suppresses the expression of RELA, CTNNB1, and STAT3 in human cervical and colorectal cancer cells. Cell Biochem Funct 2017; 35(7): 464-71. http://dx.doi.org/10.1002/cbf.3304 PMID: 29023799
- [181] Peng R, Men J, Ma R, et al. miR-214 down-regulates ARL2 and suppresses growth and invasion of cervical cancer cells. Biochem Biophys Res Commun 2017; 484(3): 623-30. http://dx.doi.org/10.1016/j.bbrc.2017.01.152 PMID: 28137590

- [182] Wang JM, Ju BH, Pan CJ, et al. MiR-214 inhibits cell migration, invasion and promotes the drug sensitivity in human cervical cancer by targeting FOXM1. Am J Transl Res 2017; 9(8): 3541-57. PMID: 28861147
- [183] Yang Y, Liu Y, Li G, Li L, Geng P, Song H. microRNA-214 suppresses the growth of cervical cancer cells by targeting EZH2. Oncol Lett 2018; 16(5): 5679-86.
- http://dx.doi.org/10.3892/ol.2018.9363 PMID: 30344723 [184] Gai H. Preliminary research on the correlation between miR-218
- down-regulation and cervical cancer. Clin Res 2012; 50: 28-9.
  [185] Yuan W, Xiaoyun H, Haifeng Q, *et al.* MicroRNA-218 enhances the radiosensitivity of human cervical cancer *via* promoting radiation induced apoptosis. Int J Med Sci 2014; 11(7): 691-6. http://dx.doi.org/10.7150/ijms.8880 PMID: 24843318
- [186] Kogo R, How C, Chaudary N, et al. The microRNA-218~Survivin axis regulates migration, invasion, and lymph node metastasis in cervical cancer. Oncotarget 2015; 6(2): 1090-100. http://dx.doi.org/10.18632/oncotarget.2836 PMID: 25473903
- [187] Tang BB, Liu SY, Zhan YU, et al. microRNA-218 expression and its association with the clinicopathological characteristics of patients with cervical cancer. Exp Ther Med 2015; 10(1): 269-74. http://dx.doi.org/10.3892/etm.2015.2455 PMID: 26170947
- [188] Jiménez-Wences H, Martínez-Carrillo DN, Peralta-Zaragoza O, et al. Methylation and expression of miRNAs in precancerous lesions and cervical cancer with HPV16 infection. Oncol Rep 2016; 35(4): 2297-305.

http://dx.doi.org/10.3892/or.2016.4583 PMID: 26797462

- [189] Xu Y, He Q, Lu Y, Tao F, Zhao L, Ou R. MicroRNA-218-5p inhibits cell growth and metastasis in cervical cancer via LYN/NF-κB signaling pathway. Cancer Cell Int 2018; 18: 198. http://dx.doi.org/10.1186/s12935-018-0673-1 PMID: 30524205
- [190] Zhang J, Li S, Li Y, Liu H, Zhang Y, Zhang Q. miRNA-218 regulates the proliferation and apoptosis of cervical cancer cells *via* targeting Gli3. Exp Ther Med 2018; 16(3): 2433-41. http://dx.doi.org/10.3892/etm.2018.6491 PMID: 30210595
- Zhu L, Tu H, Liang Y, Tang D. MiR-218 produces anti-tumor effects on cervical cancer cells *in vitro*. World J Surg Oncol 2018; 16(1): 204. http://dx.doi.org/10.1186/s12957-018-1506-3 PMID: 30314496
- [192] Wang F, Li Y, Zhou J, *et al.* miR-375 is down-regulated in squamous cervical cancer and inhibits cell migration and invasion *via* targeting transcription factor SP1. Am J Pathol 2011; 179(5): 2580-8.

http://dx.doi.org/10.1016/j.ajpath.2011.07.037 PMID: 21945323

[193] Bierkens M, Krijgsman O, Wilting SM, et al. Focal aberrations indicate EYA2 and hsa-miR-375 as oncogene and tumor suppressor in cervical carcinogenesis. Genes Chromosomes Cancer 2013; 52(1): 56-68.

http://dx.doi.org/10.1002/gcc.22006 PMID: 22987659

- [194] Shen Y, Li Y, Ye F, et al. Identification of miR-23a as a novel microRNA normalizer for relative quantification in human uterine cervical tissues. Exp Mol Med 2011; 43(6): 358-66. http://dx.doi.org/10.3858/emm.2011.43.6.039 PMID: 21519184
- [195] Xu J, Li Y, Wang F, et al. Suppressed miR-424 expression via upregulation of target gene Chk1 contributes to the progression of cervical cancer. Oncogene 2013; 32(8): 976-87. http://dx.doi.org/10.1038/onc.2012.121 PMID: 22469983
- [196] Gao YL, Zhao ZS, Zhang MY, Han LJ, Dong YJ, Xu B. Long noncoding RNA PVT1 facilitates cervical cancer progression *via* negative regulating of miR-424. Oncol Res 2017; 25(8): 1391-8. http://dx.doi.org/10.3727/096504017X14881559833562 PMID: 28276314
- [197] Dong J, Wang Q, Li L, Xiao-Jin Z. Upregulation of long noncoding RNA small nucleolar RNA host gene 12 contributes to cell growth and invasion in cervical cancer by acting as a sponge for MiR-424-5p. Cell Physiol Biochem 2018; 45(5): 2086-94. http://dx.doi.org/10.1159/000488045 PMID: 29533945
- [198] Hong S, Cheng S, Songock W, Bodily J, Laimins LA. Suppression of microRNA 424 levels by human papillomaviruses is necessary for differentiation-dependent genome amplification. J Virol 2017; 91(24): e01712-7.

http://dx.doi.org/10.1128/JVI.01712-17 PMID: 28978708

[199] Luo M, Shen D, Zhou X, Chen X, Wang W. MicroRNA-497 is a potential prognostic marker in human cervical cancer and functions as a tumor suppressor by targeting the insulin-like growth factor 1 receptor. Surgery 2013; 153(6): 836-47. http://dx.doi.org/10.1016/j.surg.2012.12.004 PMID: 23453369

- [200] Tao L, Zhang CY, Guo L, et al. MicroRNA-497 accelerates apoptosis while inhibiting proliferation, migration, and invasion through negative regulation of the MAPK/ERK signaling pathway via RAF-1. J Cell Physiol 2018; 233(10): 6578-88. http://dx.doi.org/10.1002/jcp.26272 PMID: 29150931
- [201] Hildesheim A, Gonzalez P, Kreimer AR, et al. Costa Rica HPV Vaccine Trial (CVT) group. Impact of Human Papillomavirus (HPV) 16 and 18 vaccination on prevalent infections and rates of

cervical lesions after excisional treatment. Am J Obstet Gynecol 2016; 215(2): 212.e1-212.e15. http://dx.doi.org/10.1016/j.ajog.2016.02.021 PMID: 26892991

- [202] Drury RE, O'Connor D, Pollard AJ. The clinical application of microRNAs in infectious disease. Front Immunol 2017; 8: 1182. http://dx.doi.org/10.3389/fimmu.2017.01182 PMID: 28993774
- [203] Chabeda A, Yanez RJR, Lamprecht R, Meyers AE, Rybicki EP, Hitzeroth II. Therapeutic vaccines for high-risk HPV-associated diseases. Papillomavirus Res 2018; 5: 46-58. http://dx.doi.org/10.1016/j.pvr.2017.12.006 PMID: 29277575