

The complete mitochondria genome of *Chrysomya phaonis* (Seguy, 1928) (Diptera: Calliphoridae)

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ABSTRACT

Chrysomya phaonis (Seguy, 1928) is one of the blowflies of great medical and forensic importance. In this paper, we report that the entire genome of mitochondrial DNA of *C. phaonis* is 15,831 bp in length, which consists of 39 genes including 13 protein-coding genes, 24 tRNA genes, 2 mitochondrial ribosomal RNA genes, and a 992 bp non-coding A + T-rich region. The overall base compositions of A, G, C, and T are 38.79%, 9.75%, 14.15%, and 37.31%, respectively. We provide the first complete mitochondrial genome of *C. phaonis*, and should provide useful information for phylogenetic and species identification for *C. phaonis*.

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Blowflies (Diptera: Calliphoridae) are distributed worldwide, some species play important role in the mechanical transfer of disease to human beings and animals (Norris 1965; Zumpt 1965; Greenberg 1971; Greenberg 1973; Kuhlhorn 1983; Ghandour 1988). *Chrysomya phaonis* (Seguy, 1928) is one of the most important oriental species of the blowflies, *C. phaonis* distributes in China, India, Nepal, and Afghanistan (Fan 1992; Fan et al. 1997; Kurahashi et al. 1994). However, there is limited molecular biology information about this species. Viewing it is a potential medical vector and indicator in forensic science, we report here the complete mitochondrial genome of *C. phaonis* for species identification and phylogenetic analysis.

Examined sample of *C. phaonis* were obtained in Baiyi County, Tibet, China (N29°37'31.39"; E94°23'25.01") in July 2013. The studied specimen is stored in the medical vector collections of Zhongshan Entry-Exit Inspection and Quarantine technology center, and the accession number to the specimen is 20130728-232F-Baiyi. We designed 10 pairs of oligo-nucleotide primers according to the conserved regions from reported mitochondria genome sequences of its most related species *C. pinguis*.

The complete mitochondrial genome of *C. phaonis* (GenBank accession KX500359) is 15,831 bp in length, which consists of 39 genes (Table 1) including 13 protein-coding genes, 24 tRNA genes, 2 mitochondrial ribosomal RNA genes (12S rRNA and 16S rRNA), and a 992 bp non-coding A + T-rich region. The 13 protein-coding genes include seven NADH

dehydrogenase (*ND1-6* and *ND4L*), three subunits of cytochrome oxidase (*COI-III*), two subunits of ATP synthase (*ATP4* and *ATP6*), one subunit of cytochrome b (*Cytb*). Twelve of the 13 protein-coding genes were identified with ATN as start codon coding for M except for *COI*, which is similar to the former studies (Yan et al. 2014; Zhong et al. 2016).

Chrysomya phaonis not only could cause harm to human health as medical vectors (Norris 1965; Zumpt 1965; Greenberg 1971; Kuhlhorn 1983; Ghandour 1988), but also are significant in forensic science (Harvey et al. 2008). DNA typing of forensic insect specimens offers a quick and reliable alternative, so more and more researchers have started using the whole sequences of mitochondrial genome or part of the genes to identify species of Blowflies (Wells & Williams 2007; Harvey et al. 2008; Desmyter & Gosselin 2009; DeBry et al. 2013).

Here, we provide the entire genome of mitochondrial DNA of *C. phaonis*. The phylogenetic analysis of *C. phaonis* was performed by comparison with other 39 Diptera species mitochondrial genomes (Figure 1). Phylogenetic tree was generated by a neighbour-joining analysis of MEGA 6.0 program (Tamura et al. 2013) using 1000 bootstrap replicates. The N-J tree revealed that *C. phaonis* was placed mostly close to *C. pinguis*, which could not be distinguished with the common adopted COI DNA barcode sequences. So, it may provide some help for the molecular identification of *C. phaonis*, in particular to distinguish *C. phaonis* from its closely related species *C. pinguis*.

Table 1. Mitochondrial gene profile of *C. phaonis*.

Gene	Direction	Nucleotide number	Size		Anticodon	Codon	
			OL	Non		Start	Stop
tRNA ^{Ile}	J	1–64	64	4	GAT		
tRNA ^{Gln}	N	69–137	69	8	TTG		
tRNA ^{Met}	J	146–214	69		CAT		
ND2	J	215–1229	1015			ATT	T
tRNA ^{Trp}	J	1230–1297	68	8	TCA		
tRNA ^{Cys}	N	1290–1353	64	7	GCA		
tRNA ^{Tyr}	N	1361–1426	66	2	GTA		
COI	J	1425–2958	1534			TCG	T
tRNA ^{Leu(UUR)}	J	2959–3024	66	5	TAA		
COII	J	3030–3717	688			ATG	T
tRNA ^{Lys}	J	3718–3788	71	1	CTT		
tRNA ^{Asp}	J	3788–3854	67		GTC		
ATP8	J	3855–4019	165	7		ATT	TAA
ATP6	J	4013–4690	678	4		ATG	TAA
COIII	J	4695–5483	789	6		ATG	TAA
tRNA ^{Gly}	J	5490–5554	65		TCC		
ND3	J	5555–5908	354	2		ATT	TAA
tRNA ^{Ala}	J	5911–5975	65	1	TGC		
tRNA ^{Arg}	J	5975–6038	64	6	TCG		
tRNA ^{Asn}	J	6045–6110	66	1	GTT		
tRNA ^{Ser(AGN)}	J	6110–6179	70		GCT		
tRNA ^{Glu}	J	6180–6247	68	18	TTC		
tRNA ^{Phe}	N	6266–6332	67		GAA		
ND5	N	6333–8052	1720	15		ATT	T
tRNA ^{His}	N	8068–8133	66		GTG		
ND4	N	8134–9472	1339	7		ATG	T
ND4L	N	9466–9762	297	2		ATG	TAA
tRNA ^{Thr}	J	9765–9829	65		TGT		
tRNA ^{Pro}	N	9830–9895	66	2	TGG		
ND6	J	9898–10,422	525	1		ATT	TAA
Cytb	J	10,422–11,556	1135			ATG	T
tRNA ^{Ser(UCN)}	J	11,557–11,624	68	16	TGA		
ND1	N	11,641–12,579	939	10		ATA	TAA
tRNA ^{Leu(CUN)}	N	12,590–12,654	65	1	TAG		
IrRNA	N	12,656–13,983	1328				
tRNA ^{Val}	N	13,984–14,055	72		TAC		
srRNA	N	14,056–14,839	784				
A + T rich		14,840–15,831	992	66			
tRNA ^{Ile}	J	14,929–14,994	66		GAT		

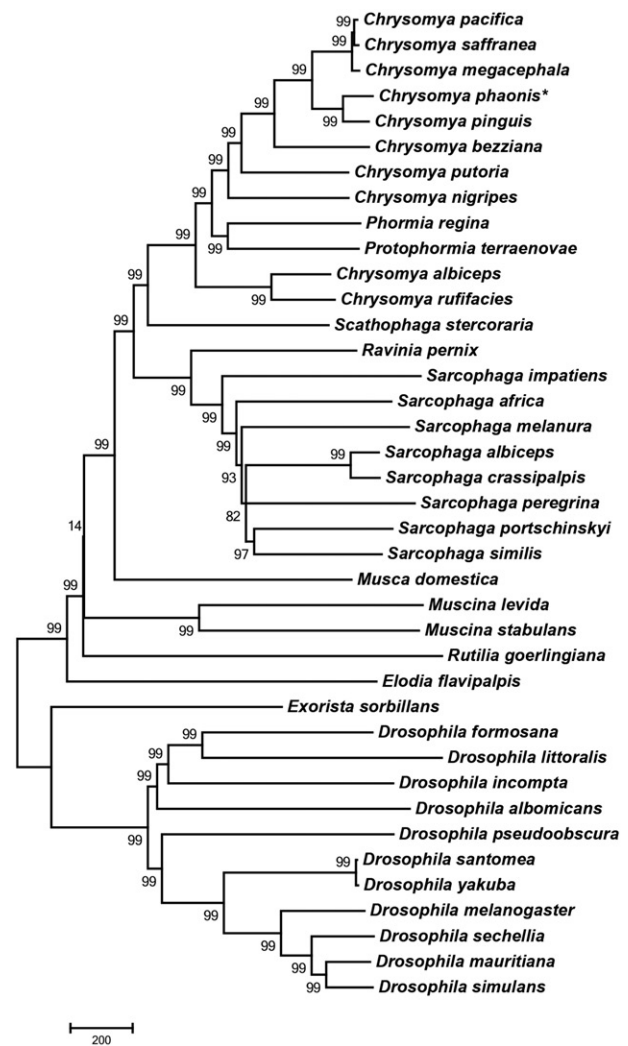


Figure 1. The neighbour-joining (NJ) tree of *C. phaonis* with other 39 Diptera species based on mitochondrial genomes. For each node, the bootstrap support was calculated using 1000 replicates. GenBank accession numbers of mitochondrial genomes used in this phylogeny analysis were listed: *C. albiceps* (NC_019631.1); *C. bezziana* (JX913737.1); *C. megacephala* (KT272775.1); *C. nigripes* (KT444441.1); *C. pacifica* (KP861632.1); *C. pinguis* (KM244730.1); *C. putoria* (AF352790.1); *C. saffranaea* (JX913742.1); *C. saffranaea* (JX913742.1); *Protophormia terraenovae* (JX913743.1); *Sarcophaga africa* (KM881633.1); *S. albiceps* (NC_028413.1); *S. crassipalpis* (KP861920.1); *S. impatiens* (NC_017605.1); *S. melanura* (NC_026112.1); *S. peregrina* (NC_023532.1); *S. portschinskyi* (NC_025574.1); *S. similis* (NC_025573.1); *Scathophaga stercoraria* (KM200724.1); *Musca domestica* (KT444442.1); *Muscina stabulans* (NC_029487.1); *M. stabulans* (NC_026292.1); *Elodia flavipalpis* (NC_018118.1); *Exorista sorbillans* (HQ322500.1); *Ravinia pernix* (NC_026196.1); *Rutilla goerlingiana* (NC_019640.1); *Phormia regina* (KC005712.1); *Drosophila albomicans* (NC_027937.1); *Drosophila formosana* (NC_028518.1); *D. incompta* (NC_025936.1); *D. littoralis* (FJ447340.1); *D. mauritiana* (AF200831.1); *D. mauritiana* (NC_005779.1); *D. melanogaster* (KT174474.1); *D. pseudoobscura* (NC_018348.1); *D. santomea* (KF824869.1); *D. sechellia* (AF200832.1); *D. simulans* (AY518674.1); *D. yakuba* (KF824899.1).

Disclosure statement

The authors report no conflicts of interest. The authors alone are responsible for the content and writing of the paper.

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