

The synthesis of new potential photosensitizers. 1. Mono-carboxylic acid derivatives of tetraphenylporphyrin

Grzegorz Zięba · Marcin Rojkiewicz · Violetta Kozik ·
Krystyna Jarzembek · Anna Jarczyk · Aleksander Sochanik ·
Piotr Kuś

Received: 5 October 2010 / Accepted: 12 July 2011 / Published online: 24 August 2011
© The Author(s) 2011. This article is published with open access at Springerlink.com

Abstract A series of mono-alkylcarboxylic acid derivatives of tetraphenylporphyrin have been prepared. All the porphyrins were completely characterized by use of mass, ^1H NMR, UV–visible, and fluorescence spectroscopy. Experimental $\log P$ were determined by use of reversed-phase thin-layer chromatography with use of $\log P_{\text{Rekker}}$. These porphyrins are potential photosensitizers in photodynamic therapy.

Keywords Porphyrins · Sensitizers · Hydrophobicity · Fluorescence spectroscopy

Introduction

Hematoporphyrin (**1**), the first compound that was tried in photodynamic therapy (PDT), contains, besides the basic porphyrin ring, two carboxyethylene groups. Photofrin, currently the most frequently used porphyrin-type compound in anticancer therapy, was obtained from hematoporphyrin (**1**) (Fig. 1) [1]. Photofrin molecules contain carboxyl (carboxyethylene) groups which give the compound its hydrophobic and hydrophilic properties. Photofrin preparations are, however, a mixture of several porphyrin compounds. Numerous research teams around

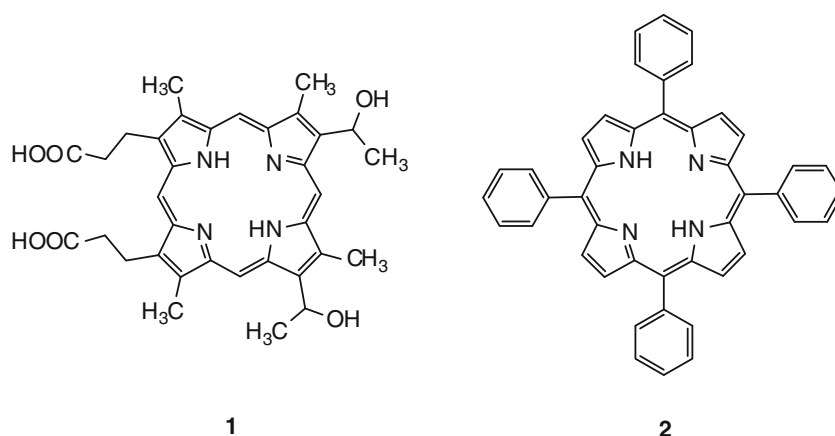
the world have been trying to obtain novel compounds with precisely determined structure and controlled biological properties [2–4]. Several of these compounds are derivatives of tetraphenylporphyrin (**2**) (tetraarylporphyrin, Fig. 1). The aryl fragments in these porphyrins are modified by substituting hydrogen atoms with various groups altering the hydrophobic–hydrophilic character of the whole molecule, most often without changing its photochemical properties.

Simple tetraarylporphyrins are highly soluble in slightly polar solvents, most often halogenated or DMF; these however cannot be used in biological investigations except at the stage of preparing photosensitizer–carrier systems, e.g. liposomes. Most synthetic porphyrins are not very soluble in simple alcohols. Some porphyrins are soluble in water which makes them suitable for basic biological studies. Introduction of strongly polar groups, for example ammonium or sulfone groups, enables dissolution of tetraarylporphyrins in water. Suitably modified porphyrins can be incorporated into lipid membranes, micelles, or liposomes; in this form they are delivered into cells [5–13]. Porphyrin carriers of this type are very promising, because most of the porphyrins obtained synthetically are strongly hydrophobic which makes them useless in investigations of direct PDT or other therapeutic applications. Derivatives with acidic moieties at the ends of alkyl chains have not yet been studied for their applicability in photodynamic therapy. Compounds with alkyl chains of different lengths separating the porphyrin fragment from the carboxyl group were chosen for study. Hydrophobic–hydrophilic character is responsible for a variety of the properties of organic compounds, for example solubility, permeability through semi-permeable membranes (for example cell membranes), and ability to anchor in lipid layers. Most importantly, for PDD and PDT it is responsible for the manner of transport

G. Zięba · M. Rojkiewicz · V. Kozik · K. Jarzembek ·
A. Jarczyk · P. Kuś (✉)
Department of Chemistry, University of Silesia,
9, Szkolna Street, 40-006 Katowice, Poland
e-mail: pkus@ich.us.edu.pl

A. Sochanik
Department of Molecular Biology, Maria Skłodowska-Curie
Memorial Cancer Centre and Institute of Oncology,
44-100 Gliwice, Poland

Fig. 1 Hematoporphyrin and tetraphenylporphyrin



of photosensitizer molecules into the living organism. The easiest way to verify the assumed properties of a compound is by synthesis of the compound.

In search of novel potential photosensitizers we obtained a series of carboxyalkoxy tetraphenylporphyrin derivatives **4–8** (Fig. 2). Carboxyl groups are attached to one of the phenyl rings of tetraphenylporphyrin via alkyl chains of different lengths. Previously, compounds with $n = 1$ and $n = 3$ were obtained as intermediates during the synthesis of more complex tetraphenylporphyrin derivatives. In particular, cholesterol derivatives of carboxymethylporphyrin ($n = 1$) were synthesized and their gelator properties described [14]. Butyric acid ($n = 3$) was attached to arylporphyrin via an ether bond in order to obtain a porphyrin with an easily modifiable fragment (in this case a COOH group). Next, it was substituted using either lipophilic guanosine in order to examine the self-assembly of this group [15] or with a tyrosine group when obtaining modified TiO₂ clusters in which photoinduced electron transfer occurred [16]. Amide derivatives of this compound were also obtained as potential agents for photodynamic therapy of cancers [17]. Glycosidic porphyrins obtained from the butyric acid derivative of tritolyphenylporphyrin were also designed as potential PDT agents [18].

Results and discussion

To the best of our knowledge no derivatives with carboxyl groups at the distal ends of alkyl chains have been investigated for their usefulness in PDT. We thus decided to investigate compounds with alkyl chains of different lengths separating the porphyrin fragment from the carboxyl group. To synthesize alkyl precursors with $n = 4, 5, 6, 7,$ and 11 we used the classic method of obtaining carboxyl derivatives starting from suitable dibromoalkyl derivatives and malonic acid diethyl ester [19, 20]. The yields of the obtained diethyl (bromoalkyl)malonates were not very high but this could be

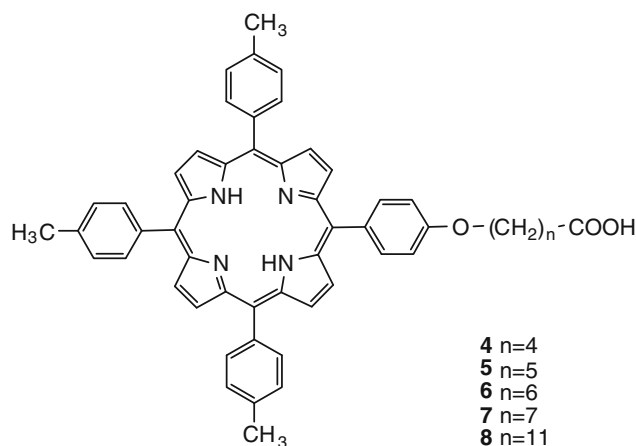
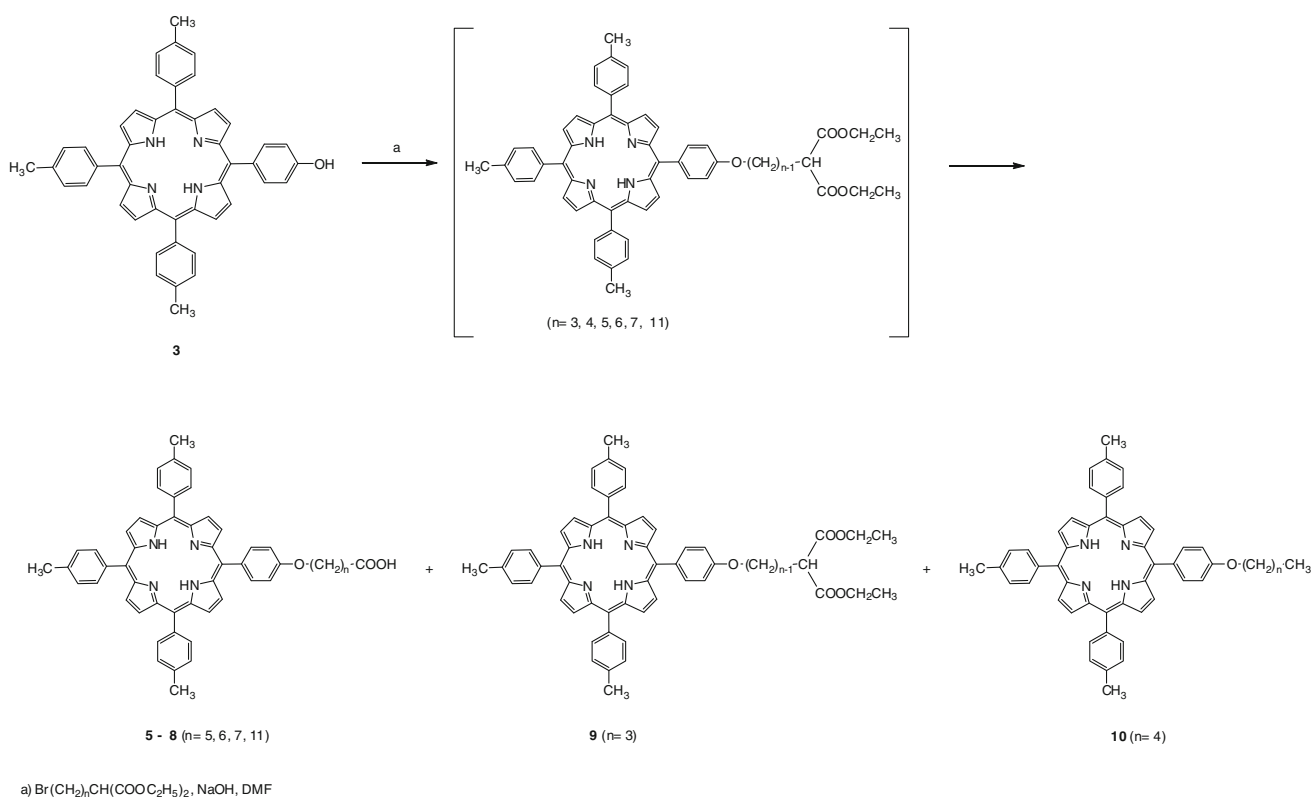


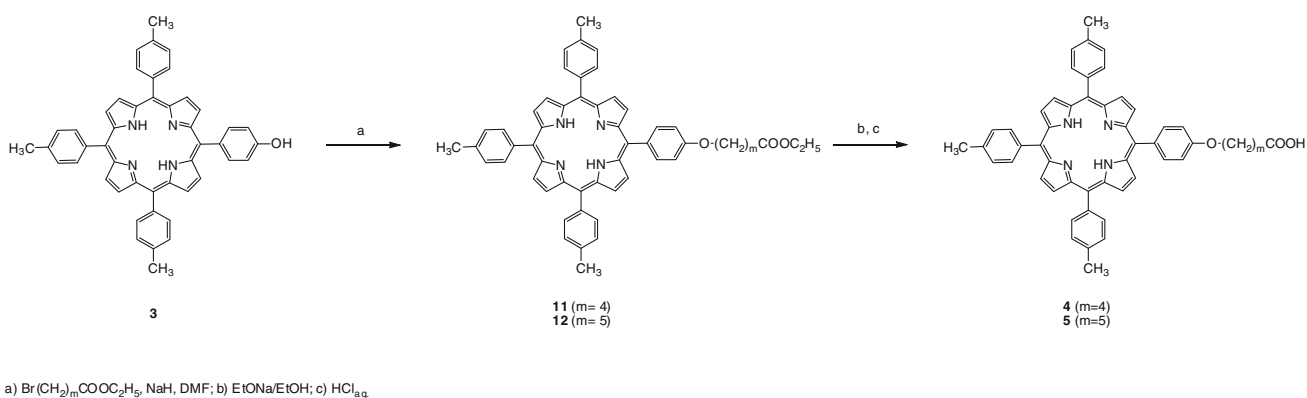
Fig. 2 Synthesized compounds

expected, especially for shorter alkyl chains. These compounds quite easily undergo intramolecular cyclization reaction under the reaction conditions [21, 22]. Hydroxyphenyltritolymporphyrin (**3**) was obtained according to Ref. [23]. Diethyl (ω -bromoalkyl)malonates were coupled to a hydroxyl fragment of porphyrin using NaOH in DMF (Scheme 1). Under the reaction conditions the monocarboxy derivatives were obtained directly; in the presence of NaOH hydrolysis of the ester took place, whereas DMF distillation at an elevated temperature led to simultaneous decarboxylation of the malonic acid derivative. Exceptional for this synthetic route were the reactions of diethyl (3-bromopropyl)malonate leading to a malonic acid derivative and of diethyl (4-bromobutyl)malonate which gave a pentyloxy derivative of tetraphenylporphyrin. The properties of the obtained derivative were compared with those of the similar compound obtained by direct alkylation of the hydroxyphenyl derivative of porphyrin with 1-bromopentane.

Porphyrins **4** and **5** were obtained by a different method, direct reaction of a suitable ethyl ester of ω -bromoalkyl acid with hydroxyl derivative of porphyrin (Scheme 2). The esters were hydrolysed to the corresponding carboxylic derivatives.



Scheme 1



Scheme 2

All of the obtained porphyrins **4–8** had characteristic common bands in the visible part of the UV–visible spectrum. They have very strong absorption peaks centred at 422 nm (Soret band) and four other peaks centred at 519, 555, 594, and 650 nm (Q bands). The spectra clearly identify the porphyrinic chromophores of compounds **4–8**. The minor differences between the absorption maxima of the porphyrins implied their photosensitizing properties are also comparable. UV–visible data for all the compounds are listed in Table 1.

Fluorescence spectra of all porphyrins and TPP as a reference are similar in shape, and consist of two bands at

654 and 720 nm. From Table 2 we can see that the quantum yields of all the porphyrins are almost identical with that of the reference (TPP). The photostability of compounds **4–9**, **11**, and **12** was determined for their chloroform solutions at 25 °C. The results presented in Table 3 show good photostability of the compounds which indicates their potential usefulness for PDT.

Mass spectrometric analysis of all the synthesized porphyrins was performed by use of the ESI–MS technique. The mass spectra (positive mode) of all the porphyrins contained a base peak corresponding to the intact porphyrin—no fragment ions were detected. For porphyrins **6–8**

Table 1 UV–visible spectra of compounds **4–9**, **11**, and **12** [$\lambda(\log \varepsilon)$] in dichloromethane solution

Derivative	Soret/nm	Q _{IV} /nm	Q _{III} /nm	Q _{II} /nm	Q _I /nm
4	422 (5.38)	517 (3.95)	554 (3.77)	592 (3.51)	650 (3.63)
5	422 (5.54)	518 (4.04)	555 (3.84)	592 (3.57)	651 (3.58)
6	422 (5.57)	519 (4.10)	555 (3.86)	594 (3.60)	650 (3.58)
7	422 (5.58)	519 (4.17)	555 (3.94)	594 (3.67)	650 (3.66)
8	422 (5.57)	519 (4.13)	555 (3.90)	594 (3.63)	650 (3.67)
9	422 (5.70)	519 (4.30)	554 (4.07)	593 (3.81)	649 (3.77)
11	422 (5.46)	518 (4.05)	555 (4.05)	594 (3.55)	650 (3.52)
12	422 (5.11)	517 (3.67)	554 (3.47)	594 (3.21)	651 (3.31)

Table 2 Fluorescence emission maxima, wavelengths (λ_{\max}), and fluorescence quantum yields (Φ_F) for compounds **4–9** and **11** and tetraphenylporphyrin (TPP) as reference; $\log P_{\text{Rekker}}$ and $\log P_{\text{exp}}$ for compounds **5–9** and **11**

Photosensitizer	$\lambda_{\max}1/\text{nm}$	$\lambda_{\max}2/\text{nm}$	Φ_F	$\log P_{\text{Rekker}}$	$\log P_{\text{exp}}$
4	653	720	0.11	–	–
5	652	718	0.11	9.08	8.25
6	654	719	0.13	9.60	10.79
7	654	721	0.11	10.12	12.05
8	654	720	0.11	12.19	18.48
9	654	720	0.11	9.39	8.85
11	654	720	0.11	9.55	8.92
TPP	652	718	0.1	–	–

Table 3 Photostability of compounds **4–9**, **11**, and **12**

Photosensitizer	0	15	30	45	60	75	90
4	100	99	99	98	97	97	95
5	100	99	96	96	95	95	95
6	100	100	99	99	97	96	96
7	100	100	99	98	98	97	97
8	100	99	98	99	96	95	95
9	100	98	98	97	96	96	96
11	100	99	99	98	97	97	95
12	100	100	98	97	96	96	96

For irradiation, visible light from two 36-W fluorescent lamps was used and the light intensity was adjusted to 16.5 W/m². Photostability was determined by monitoring the intensity of the Soret band after a defined irradiation time (0–90 min)

and **10** the most intense ions appeared when sodium cations were present in the solutions.

Lipophilicity is an important molecular descriptor very often correlated with the biological activity of drugs and other compounds. It is characterized, for example, by the logarithm of a partition coefficient ($\log P$). We determined $\log P$ values of all the compounds by a TLC method. Table 2 shows values calculated by use of the $\log P_{\text{Rekker}}$ method and the experimental $\log P$ values. $\log P_{\text{exp}}$ was calculated by use of the equation [24]:

$$\log P_{\text{exp}} = \frac{\log P_{\text{Rekker}}}{2(R_{F_1} + R_{F_2})}$$

where R_{F_1} and R_{F_2} are retardation factors for methanol–chloroform 8:2 and 7:3 (v/v), respectively, as mobile phase. $\log P_{\text{Rekker}}$ was calculated, in compliance with the constant fragmentations method proposed by Rekker, by use of the equation [25, 26]:

$$\log P_{\text{Rekker}} = \sum_1^n a_n f_n$$

where f is a constant representing the lipophilicity of a defined structural fragment and a describes how many times a specific fragment is repeated in a molecule. Experimental results show that additional methylene units in the carbon chain result in an increase of the compound's lipophilicity. We obtained $\log P_{\text{exp}} = 1.49$ for tetrakis(3-hydroxyphenyl)porphyrin as reference compound. The literature shake-flask $\log P$ value for this compound is 2.02 [27].

Solubility in polar solvents is a serious obstacle for the synthesized compounds. As could be expected, elongation of the alkyl chain led to increased hydrophobicity of the whole molecule. As a result, this often leads to aggregation and precipitation of porphyrins from solutions used for biological investigations. Currently, all these compounds are undergoing biological investigation using liposomal formulations which can substantially reduce the extent of photosensitizer aggregation and can be valuable carriers and delivery systems.

Conclusions

Novel mono-carboxyalkoxyphenyl derivatives of porphyrin were synthesized and their physicochemical properties were examined. Quantum yields of all the obtained porphyrins are almost identical with that of the reference compound (TPP). Information about the lipophilicity of the examined compounds can prove helpful in further research on their potential medical uses. Their balanced hydrophobic–hydrophilic properties make it

possible to place them in systems (e.g. micelles, liposomes) similar to natural biological membranes.

Experimental

All chemical reagents were purchased from Aldrich or Acros and were used without further purification. ^1H NMR and ^{13}C NMR spectra were recorded in CDCl_3 using a Varian spectrometer (400 MHz) with TMS as reference. UV–visible spectra were recorded in dichloromethane solutions using a Genesys 6 (ThermoSpectronic) spectrophotometer. Fluorescence spectra of the samples were recorded on a Varian Eclipse Cary-fluorescence spectrophotometer. ESI MS spectra were acquired using an LCQ DUO Finnigan Thermoquest or a Varian MS-500 instrument. Porphyrin solutions were directly injected into the ion source by use of a syringe pump.

General procedure for diethyl (ω -bromoalkyl)malonates

Diethyl malonate (0.05 mol), 1, ω -dibromoalkane (0.05 mol), 0.375 g tetrabutylammonium iodide (1 mmol), and 15 cm^3 toluene were placed in a flask. A solution of 17.25 g K_2CO_3 in 15 cm^3 water was added. The obtained mixture was stirred and heated to reflux for 20 h. After cooling to room temperature the solid was removed by suction. The organic phase was separated and the aqueous phase was extracted with toluene (3 \times 10 cm^3). The combined organic layers were washed with water and dried over anhydrous magnesium sulfate. After evaporating the solvent the product was distilled under reduced pressure.

Diethyl (3-bromopropyl)malonate

Yield 16%; b.p.: 140–145 $^\circ\text{C}$ (7–8 mbar), lit. 135 $^\circ\text{C}$ (4 mbar) [28], 121–123 $^\circ\text{C}$ (2 mbar) [19].

Diethyl (4-bromobutyl)malonate

Yield 16%; b.p.: 104–106 $^\circ\text{C}$ (3–4 mbar), lit. 133–136 $^\circ\text{C}$ (2 mbar) [29].

Diethyl (5-bromopentyl)malonate

Yield 21%; b.p.: 178–180 $^\circ\text{C}$ (8 mbar), lit. 153 $^\circ\text{C}$ (3 mbar) [30]; 159–161 $^\circ\text{C}$ (3 mbar) [31].

Diethyl (6-bromohexyl)malonate

Yield 17%; b.p.: 175–178 $^\circ\text{C}$ (9 mbar), lit. 185–187 $^\circ\text{C}$ (10 mbar) [32]; 193 $^\circ\text{C}$ (12 mbar) [31].

Diethyl (10-bromodecyl)malonate

Yield 20%; b.p.: 222–224 $^\circ\text{C}$ (7–8 mbar), lit. 123 $^\circ\text{C}$ (0.02 mbar) [20, 33].

General procedure for 5-[4-(carboxyalkyloxy)phenyl]-10,15,20-tritolylporphyrins

5-(4-Hydroxyphenyl)-10,15,20-tritolylporphyrin (0.24 mmol), solid NaOH (0.8 mmol), and 20 cm^3 DMF were placed in a round-bottomed flask. The mixture was stirred at room temperature until NaOH was dissolved. Then 2 mmol diethyl (ω -bromoalkyl)malonate were added and the resulting mixture was stirred at room temperature for 24 h. After evaporation of the DMF under reduced pressure 50 cm^3 dichloromethane was added to the residue. The resulting solution was washed with water (5 \times 50 cm^3) and dried over MgSO_4 . After evaporation of the solvent the crude product was washed with petroleum ether (20 cm^3) and chromatographed on silica with dichloromethane.

5-[4-(5-Carboxypentyloxy)phenyl]-10,15,20-tritolylporphyrin (5, $\text{C}_{53}\text{H}_{46}\text{N}_4\text{O}_3$)

Yield 14%; ^1H NMR (400 MHz, CDCl_3): δ = 8.86 (m, 8H), 8.10 (d, 2H, J = 8.4 Hz), 8.09 (d, 6H, J = 8.0 Hz), 7.54 (d, 6H, J = 8.0 Hz), 7.25 (d, 2H, J = 8.4 Hz), 4.24 (t, 2H), 2.70 (s, 9H), 2.51 (t, 2H), 2.01 (q, 2H), 1.86 (q, 2H), 1.72 (q, 2H), -2.76 (bs, 2H) ppm; ESI–MS: m/z = 787 (100) $[\text{M} + \text{H}]^+$.

5-[4-(6-Carboxyhexyloxy)phenyl]-10,15,20-tritolylporphyrin (6, $\text{C}_{54}\text{H}_{48}\text{N}_4\text{O}_3$)

Yield 57%; ^1H NMR (400 MHz, CDCl_3): δ = 8.86 (m, 8H), 8.11 (d, 2H, J = 8.4 Hz), 8.10 (d, 6H, J = 8.0 Hz), 7.55 (d, 6H, J = 8.0 Hz), 7.27 (d, 2H, J = 8.4 Hz), 4.27 (t, 2H), 3.55 (t, 2H), 2.71 (s, 9H), 2.10–2.00 (m, 4H), 1.77 (q, 2H), 1.53 (q, 2H), -2.76 (bs, 2H) ppm; ESI–MS: m/z = 823 (100) $[\text{M} + \text{Na}]^+$.

5-[4-(7-Carboxyheptyloxy)phenyl]-10,15,20-tritolylporphyrin (7, $\text{C}_{55}\text{H}_{50}\text{N}_4\text{O}_3$)

Yield 14%; ^1H NMR (400 MHz, CDCl_3): δ = 8.85 (m, 8H), 8.10 (d, 2H, J = 8.4 Hz), 8.09 (d, 6H, J = 8.0 Hz), 7.54 (d, 6H, J = 8.0 Hz), 7.27 (d, 2H, J = 8.4 Hz), 4.24 (t, 2H), 3.50 (t, 2H), 2.70 (s, 9H), 1.99 (q, 4H), 1.66 (q, 4H), 1.52 (m, 2H), -2.76 (bs, 2H) ppm; ESI–MS: m/z = 837 (100) $[\text{M} + \text{Na}]^+$.

5-[4-(11-Carboxyundecyloxy)phenyl]-10,15,20-tritolylporphyrin (8, $\text{C}_{59}\text{H}_{58}\text{N}_4\text{O}_3$)

Yield 18%; ^1H NMR (400 MHz, CDCl_3): δ = 8.85 (m, 8H), 8.10 (d, 2H, J = 8.4 Hz), 8.09 (d, 6H, J = 8.0 Hz), 7.55 (d, 6H, J = 8.0 Hz), 7.27 (d, 2H, J = 8.4 Hz), 4.25 (t, 2H), 3.43 (t, 2H), 2.70 (s, 9H), 2.00 (q, 2H), 1.90 (q, 2H), 1.62 (q, 2H), 1.53–1.20 (m, 12H), -2.76 (bs, 2H) ppm; ESI–MS: m/z = 894 (100) $[\text{M} + \text{Na}]^+$.

5-[4-[4,4-Bis(ethoxycarbonyl)butyloxy]phenyl]-10,15,20-tritolyldiporphyrin (**9**, C₅₇H₅₂N₄O₅)

Yield 27%; ¹H NMR (400 MHz, CDCl₃): δ = 8.84 (m, 8H), 8.10 (d, 2H, *J* = 8.4 Hz), 8.09 (d, 6H, *J* = 7.6 Hz), 7.54 (d, 6H, *J* = 7.6 Hz), 7.24 (d, 2H, *J* = 8.4 Hz), 4.27 (q, 4H), 3.56 (t, 1H), 2.69 (s, 9H), 2.27 (q, 2H), 2.04 (q, 2H), 1.51 (bs, 2H), 1.33 (t, 6H), -2.76 (bs, 2H) ppm; ESI-MS: *m/z* = 873 (100) [M + H]⁺.

5-(4-Pentyloxyphenyl)-10,15,20-tritolyldiporphyrin (**10**, C₅₂H₄₆N₄O)

Yield 14%; ¹H NMR (400 MHz, CDCl₃): δ = 8.87 (m, 8H), 8.11 (d, 2H, *J* = 8.4 Hz), 8.10 (d, 6H, *J* = 8.0 Hz), 7.55 (d, 6H, *J* = 8.0 Hz), 7.27 (d, 2H, *J* = 8.4 Hz), 4.23 (t, 2H), 2.71 (s, 9H), 1.99 (q, 2H), 1.62 (q, 2H), 1.53 (q, 2H), 1.05 (t, 3H), -2.73 (bs, 2H) ppm; ESI-MS: *m/z* = 765 (100) [M + Na]⁺, 743 (100) [M + H]⁺; UV-Vis (CH₂Cl₂): λ_{max} (log ε) = 422 (5.54), 519 (4.13), 555 (3.89), 593 (3.63), 650 (3.60) nm.

General procedure for 5-[4-[ω-(ethoxycarbonyl)alkyloxy]phenyl]-10,15,20-tritolyldiporphyrins

5-(4-Hydroxyphenyl)-10,15,20-tritolyldiporphyrin (100 mg, 0.15 mmol), 5 mg NaH (0.2 mmol), and 50 cm³ DMF were placed in a round-bottomed flask and stirred at room temperature for 30 min. Then 1 mmol ethyl ω-bromoalkylcarboxylate was added and the resulting mixture was stirred at 80 °C for 48 h. After evaporation of the DMF under reduced pressure 50 cm³ dichloromethane was added to the residue. The resulting solution was washed with water (3 × 50 cm³) and dried over MgSO₄. After evaporation of the solvent the crude product was washed with petroleum ether (20 cm³) and chromatographed on silica with dichloromethane-ethanol 30:1 (v/v).

5-[4-[4-(Ethoxycarbonyl)butyloxy]phenyl]-10,15,20-tritolyldiporphyrin (**11**, C₅₄H₄₈N₄O₃)

Yield 33%; ¹H NMR (400 MHz, CDCl₃): δ = 8.85 (m, 8H), 8.11 (d, 2H, *J* = 8.0 Hz), 8.09 (d, 6H, *J* = 7.6 Hz), 7.55 (d, 6H, *J* = 7.6 Hz), 7.27 (d, 2H, *J* = 8.0 Hz), 4.28 (t, 2H), 4.21 (q, 2H), 2.71 (s, 9H), 2.52 (q, 2H), 2.03 (m, 4H), 1.33 (t, 3H), -2.76 (bs, 2H) ppm; ¹³C NMR (125 MHz, CDCl₃): δ = 173.19, 155.75, 139.33, 137.32, 135.68, 134.57, 134.53, 130.89, 127.42, 120.09, 120.05, 113.67, 60.44, 33.36, 33.03, 32.04, 23.55, 21.53, 14.26 ppm; ESI-MS: *m/z* = 801 (100) [M + H]⁺.

5-[4-[5-(Ethoxycarbonyl)pentyloxy]phenyl]-10,15,20-tritolyldiporphyrin (**12**, C₅₅H₅₀N₄O₃)

Yield 29%; ¹H NMR (400 MHz, CDCl₃): δ = 8.85 (m, 8H), 8.11 (d, 2H, *J* = 8.0 Hz), 8.10 (d, 6H, *J* = 7.6 Hz), 7.55 (d, 6H, *J* = 7.6 Hz), 7.27 (d, 2H, *J* = 8.0 Hz), 4.26 (t, 2H), 4.19 (q, 2H), 2.71 (s, 9H), 2.44 (t, 2H), 2.01 (q, 2H),

1.84 (q, 2H), 1.69 (q, 2H), 1.31 (t, 3H), -2.76 (bs, 2H) ppm; ¹³C NMR (125 MHz, CDCl₃): δ = 173.75, 158.89, 139.36, 137.32, 135.61, 134.54, 129.60, 129.07, 127.43, 120.10, 112.72, 67.98, 60.35, 34.40, 29.23, 25.90, 24.90, 21.55, 14.35 ppm; ESI-MS: *m/z* = 815 (100) [M + H]⁺.

General procedure for 5-(4-carboxyalkyloxyphenyl)-10,15,20-tritolyldiporphyrins from ethyl ester

5-[4-(ω-Ethoxycarbonylalkyloxy)phenyl]-10,15,20-tritolyldiporphyrin (0.03 g, ~0.04 mmol) dissolved in 10 cm³ dichloromethane was added to a solution of sodium ethoxide (1 g sodium dissolved in 10 cm³ ethanol). One drop of water was added to the mixture. The reaction mixture was stirred for 3 h at room temperature. The solution was then acidified to pH 7 with 1 M HCl, washed with water (3 × 30 cm³), dried over Na₂SO₄, and concentrated. Purification by column chromatography on silica gel (elution with CH₂Cl₂-EtOH, 25:1 (v/v)) afforded the product.

5-[4-(4-Carboxybutyloxy)phenyl]-10,15,20-tritolyldiporphyrin (**4**, C₅₂H₄₄N₄O₃)

Yield 24%; ¹H NMR (400 MHz, CDCl₃): δ = 8.85 (d, 8H), 8.09 (d, 8H, *J* = 8.0 Hz), 7.54 (d, 6H, *J* = 8.0 Hz), 7.24 (d, 2H, *J* = 8.0 Hz), 4.26 (t, 2H), 2.76 (s, 9H), 2.59 (q, 2H), 2.03 (q, 2H), 1.62 (m, 2H), -2.76 (bs, 2H) ppm; ESI-MS: *m/z* = 773 (100) [M + H]⁺.

5-[4-(5-Carboxypentyloxy)phenyl]-10,15,20-tritolyldiporphyrin (**5**) was obtained by this method in 19% yield.

Acknowledgments Financial support by the Polish State Committee for Scientific Research (grant no. R 05 043 03) is gratefully acknowledged. The work of two authors (Marcin Rojkiewicz and Grzegorz Zięba) was partially supported by a PhD scholarship within the framework of the "University as a Partner of the Economy Based on Science" (UPGOW) project, subsidized by the European Social Fund (EFS) of the European Union.

Open Access This article is distributed under the terms of the Creative Commons Attribution Noncommercial License which permits any noncommercial use, distribution, and reproduction in any medium, provided the original author(s) and source are credited.

References

- Schaffer M, Schaffer PM, Hofstetter A, Dühmke E, Jori G (2002) Photochem Photobiol Sci 1:438
- Nyman ES, Hynninen PH (2004) J Photochem Photobiol B 73:1
- Bonnett R (1995) Chem Soc Rev 24:19
- Sternberg ED, Dolphin D, Brückner C (1998) Tetrahedron 54:4151
- Ballut S, Makky A, Loock B, Michael J-P, Maillard P, Rosilio V (2009) Chem Commun 8:224
- Ricchelli F (1995) J Photochem Photobiol B 29:109
- Voszka I, Galántai R, Maillard P, Csik G (1999) J Photochem Photobiol B 52:92

8. Tsuchida E, Komatsu T, Arai K, Nishide H (1993) *Chem Commun* 8:730
9. Nagata N, Kugimiya S, Kobuke Y (2001) *Chem Commun* 8:689
10. Lavi A, Weitman H, Holmes RT, Smith KM, Ehrenberg B (2002) *Biophys J* 82:2101
11. Postigo F, Mora M, De Madariaga MA, Nonell S, Sagristá ML (2004) *Int J Pharm* 278:239
12. Voszka I, Szabó Z, Csik G, Maillard P, Gróf P (2005) *J Photochem Photobiol B* 79:83
13. Ben-Dror S, Bronshtein I, Wiehe A, Röder B, Senge MO, Ehrenberg B (2006) *J Photochem Photobiol* 82:695
14. Tian HJ, Inoue K, Yoza K, Ishi-I T, Shinkai S (1998) *Chem Lett* 871
15. Masiero S, Gottarelli G, Pieraccini S (2000) *Chem Commun* 1995
16. Yu J, Chen J, Wang X, Zhang B, Cao Y (2003) *Chem Commun* 1856
17. Sol V, Lamarche F, Enache M, Garcia G, Granet R, Guilloton M, Blais JC, Krausz P (2006) *Bioorg Med Chem* 14:1364
18. Chaleix V, Sol V, Huang Y-M, Guilloton M, Granet R, Blais JC, Krausz P (2003) *Eur J Org Chem* 1486
19. Ikegami S, Asai T, Tsuneoka K, Matsumara S, Akaboshi S (1974) *Tetrahedron* 30:2087
20. Matsubara H, Seto K, Tabuchi H, Imazaki H, Takahashi S (1993) *Bull Chem Soc Jpn* 66:578
21. Cort AD, Mandolini L, Masci B (1983) *J Org Chem* 48:3979
22. Casadei MA, Galli C, Mandolini L (1981) *J Org Chem* 46:3127
23. Little RG, Anton JA, Loach PA, Ibers JA (1975) *J Heterocycl Chem* 12:343
24. Nistrój A (2007) *J Planar Chromatogr-Mod TLC* 20:483
25. Rekker RF, Mannhold R (1992) In: *Calculation of drug lipophilicity*. VCH, Weinheim
26. Mannhold R, Rekker RF (2000) *Perspect Drug Discov* 18:1
27. Serra AC, Pineiro M, Rocha Gonsalves AM, Abrantes M, Laranjo M, Santos AC, Botelho MF (2008) *J Photochem Photobiol B* 92:59
28. van Tamelen EE, Foltz RL (1969) *J Am Chem Soc* 91:7372
29. Tatarinova VI, Vasiliev AA, Petrosyan VA (1990) *Izv Akad Nauk SSSR Ser Khim* 2646
30. Knipe AC, Stirling CJM (1968) *J Chem Soc B* 67
31. Salmon-Legagneur F, Nevéu C (1956) *Bull Soc Chim* 1345
32. Buchta E, Fischer M (1966) *Chem Ber* 99:1509
33. Zieger K, Weber K (1934) *Liebigs Ann Chem* 512:164