



Propionate and Alzheimer's Disease

Jessica Killingsworth*, Darrell Sawmiller and R. Douglas Shytle

Department of Neurosurgery and Brain Repair, Morsani College of Medicine, University of South Florida, Tampa, FL, United States

Propionate, a short-chain fatty acid, serves important roles in the human body. However, our review of the current literature suggests that under certain conditions, excess levels of propionate may play a role in Alzheimer's disease (AD). The cause of the excessive levels of propionate may be related to the *Bacteroidetes* phylum, which are the primary producers of propionate in the human gut. Studies have shown that the relative abundance of the *Bacteroidetes* phylum is significantly increased in older adults. Other studies have shown that levels of the *Bacteroidetes* phylum are increased in persons with AD. Studies on the diet, medication use, and propionate metabolism offer additional potential causes. There are many different mechanisms by which excess levels of propionate may lead to AD, such as hyperammonemia. These mechanisms offer potential points for intervention.

Keywords: gut microbiome, valproate, short chain fatty acids, Alzheimer's disease, propionate

INTRODUCTION

OPEN ACCESS

Edited by:

Gjumrakch Aliev, GALLY International Biomedical Research, United States

Reviewed by:

Gang Wang, Jiangnan University, China Edward Chambers, Imperial College London, United Kingdom

*Correspondence:

Jessica Killingsworth jkillingswor@mail.usf.edu

Received: 04 July 2020 Accepted: 15 December 2020 Published: 11 January 2021

Citation:

Killingsworth J, Sawmiller D and Shytle RD (2021) Propionate and Alzheimer's Disease. Front. Aging Neurosci. 12:580001. doi: 10.3389/fnagi.2020.580001 Dietary fibers are carbohydrate polymers that have at least 10 monomeric units (Cummings et al., 2009). The enzymes needed to digest most dietary fibers are lacking in the human body (den Besten et al., 2013). Therefore, the microbiota in the intestine is tasked with fermenting dietary fibers. Fermentation results in the production of short-chain fatty acids (SCFAs), which serve several important functions. In the gut, they aid in microbial growth (Alexander et al., 2019). They are also second messengers that can modulate gene expression and initiate the synthesis of gut peptides and hormones. One of the major SCFAs is propionate, which is three carbons in length (Alexander et al., 2019). It is estimated that in a human being who weighs 85 kg, the gut microbiota produce approximately 29.5 mg/kg of propionate each day *via* fermentation (Morrison and Preston, 2016).

In addition to fermentation, two other sources of propionate are food and the oral microbiome. In 1984, the Food and Drug Administration (FDA) labeled propionate as generally recognized as safe (GRAS) and approved its use for food preservation (U.S. Department of Agriculture, 2008). It is found in a concentration of 0.1 to 0.4% in various foods, including baked goods, dairy products, meat products, puddings, gelatins, and jams (Mani-López et al., 2012; Reis et al., 2012; Tirosh et al., 2019; U.S. Department of Agriculture Technical Advisory Committee, 2002). Therefore, most persons are exposed to dietary sources of propionate every day. It is estimated that in a single meal consisting of processed food, propionate is 0.3% (w/w; Tirosh et al., 2019). Dietary choices thus could potentially impact the amount of propionate in the peripheral circulation. Indeed, Chambers et al. (2017) found an increase in the levels of propionate after an overnight fast. Similarly, Tirosh et al. (2019) found that a meal containing a low dose of propionate led to a significant increase in postprandial plasma levels of propionate in healthy humans. As for the oral microbiome, oral microbiota can produce propionate (Takahashi, 2015). Increased propionate levels are associated with gingivitis and periodontal disease.

1

When propionate is ingested or generated in the intestine, it makes its way to the liver through the hepatic portal vein (Hoyles et al., 2018). Approximately 90-95% of propionate is used by the liver. The remaining propionate enters the peripheral circulation. Tian et al. (2020) found that the average serum levels of propionate were 2.843 mmol/L (2,843 µM) directly following a 12 h fast in healthy adults between the ages of 20 and 40. In an interesting study, Wolever et al. (1997) found that the average propionate serum levels over 12 h were 3.8 μ mol/L (3.8 μ M) in young adults and 4.6 μ mol/L (4.6 μ M) in middle-aged adults. Propionate can cross the blood-brain barrier (BBB). Hoyles et al. (2018) found that there is a propionate free fatty acid receptor 3 (FFAR3) on the endothelium of the human brain. According to the Human Metabolome Database, typical values of propionate in the cerebrospinal fluid (CSF) are 2.8–3.2 μ M in adults (Wishart et al., 2017). Propionate is also found in the saliva. According to the Human Metabolome Database, different studies have found different ranges of resting propionate saliva levels, with levels ranging from 1 to 1,089.82 µM (Wishart et al., 2017). Differences in the levels of propionate have been attributed to oral health, gender, and smoking status (Takeda et al., 2009).

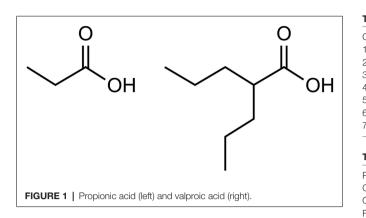
Propionate serves several functions in the human body. For instance, propionate promotes enteric smooth muscle contractions and stimulates host defense peptide expression (Mitsui et al., 2005; Sunkara et al., 2012; De Vadder et al., 2014). Deficient levels of propionate have been associated with increased risk for asthma and allergies, highlighting the positive role of propionate in the immune system (Böttcher et al., 2000; Roduit et al., 2018; Ivashkin et al., 2019). Additionally, the metabolism of propionate is associated with glucose production and energy metabolism (Ringer, 1912; Tirosh et al., 2019). Through a series of reactions, propionate is first converted to propionyl-CoA before ultimately being converted to succinyl-CoA (Berg et al., 2002). Succinyl-CoA is a substrate in the TCA cycle. Thus, dietary propionate could perhaps impact the TCA cycle. Perry et al. (2016) found that ingestion of a bolus of propionate increased the concentrations of propionyl-CoA by 100-fold in rodents. Additionally, propionate can participate in the gut-brain axis (Chambers et al., 2015). There is evidence that propionate can affect satiety by stimulating the release of peptide YY (PYY) and glucagon-like peptide-1 (GLP-1). PYY and GLP-1 function to provide a short-term signal of satiety to the brain.

Excess levels of propionate appear to be problematic. One example of the effects of excess propionate is propionic acidemia (PA). This metabolic disorder has been associated with motor impairments, brain atrophy, cognitive impairments, and dementia (Sethi et al., 1989; Morland et al., 2018; Schwoerer et al., 2018). Furthermore, persons with periodontal disease have increased levels of propionate in their saliva and appear to be at an increased risk for developing Alzheimer's disease (AD; Aimetti et al., 2011; Kamer et al., 2015; Chen et al., 2017). In line with those findings, there is emerging evidence that suggests that excess propionate may play a role in dementia, particularly in AD. Dementia is an age-related disease that is associated with cognitive decline. AD is the most common type of dementia and is characterized by neurofibrillary tangles and β -amyloid plaques (Delacourte, 1994; Hardy and Duff, 1994). According to the Alzheimer's Association, an estimated 5.8 million individuals currently have AD in the United States (Alzheimer's Association, 2020). This number is projected to increase to 13.8 million by the year 2050 (Alzheimer's Association, 2020). In this review article, we will explore the literature that supports the potential role of excess propionate in AD.

PROPIONATE, VALPROATE, AND ALZHEIMER'S DISEASE

Several recent studies offer evidence for a link between propionate and AD. For instance, Figueira et al. (2016) analyzed saliva samples of persons with dementia and healthy controls. They found a 1.35-fold increase in propionate levels in persons with dementia when compared to healthy controls. Yilmaz et al. (2017) analyzed saliva samples from persons with mild cognitive impairment, persons with AD, and healthy controls. They found that the levels of propionate were significantly increased in persons with AD in comparison to healthy controls. Both studies were limited in that they only evaluated saliva samples; however, there is evidence for the validity of using saliva samples. Martin et al. (2018) found that in comparison to plasma levels, saliva levels of oxytocin better correlated with CSF levels. Similarly, Adamashvili et al. (2005) found that saliva levels of human major histocompatibility antigens (HLA) correlated with CSF HLA levels in persons with Multiple Sclerosis. Kennedy et al. (2001) found that epinephrine in saliva appears to originate from both the salivary sympathetic nerves and peripheral circulation. Moreover, Valstar et al. (2020) recently discovered the existence of what they deemed the tubarial glands, which are salivary glands located within the nasopharynx. Several rodent studies link fecal and circulating levels of propionate to AD. Fujii et al. (2019) found that mice given a fecal microbiota transplant from patients with AD had significantly higher levels of propionate in comparison to controls. There is evidence that fecal levels of propionate are positively correlated with circulating levels of propionate in humans (Müller et al., 2019). Additionally, González-Domínguez et al. (2015) found a 1.23-fold increase in propionate in the hippocampus of AD transgenic mice. Similarly, Syeda et al. (2018) found a significantly higher concentration of propionate in the prefrontal cortex of AD transgenic mice in comparison to wild type mice. They also found that AD transgenic mice had significantly higher fecal concentrations of propionate at 6 months of age in comparison to wild type mice.

Valproate provides further evidence for the role of excess propionate in AD. As illustrated by **Figure 1**, valproate is structurally similar to propionate. Valproate (VPA) and its conjugate acid (valproic acid) are FDA approved medications (under trade names Divalproex, Depakote, Depakote ER, Depakene, Depacon, and Stavzor) for the treatment of epilepsy and bipolar disorder and are prescribed off label for agitation for persons with dementia (Chiu et al., 2013; Baillon et al., 2018). However, a growing body of evidence



suggests that VPA causes neurotoxicity that is associated with its metabolism into propionic acid (Farooq et al., 2017). In excessive amounts, propionic acid inhibits a urea cycle enzyme, carbamoyl phosphate synthase, which impairs the body's ability to excrete ammonia and thus results in hyperammonemia. Once considered only a rare side effect, VPA induced hyperammonemic encephalopathy is now reported frequently in the literature. It produces clinical signs and symptoms that appear to mirror those found in dementia (see Table 1, Kowalski et al., 2013). In a study investigating VPA's effects on fatty acid metabolism and the urea cycle in schizophrenic patients, Ando et al. (2017) found that 30% developed hyperammonemia (defined as ammonia greater than 47 µmol/L). In a follow-up study by Baddour et al. (2018) on 347 patients treated at a community teaching hospital, the reported incidence of hyperammonemia was found to be 36%, with 43.2% of those patients presenting with clinical symptoms.

As further evidence of the neurotoxicity associated with VPA, Armon et al. (1996) found that patients with epilepsy who had been on VPA therapy for at least 1-year experienced reversible cognitive impairments and brain atrophy. Tsai et al. (2016) found that valproic acid treatment increased the risk of developing dementia by 73-95% in patients with bipolar disorder in comparison to patients who were not on valproic acid treatment. Pardoe et al. (2013) compared the brain volume, white matter volume, and parietal lobe thickness of persons with epilepsy who were taking sodium valproate to persons with epilepsy not on sodium valproate treatment and to healthy controls. They found that sodium valproate treatment resulted in brain atrophy, white matter volume loss, and a reduction in parietal lobe thickness. Tariot et al. (2011) explored the effects of VPA on patients with AD. They found that valproate treatment caused greater brain volume loss in comparison to the placebo. In a second study with the same AD patients from the Tariot et al.'s (2011) study, Fleisher et al. (2011) found that VPA caused increased brain volume loss and ventricular expansion in comparison to controls. Furthermore, through the first year of the study, the Mini-Mental State Examination scores implied that VPA treatment led to an accelerated decline in cognition. Taken together, these studies provide further evidence for the role of propionate in AD.

TABLE 1	Symptoms of valproate-induced hyperammonemia.
---------	---

Changes in
1. Orientation
2. Memory
3. Motor function
4. Fatigue
5. Onset or worsening of psychosis
6. Delirium
7. Mood

TABLE 2 | Bacteroidetes.

Phylum	Bacteroidetes	
Class	Bacteroidia	
Order	Bacteroidales	
Family	Bacteroidaceae	
Genus	Bacteroides	

THE COMMENSAL MICROBIOTA, PROPIONATE, AND ALZHEIMER'S DISEASE

The commensal microbiota is also relevant to the discussion on propionate and AD. Aguirre et al. (2016) and Salonen et al. (2014) found that one type of bacteria called *Bacteroidetes* serves as the principal producers of propionate in the human gut. Specifically, *Bacteroidetes* convert hexose sugars into propionate *via* a succinate pathway (Reichardt et al., 2014). **Table 2** illustrates the class, order, family, and genus relevant to the discussion on propionate. There appear to be age-related changes in the human microbiome, particularly changes in the relative abundance of *Bacteroidetes*. For instance, Claesson et al. (2011) found that elderly participants had a greater proportion of *Bacteroides* species than younger adults. Likewise, Odamaki et al. (2016) found that the elderly participants had a significantly higher relative abundance of *Bacteroidetes*, in comparison to infant and adult participants.

Bacteroidetes appear to play a role in AD and appear to potentially account for the excess levels of propionate in AD. For instance, Vogt et al. (2017) found that the Bacteroidetes phylum was increased in persons with AD when compared to healthy controls. They also found that the levels of several AD markers in CSF were significantly correlated with the relative abundance of the Bacteroides genus. Haran et al. (2019) found the Bacteroides genus to be increased in persons with AD. Although Liu et al. (2019) did not find the Bacteroidetes phylum to be significantly increased in persons with AD, they did find that the level of Bacteroidetes was significantly increased in persons at the predementia stage of AD. Wang et al. (2016) found that the Bacteroidales order was negatively correlated with spatial learning and memory ability, active avoidance response, and object recognition memory capability in a rodent model of AD. Both Harach et al. (2017) and Kaur et al. (2020) also found that the Bacteroidetes phylum was increased in rodent models of AD, while Zhan et al. (2018) found that the Bacteroidales order was increased in a rodent model of AD. Harach et al. (2017) found that germ-free generated APPPS1 mice displayed decreased levels of cerebral Aβ42 in comparison to conventionally raised

APPPS1 mice. This finding supports a possible role of the gut microbiome in amyloid precursor protein (APP) expression.

In addition to *Bacteriales*, *Actinobacteria* may also play a role in AD. Zhuang et al. (2018) found that the relative abundance of this phylum was increased in the gut of persons with AD, in comparison to healthy controls. *Propionibacterium acnes* (*P. acnes*), which is part of the *Actinobacteria* phylum, is named after its ability to produce propionic acid (Kirschbaum and Kligman, 1963). It is part of the skin, oral, and gut microbiome. It can also cross the BBB (Lu et al., 2019). Concerning AD, *P. acnes* was reportedly found in the cortex of three patients with AD (Kornhuber, 1996). Also, Emery et al. (2017) employing 16S rRNA sequencing analysis to investigate possible bacterial infections in AD brains, consistently found high levels of *P. acnes* in AD samples compared to normal brains under methodological conditions that would make contamination an unlikely explanation for their findings.

POTENTIAL MECHANISMS

There is evidence for such a wide array of different mechanisms that excess propionate likely leads to AD by way of a combination of multiple different mechanisms. Probably the most well-studied mechanism of propionate induced neurotoxicity is related to its ability to impair the urea cycle, the principal pathway for nitrogen metabolism. This condition, known as hyperammonemia, occurs in propionic acidemia (PA), an autosomal recessive genetic disease characterized by an abnormal accumulation of propionic acid (Haijes et al., 2019). As aforementioned, hyperammonemia can also occur in patients who are prescribed VPA. In cases of PA with hyperammonemia of \geq 360 µmol/L, significant encephalopathy and intellectual disability can occur (Kido et al., 2011). Abnormal accumulation of propionic acid results in excessive propionyl-CoA production, which inhibits N-acetyl-glutamate (NAG) formation (Coude et al., 1979). NAG is important because it activates carbamoyl phosphate synthetase I, which is a key enzyme in the first step of the urea cycle. Propionyl-CoA also inhibits this pathway by depleting hepatic acetyl CoA, which is responsible for NAG synthesis. Propionyl-CoA has a broad impact on metabolism, influencing not only the urea cycle, but also the citric acid cycle and related enzymes, the respiratory chain complex, and the glycine cleavage system. Considering that L-carnitine plays a crucial role in propionic acid metabolism, excessive propionic acid levels inevitably result in L-carnitine deficiency (Maldonado et al., 2016). This further potentiates propionicacid-mediated neurotoxicity by disrupting β -oxidation pathways and preventing the conversion of propionyl-CoA into the nontoxic and beneficial propionyl carnitine (Roe et al., 1984).

Although acute hyperammonemia can cause encephalopathy, the clinical manifestations of chronic, slightly elevated blood ammonia levels have received relatively little research interest within the field of dementia research (Jin et al., 2018). However, considering the well-known neurotoxic nature of ammonia, it is reasonable to speculate that chronically elevated levels of ammonia might be associated with the development of AD. Indeed, some small clinical studies have reported an association between AD and elevated blood ammonia levels (Fisman et al., 1985, 1989; Branconnier et al., 1986). While ammonia is a normal end product of human tissue metabolism, it is a highly neurotoxic compound at even sub-millimolar concentrations (Marcaida et al., 1992; Roquilly et al., 2013). Thus, ammonia detoxification in organisms is indispensable. In the brain, under normal or hyperammonemic conditions, ATP-dependent formation of glutamine by glutamine synthetase [L-glutamate:ammonia ligase (ADP-forming; E.C.6.3.1.2); GS] is predominantly used for ammonia removal (Norenberg and Martinez-Hernandez, 1979; Cooper and Plum, 1987). In hyperammonemia, astroglia located in proximity to blood-vessels in glutamatergic areas show increased GS protein content in their perivascular processes. Since ammonia freely crosses the BBB and astrocytes are responsible for maintaining the BBB, the presence of GS in the perivascular processes can produce a rapid glutamine synthesis and subsequent release into the blood to limit excess ammonia from circulation. The changes in the distribution of this critical enzyme suggest that the glutamate-glutamine cycle may be differentially impaired in hyperammonemic states (Robinson, 2000; Suárez et al., 2002). Combining a genomic and transcriptomic approach, Bensemain et al. (2009) characterized the induction of the urea cycle metabolic pathway in the brains of AD cases. They found that the expression of the ornithine transcarbamylase (OTC) protein, another key enzyme of the urea cycle, in endothelial cells of AD brain vessels was increased 880% in the CSF of probable AD cases compared with controls. Future studies investigating the relationship between chronically low-grade hyperammonemia and AD should also concurrently measure propionic acid levels in saliva and blood to determine if there is a causal relationship between excess propionic acid levels and hyperammonaemia, as seen in PA and patients treated with VPA.

Another potential mechanism may involve insulin. Studies have shown that SCFAs, especially butyrate, may improve insulin sensitivity (Henagan et al., 2015). However, there is evidence that propionate is not beneficial for insulin sensitivity. Tirosh et al. (2019) investigated the role of propionate in glucose production in humans and in rodents. This study, unlike most other studies on propionate and insulin sensitivity, included participants that were healthy and lean. Other studies have found that propionate can improve insulin sensitivity; however, they are limited in that they either: (1) included unrepresentative delivery or quantities of propionate; and/or (2) included participants who were only overweight or prediabetic (Pingitore et al., 2016; Chambers et al., 2019; Müller et al., 2019). In the Tirosh et al.'s (2019) study, the participants consumed a meal containing an amount of calcium propionate representative of that found in a typical meal consisting of processed foods. The rodents were given a similarly representative amount of propionate in their diet. As for the results, the results imply that orally delivered propionate does not have the same positive effects on insulin sensitivity that are associated with the SCFAs derived from the gut microbiota. In fact, the results imply that orally delivered propionate may instead lead to insulin resistance and glucose intolerance. In the human participants, the propionate-enriched meal leads to increased postprandial levels of insulin. In the rodents, they studied the long term results of orally delivered propionate. The results also imply a role of propionate in insulin resistance. However, it is worth noting that the study is limited in that it only included 14 middle-aged men. Thus, a larger study would be necessary to confirm their results and to elucidate the long-term effects of orally delivered propionate. Darzi et al. (2012) studied the effects of the consumption of bread containing propionate in lean, healthy women, and men. Consistent with the Tirosh et al.'s (2019) study, they found that propionate caused increased postprandial levels of insulin. Moreover, Sanna et al. (2019) found increased fecal levels of propionate to be associated with an increased risk for Type 2 diabetes mellitus. These findings are notable as there is evidence that persons with Type 2 diabetes are at an increased risk for developing AD (Cheng et al., 2011; Madmoli et al., 2019). Additionally, Ciudin et al. (2017) found that Type 2 diabetes was an independent risk factor for the progression of MCI to dementia. Thus, taken together, these studies suggest that insulin resistance may be one mechanism by which excess propionate leads to AD. However, further studies are necessary to clarify this potential mechanism.

As for other mechanisms, propionate has been found to have several other neurotoxic effects, including mitochondrial dysfunction, neuroinflammation, glutamate excitotoxicity, DNA damage, inhibition of Na⁺/K⁺-ATPase, apoptosis of neuronal cells, an increase in oxidative stress, and a decrease in superoxide dismutase activity and both glutathione and serotonin levels (Wyse et al., 1998; Rigo et al., 2006; MacFabe et al., 2008; El-Ansary et al., 2013, 2017; Khalil et al., 2015; Al-Orf et al., 2018). These neurotoxic effects have been associated with AD. For instance, Mandal et al. (2015) found that the antioxidant glutathione was significantly decreased in the brains of persons with MCI and AD. Glutathione levels were also inversely correlated with the severity of the cognitive impairments in the participants. However, future studies are necessary to further clarify the mechanisms by which excess propionate leads to AD.

POTENTIAL INTERVENTIONS

It is well established that L-carnitine (CAR) supplementation as an adjuvant therapy contributes to the amelioration of blood markers of oxidative damage in patients affected by disorders of excess propionate levels, as well as in the treatment of VPA-induced hyperammonemia (Roe et al., 1984; Ribas et al., 2010; Maldonado et al., 2016; Cutshall et al., 2017). As Maldonado et al. (2016) hypothesized from their study, "In patients treated with VPA, CAR depletion followed by [acetyl-lcarnitine] ALCAR decrease could be responsible for the increase in the ammonia levels. On the other hand, in the elderly population, serum CAR could be increased due to impaired access to tissues which in turn could result in an ALCAR decrease. This last fact could lead to ammonia impaired elimination. Perhaps higher ammonia levels and ALCAR deficit could be responsible for the cognitive and neurodegenerative diseases found in the elderly." Indeed, numerous clinical trial studies have investigated the cognitive therapeutic benefits of L-carnitine and acetyl-lcarnitine treatment in AD (Spagnoli et al., 1991; Montgomery et al., 2003). However, only recently have investigators considered explaining their putative therapeutic benefits in the context of reducing hyperammonemia in neurological disorders of the elderly (Maldonado et al., 2016, 2020). And, to our knowledge, no one until now has considered excess propionate production *via* a bacterial infection as a possible causal process resulting in prolonged low-grade hyperammonemia.

The metabolic pathways associated with the breakdown of propionate may also offer points for intervention. For example, vitamin B-12 is a cofactor in the conversion of propionate to succinyl-CoA (Berg et al., 2002). Revtovich et al. (2019) studied vitamin B-12 and propionate levels in Caenorhabditis elegans. Their results support vitamin B-12 playing a role in the breakdown of propionate. They also explored vitamin B-12 supplementation concerning mitochondrial health, since excess propionate leads to mitochondrial dysfunction. They found that vitamin B-12 supplementation in C. elegans resulted in improved mitochondrial health. Concerning AD, decreased vitamin B-12 levels appear to be linked to AD (Ma et al., 2017). Thus, perhaps decreased vitamin B-12 could be another potential cause of the excess propionate. Douaud et al. (2013) found that vitamin B-12 supplementation was able to reduce cerebral atrophy in the gray matter regions that are affected by AD. This reduction was significant, as it was a 7-fold reduction. In addition to vitamin B-12, propionyl-CoA carboxylase may also be a viable target for intervention. Like vitamin B-12, this enzyme is also involved in the conversion of propionate to succinyl-coenzyme A (Berg et al., 2002). Dysregulated propionyl-CoA carboxylase can lead to increased levels of propionate (Morland et al., 2018). Cuadrado-Tejedor et al. (2013) found that older transgenic AD mice displayed deregulated propionyl-CoA carboxylase. Therefore, propionyl-CoA carboxylase, along with vitamin B-12, warrants future study.

Another potential intervention may be the antioxidant TEMPOL(4-hydroxy-2,2,6,6-tetramethylpiperidine-N-oxyl), which is a small (MW 172 Da) stable nitroxide radical that can readily permeate biological membranes. Cai et al. (2016) found that TEMPOL could reduce cecal and fecal concentrations of propionate in obese rodents. In relation to AD, Ali et al. (2016) and Khallaf et al. (2017) found that TEMPOL was able to reduce neuroinflammation, cognitive impairments, and amyloidogenesis in the rodents. Ali et al. (2016) also found that TEMPOL was able to reduce oxido-nitrosative stress, which has been found to play a role in ammonia toxicity (Skowrońska and Albrecht, 2013). This is of particular interest as propionate can impair the urea cycle and lead to hyperammonemia. Thus, this antioxidant warrants further study, especially concerning its effects on propionate and its effects on oxido-nitrosative stress.

Butyrate, an SCFA, may be another potential intervention. Butyrate appears to have neuroprotective effects and has been indicated as a viable treatment for neurological disorders, such as Parkinson's disease (Liu et al., 2017). Concerning AD, Govindarajan et al. (2011) found that butyrate improved cognition in a rodent model of AD. Cleophas et al. (2019) found that butyrate supplementation reduced fecal levels of propionate in both lean and obese men. Thus, like vitamin B-12 and potentially TEMPOL, butyrate supplementation may be a viable method to reduce propionate levels. Yet, further research is needed to clarify this potential role of butyrate. Further work would especially need to study the effect of butyrate on circulating levels of propionate.

Diet may be a target for intervention, as diet could directly impact one's intake of propionate. Furthermore, diet can impact the levels of the Bacteroidetes phylum. In comparison to non-Western diets, a Western diet, which consists of high protein and fat, was found to increase the levels of Bacteroidetes or Bacteroides in several studies (Filippo et al., 2010; Wu et al., 2011; David et al., 2013; Yeagle, 2015; Heinritz et al., 2016). Furthermore, keeping in mind that propionate is used as a food preservative, a diet that features low consumption of foods with propionate may be another viable intervention. Some promising dietary interventions on autism spectrum disorders (ASD) show some evidence for dietary interventions for propionate. ASD appears to be another neurological disease associated with excess propionate. Multiple studies have found that propionate causes ASD-like behaviors in rodents (MacFabe et al., 2011; Kamen et al., 2018; Mepham et al., 2019; Shams et al., 2019). Additionally, Angelis et al. (2013) and Wang et al. (2012) found that children with ASD had higher levels of propionate in comparison to healthy controls. As for dietary interventions, in several studies, participants with ASD experienced improvements in their symptoms when placed on a dairy-free and gluten-free diet (Knivsberg et al., 2002; Whiteley et al., 2010; Ghalichi et al., 2016; El-Rashidy et al., 2017).

REFERENCES

- Adamashvili, I., Minagar, A., Gonzalez-Toledo, E., Featherston, L., and Kelley, R. E. (2005). Soluble HLA measurement in saliva and cerebrospinal fluid in Caucasian patients with multiple sclerosis: a preliminary study. *J. Neuroinflammation* 2:13. doi: 10.1186/1742-2094-2-13
- Aguirre, M., Eck, A., Koenen, M. E., Savelkoul, P. H., Budding, A. E., and Venema, K. (2016). Diet drives quick changes in the metabolic activity and composition of human gut microbiota in a validated *in vitro* gut model. *Res. Microbiol.* 167, 114–125. doi: 10.1016/j.resmic.2015. 09.006
- Aimetti, M., Cacciatore, S., Graziano, A., and Tenori, L. (2011). Metabonomic analysis of saliva reveals generalized chronic periodontitis signature. *Metabolomics* 8, 465–474. doi: 10.1007/s11306-011-0331-2
- Alexander, C., Swanson, K. S., Fahey, G. C., and Garleb, K. A. (2019). Perspective: physiologic importance of short-chain fatty acids from nondigestible carbohydrate fermentation. *Adv. Nutr.* 10, 576–589. doi: 10.1093/advances/nmz004
- Ali, M. R. A.-A., Abo-Youssef, A. M. H., Messiha, B. A. S., and Khattab, M. M. (2016). Tempol and perindopril protect against lipopolysaccharide-induced cognition impairment and amyloidogenesis by modulating brain-derived neurotropic factor, neuroinflammation and oxido-nitrosative stress. *Naunyn Schmiedebergs Arch. Pharmacol.* 389, 637–656. doi: 10.1007/s00210-016 -1234-6
- Al-Orf, N., El-Ansary, A., Bjørklund, G., Moubayed, N., Bhat, R. S., and Bacha, A. B. (2018). Therapeutic effects of probiotics on neurotoxicity induced by clindamycin and propionic acid in juvenile hamsters. *Metab. Brain Dis.* 33, 1811–1820. doi: 10.1007/s11011-018-0284-5
- Alzheimer's Association. (2020). 2020 Alzheimer's disease facts and figures. Alzheimers Dement. 16, 391–460. doi: 10.1002/alz.12068
- Ando, M., Amayasu, H., Itai, T., and Yoshida, H. (2017). Association between the blood concentrations of ammonia and carnitine/amino acid of schizophrenic patients treated with valproic acid. *Biopsychosoc. Med.* 11:19. doi: 10.1186/s13030-017-0101-0

Additionally, as a preventative measure, replacing propionate with other compounds for food preservation may be beneficial.

CONCLUSION

Propionate serves important functions in the body. However, excess levels of propionate may play a role in AD. The cause of the excessive levels of propionate could be related to diet, medication use, the commensal microbiota, or potentially related to propionate metabolism. Future studies should aim to clarify the cause of the excess levels. There are multiple mechanisms by which propionate may lead to AD, including glutamate excitotoxicity and hyperanmonemia. The mechanisms offer potential points for intervention.

AUTHOR CONTRIBUTIONS

JK reviewed the literature and wrote this article. DS and RS edited this article. All authors contributed to the article and approved the submitted version.

FUNDING

The authors declare that this study received funding from Alzamend Neuro Inc. The funder was not involved in the study design, collection, analysis, interpretation of data, the writing of this article or the decision to submit it for publication.

- Angelis, M. D., Piccolo, M., Vannini, L., Siragusa, S., Giacomo, A. D., Serrazzanetti, D. I., et al. (2013). Fecal microbiota and metabolome of children with autism and pervasive developmental disorder not otherwise specified. *PLoS One* 8:e76993. doi: 10.1371/journal.pone.0076993
- Armon, C., Shin, C., Miller, P., Carwile, S., Brown, E., Edinger, J. D., et al. (1996). Reversible Parkinsonism and cognitive impairment with chronic valproate use. *Neurology* 47, 626–635. doi: 10.1212/wnl.47.3.626
- Baddour, E., Tewksbury, A., and Stauner, N. (2018). Valproic acid-induced hyperammonemia: incidence, clinical significance, and treatment management. *Ment. Health Clin.* 8, 73–77. doi: 10.9740/mhc.2018. 03.073
- Baillon, S. F., Narayana, U., Luxenberg, J. S., and Clifton, A. V. (2018). Valproate preparations for agitation in dementia. *Cochrane Database Syst. Rev.* 10:CD003945. doi: 10.1002/14651858.CD003945.pub4
- Bensemain, F., Hot, D., Ferreira, S., Dumont, J., Bombois, S., Maurage, C.-A., et al. (2009). Evidence for induction of the ornithine transcarbamylase expression in Alzheimer's disease. *Mol. Psychiatry* 14, 106–116. doi: 10.1038/sj.mp.4002089
- Berg, J. M., Tymoczko, J. L., and Stryer, L. (2002). *Biochemistry*, 5th edition. New York, NY: W H Freeman.
- Böttcher, M. F., Nordin, E. K., Sandin, A., Midtvedt, T., and Bjorksten, B. (2000). Microflora-associated characteristics in faeces from allergic and nonallergic infants. *Clin. Exp. Allergy* 30, 1591–1596. doi: 10.1046/j.1365-2222.2000. 00982.x
- Branconnier, R. J., Dessain, E. C., McNiff, M. E., and Cole, J. O. (1986). Blood ammonia and Alzheimer's disease. Am. J. Psychiatry 143, 1313–1314. doi: 10.1176/ajp.143.10.1313
- Cai, J., Zhang, L., Jones, R. A., Correll, J. B., Hatzakis, E., Smith, P. B., et al. (2016). Antioxidant drug tempol promotes functional metabolic changes in the gut microbiota. *J. Proteome Res.* 15, 563–571. doi: 10.1021/acs.jproteome. 5b00957
- Chambers, E. S., Byrne, C. S., Aspey, K., Chen, Y., Khan, S., Morrison, D. J., et al. (2017). Acute oral sodium propionate supplementation raises resting energy expenditure and lipid oxidation in fasted humans. *Diabetes Obes. Metab.* 20, 1034–1039. doi: 10.1111/dom.13159

- Chambers, E. S., Byrne, C. S., Morrison, D. J., Murphy, K. G., Preston, T., Tedford, C., et al. (2019). Dietary supplementation with inulin-propionate ester or inulin improves insulin sensitivity in adults with overweight and obesity with distinct effects on the gut microbiota, plasma metabolome and systemic inflammatory responses: a randomized cross-over trial. *Gut* 68, 1430–1438. doi: 10.1136/gutjnl-2019-318424
- Chambers, E. S., Viardot, A., Psichas, A., Morrison, D. J., Murphy, K. G., Zac-Varghese, S. E., et al. (2015). Effects of targeted delivery of propionate to the human colon on appetite regulation, body weight maintenance and adiposity in overweight adults. *Gut* 64, 1744–1754. doi: 10.1136/gutjnl-2014-307913
- Chen, C.-K., Wu, Y.-T., and Chang, Y.-C. (2017). Association between chronic periodontitis and the risk of Alzheimer's disease: a retrospective, populationbased, matched-cohort study. *Alzheimers Res. Ther.* 9:56. doi: 10.1186/s13195-017-0282-6
- Cheng, D., Noble, J., Tang, M. X., Schupf, N., Mayeux, R., and Luchsinger, J. A. (2011). Type 2 diabetes and late-onset Alzheimer's disease. *Dement. Geriatr. Cogn. Disord.* 31, 424–430. doi: 10.1159/000324134
- Chiu, C.-T., Wang, Z., Hunsberger, J. G., and Chuang, D.-M. (2013). Therapeutic potential of mood stabilizers lithium and valproic acid: beyond bipolar disorder. *Pharmacol. Rev.* 65, 105–142. doi: 10.1124/pr.111.005512
- Ciudin, A., Espinosa, A., Simó-Servat, O., Ruiz, A., Alegret, M., Hernández, C., et al. (2017). Type 2 diabetes is an independent risk factor for dementia conversion in patients with mild cognitive impairment. *J. Diabetes Complicat.* 31, 1272–1274. doi: 10.1016/j.jdiacomp.2017.04.018
- Claesson, M. J., Cusack, S., O'Sullivan, O., Greene-Diniz, R., de Weerd, H., Flannery, E., et al. (2011). Composition, variability and temporal stability of the intestinal microbiota of the elderly. *Proc. Natl. Acad. Sci. USA* 108, 4586–4591. doi: 10.1073/pnas.1000097107
- Cleophas, M. C. P., Ratter, J. M., Bekkering, S., Quintin, J., Schraa, K., Stroes, E. S., et al. (2019). Effects of oral butyrate supplementation on inflammatory potential of circulating peripheral blood mononuclear cells in healthy and obese males. *Sci. Rep.* 9:775. doi: 10.1038/s41598-018-37246-7
- Cooper, A. J., and Plum, F. (1987). Biochemistry and physiology of brain ammonia. *Physiol. Rev.* 67, 440–519. doi: 10.1152/physrev.1987.67.2.440
- Coude, F. X., Sweetman, L., and Nyhan, W. L. (1979). Inhibition by propionylcoenzyme A of N-acetylglutamate synthetase in rat liver mitochondria. A possible explanation for hyperammonemia in propionic and methylmalonic acidemia. J. Clin. Invest. 64, 1544–1551. doi: 10.1172/JCI109614
- Cuadrado-Tejedor, M., Cabodevilla, J. F., Zamarbide, M., Gomez-Isla, T., Franco, R., and Perez-Mediavilla, A. (2013). Age-related mitochondrial alterations without neuronal loss in the hippocampus of a transgenic model of Alzheimer's disease. *Curr. Alzheimer Res.* 10, 390–405. doi: 10.2174/1567205011310040005
- Cummings, J., Mann, J., Nishida, C., and Vorster, H. (2009). Dietary fibre: an agreed definition. *Lancet* 373, 365–366. doi: 10.1016/S0140-6736(09)60117-3
- Cutshall, B. T., Shah, S. P., Van Berkel, M. A., Patterson, S., Harris, L. J., and Rivera, J. V. (2017). Should pharmacies be included in medication reconciliation? A report of recurrent valproic acid toxicity. *Clin. Pract. Cases Emerg. Med.* 1:122. doi: 10.5811/cpcem.2016.12.33002
- Darzi, J., Frost, G. S., and Robertson, M. D. (2012). Effects of a novel propionate-rich sourdough bread on appetite and food intake. *Eur. J. Clin. Nutr.* 66, 789–794. doi: 10.1038/ejcn.2012.1
- David, L. A., Maurice, C. F., Carmody, R. N., Gootenberg, D. B., Button, J. E., Wolfe, B. E., et al. (2013). Diet rapidly and reproducibly alters the human gut microbiome. *Nature* 505, 559–563. doi: 10.1038/nature12820
- Delacourte, A. (1994). Pathological Tau proteins of Alzheimer's disease as a biochemical marker of neurofibrillary degeneration. *Biomed. Pharmacother*. 48, 287–295. doi: 10.1016/0753-3322(94)90174-0
- den Besten, G., van Eunen, K., Groen, A. K., Venema, K., Reijngoud, D. J., and Bakker, B. M. (2013). The role of short-chain fatty acids in the interplay between diet, gut microbiota, and host energy metabolism. *J. Lipid Res.* 54, 2325–2340. doi: 10.1194/jlr.R036012
- De Vadder, F., Kovatcheva-Datchary, P., Goncalves, D., Vinera, J., Zitoun, C., Duchampt, A., et al. (2014). Microbiota-generated metabolites promote metabolic benefits *via* gut-brain neural circuits. *Cell* 156, 84–96. doi: 10.1016/j. cell.2013.12.016
- Douaud, G., Refsum, H., Jager, C. A. D., Jacoby, R., Nichols, T. E., Smith, S. M., et al. (2013). Preventing Alzheimer's disease-related gray matter atrophy

by B-vitamin treatment. Proc. Natl. Acad. Sci. U S A 110, 9523–9528. doi: 10.1073/pnas.1301816110

- El-Ansary, A., Al-Salem, H. S., Asma, A., and Al-Dbass, A. (2017). Glutamate excitotoxicity induced by orally administered propionic acid, a short chain fatty acid can be ameliorated by bee pollen. *Lipids Health Dis.* 16:96. doi: 10.1186/s12944-017-0485-7
- El-Ansary, A., Shaker, G. H., El-Gezeery, A. R., and Al-Ayadhi, L. (2013). The neurotoxic effect of clindamycin-induced gut bacterial imbalance and orally administered propionic acid on DNA damage assessed by the comet assay: protective potency of carnosine and carnitine. *Gut Pathog.* 5:9. doi: 10.1186/1757-4749-5-9
- El-Rashidy, O., El-Baz, F., El-Gendy, Y., Khalaf, R., Reda, D., and Saad, K. (2017). Ketogenic diet versus gluten free casein free diet in autistic children: a case-control study. *Metab. Brain Dis.* 32, 1935–1941. doi: 10.1007/s11011-017-0088-z
- Emery, D. C., Shoemark, D. K., Batstone, T. E., Waterfall, C. M., Coghill, J. A., Cerajewska, T. L., et al. (2017). 16S rRNA next generation sequencing analysis shows bacteria in Alzheimer's post-mortem brain. *Front. Aging Neurosci.* 9:195. doi: 10.3389/fnagi.2017.00195
- Farooq, F. S., Din, J. S., Khan, A. M., Naqvi, S., Shagufta, S., and Mohit, A. (2017). Valproate-induced hyperammonemic encephalopathy. *Cureus* 9:e1593. doi: 10.7759/cureus.1593
- Figueira, J., Jonsson, P., Adolfsson, A. N., Adolfsson, R., Nyberg, L., and Öhman, A. (2016). NMR analysis of the human saliva metabolome distinguishes dementia patients from matched controls. *Mol. Biosyst.* 12, 2562–2571. doi: 10.1039/c6mb00233a
- Filippo, C. D., Cavalieri, D., Paola, M. D., Ramazzotti, M., Poullet, J. B., Massart, S., et al. (2010). Impact of diet in shaping gut microbiota revealed by a comparative study in children from Europe and rural Africa. *Proc. Natl. Acad. Sci. U S A* 107, 14691–14696. doi: 10.1073/pnas.1005963107
- Fisman, M., Ball, M., and Blume, W. (1989). Hyperammonemia and Alzheimer's disease. J. Am. Geriatr. Soc. 37:1102. doi: 10.1111/j.1532-5415.1989.tb06935.x
- Fisman, M., Gordon, B., Feleki, V., Helmes, E., Appell, J., and Rabheru, K. (1985). Hyperammonemia in Alzheimer's disease. *Am. J. Psychiatry* 142, 71–73. doi: 10.1176/ajp.142.1.71
- Fleisher, A. S., Truran, D., Mai, J. T., Langbaum, J. B. S., Aisen, P. S., Cummings, J. L., et al. (2011). Chronic divalproex sodium use and brain atrophy in Alzheimer disease. *Neurology* 77, 1263–1271. doi: 10.1212/wnl.0b013e318230a16c
- Fujii, Y., Nguyen, T. T. T., Fujimura, Y., Kameya, N., Nakamura, S., Arakawa, K., et al. (2019). Fecal metabolite of a gnotobiotic mouse transplanted with gut microbiota from a patient with Alzheimer's disease. *Biosci. Biotech. Biochem.* 83, 2144–2152. doi: 10.1080/09168451.2019.1644149
- Ghalichi, F., Ghaemmaghami, J., Malek, A., and Ostadrahimi, A. (2016). Effect of gluten free diet on gastrointestinal and behavioral indices for children with autism spectrum disorders: a randomized clinical trial. *World J. Pediatr.* 12, 436–442. doi: 10.1007/s12519-016-0040-z
- González-Domínguez, R., García-Barrera, T., Vitorica, J., and Gómez-Ariza, J. L. (2015). Metabolomic screening of regional brain alterations in the APP/PS1 transgenic model of Alzheimer's disease by direct infusion mass spectrometry. *J. Pharm. Biomed. Anal.* 102, 425–435. doi: 10.1016/j. ejogrb.2020.06.039
- Govindarajan, N., Agis-Balboa, R. C., Walter, J., and Fischer, A. (2011). Sodium butyrate improves memory function in an Alzheimer's disease mouse model when administered at an advanced stage of disease progression. J. Alzheimers Dis. 26, 187–197. doi: 10.3233/JAD-2011-110080
- Haijes, H. A., Hasselt, P. M., Jans, J. J. M., and Verhoeven-Duif, N. M. (2019). Pathophysiology of propionic and methylmalonic acidemias. Part 2: treatment strategies. J. Inherit. Metab. Dis. 42, 745–761. doi: 10.1002/jimd.12128
- Harach, T., Marungruang, N., Duthilleul, N., Cheatham, V., Coy, K. D. M., Frisoni, G., et al. (2017). Reduction of Aβ amyloid pathology in APPPS1 transgenic mice in the absence of gut microbiota. *Sci. Rep.* 7:41802. doi: 10.1038/srep41802
- Haran, J. P., Bhattarai, S. K., Foley, S. E., Dutta, P., Ward, D. V., Bucci, V., et al. (2019). Alzheimer's disease microbiome is associated with dysregulation of the anti-inflammatory p-glycoprotein pathway. *mBio* 10:e00632-19. doi: 10.1128/mBio.00632-19
- Hardy, J., and Duff, K. (1994). "Amyloid deposition as the central event in the etiology and pathogenesis of Alzheimer's disease," in *Alzheimer Disease*.

Advances in Alzheimer Disease Therapy, eds E. Giacobini and R. E. Becker (Boston, MA: Birkhäuser), 23–27. doi: 10.1007/978-1-4615-8149-9_4

- Heinritz, S., Weiss, E., Eklund, M., Aumiller, T., Heyer, C., Messner, S., et al. (2016). Impact of a high-fat or high-fiber diet on intestinal microbiota and metabolic markers in a pig model. *Nutrients* 8:317. doi: 10.3390/nu80 50317
- Henagan, T. M., Stefanska, B., Fang, Z., Navard, A. M., Ye, J., Lenard, N. R., et al. (2015). Sodium butyrate epigenetically modulates high-fat diet-induced skeletal muscle mitochondrial adaptation, obesity and insulin resistance through nucleosome positioning. *Br. J. Pharmacol.* 172, 2782–2798. doi: 10.1111/bph.13058
- Hoyles, L., Snelling, T., Umlai, U.-K., Nicholson, J. K., Carding, S. R., Glen, R. C., et al. (2018). Microbiome-host systems interactions: protective effects of propionate upon the blood-brain barrier. *Microbiome* 6:55. doi: 10.1186/s40168-018-0439-y
- Ivashkin, V., Zolnikova, O., Potskherashvili, N., Trukhmanov, A., Sedova, A., and Bueverova, E. (2019). Metabolic activity of intestinal microflora in patients with bronchial asthma. *Clin. Pract.* 9:1126. doi: 10.4081/cp.2019.1126
- Jin, Y., Singh, P., Chung, H.-J., and Hong, S.-T. (2018). blood ammonia as a possible etiological agent for Alzheimer's disease. *Nutrients* 10:564. doi: 10.3390/nu10050564
- Kamen, C. L., Zevy, D. L., Ward, J. M., Bishnoi, I. R., Kavaliers, M., and Ossenkopp, K.-P. (2018). Systemic treatment with the enteric bacterial fermentation product, propionic acid, reduces acoustic startle response magnitude in rats in a dose-dependent fashion: contribution to a rodent model of ASD. *Neurotox. Res.* 35, 353–359. doi: 10.1007/s12640-018-9960-9
- Kamer, A. R., Pirraglia, E., Tsui, W., Rusinek, H., Vallabhajosula, S., Mosconi, L., et al. (2015). Periodontal disease associates with higher brain amyloid load in normal elderly. *Neurobiol. Aging* 36, 627–633. doi: 10.1016/j.neurobiolaging. 2014.10.038
- Kaur, H., Nagamoto-Combs, K., Golovko, S., Golovko, M. Y., Klug, M. G., and Combs, C. K. (2020). Probiotics ameliorate intestinal pathophysiology in a mouse model of Alzheimer's disease. *Neurobiol. Aging* 92, 114–134. doi: 10.1016/j.neurobiolaging.2020.04.009
- Kennedy, B., Dillon, E., Mills, P. J., and Ziegler, M. G. (2001). Catecholamines in human saliva. *Life Sci.* 69, 87–99. doi: 10.1016/s0024-3205(01)01111-0
- Khalil, S. R., Abd-Elhakim, Y. M., Selim, M. E., and Al-Ayadhi, L. Y. (2015). Apitoxin protects rat pups brain from propionic acid-induced oxidative stress: the expression pattern of Bcl-2 and Caspase-3 apoptotic genes. *Neurotoxicology* 49, 121–131. doi: 10.1016/j.neuro.2015.05.011
- Khallaf, W. A., Messiha, B. A., Abo-Youssef, A. M., and El-Sayed, N. S. (2017). Protective effects of telmisartan and tempol on lipopolysaccharide-induced cognitive impairment, neuroinflammation and amyloidogenesis: possible role of brain-derived neurotrophic factor. *Can. J. Physiol. Pharmacol.* 95, 850–860. doi: 10.1139/cjpp-2017-0042
- Kido, J., Nakamura, K., Mitsubuchi, H., Ohura, T., Takayanagi, M., Matsuo, M., et al. (2011). Long-term outcome and intervention of urea cycle disorders in Japan. J. Inherit. Metab. Dis. 35, 777–785. doi: 10.1007/s10545-011-9427-0
- Kirschbaum, J., and Kligman, A. (1963). The pathogenic role of Corynebacterium acnes in acne vulgaris. Arch. Dermatol. 88, 832–833. doi: 10.1001/archderm. 1963.01590240156026
- Knivsberg, A., Reichelt, K., Høien, T., and Nødland, M. (2002). A randomized, controlled study of dietary intervention in autistic syndromes. *Nutr. Neurosci.* 5, 251–261. doi: 10.1080/10284150290028945
- Kornhuber, H. (1996). Propionibacterium acnes in the cortex of patients with Alzheimer's disease. Eur. Arch. Psychiatry Clin. Neurosci. 246, 108–109. doi: 10.1007/BF02274902
- Kowalski, P. C., Dowben, J. S., and Keltner, N. L. (2013). Ammonium: the deadly toxin you don't want to miss when using mood stabilizers. *Perspect. Psychiatr. Care* 49, 221–225. doi: 10.1111/ppc.12040
- Liu, J., Wang, F., Liu, S., Du, J., Hu, X., Xiong, J., et al. (2017). Sodium butyrate exerts protective effect against Parkinson's disease in mice *via* stimulation of glucagon like peptide-1. *J. Neurol. Sci.* 381, 176–181. doi: 10.1016/j.jns.2017. 08.3235
- Liu, P., Wu, L., Peng, G., Han, Y., Tang, R., Ge, J., et al. (2019). Altered microbiomes distinguish Alzheimer's disease from amnestic mild cognitive impairment and health in a Chinese cohort. *Brain Behav. Immun.* 80, 633–643. doi: 10.1016/j.bbi.2019.05.008

- Lu, X., Qi, X., Yi, X., Jian, Z., and Gao, T. (2019). Transcellular traversal of the blood-brain barrier by the pathogenic Propionibacterium acnes. J. Cell. Biochem. 120, 8457–8465. doi: 10.1002/jcb.28132
- Ma, F., Wu, T., Zhao, J., Ji, L., Song, A., Zhang, M., et al. (2017). Plasma homocysteine and serum folate and vitamin B12 levels in mild cognitive impairment and Alzheimer's disease: a case-control study. *Nutrients* 9:725. doi: 10.3390/nu9070725
- MacFabe, D. F., Cain, N. E., Boon, F., Ossenkopp, K.-P., and Cain, D. P. (2011). Effects of the enteric bacterial metabolic product propionic acid on object-directed behavior, social behavior, cognition and neuroinflammation in adolescent rats: relevance to autism spectrum disorder. *Behav. Brain Res.* 217, 47–54. doi: 10.1016/j.bbr.2010.10.005
- MacFabe, D., Rodriguez-Capote, K., Hoffman, J. E., Franklin, A. E., Mohammad-Asef, Y., Taylor, A. R., et al. (2008). A novel rodent model of autism: intraventricular infusions of propionic acid increase locomotor activity and induce neuroinflammation and oxidative stress in discrete regions of adult rat brain. Am. J. Biochem. Biotechnol. 4, 146–166. doi: 10.3844/ajbbsp.2008. 146.166
- Madmoli, M., Modheji, Y., Rafi, A., Feyzi, R., Darabiyan, P., and AfsharNia, A. (2019). Diabetes and its predictive role in the incidence of Alzheimer's disease. *Med. Sci.* 23, 30–34.
- Maldonado, C., Guevara, N., Queijo, C., González, R., Fagiolino, P., and Vázquez, M. (2016). Carnitine and/or acetylcarnitine deficiency as a cause of higher levels of ammonia. *Biomed Res. Int.* 2016:2920108. doi: 10.1155/2016/2920108
- Maldonado, C., Vázquez, M., and Fagiolino, P. (2020). Potential therapeutic role of carnitine and acetylcarnitine in neurological disorders. *Curr. Pharm. Des.* 26, 1277–1285. doi: 10.2174/1381612826666200212114038
- Mandal, P. K., Saharan, S., Tripathi, M., and Murari, G. (2015). Brain glutathione levels—a novel biomarker for mild cognitive impairment and Alzheimer's disease. *Biol. Psychiatry* 78, 702–710. doi: 10.1016/j.biopsych.2015.04.005
- Mani-López, E., García, H. S., and López-Malo, A. (2012). Organic acids as antimicrobials to control Salmonella in meat and poultry products. *Food Res. Int.* 45, 713–721. doi: 10.1016/j.foodres.2011.04.043
- Marcaida, G., Felipo, V., Hermenegildo, C., Minana, M. D., and Grisolia, S. (1992). Acute ammonia toxicity is mediated by the NMDA type of glutamate receptors. *FEBS Lett.* 296, 67–68. doi: 10.1016/0014-5793(92)80404-5
- Martin, J., Kagerbauer, S. M., Gempt, J., Podtschaske, A., Hapfelmeier, A., and Schneider, G. (2018). Oxytocin levels in saliva correlate better than plasma levels with concentrations in the cerebrospinal fluid of patients in neurocritical care. J. Neuroendocrinol. 30:e12596. doi: 10.1111/jne.12596
- Mepham, J. R., Boon, F. H., Foley, K. A., Cain, D. P., Macfabe, D. F., and Ossenkopp, K.-P. (2019). Impaired spatial cognition in adult rats treated with multiple intracerebroventricular (ICV) infusions of the enteric bacterial metabolite, propionic acid and return to baseline after 1 week of no treatment: contribution to a rodent model of ASD. *Neurotox. Res.* 35, 823–837. doi: 10.1007/s12640-019-0002-z
- Mitsui, R., Ono, S., Karaki, S. I., and Kuwahara, A. (2005). Propionate modulates spontaneous contractions via enteric nerves and prostaglandin release in the rat distal colon. Jpn. J. Physiol. 55, 331–338. doi: 10.2170/jjphysiol.RP000205
- Montgomery, S. A., Thal, L., and Amrein, R. (2003). Meta-analysis of double blind randomized controlled clinical trials of acetyl-L-carnitine versus placebo in the treatment of mild cognitive impairment and mild Alzheimer's disease. *Int. Clin. Psychopharmacol.* 18, 61–71. doi: 10.1097/00004850-200303000-00001
- Morland, C., Frøland, A.-S., Pettersen, M. N., Storm-Mathisen, J., Gundersen, V., Rise, F., et al. (2018). Propionate enters GABAergic neurons, inhibits GABA transaminase, causes GABA accumulation and lethargy in a model of propionic acidemia. *Biochem. J.* 475, 749–758. doi: 10.1042/BCJ20170814
- Morrison, D. J., and Preston, T. (2016). Formation of short chain fatty acids by the gut microbiota and their impact on human metabolism. *Gut Microbes* 7, 189–200. doi: 10.1080/19490976.2015.1134082
- Müller, M., Hernández, M. A. G., Goossens, G. H., Reijnders, D., Holst, J. J., Jocken, J. W. E., et al. (2019). Circulating but not faecal short-chain fatty acids are related to insulin sensitivity, lipolysis and GLP-1 concentrations in humans. *Sci. Rep.* 9:12515. doi: 10.1038/s41598-019-48775-0
- Norenberg, M. D., and Martinez-Hernandez, A. (1979). Fine structural localization of glutamine synthetase in astrocytes of rat brain. *Brain Res.* 161, 303–310. doi: 10.1016/0006-8993(79)90071-4

- Odamaki, T., Kato, K., Sugahara, H., Hashikura, N., Takahashi, S., Xiao, J.-Z., et al. (2016). Age-related changes in gut microbiota composition from newborn to centenarian: a cross-sectional study. *BMC Microbiol.* 16:90. doi: 10.1186/s12866-016-0708-5
- Pardoe, H. R., Berg, A. T., and Jackson, G. D. (2013). Sodium valproate use is associated with reduced parietal lobe thickness and brain volume. *Neurology* 80, 1895–1900. doi: 10.1212/WNL.0b013e318292a2e5
- Perry, R. J., Borders, C. B., Cline, G. W., Zhang, X. M., Alves, T. C., Petersen, K. F., et al. (2016). Propionate increases hepatic pyruvate cycling and anaplerosis and alters mitochondrial metabolism. *J. Biol. Chem.* 291, 12161–12170. doi: 10.1074/jbc.M116.720631
- Pingitore, A., Chambers, E. S., Hill, T., Maldonado, I. R., Liu, B., Bewick, G., et al. (2016). The diet-derived short chain fatty acid propionate improves β -cell function in humans and stimulates insulin secretion from human islets *in vitro*. *Diabetes Obes. Metab.* 19, 257–265. doi: 10.1111/dom.12811
- Reichardt, N., Duncan, S. H., Young, P., Belenguer, A., Leitch, C. M., Scott, K. P., et al. (2014). Erratum: phylogenetic distribution of three pathways for propionate production within the human gut microbiota. *ISME J.* 8, 1352–1352. doi: 10.1038/ismej.2014.14
- Reis, J. A., Paula, A. T., Casarotti, S. N., and Penna, A. L. B. (2012). Lactic acid bacteria antimicrobial compounds: characteristics and applications. *Food Eng. Rev.* 4, 124–140. doi: 10.1007/s12393-012-9051-2
- Revtovich, A. V., Lee, R., and Kirienko, N. V. (2019). Interplay between mitochondria and diet mediates pathogen and stress resistance in *Caenorhabditis elegans*. *PLoS Genet*. 15:e1008011. doi: 10.1371/journal. pgen.1008011
- Ribas, G. S., Manfredini, V., de Mari, J. F., Wayhs, C. Y., Vanzin, C. S., Biancini, G. B., et al. (2010). Reduction of lipid and protein damage in patients with disorders of propionate metabolism under treatment: a possible protective role of L-carnitine supplementation. *Int. J. Dev. Neurosci.* 28, 127–132. doi: 10.1016/j.ijdevneu.2010.01.002
- Rigo, F. K., Pasquetti, L., Malfatti, C. R. M., Fighera, M. R., Coelho, R. C., Petri, C. Z., et al. (2006). Propionic acid induces convulsions and protein carbonylation in rats. *Neurosci. Lett.* 408, 151–154. doi: 10.1016/j.neulet.2006. 08.075
- Ringer, A. (1912). The chemistry of gluconeogenesis: I. The quantitative conversion of propionic acid into glucose. J. Biol. Chem. 12, 511–515. doi: 10.1016/j.mehy.2018.10.001
- Robinson, S. R. (2000). Neuronal expression of glutamine synthetase in Alzheimer's disease indicates a profound impairment of metabolic interactions with astrocytes. *Neurochem. Int.* 36, 471–482. doi: 10.1016/s0197-0186(99)00150-3
- Roduit, C., Frei, R., Ferstl, R., Loeliger, S., Westermann, P., Rhyner, C., et al. (2018). High levels of butyrate and propionate in early life are associated with protection against atopy. *Allergy* 74, 799–809. doi: 10.1111/all.13660
- Roe, C. R., Millington, D. S., Maltby, D. A., Bohan, T. P., and Hoppel, C. L. (1984). L-carnitine enhances excretion of propionyl coenzyme A as propionylcarnitine in propionic acidemia. J. Clin. Invest. 73, 1785–1788. doi: 10.1172/JCI111387
- Roquilly, A., Perbet, S., Simonneau, F., Cinotti, R., Sebille, V., Volteau, C., et al. (2013). Ammonia plasma concentration and prolonged infusion of remifentanil in patients with acute kidney injury. *Miner. Anestesiol.* 79, 884–890.
- Salonen, A., Lahti, L., Salojärvi, J., Holtrop, G., Korpela, K., Duncan, S. H., et al. (2014). Impact of diet and individual variation on intestinal microbiota composition and fermentation products in obese men. *ISME J.* 8, 2218–2230. doi: 10.1038/ismej.2014.63
- Sanna, S., Zuydam, N. R. V., Mahajan, A., Kurilshikov, A., Vila, A. V., Vôsa, U., et al. (2019). Causal relationships among the gut microbiome, short-chain fatty acids and metabolic diseases. *Nat. Genet.* 51, 600–605. doi: 10.1038/s41588-019-0350-x
- Schwoerer, J. S., Candadai, S. C., and Held, P. K. (2018). Long-term outcomes in Amish patients diagnosed with propionic acidemia. *Mol. Genet. Metab. Rep.* 16, 36–38. doi: 10.1016/j.ymgmr.2018.05.004
- Sethi, K. D., Ray, R., Roesel, R. A., Carter, A. L., Gallagher, B. B., Loring, D. W., et al. (1989). Adult-onset chorea and dementia with propionic acidemia. *Neurology* 39, 1343–1343. doi: 10.1212/wnl.39.10.1343
- Shams, S., Foley, K. A., Kavaliers, M., Macfabe, D. F., and Ossenkopp, K.-P. (2019). Systemic treatment with the enteric bacterial metabolic product propionic

acid results in reduction of social behavior in juvenile rats: contribution to a rodent model of autism spectrum disorder. *Dev. Psychobiol.* 61, 688–699. doi: 10.1002/dev.21825

- Skowrońska, M., and Albrecht, J. (2013). Oxidative and nitrosative stress in ammonia neurotoxicity. *Neurochem. Int.* 62, 731–737. doi: 10.1016/j.jid.2018. 04.042
- Spagnoli, A., Lucca, U., Menasce, G., Bandera, L., Cizza, G., Forloni, G., et al. (1991). Long-term acetyl-L-carnitine treatment in Alzheimer's disease. *Neurology* 41, 1726–1732. doi: 10.1212/wnl.41.11.1726
- Suárez, I., Bodega, G., and Fernández, B. (2002). Glutamine synthetase in brain: effect of ammonia. *Neurochem. Int.* 41, 123–142. doi: 10.1016/s0197-0186(02)00033-5
- Sunkara, L. T., Jiang, W., and Zhang, G. (2012). Modulation of antimicrobial host defense peptide gene expression by free fatty acids. *PLoS One* 7:e49558. doi: 10.1371/journal.pone.0049558
- Syeda, T., Sanchez-Tapia, M., Pinedo-Vargas, L., Granados, O., Cuervo-Zanatta, D., Rojas-Santiago, E., et al. (2018). Bioactive food abates metabolic and synaptic alterations by modulation of gut microbiota in a mouse model of Alzheimer's disease. J. Alzheimers Dis. 66, 1657–1682. doi: 10.3233/JAD-180556
- Takahashi, N. (2015). Oral microbiome metabolism: from "who are they?" to "what are they doing?" *J. Dent. Res.* 94, 1628–1637. doi: 10.1177/0022034515606045
- Takeda, I., Stretch, C., Barnaby, P., Bhatnager, K., Rankin, K., Fu, H., et al. (2009). Understanding the human salivary metabolome. NMR Biomed. 22, 577–584. doi: 10.1002/nbm.1369
- Tariot, P. N., Schneider, L. S., Cummings, J., Thomas, R. G., Raman, R., Jakimovich, L. J., et al. (2011). Chronic divalproex sodium to attenuate agitation and clinical progression of Alzheimer disease. *Arch. Gen. Psychiatry* 68, 853–861. doi: 10.1001/archgenpsychiatry. 2011.72
- Tian, Z., Zhuang, X., Luo, M., Yin, W., and Xiong, L. (2020). The propionic acid and butyric acid in serum but not in feces are increased in patients with diarrhea-predominant irritable bowel syndrome. *BMC Gastroenterol.* 20:73. doi: 10.1186/s12876-020-01212-3
- Tirosh, A., Calay, E. S., Tuncman, G., Claiborn, K. C., Inouye, K. E., Eguchi, K., et al. (2019). The short-chain fatty acid propionate increases glucagon and FABP4 production, impairing insulin action in mice and humans. *Sci. Transl. Med.* 11:eaav0120. doi: 10.1126/scitranslmed.aav0120
- Tsai, P.-S., Liu, I.-C., Chiu, C.-H., Huang, C.-J., and Wang, M.-Y. (2016). Effect of valproic acid on dementia onset in patients with bipolar disorder. J. Affect. Dis. 201, 131–136. doi: 10.1016/j.jad.2016.05.010
- U.S. Department of Agriculture. (2008). *Propionic Acid.* Available online at: https://www.ams.usda.gov/sites/default/files/media/Prop% 20acid%20report.pdf. Accessed June 10, 2020.
- U.S. Department of Agriculture Technical Advisory Committee. (2002). *Calcium Propionate.* Available online at: https://www.ams.usda.gov/sites/default/files/ media/Calcium%20Propionate%20TR.pdf. Accessed August 30, 2020.
- Valstar, M. H., de Bakker, B. S., Steenbakkers, R. J., de Jong, K. H., Smit, L. A., Nulent, T. J. K., et al. (2020). The tubarial salivary glands: a potential new organ at risk for radiotherapy. *Radiother. Oncol.* doi: 10.1016/j.radonc.2020.09.034. [Epub ahead of print].
- Vogt, N. M., Kerby, R. L., Dill-McFarland, K. A., Harding, S. J., Merluzzi, A. P., Johnson, S. C., et al. (2017). Gut microbiome alterations in Alzheimer's disease. *Sci. Rep.* 7:13537. doi: 10.1038/s41598-017-13601-y
- Wang, L., Christophersen, C. T., Sorich, M. J., Gerber, J. P., Angley, M. T., and Conlon, M. A. (2012). Elevated fecal short chain fatty acid and ammonia concentrations in children with autism spectrum disorder. *Dig. Dis. Sci.* 57, 2096–2102. doi: 10.1007/s10620-012-2167-7
- Wang, J., Ye, F., Cheng, X., Zhang, X., Liu, F., Liu, G., et al. (2016). The effects of LW-AFC on intestinal microbiome in senescence-accelerated mouse prone 8 strain, a mouse model of Alzheimer's disease. J. Alzheimers Dis. 53, 907–919. doi: 10.3233/JAD-160138
- Whiteley, P., Haracopos, D., Knivsberg, A.-M., Reichelt, K. L., Parlar, S., Jacobsen, J., et al. (2010). The ScanBrit randomized, controlled, singleblind study of a gluten- and casein-free dietary intervention for children with autism spectrum disorders. *Nutr. Neurosci.* 13, 87–100. doi: 10.1179/147683010X12611460763922

- Wishart, D. S., Feunang, Y. D., Marcu, A., Guo, A. C., Liang, K., Vázquez-Fresno, R., et al. (2017). HMDB 4.0: the human metabolome database for 2018. *Nucleic Acids Res.* 46, D608–D617. doi: 10.1093/nar/gkx1089
- Wolever, T. M., Josse, R. G., Leiter, L. A., and Chiasson, J. (1997). Time of day and glucose tolerance status affect serum short-chain fatty concentrations in humans. *Metabolism* 46, 805–811. doi: 10.1016/s0026-0495(97)90127-x
- Wu, G. D., Chen, J., Hoffmann, C., Bittinger, K., Chen, Y.-Y., Keilbaugh, S. A., et al. (2011). Linking long-term dietary patterns with gut microbial enterotypes. *Science* 334, 105–108. doi: 10.1126/science.1208344
- Wyse, A. T. S., Brusque, A. M., Silva, C. G., Streck, E. L., Wajner, M., and Wannmacher, C. M. D. (1998). Inhibition of Na,K-ATPase from rat brain cortex by propionic acid. *Neuroreport* 9, 1719–1721. doi: 10.1097/00001756-199806010-00009
- Yeagle, P. (2015). Microbiome of uncontacted Amerindians. *Science* 348, 298–298. doi: 10.1126/science.348.6232.298-a
- Yilmaz, A., Geddes, T., Han, B., Bahado-Singh, R. O., Wilson, G. D., Imam, K., et al. (2017). Diagnostic biomarkers of Alzheimer's disease as identified in saliva using 1H NMR-based metabolomics. *J. Alzheimers Dis.* 58, 355–359. doi: 10.3233/JAD-161226

- Zhan, G., Yang, N., Li, S., Huang, N., Fang, X., Zhang, J., et al. (2018). Abnormal gut microbiota composition contributes to cognitive dysfunction in SAMP8 mice. *Aging* 10, 1257–1267. doi: 10.18632/aging. 101464
- Zhuang, Z.-Q., Shen, L.-L., Li, W.-W., Fu, X., Zeng, F., Gui, L., et al. (2018). Gut microbiota is altered in patients with Alzheimer's disease. *J. Alzheimers Dis.* 63, 1337–1346. doi: 10.3233/JAD-180176

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2021 Killingsworth, Sawmiller and Shytle. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.