



Draft Metagenomes of Endolithic Cyanobacteria and Cohabitants from Hyper-Arid Deserts

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ABSTRACT Cyanobacteria are essential to microbial communities inhabiting translucent rocks in hyper-arid deserts. Metagenomic studies revealed unique adaptations of these cyanobacteria, but validation of the corresponding metabolic pathways remained challenging without access to isolates. Here, we present high-quality metagenome-assembled genomes for cyanobacteria, and their heterotrophic companions, isolated from endolithic substrates.

In the most arid deserts, where environmental conditions are extreme, microbial communities find refuge inside rocks as a survival strategy (1). The rock habitat protects microorganisms from high UV radiation and drastic temperature fluctuations and promotes water retention within the rock matrix (2). Molecular studies of endolithic communities (within rock) revealed ecosystems spanning all domains of life and multiple trophic levels (3–5). The communities are based on the primary production of cyanobacteria, and sometimes algae, and are constituted of an assemblage of heterotrophic bacteria and/or archaea and viruses (6–10). Endolithic communities are highly specific to their lithic substrate, with fine-scale diversification of the microbial reservoir driven by substrate properties (3, 10).

Cyanobacteria inhabiting endolithic substrates in arid deserts are mostly members of the orders *Chroococcales* (*Chroococcidiopsis* and *Gloeocapsa*), *Nostocales*, and *Oscillatoriales* (1). Metagenomic studies of endolithic communities revealed unique adaptations of these cyanobacteria, and a large number of pathways for secondary metabolites, nonribosomal peptides, and polyketides are encoded in their genomes (7, 10). However, validation of these pathways remained challenging without access to isolates. Here, we present the metagenome-assembled genomes (MAGs) of cyanobacteria isolated from endolithic substrates collected in the Atacama and Negev Deserts (Table 1). Because these isolates are not purified cultures, their companions—heterotrophic bacteria—were also sequenced.

Cyanobacterial isolates were obtained by incubating ground colonized rock samples collected in the Atacama and Negev Deserts (3, 4) in Bold's basal medium (11) and in BG11 liquid medium (12) for 5 weeks at 25°C under 24 μM photons/m²/s of white light (WL) using Philips daylight deluxe linear fluorescent T12 40-W light bulbs and a combination of neutral-density filters (299 1.2ND and 298 0.15ND; Lee Filters, Burbank, CA). Single colonies from 1% agar BG11 plates were then transferred to liquid BG11 medium and grown under WL; it is important to note that these were not anoxic cyanobacterial cultures but, rather, a mixture of cyanobacteria and heterotrophic bacteria. Total DNA was extracted from cell pellets using the PowerSoil DNA extraction kit (MoBio Laboratories, Inc., Solana Beach, CA). Nextera libraries, with Ranger size technology, were made with total DNA and sequenced to a 2-Gb depth using 2 \times 150-nucleotide (nt) reads on an Illumina NovaSeq instrument at the Department of Energy (DOE) Joint Genome Institute (JGI). Sequence quality control was performed with the BBTools package (<https://jgi.doe.gov/data-and-tools/bbtools/>), and sequence reads were assembled with metaSPAdes version 3.13.0 using the “metagenome” flag and running the assembly module without error correction and with kmer sizes 33, 55, 77, 99, and 127 (13).

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TABLE 1 Metagenome and MAG statistics of endolithic cyanobacterial isolates from the Atacama Desert, Chile, and the Negev Desert, Israel

| Sample name | Substrate | IMG taxon ID | Metagenome size (Mbp) | Bin ID | Taxon/genus | MAG completion (%) | MAG contamination (%) | MAG size (Mbp) | MAG gene count | MAG scaffold count |
|-------------|------------|--------------|-----------------------|--------------|--------------------------|--------------------|-----------------------|----------------|----------------|--------------------|
| C-VL-3P3 | Calcite | 3300039404 | 11 | 3300039404_1 | <i>Chroococcidiopsis</i> | 99.48 | 1.63 | 6.6 | 6,630 | 157 |
| | | | | 3300039404_2 | <i>Deinococcus</i> | 97.67 | 0.99 | 4.1 | 4,214 | 68 |
| G-Km37-3P1 | Gypsum | 3300039405 | 18.2 | 3300039405_1 | <i>Methylobacterium</i> | 100 | 0 | 6.9 | 6,942 | 64 |
| | | | | 3300039405_2 | <i>Deinococcus</i> | 97.67 | 0.99 | 4.1 | 4,212 | 67 |
| G-Km37-3P3 | Gypsum | 3300039416 | 42.9 | 3300039416_1 | <i>Chroococcidiopsis</i> | 99.48 | 1.63 | 6.6 | 6,618 | 153 |
| | | | | 3300039416_2 | <i>Deinococcus</i> | 97.67 | 0.99 | 4.1 | 4,213 | 66 |
| G-MTQ-3P1 | Gypsum | 3300038622 | 16.2 | 3300038622_1 | <i>Chroococcidiopsis</i> | 99.48 | 1.63 | 6.6 | 6,601 | 163 |
| | | | | 3300038622_2 | <i>Methylobacterium</i> | 52.45 | 1.25 | 4.2 | 4,745 | 668 |
| G-MTQ-3P2 | Gypsum | 3300037877 | 9.9 | 3300037877_1 | <i>Chroococcidiopsis</i> | 99.48 | 1.63 | 6.6 | 6,608 | 161 |
| H-SG-1P1 | Gypsum | 3300039034 | 38.8 | 3300039034_1 | <i>Chroococcidiopsis</i> | 99.48 | 1.63 | 6.6 | 6,605 | 160 |
| H-SG-2P1 | Gypsum | 3300039035 | 43.1 | 3300039035_2 | <i>Chroococcidiopsis</i> | 99.48 | 1.63 | 6.6 | 6,619 | 155 |
| | | | | 3300039035_3 | <i>Deinococcus</i> | 97.67 | 0.99 | 4.1 | 4,230 | 70 |
| I-MTQ-2P3 | Ignimbrite | 3300039417 | 20 | 3300039417_1 | <i>Chroococcidiopsis</i> | 97.11 | 4.52 | 7.6 | 7,825 | 531 |
| | | | | 3300039417_2 | <i>Deinococcus</i> | 98.52 | 0.99 | 4.2 | 4,428 | 95 |
| | | | | 3300039417_3 | <i>Thermomicrobiales</i> | 63.91 | 1.89 | 2.4 | 2,800 | 503 |
| I-MTQ-3P1 | Ignimbrite | 3300039418 | 28.2 | 3300039418_3 | <i>Deinococcus</i> | 97.67 | 0.99 | 4.1 | 4,241 | 70 |
| I-MTQ-3P3 | Ignimbrite | 3300039424 | 30.6 | 3300039424_2 | <i>Aquamicrobium</i> | 99.59 | 0.75 | 4.4 | 4,417 | 7 |
| | | | | 3300039424_3 | <i>Deinococcus</i> | 97.25 | 0.99 | 4.1 | 4,315 | 99 |
| | | | | 3300039424_4 | <i>Microcella</i> | 99.38 | 0.25 | 2.5 | 2,464 | 5 |
| I-MTQ-4P3 | Ignimbrite | 3300039425 | 10.3 | 3300039425_1 | <i>Deinococcus</i> | 97.67 | 0.99 | 4.1 | 4,237 | 70 |
| S-NGV-2P1 | Sandstone | 3300039401 | 43 | 3300039401_1 | <i>Chroococcidiopsis</i> | 99.48 | 1.63 | 6.6 | 6,617 | 153 |
| | | | | 3300039401_2 | <i>Deinococcus</i> | 97.67 | 0.99 | 4.1 | 4,214 | 67 |
| S-NGV-2P2 | Sandstone | 3300039032 | 6.8 | 3300039032_1 | <i>Chroococcidiopsis</i> | 99.48 | 1.63 | 6.5 | 6,596 | 163 |
| S-NGV-3P2 | Sandstone | 3300039033 | 6.8 | 3300039033_1 | <i>Chroococcidiopsis</i> | 99.48 | 1.63 | 6.6 | 6,618 | 158 |

MetaBAT v2.12.1 (14) was used for binning. MAGs were evaluated with CheckM v1.0.12 (15) and annotated with GTDB-Tk version v0.2.2 and the GTDB database release 86 (16). Default parameters were used for all software unless otherwise noted. Only high-quality (HQ) and medium-quality (MQ) bins were reported based on Minimum Information about a Metagenome-Assembled Genome (MIMAG) standards (17).

High-quality MAGs of cyanobacteria, together with MAGs of heterotrophic bacteria, were recovered from most samples (Table 1). All cyanobacteria belonged to the *Chroococcidiopsis* genus; *Deinococcus* was the most common heterotrophic bacterium, but we also found members of the *Proteobacteria*, *Actinobacteria*, and *Chloroflexi*, illustrating the diversity of these communities.

Data availability. The raw sequencing data are available from the National Centre for Biotechnology Information under BioProject numbers [PRJNA654119](https://ncbi.nlm.nih.gov/bioproject/PRJNA654119), [PRJNA654120](https://ncbi.nlm.nih.gov/bioproject/PRJNA654120), [PRJNA654121](https://ncbi.nlm.nih.gov/bioproject/PRJNA654121), [PRJNA654122](https://ncbi.nlm.nih.gov/bioproject/PRJNA654122), [PRJNA654123](https://ncbi.nlm.nih.gov/bioproject/PRJNA654123), [PRJNA654124](https://ncbi.nlm.nih.gov/bioproject/PRJNA654124), [PRJNA677471](https://ncbi.nlm.nih.gov/bioproject/PRJNA677471), [PRJNA677472](https://ncbi.nlm.nih.gov/bioproject/PRJNA677472), [PRJNA677473](https://ncbi.nlm.nih.gov/bioproject/PRJNA677473), [PRJNA677474](https://ncbi.nlm.nih.gov/bioproject/PRJNA677474), [PRJNA677475](https://ncbi.nlm.nih.gov/bioproject/PRJNA677475), [PRJNA677476](https://ncbi.nlm.nih.gov/bioproject/PRJNA677476), [PRJNA677477](https://ncbi.nlm.nih.gov/bioproject/PRJNA677477), and [PRJNA677478](https://ncbi.nlm.nih.gov/bioproject/PRJNA677478). The metagenome coassembly and functional annotation are available from the JGI Genome Portal under the IMG taxon IDs reported in Table 1. To obtain cultures of cyanobacterial isolates, please contact the corresponding author.

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REFERENCES

- Meslier V, DiRuggiero J. 2019. Endolithic microbial communities as model systems for ecology and astrobiology, p 145–168. In Seckbach J, Rampelotto P (ed), *Model ecosystems in extreme environments*. Academic Press, San Diego, CA. <https://doi.org/10.1016/B978-0-12-812742-1.00007-6>.
- Walker JJ, Pace NR. 2007. Endolithic microbial ecosystems. *Annu Rev Microbiol* 61:331–347. <https://doi.org/10.1146/annurev.micro.61.080706.093302>.
- Meslier V, Casero MC, Dailey M, Wierzchos J, Ascaso C, Artieda O, McCullough PR, DiRuggiero J. 2018. Fundamental drivers for endolithic microbial community assemblies in the hyperarid Atacama Desert. *Environ Microbiol* 20:1765–1781. <https://doi.org/10.1111/1462-2920.14106>.
- Qu EB, Omelon CR, Oren A, Meslier V, Cowan DA, Maggs-Kolling G, DiRuggiero J. 2019. Trophic selective pressures organize the composition of endolithic microbial communities from global deserts. *Front Microbiol* 10:2952. <https://doi.org/10.3389/fmicb.2019.02952>.
- Wierzchos J, DiRuggiero J, Vitek P, Artieda O, Souza-Egipsy V, Skaloud P, Tisza M, Davila AF, Vilchez C, Garbayo I, Ascaso C. 2015. Adaptation strategies of endolithic chlorophototrophs to survive the hyperarid and extreme solar radiation environment of the Atacama Desert. *Front Microbiol* 6:934. <https://doi.org/10.3389/fmicb.2015.00934>.
- Crits-Christoph A, Gelsinger DR, Ma B, Wierzchos J, Ravel J, Ascaso C, Artieda O, Davila A, DiRuggiero J. 2016. Functional interactions of archaea, bacteria and viruses in a hypersaline endolithic community. *Environ Microbiol* 18:2064–2077. <https://doi.org/10.1111/1462-2920.13259>.
- Crits-Christoph A, Robinson CK, Ma B, Ravel J, Wierzchos J, Ascaso C, Artieda O, DiRuggiero J. 2016. Phylogenetic and functional substrate specificity for endolithic microbial communities in hyper-arid environments. *Frontiers Microbiol* 7:301. <https://doi.org/10.3389/fmicb.2016.00301>.
- Uritskiy G, Getsin S, Munn A, Gomez-Silva B, Davila A, Glass B, Taylor J, DiRuggiero J. 2019. Halophilic microbial community compositional shift after a rare rainfall in the Atacama Desert. *ISME J* 13:2737–2749. <https://doi.org/10.1038/s41396-019-0468-y>.
- Uritskiy G, Tisza MJ, Gelsinger DR, Munn A, Taylor J, DiRuggiero J. 2020. Cellular life from the three domains and viruses are transcriptionally active in a hypersaline desert community. *Environ Microbiol* <https://doi.org/10.1111/1462-2920.15023>.
- Ertekin E, Meslier V, Browning A, Treadgold J, DiRuggiero J. 2021. Rock structure drives the taxonomic and functional diversity of endolithic microbial communities in extreme environments. *Environ Microbiol* <https://doi.org/10.1111/1462-2920.15287>.
- Cox ER, Bold HC. 1966. Taxonomic investigation of *Stigeoclonium*. In *Phycological studies VII*, vol 10. University of Texas, Austin, Texas.
- Rippka R, Deruelles J, Waterbury JB, Herdman M, Stainer RY. 1979. Generic assignments, strain histories and properties of pure cultures of cyanobacteria. *J Gen Microbiol* 111:1–61. <https://doi.org/10.1099/00221287-111-1-1>.
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Pribelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner P. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J Comput Biol* 19:455–477. <https://doi.org/10.1089/cmb.2012.0021>.
- Kang DD, Froula J, Egan R, Wang Z. 2015. MetaBAT, an efficient tool for accurately reconstructing single genomes from complex microbial communities. *PeerJ* 3:e1165. <https://doi.org/10.7717/peerj.1165>.
- Parks DH, Imelfort M, Skennerton CT, Hugenholtz P, Tyson GW. 2015. CheckM: assessing the quality of microbial genomes recovered from isolates, single cells, and metagenomes. *Genome Res* 25:1043–1055. <https://doi.org/10.1101/gr.186072.114>.
- Chaumeil P-A, Mussig AJ, Hugenholtz P, Parks DH. 2019. GTDB-Tk: a toolkit to classify genomes with the Genome Taxonomy Database. *Bioinformatics* 36:1925–1927. <https://doi.org/10.1093/bioinformatics/btz848>.
- Bowers RM, Kyrpides NC, Stepanauskas R, Harmon-Smith M, Doud D, Reddy TBK, Schulz F, Jarett J, Rivers AR, Eloë-Fadrosch EA, Tringe SG, Ivanova NN, Copeland A, Clum A, Becraft ED, Malmstrom RR, Birren B, Podar M, Bork P, Weinstock GM, Garrity GM, Dodsworth JA, Yooshep S, Sutton G, Glöckner FO, Gilbert JA, Nelson WC, Hallam SJ, Jungbluth SP, Ettema TJG, Tighe S, Konstantinidis KT, Liu W-T, Baker BJ, Rattei T, Eisen JA, Hedlund B, McMahon KD, Fierer N, Knight R, Finn R, Cochrane G, Karsch-Mizrachi I, Tyson GW, Rinke C, Lapidus A, Meyer F, Yilmaz P, Parks DH, Eren AM, Genome Standards Consortium, et al. 2017. Minimum information about a single amplified genome (MISAG) and a metagenome-assembled genome (MIMAG) of bacteria and archaea. *Nat Biotechnol* 35:725–731. <https://doi.org/10.1038/nbt.3893>.