## Letter

**RESEARCH LETTER** 

Metabolomic Signatures of Myocardial Glucose Uptake on Fluorine-18 Fluorodeoxyglucose Positron Emission Tomography

Fluorine-18 fluorodeoxyglucose positron emission tomography (FDG-PET) plays a critical role in evaluating myocardial inflammation. However, to highlight pathologic myocardial glucose uptake (MGU), physiological MGU must be suppressed, often accomplished through a ketogenic diet (KD).<sup>1</sup> Failure to suppress physiological MGU occurs in ~20% of subjects even following strict, highly supervised dietary interventions,<sup>1,2</sup> potentially leading to misdiagnosis, repeat scans, and unnecessary costs.

In a crossover trial (KEETO-CROSS [Ketogenic Endogenous versus Exogenous Therapies for Myocardial Glucose Suppression]; NCT04275453), we showed that ketone ester (KE) drink (exogenous ketosis) was inferior to the KD (endogenous ketosis) for suppressing MGU.<sup>2</sup> However, circulating ketone (beta-hydroxybutyrate [BHB]) levels strongly predicted myocardial glucose suppression (MGS) within each arm and could be used upstream of FDG-PET to assess myocardial preparation. Furthermore, the ketosis required to suppress the myocardium varied by technique (with significantly higher levels required for KE). To understand whether a common metabolite signature might underlie MGS irrespective of technique, we performed targeted metabolomic plasma profiling paired with 57 FDG-PET scans from 20 participants in KEETO-CROSS. We hypothesized that ketone and fatty acid oxidative markers would associate with MGS, suggesting a mechanism of substrate competition for successful suppression.

In KEETO-CROSS, participants free of cardiovascular disease (in whom any MGU could be assumed physiological) were randomly assigned to the KE (one visit) or KD (2 visits at 24 and 72 hours) arm, and

crossed over after a >1-week washout period.<sup>2</sup> All 20 participants completed the KE arm, whereas 19 and 18 participants completed the 24- and 72-hour KD visits, respectively. The study was approved by an institutional review board, and informed consent was obtained. At these 3 visits, FDG-PET was performed and MGU was analyzed according to a protocol previously described.<sup>2</sup>

Quantitative metabolomic profiling of 30 amino acids and 57 acylcarnitines was performed in fasting, frozen plasma collected at these visits by using liquid chromatography/mass spectrometry (1290/6495 Triple Quadrupole LC/MS, Agilent Technologies). Absolute metabolite quantification was achieved by the addition of stable isotope-labeled internal standards. Twenty-six metabolites with >50% of samples below limits of detection were not analyzed, leaving 61 metabolites for analysis. Metabolite levels below assay limits were analyzed as the lower limit of detection.

Box-Cox transformation and standardization of metabolites into z scores were performed. Linear mixed effects regression models<sup>3</sup> were used to assess the association between each metabolite and MGU, controlling for study visit as a fixed effect, modeling participant as a random effect, and using an independent covariance structure, where applicable. A Benjamini-Hochberg false discovery rate cutoff of 0.05 defined significance. C statistics were calculated to assess the discriminative capability of metabolites for achieving MGS, accounting for clustering at the participant level. Analyses were performed by using STATA version 14 (StataCorp).

The mean age was  $30 \pm 7$  years, 50% were female, 45% were non-White, and the mean fasting time before FDG injection was 16.0  $\pm$  0.9 hours.<sup>2</sup> MGS failure occurred in 11 of 20 scans with the KE and 8 of 37 with the KD. **Figure 1A** depicts volcano plots for the relationship between 61 metabolites and MGU for the 2 KD visits (combined) and the KE visit. During the KD, two C4-OH carnitine species (hydroxybutyrylcarnitine and hydroxy-isobutyrylcarnitine) were negatively associated with MGU. Hydroxybutyrylcarnitines are generated by ketone and fatty acid oxidation, whereas hydroxy-isobutyrylcarnitines are generated during breakdown of the branchedchain amino acid (BCAA) valine. In addition, the long-chain acylcarnitines C16:1-OH and C18:1-OH



(markers of long-chain fatty acid oxidation) were inversely related to MGU. With KE, both C4-OH carnitine species, as well as the short-chain acylcarnitine C6-carnitine, were negatively associated with MGU, whereas proline was positively associated with MGU. Medium-chain acylcarnitines, dicarboxylated acylcarnitine species, and most amino acids (including BCAAs) were not associated with MGU.

Given their common relationship to MGS with both techniques, we assessed C statistics for MGS with C4-OH isobutyrylcarnitine and C4-OH butyrylcarnitine. Both were predictive of MGS (C statistics of 0.83 [95% CI: 0.65-1.00] and 0.81 [0.70-0.92], respectively) in all 57 FDG-PET scans, and corresponding odds ratios were 4.93 (95% CI: 0.62-39.43) and 9.59 (95% CI: 3.54-25.97) for MGS per SD increase. Although we previously found that BHB achieved comparable C statistics in the KD and KE arms separately,<sup>2</sup> BHB was not predictive in a combined model (C statistic: 0.57; 95% CI: 0.41-0.74). Furthermore, although BHB levels were significantly higher on KE than the 24- and 72-hour KD visits (P < 0.001 and P = 0.001), C4-OH isobutyrylcarnitine (although not butyrylcarnitine) levels were higher during KD visits than during KE (P = 0.018 and P = 0.003).

In KEETO-CROSS, combined markers of enhanced ketone/fatty acid oxidation and BCAA catabolism strongly predicted MGS, had robust discriminatory value, and demonstrated utility in measuring before FDG-PET to reduce the risk of false-positive scans. These findings suggest that strategies to enhance these oxidative pathways, rather than augmenting circulating metabolite levels alone,<sup>1</sup> may be more effective in improving MGS. In addition, our results showing higher C4-OH isobutyrylcarnitine levels with KD explain why, despite achieving substantially higher BHB levels, KE was inferior to KD for MGS (Figure 1B). Finally, these biomarkers may act as novel surrogate endpoints during early-phase studies to prioritize high-yield MGS techniques. Limitations include study of a healthy cohort and study of only 2 MGS strategies.

Mahesh K. Vidula, MD Daniel P. Kelly, MD Zoltan Arany, MD, PhD Kenneth B. Margulies, MD Svati H. Shah, MD, MS, MHS Thomas P. Cappola, MD Paco E. Bravo, MD \*Senthil Selvaraj, MD, MS, MA \*Division of Cardiology, Department of Medicine Duke University School of Medicine Duke Molecular Physiology Institute 300 North Duke Street, Carmichael Building Durham, North Carolina 27701, USA E-mail: Senthil.Selvaraj@duke.edu

https://doi.org/10.1016/j.jacbts.2022.09.011

© 2022 The Authors. Published by Elsevier on behalf of the American College of Cardiology Foundation. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/).

Kathryn Chatfield, MD, served as Guest Associate Editor for this paper. Michael Bristow, MD, PhD, served as Guest Editor-in-Chief for this paper.

KEETO-CROSS was funded by the American Society of Nuclear Cardiology Institute for the Advancement of Nuclear Cardiology award (to Dr Selvaraj) and the Department of Radiology at the University of Pennsylvania. Dr Kelly is supported by the National Institutes of Health (R01 HL151345). Dr Selvarai receives, or has recently received, related research support from the National Heart, Lung, and Blood Institute (K23HL161348), Doris Duke Charitable Foundation (#2020061), American Heart Association (#935275), Institute for Translational Medicine and Therapeutics, and the American Society of Nuclear Cardiology (Institute for the Advancement of Nuclear Cardiology award). All other authors have reported that they have no relationships relevant to the contents of this paper to disclose. Research reported in the publication was supported by the Institute for Translational Medicine and Therapeutics of the Perelman School of Medicine at the University of Pennsylvania. The metabolite assays were performed by the Penn Metabolomics Core at the Perelman School of Medicine, University of Pennsylvania, Figures were constructed by using GraphPad Prism and BioRender. The authors thank the staff of the Penn Nuclear Medicine Laboratory and PET Center for their assistance.

The authors attest they are in compliance with human studies committees and animal welfare regulations of the authors' institutions and Food and Drug Administration guidelines, including patient consent where appropriate. For more information, visit the Author Center.

## REFERENCES

**1.** Osborne MT, Hulten EA, Murthy VL, et al. Patient preparation for cardiac fluorine-18 fluorodeoxyglucose positron emission tomography imaging of inflammation. *J Nucl Cardiol.* 2017;24:86–99.

**2.** Selvaraj S, Margulies KB, Dugyala S, et al. Comparison of exogenous ketone administration versus dietary carbohydrate restriction on myocardial glucose suppression: a crossover clinical trial. *J Nucl Med.* 2022;63:770-776.

**3.** Selvaraj S, Seidelmann SB, Soni M, et al. Comprehensive nutrient consumption estimation and metabolic profiling during ketogenic diet and relationship with myocardial glucose uptake on FDG-PET. *Eur Heart J Cardiovasc Imaging.* 2022;23(12):1690–1697. https://doi.org/10.1093/ehjci/ jeac031