



Article Labdane Diterpenes from the Fruits of Sinopodophyllum emodi

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Abstract: Two new labdane diterpenes, sinoditerpene A (1) and B (2), were isolated from the fruits of *Sinopodophyllum emodi*, along with two known analogues **3** and **4**. Their structures were established on the basis of extensive spectroscopic analysis. The isolation of compounds **1**–**4** represents the first report of diterpenes from the genus *Sinopodophyllum*. The cytotoxic activities of all isolated compounds were evaluated in comparison with 5-fluorouracil against the MCF-7 and HepG2 cell lines, towards which **3** showed more potent cytotoxicity.

Keywords: Sinopodophyllum emodi; labdane diterpene; cytotoxic activity

1. Introduction

Sinopodophyllum emodi is an important medicinal plant that has been described in the Chinese Pharmacopoeia and Tibetan medicine [1]. Its dried roots and rhizomes (called Taoerqi in Chinese) are frequently used for the treatment of certain cancers, various verrucosis [2], constipation, verminosis [3], rheumatoid pain [4], and pyogenic skin tissue infections [5]. The dried ripe fruits (called "Xiaoyelian" in Chinese) are clinically applied to the treatment of amenorrhea, dead fetus, and placental retention [1]. Previous phytochemical and pharmacological investigations revealed that *S. emodi* is particularly rich in aryltetralin lactone lignans and prenylated flavonoids, and has attracted wide attention due to their cytotoxic properties [1-3,6-9]. In our search for cytotoxic natural products, we previously reported the isolation, identification and cytotoxic activity of aryltetralin lactone and tetrahydrofuranoid lignans, and prenylated flavonoids from *S. emodi* [10–12]. In a further examination of the fruits of this plant, two new labdane diterpenes, sinoditerpene A (1) and B (2), were obtained together with two known analogues 3 and 4 (Figure 1). For the first time, the NMR signals for compound 4 were completely assigned by 2D NMR spectra (¹H-¹H COSY, HSQC, HMBC, NOESY). Details of the isolation, structure elucidation, and cytotoxicity of all isolated compounds against MCF-7 and HepG2 cell lines are described here.



Figure 1. The chemical structures of compounds 1–4 from *S. emodi*.

2. Results and Discussion

The EtOH extract of the fruits of *Sinopodophyllum emodi* was partitioned between PE, CH₂Cl₂, EtOAc, *n*-BuOH and water, respectively. The CH₂Cl₂ layer was fractionated and purified by repeated column chromatography, allowing the isolation of four labdane diterpenes **1–4**. By comparing their physical and spectroscopic data (¹H-NMR, ¹³C-NMR, ¹H-¹H COSY, HSQC, HMBC, and NOESY in Supplementary Materials) with literature values [13], the two known metabolites were identified as labda-7,13-diene-3,15-diol (**3**) and 14,15-dinor-3β-hydroxy-7-labden-13-one (**4**). The chemical structures of two new labdane diterpenes were determined on the basis of spectroscopic evidences (¹H-NMR, ¹³C-NMR, ¹H-¹H COSY, HSQC, HMBC, DEPT, and NOESY in Supplementary Materials), and their absolute configurations were elucidated by [α]_D and NOESY analysis.

Compound 1 was obtained as a sticky oil and possessed a molecular formula $C_{25}H_{40}O_5$ with six degrees of unsaturation, as revealed from its HR-ESI-MS analysis (m/z 443.2771 [M + Na]⁺, calcd 443.2773). The IR spectrum displayed the presence of carbonyl (1736 cm⁻¹) and hydroxy (3464 cm⁻¹) groups. The ¹³C-NMR and DEPT spectra showed twenty-five carbon signals, including six quaternary carbons, six methyls, eight methylenes and five methines. The ¹H- and ¹³C-NMR spectra (Tables 1 and 2) revealed the presence of malonic acid monoethyl ester [14] and a labdane skeleton [13,15,16]. One oxygenated methylene [δ_H 4.18 (2H, q, J = 7.2 Hz), δ_C 62.3], one methyl [δ_H 1.26 (3H, t, J = 7.2 Hz), $\delta_{\rm C}$ 14.1], one methylene [$\delta_{\rm H}$ 3.22 (2H, s), $\delta_{\rm C}$ 41.7], and two ester carbonyls [$\delta_{\rm C}$ 166.7, 166.6] were observed, suggesting the presence of malonic acid monoethyl ester [14]. Two ester carbonyl [δ_{C} 166.7, 166.6], and four olefinic carbons $\delta_{\rm C}$ 117.9, 122.3, 135.0, 143.3 accounted for four out of the six degrees of unsaturation, and the remaining two indicated that compound 1 was bicyclic. Five methyls [$\delta_{\rm H}$ 0.74 (3H, s), 0.83 (3H, s), 0.95 (3H, s), 1.67 (3H, br.s), 1.70 (3H, br.s), δ_C 13.6, 15.0, 16.6, 21.9, 27.9], six methylenes [one oxygenated δ_H 4.64 (2H, d, J = 7.2 Hz), δ_C 61.5], five methine [one oxygenated $\delta_{\rm H}$ 3.22 (1H, dd, J = 11.2, 4.5 Hz), $\delta_{\rm C}$ 79.1; two olefinic $\delta_{\rm H}$ 5.38 (1H, br.s), 5.32 (1H, t, J = 7.2 Hz)], and four olefinic carbons [δ_C 117.9, 122.3, 135.0, 143.3], suggested that compound 1 possessed a 3,15-dihydroxy-7,13-labdadien skeleton [13]. The HMBC correlation (Figure 2) between the ester carbonyl $\delta_{\rm C}$ 166.7 (C-1') and the methylene $\delta_{\rm H}$ 4.64 (2H, d, J = 7.2 Hz, H-15), indicated that the 15-OH was esterified by malonic acid monoethyl ester.

The relative configuration of the ring substituents of compound **1** was determined by analyzing the NOESY spectrum (Figure 3). The NOESY correlations from H-3 to Me-18, H-5, and H-1 α indicated that they were cofacial and were assigned α -orientation. The NOESY cross peak of Me-19/Me-20

and Me-20/H-11 showed that these protons were β -oriented. Additionally, the stereochemistry of the side-chain double bond was established as *E* by virtue of the Me-16 chemical shifts (δ_H 1.70; δ_C 16.6) [13]. This was also supported by the cross peak of H-12/H-14 in the NOESY spectrum. The absolute configuration is the same as (–)-labda-7,13-diene-3,15-diol [17,18], which was determined by a microhydrolysis method and comparison of [α]_D values. Thus, compound **1** was established as 3 β -hydroxy-15-(3'-ethoxy-3'-oxopropionyloxy)-7,13*E*-labdadiene, and named sinoditerpene A.

No.	1	2	3	4
4	1.07 (1H, td, 13.2, 4.2)	1.06 (1H, td, 13.1, 4.5)	1.06 (1H, td, 13.2, 4.4)	1.10 (1H, td, 13.3, 4.0)
1	1.84 (1H, dt, 13.2, 3.4)	1.84 (1H, dt, 13.1, 3.4)	1.84 (1H, dt, 13.2, 3.4)	1.91 (1H, dt, 13.3, 3.6)
2	1.60 (2H, m)	1.60 (2H, m)	1.60 (2H, m)	1.60 (2H, m)
3	3.22 (1H, dd, 11.2, 4.5)	3.22 (1H, dd, 11.0, 4.8)	3.20 (1H, dd, 11.2, 4.5)	3.21 (1H, dd, 11.4, 4.4)
5	1.16 (1H, dd, 10.9, 6.1)	1.17 (1H, dd, 10.9, 6.1)	1.15 (1H, dd, 10.9, 6.1)	1.15 (1H, dd, 10.9, 6.1)
6	1.95 (2H, m)	1.95 (2H, m)	1.95 (2H, m)	1.95 (2H, m)
7	5.38 (1H, br.s)	5.38 (1H, br.s)	5.37 (1H, br.s)	5.39 (1H, br.s)
9	1.57 (1H, m)	1.58 (1H, m)	1.58 (1H, m)	1.57 (1H, m)
11	1.47 (1H, m)	1.50 (1H, m)	1.50 (1H, m)	1.78 (1H, m)
	1.25 (1H, m)	1.25 (1H, m)	1.28 (1H, m)	1.45 (1H, m)
12	1.95 (1H, m)	1.95 (1H, m)	1.95 (1H, m)	2.40 (1H, m)
	2.22 (1H, m)	2.20 (1H, m)	2.20 (1H, m)	2.63 (1H, m)
14	5.32 (1H, t, 7.2)	5.32 (1H, t, 7.5)	5.40 (1H, t, 7.0)	
15	4.64 (2H, d, 7.2)	4.58 (2H, d, 7.5)	4.13 (2H, d, 7.0)	
16	1.70 (3H, s)	1.68 (3H, s)	1.67 (3H, s)	2.11 (3H, s)
17	1.67 (3H, s)	1.68 (3H, s)	1.66 (3H, s)	1.64 (3H, s)
18	0.95 (3H, s)	0.95 (3H, s)	0.94 (3H, s)	0.94 (3H, s)
19	0.83 (3H, s)	0.83 (3H, s)	0.83 (3H, s)	0.83 (3H, s)
20	0.74 (3H, s)	0.73 (3H, s)	0.73 (3H, s)	0.76 (3H, s)
2′	3.22 (2H, s)	2.76 (1H, d, 15.6)		
		2.87 (1H, d, 15.6)		
4'		2.76 (1H, d, 15.6)		
		2.87 (1H, d, 15.6)		
CH_2	4.18 (2H, q, 7.2)	4.12 (2H, q, 7.2)		
CH_3	1.26 (3H, t, 7.2)	1.28 (3H, t, 7.2)		
CH ₂		4.26 (2H, q, 7.2)		
CH ₃		1.26 (3H, t, 7.2)		

Table 1. ¹H-NMR data (500 MHz, δ in ppm, *J* in Hz) of compounds 1–4 in CDCl₃.

Table 2. ¹³C-NMR data (125 MHz, δ in ppm) of compounds 1–4 in CDCl₃.

No.	1	2	3	4	No.	1	2	3	4
1	37.2	37.2	37.1	37.4	16	16.6	16.6	16.5	30.0
2	27.3	27.4	27.3	27.4	17	21.9	21.9	21.9	22.0
3	79.1	79.1	79.1	79.1	18	27.9	27.9	27.8	27.9
4	38.6	38.6	38.6	38.7	19	15.0	15.0	15.0	15.1
5	49.5	49.5	49.5	49.5	20	13.6	13.6	13.6	13.6
6	23.4	23.4	23.4	23.4	1'	166.7	169.80		
7	122.3	122.3	122.1	122.9	2′	41.7	43.2		
8	135.0	135.0	135.1	134.3	3′	166.6	73.2		
9	54.3	54.2	54.3	54.3	4'		43.3		
10	36.6	36.6	36.6	36.7	5'		169.76		
11	25.5	25.3	25.5	20.8	6'		173.4		
12	41.9	41.8	41.9	45.7	CH_2	62.3	62.3		
13	143.3	143.2	140.0	208.7	CH_3	14.1	14.0		
14	117.9	118.0	123.5		CH_2		61.0		
15	61.5	61.8	59.4		CH_3		14.1		



Figure 2. Key ¹H-¹H COSY, HMBC correlations of compounds 1–3.



Figure 3. Key NOESY correlations of compounds 1-3.

Compound **2** was obtained as a sticky oil and possessed a molecular formula $C_{30}H_{48}O_8$ with seven degrees of unsaturation, as revealed from its HR-ESI-MS analysis (m/z 559.3249 [M + Na]⁺, calcd 559.3247). The IR spectrum displayed the presence of carbonyl (1734, 1718 cm⁻¹) and hydroxyl (3465 cm⁻¹) groups. Its ¹H- and ¹³C-NMR data (Tables 1 and 2) were quite similar to those of compound **1**, except that citric acid-1,2-diethyl ester was observed instead of the malonic acid monoethyl ester of compound **1**. Two hydroxymethyls [δ_H 4.12 (2H, q, J = 7.2 Hz), 4.26 (2H, q, J = 7.2 Hz), δ_C 61.0, 62.3], two methyls [δ_H 1.26 (3H, t, J = 7.2 Hz), 1.28 (3H, t, J = 7.2 Hz), δ_C 14.0, 14.1], two methylenes [δ_H 2.76 (1H, d, J = 15.6 Hz), 2.87 (1H, d, J = 15.6 Hz), 2.76 (1H, d, J = 15.6 Hz), 2.87 (1H, d, J = 15.6 Hz), 2.76 (1H, d, J = 7.5 Hz, 169.80 (C-1') and the methylene δ_H 4.58 (2H, d, J = 7.5 Hz, H-15), indicated that the 15-OH was esterified by citric acid-1,2-diethyl ester. Thus, compound **2** was established as 3 β -hydroxy-15-[(3'-hydroxy-3',4'-bis(ethoxycarbonyl)-butyroxy)]-7,13*E*-labdadiene, and named sinoditerpene B.

Compound 4 was obtained as a sticky oil and possessed a molecular formula $C_{18}H_{30}O_2$ with four degrees of unsaturation, as revealed from its HR-ESI-MS analysis (m/z 301.2148 [M + Na]⁺, calcd 301.2143). The IR spectrum displayed the presence of carbonyl (1714 cm⁻¹) and hydroxy (3415 cm⁻¹) groups. Its ¹H- and ¹³C-NMR (Tables 1 and 2) were similar to those of compound **3**, except that a carbonyl group δ_C 208.7 were observed instead of two olefinic carbons δ_C 140.0, 123.5, and one oxygenated methylene δ_C 59.4 in **1**. Due to apparent differences of carbon signal number in **4** (20 carbons) and **3** (18 carbons), it was reasonably assumed that **4** was a dinor-derivative of **3**. The HMBC correlation between the carbonyl group δ_C 208.7 (C-13) and the methyl δ_H 2.11 (3H, s, H-16), the methylene δ_H 2.40 (1H, m, H-12), 2.63 (1H, m, H-12), in combination with ¹H-¹H COSY cross peak of H-11 with H-12 and H-9, indicated that 3-oxobutyl was linked to C-9. On the basis of the above evidences and related literature [13], compound **4** was established as 14,15-dinor-3 β -hydroxy-7-labden-13-one.

Chinese herbal medicines produce a wide variety of secondary metabolites, which can be exploited as potential anticancer agents. Previous chemical investigations on *S. emodi* revealed the presence of aryltetralin lactone and tetrahydrofuranoid lignans [2,3,7,8,10], flavonoids [1,6,9,11,20], steroids [21], and phenolics [22]. Cytotoxic activities of some isolated *S. emodi* constituents have been shown in various cancer cell lines. In the HeLa and KB cell lines, deoxypodphyllotoxin was about 579 and 1123 times more toxic than etoposide, respectively [10]. Sinolignan C displayed cytotoxicity against the KB cell line and was more cytotoxic than etoposide [12]. 3-Methoxyquercetin showed cytotoxicity against the MCF-7 and HepG2 cell lines, with IC₅₀ values of 3.14 and 2.08 µM, respectively [11].

All of the isolated labdane diterpenoids were evaluated for cytotoxic activities against the MCF-7 and HepG2 cell lines (Table 3). Compound **3** was more cytotoxic than 5-fluorouracil, whereas compounds **1**, **2** and **4** displayed no cytotoxicity against MCF-7 and HepG2 cell lines. Compounds **1**–4 have the same structural skeleton, so the variation in cytotoxicity between them indicates a free hydroxyl group at C-15 was structurally required for the cytotoxic activity against the MCF-7 and HepG2 cells lines. Esterification at C-15 drastically reduced the cytotoxic activity of the parent compound **3**. These results revealed the potential of compound **3** as an ideal antitumor lead compound.

Table 3. Cytotoxicities of compounds 1–4 against MCF-7 and HepG2 cell lines (IC₅₀, μ M).

Compound	MCF-7	HepG2	Compound	MCF-7	HepG2
1 2 5-Fluorouracil	$\begin{array}{c} 74.6 \pm 5.5 \\ 88.3 \pm 7.1 \\ 6.74 \pm 0.52 \end{array}$	$\begin{array}{c} 63.5 \pm 6.2 \\ 75.2 \pm 6.8 \\ 5.18 \pm 0.40 \end{array}$	3 4	$\begin{array}{c} 5.73 \pm 0.46 \\ 50.2 \pm 5.0 \end{array}$	$\begin{array}{c} 3.85 \pm 0.29 \\ 39.1 \pm 4.7 \end{array}$

3. Experimental Section

3.1. General Procedures

The IR spectra were measured on a Tensor 27 Fourier transform infrared (FTIR) spectrometer (Bruker Optics, Ettlingen, Germany) as KBr discs. The 1D and 2D NMR spectra were recorded on an AC (E)-500 spectrometer(Bruker Biospin, Fallanden, Switzerland) using TMS as an internal standard. HR-ESI-MS was determined on a microTOF-Q instrument (Bruker Daltonics, Billerica, MA, USA). Optical rotations were measured on an Autopol IV Digital Polarimeter (Rudolph Research Analytical, Hackettstoun, NJ, USA). The chromatographic silica gel (200–300 mesh) was obtained from Qingdao Ocean Chemical Factory (Qingdao, China). ODS (50 µm) was produced from YMC Co. Ltd. (Kyoto, Japan). Sephadex LH-20 was produced by Amersham Pharmacia Biotech (Uppsala, Sweden). Chemical reagents for isolation were of analytical grade and purchased from Tianjin Siyou Co., Ltd. (Tianjin, China). Biological reagents were from Sigma Company (St. Louis, MO, USA). Human heptocellular (HepG2) and breast (MCF-7) cell lines were from Institute of Materia Medica, Chinese Academy of Medical Sciences and Peking Union Medical College (Beijing, China).

3.2. Plant Material

The plant material was collected from Deqin, Yunnan Province, China, in September 2013, and identified by Prof. Cheng-Ming Dong as the fruits of *S. emodi*, according to the Chinese Traditional Medicine Dictionary [23]. A voucher specimen (SE 20130929) was deposited at the School of Pharmacy, Henan University of Traditional Chinese Medicine.

3.3. Extraction and Isolation

The dried fruits of *S. emodi* were ground into a power (9.1 kg), and refluxed with 95% EtOH(3×20 L). The filtrate was concentrated under reduced pressure to yield a dark brown residue (1.6 kg). The residue was suspended in water (3.2 L) and partitioned with petroleum ether (PE, 3.2 L × 3), CH₂Cl₂ (3.2 L × 3), EtOAc (3.2 L × 3), and *n*-BuOH (3.2 L × 3), successively. The CH₂Cl₂ extract (400.05 g) was fractionated using silica gel column chromatography (CC, 20 × 140 cm) with a gradient of PE (60–90 °C)–acetone. The fractions were combined into eleven main fractions C1–11 based on TLC results. Fraction C3 (5.02 g) was chromatographed over open ODS (2.5 × 45 cm), eluted by methanol–H₂O (20: 80 to 85: 15) to yield subfractions C–3–1–C–3–3. Subfraction C–3–3 (0.95 g) was further submitted to silica gel CC (1.0 × 25 cm) eluted by PE–acetone (100: 20) to give **1** (7.8 mg) and **2** (4.6 mg). Fraction C4 (6.95 g) was subjected to Sephadex LH-20 CC (2.0 × 90 cm) eluted by methanol to yield subfractions C4–1 and C4–2. Subfractions C4–1 (1.74 g) was separated by open ODS (2.0 × 40 cm) eluted by methanol–H₂O (40: 60 to 80: 20) to give **3** (20.8 mg) and **4** (5.2 mg).

3.4. Spectroscopic and Physical Data

Sinoditerpene A (1). Sticky oil; $[\alpha]_D^{25}$ –12.4 (*c* 0.14, CHCl₃); IR (KBr) ν_{max} 3464, 2967, 2930, 2855, 1737 cm⁻¹; HR-ESI-MS (positive): *m*/*z* 443.2771 [M + Na]⁺ (calcd for C₂₅H₄₀O₅Na, 443.2773); NMR data (CDCl₃), see Tables 1 and 2.

Sinoditerpene B (**2**). Sticky oil; $[\alpha]_D^{25}$ –9.5 (*c* 0.16, CHCl₃); IR (KBr) ν_{max} 3456, 2956, 2924, 2853, 1734, 1718; HR-ESI-MS (positive): m/z 559.3249 [M + Na]⁺ (calcd for C₃₀H₄₈O₈Na, 559.3247); NMR data (CDCl₃), see Tables 1 and 2.

3.5. Alkali Hydrolysis

Compound 1 (6 mg) was refluxed with 4N alcoholic KOH (5 mL) for 1 h. The resulting liquid was neutralized by 1N HCI until pH 7, and concentrated under reduced pressure, and then extracted with CH_2Cl_2 (10 mL \times 3) after 10 mL water was added. The organic phase was washed with water, dried over Na_2SO_4 and concentrated *in vacuo*. The residue was chromatographed on silica gel (PE–acetone 100:7–100:20) to obtain the hydrolyzed target compound.

3.6. Cytotoxicity Asssay

Tumor cells were maintained in RPMI-1640 medium containing 10% heat-inactivated fetal bovine serum, penicillin (100 units/mL), streptomycin (100 μ g/mL) under humidified air with 5% CO₂ at 37 °C. Exponentially growing cells were seeded into 96-well tissue culture-treated plates and precultured for 1 day. The cytotoxic activities of isolated compounds were tested against MCF-7 and HepG2 cell lines, using an established MTT assay protocol [10]. 5-fluorouracil was used as the positive control.

4. Conclusions

Further phytochemical studies on *S. emodi* resulted in the isolation of two new labdane diterpenes **1** and **2** and two known analogues **3** and **4**. All the labdane diterpenoids contained 7(8) and 13(14) double bonds. Isolation of diterpenoids from *S. emodi* is reported here for the first time, which also enriches our knowledge about the chemical diversity of this plant. Compound **1** is the first reported example of a labdane diterpene with 7(8) and 13(14) double bonds exhibiting cytotoxicity in the MCF-7 and HepG2 cell lines. As an antitumor lead compound, further investigations are necessary to explore the structure-activity relationship and lead optimization. Our research demonstrated that the fruits of *S. emodi* have chemopreventive potential as a herbal medicine and its labdane diterpenes are partly responsible for the observed potency. In addition, these results will broaden the application field of *S. emodi*.

Supplementary Materials: Supplementary materials can be accessed at: http://www.mdpi.com/1420-3049/21/4/434/s1.

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Sample Availability: Samples of the compounds 1–4 are available from the authors.



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