





Draft Genome Sequence of *Mobilitalea sibirica* Strain P3M-3^T, the Sole Representative of the Genus Mobilitalea

Nils Thieme, a Regina Rettenmaier, a Wolfgang Liebl, a Vladimir V. Zverlova, b

^aChair of Microbiology, TUM School of Life Sciences Weihenstephan, Technical University of Munich, Freising, Germany bInstitute of Molecular Genetics, National Research Centre, Kurchatov Institute, Moscow, Russia

ABSTRACT Mobilitalea sibirica strain P3M-3^T is a strictly anaerobic, halotolerant, organotrophic bacterium of the family Lachnospiraceae that can utilize various plantderived polysaccharides as its carbon source. The strain was originally isolated from a microbial mat in western Siberia (Russia). In this study, we present the draft genome sequence of M. sibirica P3M-3^T based on Illumina paired-end sequencing.

•he strain *Mobilitalea sibirica* P3M-3^T (phylum *Firmicutes*, order *Clostridiales*, family Lachnospiraceae) is a mesophilic and strictly anaerobic bacterium that was isolated in western Siberia (Russia) from a microbial mat (1). P3M-3^T utilizes starch, xylan, and crystalline cellulose as well as other polysaccharides as its carbon source. The main products of its fermentation are acetate, ethanol, H₂, and CO₂. The ability to depolymerize a wide array of polysaccharides makes M. sibirica an intriguing candidate for biorefinery approaches, e.g., microbial cocultures (2). Since strain P3M-3^T is the first and sole representative of the genus Mobilitalea, its genome sequence information is essential for targeted genetic engineering. Therefore, we created a draft genome sequence of M. sibirica P3M-3^T in this study using Illumina paired-end sequencing.

M. sibirica P3M-3^T (=DSM 26468^T) was obtained from the German Collection of Microorganisms and Cell Cultures (DSMZ). The strain was cultivated anaerobically in GS2 medium (3) with 0.5% (wt/vol) cellobiose (Merck, Darmstadt, Germany) as the carbon source at 37°C. High-molecular-weight (HMW) DNA was extracted from a culture using the MagAttract HMW DNA kit from Qiagen (Hilden, Germany). The 16S rRNA gene was amplified and later sequenced with the primers 27F (AGAGTTTGATCMTGGCTCAG) and 1492R (CGGTTACCTTGTTACGACTT). The clonal purity of the strain was confirmed through Sanger sequencing by GeneWiz (South Plainfield, NJ, USA). At least 1 µg of chromosomal DNA was used to prepare a DNA library with the TruSeq DNA PCR-free sample preparation kit (Illumina, San Diego, CA, USA) (4). The library was sequenced using the Illumina MiSeq system in paired-end mode, according to the manufacturer's instructions. The read length was 150 bp, and 4,859,739 forward reads as well as 4,859,739 reverse reads were generated. Read assembly was performed using SPAdes v3.13.0 (5), and default parameters were used. The BayesHammer package (6) is bundled in SPAdes v3.13.0, and it was used with default settings for read error correction. A total of 40 contigs were assembled with a 40-fold coverage and an N_{50} value of 165,485 bp. The combined length of the contigs is 3,854,872 bp, with a G+C content of 36.55 mol%. Open reading frames (ORFs) were predicted and annotated using the NCBI Prokaryotic Genome Annotation Pipeline (PGAP) (7), resulting in 3,332 coding sequences and 74 RNAs (3 5S rRNAs, 7 16S rRNAs, 7 23S rRNAs, 53 tRNAs, and 4 noncoding RNAs [ncRNAs]).

The genome sequence presented in this work is a valuable resource for evaluating M. sibirica P3M-3^T for future biotechnological applications. Furthermore, the genome sequence also allows us to gather more insights into the genetic properties and phylogeny of the genus Mobilitalea.

Citation Thieme N, Rettenmaier R, Liebl W, Zverlov W. 2021. Draft genome sequence of *Mobilitalea sibirica* strain P3M-3^T, the sole representative of the genus Mobilitalea. Microbiol Resour Announc 10:e00129-21. https://doi.org/10.1128/MRA.00129-21.

Editor J. Cameron Thrash, University of Southern California

Copyright © 2021 Thieme et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Vladimir V. Zverlov, vladimir.zverlov@tum.de.

Received 3 February 2021 Accepted 12 March 2021 Published 1 April 2021



Data availability. This whole-genome shotgun project has been deposited at DDBJ/ ENA/GenBank under the accession number JAEAGR000000000. The version described in this paper is version JAEAGR000000000.1. The raw sequencing reads are provided in the Sequence Read Archive (SRA) under the accession number SRR13181447. The whole project is summarized under the BioProject accession number PRJNA682100 and contains one BioSample under the accession number SAMN16980878.

ACKNOWLEDGMENTS

This study was in part funded by the Bundesministerium für Bildung und Forschung (BMBF) and Projektträger Jülich (PTJ) (031B0672C). The Technical University of Munich supported this work through the Open Access Publishing funding program.

We thank the group of the ZIEL-Core Facility Microbiome at the Technical University of Munich (Freising, Germany) for their support in genome sequencing.

REFERENCES

- Podosokorskaya OA, Bonch-Osmolovskaya EA, Beskorovaynyy AV, Toshchakov SV, Kolganova TV, Kublanov IV. 2014. *Mobilitalea sibirica* gen. nov., sp. nov., a halotolerant polysaccharide-degrading bacterium. Int J Syst Evol Microbiol 64:2657–2661. https://doi.org/10.1099/ijs.0.057109-0.
- Bader J, Mast-Gerlach E, Popović MK, Bajpai R, Stahl U. 2010. Relevance of microbial coculture fermentations in biotechnology. J Appl Microbiol 109:371–387. https://doi.org/10.1111/j.1365-2672.2009.04659.x.
- Koeck DE, Ludwig W, Wanner G, Zverlov VV, Liebl W, Schwarz WH. 2015. Herbinix hemicellulosilytica gen. nov., sp. nov., a thermophilic cellulosedegrading bacterium isolated from a thermophilic biogas reactor. Int J Syst Evol Microbiol 65:2365–2371. https://doi.org/10.1099/ijs.0.000264.
- 4. Huptas C, Scherer S, Wenning M. 2016. Optimized Illumina PCR-free library preparation for bacterial whole genome sequencing and analysis of

- factors influencing de novo assembly. BMC Res Notes 9:269. https://doi.org/10.1186/s13104-016-2072-9.
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. J Comput Biol 19:455–477. https://doi.org/10.1089/cmb.2012.0021.
- Nikolenko SI, Korobeynikov AI, Alekseyev MA. 2013. BayesHammer: Bayesian clustering for error correction in single-cell sequencing. BMC Genomics 14:S7. https://doi.org/10.1186/1471-2164-14-S1-S7.
- Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI Prokaryotic Genome Annotation Pipeline. Nucleic Acids Res 44:6614–6624. https://doi.org/10.1093/nar/gkw569.

Volume 10 Issue 13 e00129-21 mra.asm.org **2**