

rsob.royalsocietypublishing.org

Perspective



Cite this article: Lo YMD. 2012 Non-invasive prenatal diagnosis by massively parallel sequencing of maternal plasma DNA. Open Biol 2: 120086.

http://dx.doi.org/10.1098/rsob.120086

Received: 3 May 2012 Accepted: 14 May 2012

Subject Area:

genetics/genomics/biotechnology/ developmental biology

Keywords:

Down syndrome, trisomy 21, plasma nucleic acids, foetal DNA in maternal plasma, next-generation sequencing

Author for correspondence:

Yuk Ming Dennis Lo e-mail: loym@cuhk.edu.hk

[†]An invited Perspective to mark the election of the author to the fellowship of the Royal Society in 2011.



Non-invasive prenatal diagnosis by massively parallel sequencing of maternal plasma DNA

Yuk Ming Dennis Lo^{1,2,†}

¹Li Ka Shing Institute of Health Sciences, and ²Department of Chemical Pathology, The Chinese University of Hong Kong, Prince of Wales Hospital, Shatin, New Territories, Hong Kong SAR, People's Republic of China

1. Summary

The presence of foetal DNA in the plasma of pregnant women has opened up new possibilities for non-invasive prenatal diagnosis. The use of circulating foetal DNA for the non-invasive prenatal detection of foetal chromosomal aneuploidies is challenging as foetal DNA represents a minor fraction of maternal plasma DNA. In 2007, it was shown that single molecule counting methods would allow the detection of the presence of a trisomic foetus, as long as enough molecules were counted. With the advent of massively parallel sequencing, millions or billions of DNA molecules can be readily counted. Using massively parallel sequencing, foetal trisomies 21, 13 and 18 have been detected from maternal plasma. Recently, large-scale clinical studies have validated the robustness of this approach for the prenatal detection of foetal chromosomal aneuploidies. A proof-of-concept study has also shown that a genome-wide genetic and mutational map of a foetus can be constructed from the maternal plasma DNA sequencing data. These developments suggest that the analysis of foetal DNA in maternal plasma would play an increasingly important role in future obstetrics practice. It is thus a priority that the ethical, social and legal issues regarding this technology be systematically studied.

2. Introduction

Prenatal diagnosis is now an established part of modern obstetrics practice. However, conventional definitive methods for prenatal diagnosis involve the invasive sampling of foetal tissues, using methods such as amniocentesis and chorionic villus sampling (CVS). Such methods carry with them a small, but definite risk for the foetus [1]. Ultrasound scanning and maternal serum biochemical analysis have emerged as non-invasive methods for screening for foetal chromosomal aneuploidies, such as trisomy 21 [2]. However, for the detection of foetal chromosomal aneuploidies, such methods measure epiphenomena, which are associated with the disorders, rather than analysing the core pathology. As a result, they typically can only be used within a relatively narrow gestational age window, and despite their development over many years, their sensitivity and specificity still have much room for improvement.

Because of these limitations, there has been a search over the last few decades for safe, non-invasive methods for prenatal diagnosis that can allow the direct analysis of foetal genetic materials. Early work had focused on the isolation of foetal nucleated cells that had entered into the maternal blood. However, the concentrations of such cells are very low, typically of the order of one or a few foetal nucleated cells per millilitre of maternal blood [3]. Probably as a result of such low concentrations, prenatal testing carried out using

© 2012 The Authors. Published by the Royal Society under the terms of the Creative Commons Attribution License http://creativecommons.org/licenses/by/3.0/, which permits unrestricted use, provided the original author and source are credited.

circulating foetal cells has been found to have relatively low sensitivity and specificity [4].

In 1997, Lo et al. were inspired by the presence of tumourderived DNA in the plasma and serum of cancer patients [5,6], and wondered whether an analogous phenomenon might also be present in pregnancy. Lo et al. were able to find Y chromosomal DNA sequences in the plasma and serum of women carrying male foetuses, and concluded that cell-free foetal DNA was present in maternal plasma and serum [7]. Subsequent measurements using real-time polymerase chain reaction (PCR) have indicated that cellfree foetal DNA is present in maternal plasma at a mean fractional concentration of 3 per cent in the first and second trimesters [8]. More recent measurements using more accurate digital PCR-based techniques show that the median fractional concentration may be approximately 10 per cent [9]. Such relatively high fractional foetal DNA concentrations, when compared with the fractional concentrations of foetal nucleated cells in maternal blood, suggest that non-invasive prenatal diagnosis carried out using the former would probably be more robust. Another advantage of the use of foetal DNA in maternal plasma, compared with circulating foetal nucleated cells, is the lack of persistence of the former following delivery [10,11]. Conversely, there are numerous reports describing the persistence of foetal nucleated cells in the maternal circulation following delivery [12].

The robustness of non-invasive prenatal diagnosis using cell-free foetal DNA in maternal plasma can be seen by the many reports that describe diagnostic tests based on the detection of DNA sequences that the foetus has inherited from the father and which are absent in the maternal genome. Examples include the detection of foetal sex based on the detection of Y chromosomal DNA from maternal plasma [13] and the detection of the RHD gene of a Rhesus D-positive foetus in the plasma of a Rhesus D-negative pregnant woman [14,15].

3. The challenge for the detection of foetal chromosomal aneuploidies

The detection of foetal chromosomal aneuploidies, such as trisomy 21, is much more challenging than the determination of foetal sex and Rhesus D blood group genotype because, apart from detecting the presence of foetal DNA in maternal plasma, one also has to measure the foetal chromosome dosage involving the chromosome of interest. The latter task is made more difficult because of the fact that foetal DNA is present as a minor fraction of the DNA that is found in maternal plasma [8,9].

Early work has focused on the analysis of a subset of nucleic acids present in maternal plasma that is foetalspecific. Examples include DNA molecules bearing foetal-specific DNA methylation patterns [16-19] and RNA molecules that are specifically transcribed from the placenta [20]. All but one [19] of the above-mentioned methods involve the use of genetic polymorphisms and necessitate the use of multiple markers to achieve a broad population coverage. The method that does not require the use of genetic polymorphisms is based on chromatin immunoprecipitation and complex data normalization procedures [19]. These steps would be challenging to be reproducibly performed for a clinical diagnostic or a screening test. Another approach

that has been described involves the enrichment of the fractional concentration of foetal DNA. One way to achieve such enrichment uses the observation that foetal DNA molecules in maternal plasma are shorter than the maternally derived ones [21,22]. However, the amount of enrichment that one could achieve thus far using this approach is relatively limited. Another method that has been reported involves the use of formaldehyde on maternal blood samples. This method has been claimed to minimize the release of DNA from the maternal blood cells and thus to result in a higher fractional concentration of foetal DNA in the plasma fraction [23,24]. However, this method has not been reproduced by a number of laboratories [25,26].

4. Molecular counting approach

An alternative approach for the detection of foetal chromosomal aneuploidies is to measure the quantitative perturbations in the genomic representation of the involved chromosome in maternal plasma. However, the challenge is that such perturbations are generally very small and are related to the fractional concentration of circulating foetal DNA [27]. For example, it has been shown that when the fractional foetal DNA concentration is 10 per cent, the genomic representation of chromosome 21 in maternal plasma will be increased by 5 per cent by the presence of a trisomy 21 foetus. The detection of such a small amount of quantitative perturbation requires the use of extremely precise measurement methods. In 2007, it was demonstrated that single-molecule counting techniques using digital PCR as an example can be used for such a purpose [27,28]. Such an approach was first realized using plasma foetal (placenta-derived) RNA that is present in maternal plasma [27]. The potential extension of this approach to plasma DNA was also explored using DNA mixtures and computer simulations [27,28]. The numbers of molecules that one would need to be counted for different fractional foetal DNA concentrations have been outlined. For example, it is suggested that to achieve the detection of a trisomy 21 foetus in a maternal plasma sample containing 25 per cent foetal DNA would require the performance of 7680 digital PCRs. Furthermore, for every twofold reduction in the fractional concentration of foetal DNA, the number of molecules that one would need to count would increase by 2² (i.e. 4) times [27].

5. Non-invasive prenatal diagnosis by massively parallel sequencing

With the advent of massively parallel sequencing, it has become relatively easy to count millions (or even billions) of DNA molecules [29]. In 2008, two groups showed that the massively parallel sequencing of maternal plasma DNA would allow one to work out the genomic representations of different chromosomes in maternal plasma and to detect the perturbations of such representations when a pregnant woman is carrying a trisomic foetus [30,31]. This approach involves the random sequencing of millions of DNA molecules in maternal plasma. Individual sequence tags are aligned to the human genome to determine the chromosome of origin of a particular sequence tag. The sensitivity and specificity of these early reports, involving a relatively small number of samples, are both 100 per cent. It has been shown that both sequencing-by-synthesis [30,31] and sequencing-byligation [32] platforms would work for this approach. The method is also applicable for foetal trisomy 21 that is caused by a Robertsonian translocation [33].

The throughput of the use of massively parallel sequencing for the detection of foetal trisomy 21 has since then been validated by a number of large-scale clinical studies [34-37]. The detection of foetal trisomies 13 and 18 has been shown to be more challenging owing to the GC contents of the involved chromosomes and the analytical bias of the sequencing platform in relation to the GC contents [31,32]. Nonetheless, the use of bioinformatics algorithms that correct for such bias has been shown to allow the improved detection of trisomies 13 and 18 [38,39]. The robust detection of trisomies 13 and 18 has recently been validated by a number of large-scale studies [37,40,41]. The future use of singlemolecule sequencing platforms that do not require the use of an amplification step might further reduce the influence of the GC content and might improve the robustness of this

As a result of the various large-scale studies, the use of massively parallel sequencing of maternal plasma DNA for the prenatal detection of foetal chromosomal aneuploidies has been introduced as a clinical service in the USA and parts of Asia. It is likely that this technology will be used clinically in other regions around the world in the near future.

The cost of this technology is still relatively expensive when compared with conventional prenatal screening procedures, but is comparable with the costs of invasive testing involving amniocentesis. A number of groups have explored the possibility of reducing the sequencing costs. Random sequencing is the protocol employed by most workers in the field, whereby sequence tags will be obtained for all chromosomes [30,31]. The number of sequence tags obtained per chromosome is proportional to the size of each chromosome. In such a protocol, only a proportion of the sequenced reads will align to the chromosome of interest (e.g. chromosome 21).

In an effort to reduce the cost of sequencing, Liao et al. [43] have demonstrated that a solution-based target capture system is able to focus the sequencing power to selected genomic regions. Such a system is able to increase the sequencing coverage of the targeted region by over 200-fold. Another system has recently been described by Sparks et al. [44] in which hundreds of sets of oligonucleotides have been used to target selected regions of the genome. These targeted regions are then amplified and sequenced by massively parallel sequencing. Following a process of normalization, the authors have reported that trisomy 21 and 18 samples are distinguishable from euploid samples. However, it is important to note that the authors have only reported a training dataset, but have not tested the robustness of their algorithm using a validation dataset. In a subsequent publication by the same group, the authors have introduced a new algorithm whereby the fractional concentrations of foetal DNA in maternal plasma are incorporated into the data analysis step [45]. Nonetheless, they have not compared the performance of this new algorithm with that of their previous algorithm [44]. The new algorithm, on the other hand, has been tested in an independent cohort [46]. The relative robustness of the selective chromosome sequencing approach [46] and the previously described random sequencing approach [30,31,34,36,37,39,41] would need to be evaluated

in future studies. The cost-effectiveness issue of the targeted versus the non-targeted approach also needs to be addressed in the context of the continual and rapid reduction in the costs associated with sequencing.

6. Towards prenatal foetal whole genome sequencing

In addition to the detection of foetal chromosomal aneuploidies, the analytical precision offered by molecular counting approaches such as digital PCR has implications for the non-invasive prenatal diagnosis of monogenic diseases. For example, in the situation of a mother who is a heterozygous carrier of an autosomal recessive monogenic disease, the relative dosage of the mutant and normal versions of the gene in the mother's genome should be 1:1. However, when the mother is pregnant with a foetus, then the relative dosage of the two versions of the gene in the mother's plasma will be modified depending on the foetal genotype. If the foetus is homozygous for the mutant gene, then the addition of two doses of the mutant gene per genome-equivalent of foetal DNA into the maternal plasma will bias the relative dosage in favour of the mutant gene. If the foetus is homozygous for the normal gene, then the relative dosage of the mutant and normal genes in the mother's plasma will be biased in favour of the normal gene. Finally, if the foetus is heterozygous, then the 1:1 relative dosage of the mutant and normal genes will remain unchanged in the mother's plasma. This concept has been called the relative mutation dosage approach [47], and has been used for the non-invasive prenatal diagnosis of the haemoglobinopathies [47] and haemophilia [48].

With the advent of massively parallel sequencing, this type of concept can be applied on a genome-wide scale. Thus, Lo et al. [49] demonstrated that by sequencing maternal plasma DNA to the equivalent of 65-fold haploid genome coverage, a genome-wide genetic and mutational profile of a foetus can be assembled from the sequencing data. The authors have further used the genetic maps of the father and the mother to help assemble the foetal genetic map. The resolution of the foetal genomic map that one could assemble is limited by the resolution of the parental genetic maps. This method thus opens up the possibility that the prenatal diagnosis of multiple genetic diseases could be performed by a single test. The future refinement of targeted sequencing approaches would be expected to reduce the cost of this technology.

7. Conclusions

It is probable that non-invasive prenatal diagnosis using foetal DNA in maternal plasma would play an increasingly important role in the future practice of prenatal testing. However, it is important to address the ethical, legal and social issues surrounding such developments [50-52]. The positive side of non-invasive testing is the avoidance of harm to the foetus that would be associated with invasive testing. However, some parties may be concerned by the possibility that the availability of non-invasive testing might 'encourage' more pregnant women to undergo testing. Such issues, and others, would need specially designed studies to systematically address them.

8. Acknowledgements

The author thanks the University Grants Committee of the Government of the Hong Kong Special Administration Region, China—Areas of Excellence Scheme (AoE/M-04/ 06) for supporting this work. The author holds or has filed patent applications on aspects of non-invasive prenatal diagnosis. Part of this patent portfolio has been licensed to Sequenom. The author is a consultant to Sequenom, holds equities in and receives research support from Sequenom.

AUTHOR PROFILE



Dennis Lo is the Director of the Li Ka Shing Institute of Health Sciences, Li Ka Shing Professor of Medicine and Professor of Chemical Pathology at The Chinese University of Hong Kong. He received his undergraduate education from the University of Cambridge, and his Doctor of Medicine and Doctor of Philosophy degrees from the University of Oxford. His research interests focus on the biology and diagnostic applications of cell-free nucleic acids in plasma. In particular, he discovered the presence of cell-free foetal DNA in maternal plasma in 1997 and has since then been pioneering non-invasive prenatal diagnosis using this technology. He has received numerous awards for his research, including a State Natural Sciences Award from the State Council of China (2005), the International Federation of Clinical Chemistry and Laboratory Medicine (IFCC)-Abbott Award for Outstanding Contribution to Molecular Diagnostics (2006), the US National Academy of Clinical Biochemistry (NACB) Distinguished Scientist Award (2006), a Cheung Kong Scholars Achievement Award from the Ministry of Education of China (2006), a Silver Bauhinia Star from the Hong Kong SAR Government and the American Association for Clinical Chemistry (AACC)-NACB

Award for Outstanding Contribution to Clinical Chemistry in a Selected Area of Research (2012). He was elected a Fellow of the Royal Society of London in 2011.

References

- 1. Evans MI, Wapner RJ. 2005 Invasive prenatal diagnostic procedures 2005. Semin. Perinatol. 29, 215 – 218. (doi:10.1053/j.semperi.2005.06.004)
- Malone FD et al. 2005 First-trimester or secondtrimester screening, or both, for Down's syndrome. N. Engl. J. Med. 353, 2001-2011. (doi:10.1056/ NEJMoa043693)
- Bianchi DW, Williams JM, Sullivan LM, Hanson FW, Klinger KW, Shuber AP. 1997 PCR quantitation of fetal cells in maternal blood in normal and aneuploid pregnancies. Am. J. Hum. Genet. 61, 822-829. (doi:10.1086/514885)
- Bianchi DW et al. 2002 Fetal gender and aneuploidy detection using fetal cells in maternal blood: analysis of NIFTY I data. National Institute of Child Health and Development Fetal Cell Isolation Study. Prenat. Diagn. 22, 609-615. (doi:10. 1002/pd.347)
- Chen XQ, Stroun M, Magnenat J-L, Nicod LP, Kurt A-M, Lyautey J, Lederrey C, Anker P, 1996 Microsatellite alterations in plasma DNA of small cell lung cancer patients. Nat. Med. 2, 1033 - 1035. (doi:10.1038/nm0996-1033)
- Nawroz H, Koch W, Anker P, Stroun M, Sidransky D. 1996 Microsatellite alterations in serum DNA of head and neck cancer patients. Nat. Med. 2, 1035 - 1037. (doi:10.1038/nm0996-1035)
- 7. Lo YMD, Corbetta N, Chamberlain PF, Rai V, Sargent IL, Redman CWG, Wainscoat JS 1997 Presence of fetal DNA in maternal plasma and serum. Lancet 350, 485-487. (doi:10.1016/S0140-6736(97)02174-0)
- Lo YMD et al. 1998 Quantitative analysis of fetal DNA in maternal plasma and serum: implications

- for noninvasive prenatal diagnosis. Am. J. Hum. Genet. **62**, 768-775. (doi:10.1086/301800)
- Lun FMF, Chiu RWK, Chan KCA, Leung TY, Lau TK, Lo YMD. 2008 Microfluidics digital PCR reveals a higher than expected fraction of fetal DNA in maternal plasma. Clin. Chem. **54**, 1664-1672. (doi:10.1373/clinchem.2008.111385)
- 10. Lo YMD, Zhang J, Leung TN, Lau TK, Chang AM, Hjelm NM. 1999 Rapid clearance of fetal DNA from maternal plasma. Am. J. Hum. Genet. 64, 218-224. (doi:10.1086/302205)
- 11. Smid M et al. 2003 No evidence of fetal DNA persistence in maternal plasma after pregnancy. Hum. Genet. 112, 617-618.
- 12. Bianchi DW, Zickwolf GK, Weil GJ, Sylvester S, DeMaria MA. 1996 Male fetal progenitor cells persist in maternal blood for as long as 27 years postpartum. Proc. Natl Acad. Sci. USA 93, 705 – 708. (doi:10.1073/pnas.93.2.705)
- 13. Devaney SA, Palomaki GE, Scott JA, Bianchi DW. 2011 Noninvasive fetal sex determination using cellfree fetal DNA: a systematic review and metaanalysis. JAMA 306, 627-636. (doi:10.1001/jama. 2011.1114)
- 14. Lo YMD, Hjelm NM, Fidler C, Sargent IL, Murphy MF, Chamberlain PF, Poon PMK, Redman CWG, Wainscoat JS. 1998 Prenatal diagnosis of fetal RhD status by molecular analysis of maternal plasma. N. Engl. J. Med. 339, 1734-1738. (doi:10.1056/NEJM199812103392402)
- 15. Finning K, Martin P, Summers J, Massey E, Poole G, Daniels G. 2008 Effect of high throughput RHD typing of fetal DNA in maternal plasma on use of anti-RhD immunoglobulin in RhD negative

- pregnant women: prospective feasibility study. Br. Med. J. **336**, 816-818. (doi:10.1136/bmj.39518. 463206.25)
- 16. Poon LLM, Leung TN, Lau TK, Chow KC, Lo YMD. 2002 Differential DNA methylation between fetus and mother as a strategy for detecting fetal DNA in maternal plasma. Clin. Chem. 48, 35-41.
- 17. Chim SSC et al. 2005 Detection of the placental epigenetic signature of the maspin gene in maternal plasma. Proc. Natl Acad. Sci. USA 102, 14 753 – 14 758. (doi:10.1073/pnas.0503335102)
- 18. Tong YK et al. 2006 Noninvasive prenatal detection of fetal trisomy 18 by epigenetic allelic ratio analysis in maternal plasma: theoretical and empirical considerations. Clin. Chem. 2194-2202. (doi:10.1373/clinchem.2006.076851)
- 19. Papageorgiou EA, Karagrigoriou A, Tsaliki E, Velissariou V, Carter NP, Patsalis PC. 2011 Fetalspecific DNA methylation ratio permits noninvasive prenatal diagnosis of trisomy 21. Nat. Med. 17, 510-513. (doi:10.1038/nm.2312)
- 20. Lo YMD et al. 2007 Plasma placental RNA allelic ratio permits noninvasive prenatal chromosomal aneuploidy detection. *Nat. Med.* **13**, 218–223. (doi:10.1038/nm1530)
- 21. Chan KCA et al. 2004 Size distributions of maternal and fetal DNA in maternal plasma. Clin. Chem. 50, 88 – 92. (doi:10.1373/clinchem.2003.024893)
- 22. Li Y, Zimmermann B, Rusterholz C, Kang A, Holzgreve W, Hahn S. 2004 Size separation of circulatory DNA in maternal plasma permits ready detection of fetal DNA polymorphisms. *Clin. Chem.* **50**, 1002–1011. (doi:10.1373/ clinchem.2003.029835)

- 23. Dhallan R et al. 2004 Methods to increase the percentage of free fetal DNA recovered from the maternal circulation. JAMA 291, 1114-1119. (doi:10.1001/jama.291.9.1114)
- 24. Dhallan R et al. 2007 A non-invasive test for prenatal diagnosis based on fetal DNA present in maternal blood: a preliminary study. Lancet 369, 474-481. (doi:10.1016/S0140-6736(07) 60115-9)
- 25. Chung GT, Chiu RWK, Chan KCA, Lau TK, Leung TN, Lo YMD. 2005 Lack of dramatic enrichment of fetal DNA in maternal plasma by formaldehyde treatment. Clin. Chem. 51, 655-658. (doi:10. 1373/clinchem.2004.042168)
- 26. Chinnapapagari SK, Holzgreve W, Lapaire O, Zimmermann B, Hahn S. 2005 Treatment of maternal blood samples with formaldehyde does not alter the proportion of circulatory fetal nucleic acids (DNA and mRNA) in maternal plasma. Clin. Chem. 51, 652-655. (doi:10.1373/ clinchem.2004.042119)
- 27. Lo YMD et al. 2007 Digital PCR for the molecular detection of fetal chromosomal aneuploidy. Proc. Natl Acad. Sci. USA 104, 13 116-13 121. (doi:10. 1073/pnas.0705765104)
- 28. Fan HC, Quake SR. 2007 Detection of aneuploidy with digital polymerase chain reaction. Anal. Chem. **79**, 7576 – 7579. (doi:10.1021/ac0709394)
- 29. Schuster SC. 2008 Next-generation sequencing transforms today's biology. Nat. Meth. 5, 16-18. (doi:10.1038/nmeth1156)
- 30. Chiu RWK et al. 2008 Noninvasive prenatal diagnosis of fetal chromosomal aneuploidy by massively parallel genomic sequencing of DNA in maternal plasma. Proc. Natl Acad. Sci. USA 105, 20 458 – 20 463. (doi:10.1073/pnas.0810641105)
- 31. Fan HC, Blumenfeld YJ, Chitkara U, Hudgins L, Quake SR. 2008 Noninvasive diagnosis of fetal aneuploidy by shotgun sequencing DNA from maternal blood. Proc. Natl Acad. Sci. USA 105, 16 266 – 16 271. (doi:10.1073/pnas.0808319105)
- 32. Chiu RWK, Sun H, Akolekar R, Clouser C, Lee C, McKernan K, Zhou D, Nicolaides KH, Lo YMD 2010 Maternal plasma DNA analysis with massively parallel sequencing by ligation for noninvasive prenatal diagnosis of trisomy 21. Clin. Chem. 56, 459 – 463. (doi:10.1373/clinchem.2009.136507)

- 33. Lun FMF, Jin YY, Sun H, Leung TY, Lau TK, Chiu RWK, Lo YMD 2011 Noninvasive prenatal diagnosis of a case of Down syndrome due to robertsonian translocation by massively parallel sequencing of maternal plasma DNA. Clin. Chem. 57, 917-919. (doi:10.1373/clinchem.2011.161844)
- 34. Chiu RWK et al. 2011 Non-invasive prenatal assessment of trisomy 21 by multiplexed maternal plasma DNA sequencing: large scale validity study. Br. Med. J. 342, c7401. (doi:10. 1136/bmj.c7401)
- 35. Ehrich M et al. 2011 Noninvasive detection of fetal trisomy 21 by sequencing of DNA in maternal blood: a study in a clinical setting. Am. J. Obstet. *Gynecol.* **204,205** e1 – e11.
- 36. Palomaki GE et al. 2011 DNA sequencing of maternal plasma to detect Down syndrome: an international clinical validation study. Genet. *Med.* **13**, 913–920. (doi:10.1097/GIM.0b013e 3182368a0e)
- 37. Bianchi DW, Platt LD, Goldberg JD, Abuhamad AZ, Sehnert AJ, Rava RP. 2012 Genome-wide fetal aneuploidy detection by maternal plasma DNA sequencing. *Obstet. Gynecol.* **119**, 890 – 901. (doi:10.1097/AOG.0b013e31824fb482)
- 38. Chen EZ et al. 2011 Noninvasive prenatal diagnosis of fetal trisomy 18 and trisomy 13 by maternal plasma DNA sequencing. PLoS ONE 6, e21791. (doi:10.1371/journal.pone.0021791)
- 39. Sehnert AJ, Rhees B, Comstock D, de Feo E, Heilek G, Burke J, Rava RP 2011 Optimal detection of fetal chromosomal abnormalities by massively parallel DNA sequencing of cell-free fetal DNA from maternal blood. Clin. Chem. 57, 1042 – 1049. (doi:10.1373/clinchem.2011.165910)
- 40. Lau TK et al. In press. Noninvasive prenatal diagnosis of common fetal chromosomal aneuploidies by maternal plasma DNA sequencing. J. Matern. Fetal Neonatal Med. (doi:10.3109/14767058.2011.635730)
- 41. Palomaki GE et al. 2012 DNA sequencing of maternal plasma reliably identifies trisomy 18 and trisomy 13 as well as Down syndrome: an international collaborative study. Genet. Med. 14, 296-305. (doi:10.1038/gim.2011.73)
- 42. van den Oever JM et al. 2012 Single molecule sequencing of free DNA from maternal plasma for noninvasive trisomy 21 detection. Clin.

- *Chem.* **58**, 699 706. (doi:10.1373/clinchem.2011. 174698)
- 43. Liao GJ, Lun FMF, Zheng YWL, Chan KCA, Leung TY, Lau TK, Chiu RWK, Lo YMD. 2011 Targeted massively parallel sequencing of maternal plasma DNA permits efficient and unbiased detection of fetal alleles. Clin. Chem. 57, 92-101. (doi:10. 1373/clinchem.2010.154336)
- 44. Sparks AB et al. 2012 Selective analysis of cell-free DNA in maternal blood for evaluation of fetal trisomy. *Prenat. Diagn.* **32**, 3–9. (doi:10.1002/pd.2922)
- 45. Sparks AB, Struble CA, Wang ET, Song K, Oliphant A. 2012 Noninvasive prenatal detection and selective analysis of cell-free DNA obtained from maternal blood: evaluation for trisomy 21 and trisomy 18. Am. J. Obstet. Gynecol. **206**, 319.e1-319.e9. (doi:10.1016/j.ajog.2012.01.030)
- 46. Ashoor G, Syngelaki A, Wagner M, Birdir C, Nicolaides KH. 2012 Chromosome-selective sequencing of maternal plasma cell-free DNA for first-trimester detection of trisomy 21 and trisomy 18. Am. J. Obstet. Gynecol. **206**, 322.e1-322.e5. (doi:10.1016/j.ajog.2012.01.029)
- 47. Lun FMF et al. 2008 Noninvasive prenatal diagnosis of monogenic diseases by digital size selection and relative mutation dosage on DNA in maternal plasma. Proc. Natl Acad. Sci. USA 105, 19 920 – 19 925. (doi:10.1073/pnas.0810373105)
- 48. Tsui NB et al. 2011 Noninvasive prenatal diagnosis of hemophilia by microfluidics digital PCR analysis of maternal plasma DNA. Blood 117, 3684-3691. (doi:10.1182/blood-2010-10-310789)
- 49. Lo YMD et al. 2010 Maternal plasma DNA sequencing reveals the genome-wide genetic and mutational profile of the fetus. Sci. Transl. Med. 2, 61ra91. (doi:10.1126/scitranslmed.3001720)
- 50. Greely HT. 2011 Get ready for the flood of fetal gene screening. *Nature* **469**, 289–291. (doi:10.1038/469289a)
- 51. de Jong A, Dondorp WJ, Frints SG, de Die-Smulders CE, de Wert GM. 2011 Advances in prenatal screening: the ethical dimension. Nat. Rev. Genet. **12**, 657 – 663. (doi:10.1038/nrg3036)
- 52. Chitty LS, Hill M, White H, Wright D, Morris S. 2012 Noninvasive prenatal testing for aneuploidy—ready for prime time? Am. J. Obstet. Gynecol. 206, 269 – 275. (doi:10.1016/j.ajog.2012.02.021)