

Cumulative Inflammatory Load Is Associated with Short Leukocyte Telomere Length in the Health, Aging and Body Composition Study

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Abstract

Background: Leukocyte telomere length (LTL) is an emerging marker of biological age. Chronic inflammatory activity is commonly proposed as a promoter of biological aging in general, and of leukocyte telomere shortening in particular. In addition, senescent cells with critically short telomeres produce pro-inflammatory factors. However, in spite of the proposed causal links between inflammatory activity and LTL, there is little clinical evidence in support of their covariation and interaction.

Methodology/Principal Findings: To address this issue, we examined if individuals with high levels of the systemic inflammatory markers interleukin-6 (IL-6), tumor necrosis factor- α (TNF- α) and C-reactive protein (CRP) had increased odds for short LTL. Our sample included 1,962 high-functioning adults who participated in the Health, Aging and Body Composition Study (age range: 70–79 years). Logistic regression analyses indicated that individuals with high levels of either IL-6 or TNF- α had significantly higher odds for short LTL. Furthermore, individuals with high levels of both IL-6 and TNF- α had significantly higher odds for short LTL compared with those who had neither high (OR = 0.52, CI = 0.37–0.72), only IL-6 high (OR = 0.57, CI = 0.39–0.83) or only TNF- α high (OR = 0.67, CI = 0.46–0.99), adjusting for a wide variety of established risk factors and potential confounds. In contrast, CRP was not associated with LTL.

Conclusions/Significance: Results suggest that cumulative inflammatory load, as indexed by the combination of high levels of IL-6 and TNF- α , is associated with increased odds for short LTL. In contrast, high levels of CRP were not accompanied by short LTL in this cohort of older adults. These data provide the first large-scale demonstration of links between inflammatory markers and LTL in an older population.

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Competing Interests: The authors have read the journal's policy and have the following conflicts to declare. Drs. Epel and Blackburn are co-founders in a company measuring telomere diagnostics, Telomere Health Inc. This does not alter the authors' adherence to all the PLoS ONE policies on sharing data and materials. None of the other authors have competing interests in relation to this work.

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Introduction

Telomeres, the DNA-protein complexes that cap the ends of chromosomes and protect against genomic instability, contain variable-length tracts of telomeric DNA. Leukocyte telomere length (LTL) is increasingly recognized as an index of biological age that predicts incidence of age-related diseases [1,2], as well as all-cause and disease-specific mortality in diverse cohorts of older

adults [1,3,4,5,6]. In fact, older adults with below average LTL have more than threefold increased risk for early mortality [4,6]. Long LTL is also a predictor of years of healthy living, an outcome that integrates self-perceived functional status and duration of survival [7]. Importantly, telomere shortening is not just an index, but also a potential mechanism underlying some aspects of biological aging because short telomere length can lead to cessation of mitosis with consequent loss of ability for cell replenishment and, in some cellular settings, genomic instability, end-to-end chromosome fusion, and apoptosis as well as harmful DNA damage signaling and mitochondrial dysfunction [8,9,10,11]. Notwithstanding the utility of LTL in predicting mortality and longevity, there is little clinical evidence regarding modifiable factors associated with and potentially driving leukocyte telomere shortening.

Inflammatory activity is frequently proposed as a contributor to biological aging in general, and leukocyte telomere shortening in particular [5,12,13,14]. Accumulating evidence suggests a causal role for inflammation in the pathogenesis of multiple age-related diseases including cancer, atherosclerosis, autoimmune disorders, diabetes, and neurodegenerative diseases [15,16,17,18,19,20,21]. Elevated inflammatory activity could accelerate leukocyte telomere shortening by promoting cell turnover and replicative senescence [22], and by inducing the release of reactive oxygen species that damage telomeric DNA via oxidative stress [23]. One complication is that inflammatory cytokines including tumor necrosis factor-α (TNF-α) can both inhibit and promote activity of telomerase [24,25,26], the cellular enzyme primarily responsible for lengthening telomeres [27]. On the other hand, in vitro research on various cell types, but not including normal leukocytes, indicates that an accumulation of senescent cells with critically short LTL may produce proinflammatory factors [28,29], hich could in turn contribute to an increased inflammatory load.

Observations of short LTL in small samples of patients with inflammatory diseases including hepatitis C, liver cirrhosis, chronic kidney disease and chronic obstructive pulmonary disease provide preliminary support for the proposal that inflammation accelerates leukocyte telomere shortening [5,14,30,31]. Specifically, in these studies, patients with inflammatory disorders exhibited shorter LTL cross-sectionally compared with healthy individuals [31,32], and shorter telomeres in cells proximal to disease-related inflammatory activity [14,30]. Additionally, elevations in disease-specific and systemic inflammatory markers have been associated with short LTL in patients [5]. Although there are plausible pathways by which inflammation could accelerate telomere shortening even in adults without chronic disease, little is known about *in vivo* relationships between markers of inflammation and LTL in healthy older adults.

In the present study, we examined if elevated inflammatory activity, indexed by high levels of the systemic inflammatory cytokines interleukin-6 (IL-6) and TNF- α and the acute phase protein C-reactive protein (CRP), is associated with increased risk for short LTL in the 'Health, Aging and Body Composition' (Health ABC) cohort, a large sample of well-functioning men and women aged 70–79 years. Our sample included 1,962 Health ABC participants who had complete data available for our primary analyses. We hypothesized that participants with high levels of IL-6 and/or TNF- α and/or CRP would be more likely than participants with lower levels of these inflammatory markers to have short LTL, and that the combination of high IL-6, high TNF- α and high CRP, as an index of a high cumulative inflammatory load, would confer the greatest odds for short LTL.

Results

In spite of the restricted age range of the sample, age was significantly associated with shorter LTL (r=-.07, p=.002) and higher levels of TNF- α (r=.07, p=.003). However, age was negatively associated with levels of CRP (r=-.07, p=.001) and was not associated with levels of IL-6 (r=.03, p=.19). Inflammatory markers were significantly and positively associated with one another. However, while the effect size for the relationship between IL-6 and CRP was medium to large (r=.47, p<.001), relationships between IL-6 and TNF- α (r=.27, p<.001) as well as between TNF- α and CRP (r=.11, p<.001) had smaller effect sizes. Table 1 summarizes the baseline characteristics of the study population stratified by LTL tertile. As predicted, participants in the bottom tertile for LTL had the highest level of IL-6 and TNF- α , but there were no differences between groups in levels of CRP.

Inflammatory Activity and LTL

In our primary analytic model, we found that the odds of having short LTL (i.e., LTL in the bottom tertile or ≤4260 bp) were significantly higher for those participants who had high (i.e., top tertile) levels of either IL-6 (≥2.39 pg/mL), TNF-α (≥3.72 pg/ mL) or CRP (≥2.51 mg/L). In these analyses, the results of which are summarized in Table 2, groups were not mutually exclusive, such that the same individual could be represented in multiple groups if he or she had high levels of IL-6, TNF-α and CRP. Adjusting for a wide range of potential confounds and covariates, the odds for short LTL were significantly higher in those who had either IL-6 levels in the top tertile (OR = 1.3, 95% CI = 1.1-1.7) or TNF- α levels in the top tertile (OR = 1.5, 95% CI = 1.2–1.9). The highest odds for short LTL were observed in those participants who had high levels of both IL-6 and TNF- α (OR = 1.8, CI = 1.3– 2.4). In contrast, the addition of high levels of CRP did not confer increased odds for short LTL, as indexed by the lack of association between CRP and LTL (OR = 1.1, 95% CI = 08-1.4), and by the roughly equal odds of having short LTL for those who had high levels of CRP in addition to high levels of IL-6 and TNF-α (OR = 1.7, CI = 1.1-2.6).

We additionally ran the same models using the established highrisk clinical cutoff (>3 mg/L) for CRP [33]. While individuals who had CRP above this clinical cutoff had marginally higher odds for short LTL, this was not significant in any of the models, including when we only adjusted for telomere batch (OR = 1.3, CI = 1.0–1.7). Thus, individuals with CRP in either the top tertile in the sample or above the recognized clinical cutoff did not appear to have significantly increased odds for short LTL.

To further examine the observed associations of IL-6 and TNFα with short LTL and to compare mutually exclusive groups instead of potentially overlapping groups, we excluded CRP from our model and compared the odds for short LTL among four mutually exclusive groups of participants: those with both high IL-6 and high TNF-α; only high IL-6; only high TNF-α; and neither high IL-6 nor high TNF-α. These analyses were conducted while statistically controlling for established risk factors and potential confounds and with both high IL-6 and high TNF-α as our reference group. Results indicated that the combination of both high IL-6 and high TNF-α levels was associated with roughly double the odds for short LTL compared with having neither high IL-6 nor high TNF- α (OR = 0.52, CI = 0.37–0.72). Furthermore, the combination of high IL-6 and high TNF-α was associated with significantly higher odds for short LTL compared with having only IL-6 high (OR = 0.57, CI = 0.39–0.83) or only TNF- α high (OR = 0.67, CI = 0.46-0.99). Finally, mean LTL adjusted for site, age, gender, and race was 201 base pairs shorter in participants

Table 1. Sample characteristics by leukocyte telomere length (LTL) tertile.

		LTL Tertile				
		Short	Middle	Long	p value	
LTL (base pairs)		≤4260	4261–5280	>5280		
Age (years), M (SD)		74.0 (2.8)	73.5 (2.9)	73.4 (2.9)	<.01**	
Race (Black), n (%)		239 (36.5)	274 (41.8)	271 (41.6)	.09	
Sex (female), n (%)		255 (39.0)	330 (50.3)	398 (61.0)	<.01**	
Site (Memphis), n (%)		324 (49.5)	367 (56.0)	293 (44.9)	<.01**	
Body mass index (kg/m²)		27.6 (4.8)	27.4 (4.9)	27.3 (4.7)	.59	
Smoking status, n (%)						
	Current	72 (11.0)	73 (11.1)	66 (10.1)		
	Former	319 (48.8)	305 (46.5)	270 (41.4)		
	Never	263 (40.2)	278 (42.4)	316 (48.5)	.04*	
Sleep (hours/night)		6.9 (1.3)	6.9 (1.4)	6.8 (1.3)	.79	
Alcohol (drinks/week), n (%)						
	0	303 (46.3)	333 (50.7)	328 (50.3)		
	<1	142 (21.7)	144 (22.0)	129 (19.8)		
	1–7	160 (24.5)	135 (20.6)	142 (21.8)		
	>7	49 (7.5)	44 (6.7)	53 (8.1)	.46	
xercise (kcal/kg/week)		85.9 (73.5)	85.7 (73.1)	82.6 (64.0)	.93	
Chronic condition, n (%)		444 (67.9)	448 (68.2)	464 (71.2)	.38	
Recent infection, n (%)		43 (6.6)	46 (7.0)	52 (8.0)	.61	
Anti-inflammatory medications, n (%)		344 (52.6)	356 (54.3)	332 (50.9)	.48	
Statins, n (%)		89 (13.6)	79 (12.0)	97 (14.9)	.32	
ncome, n (%)						
	<10 k	68 (10.4)	86 (13.1)	88 (13.5)		
	10 k-25 k	262 (40.0)	272 (41.5)	242 (37.1)		
	25 k–50 k	217 (33.2)	205 (31.3)	218 (33.4)		
	>50 k	107 (16.4)	93 (14.2)	104 (16.0)	.38	
Education, n (%)						
	<hs< td=""><td>156 (23.9)</td><td>166 (25.3)</td><td>123 (18.9)</td><td></td></hs<>	156 (23.9)	166 (25.3)	123 (18.9)		
	HS	191 (29.2)	184 (28.1)	186 (28.4)		
	>HS	307 (46.9)	306 (46.7)	343 (52.6)	.05*	
Interleukin-6: Mdn (IQR)		1.9 (1.3–3.0)	1.8 (1.3–2.8)	1.7 (1.2–4.5)	.05*	
Tumor necrosis factor-α: Mdn (IQR)		3.4 (2.7–4.4)	3.2 (2.5–4.1)	3.0 (2.3–3.8)	<.01**	
C-reactive protein: Mdn (IQR)		1.6 (1.0-3.0)	1.6 (0.9-3.0)	1.74 (1.0-3.3)	.17	

Notes. HS = high school; IQR = interquartile range; LTL = leukocyte telomere length; M = mean; Mdn = Median; n = number of participants; SD = standard deviation; p-values for comparisons of means were calculated using one-way ANCOVA tests for continuous variables, and chi-squared tests for categorical variables. One-way ANCOVAs were calculated using log-transformed values because variances were not equal between groups. However, reported means and medians are based on raw data

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with high levels of both IL-6 and TNF- α (M=4701.93 bp, SE=68.07) compared with that of participants who did not have high levels of both of these (M=4902.21 bp, SE=28.20), and this difference in absolute LTL was significant, F(1,1937)=7.35, p=.007.

Discussion

Short LTL, a potential index of biological age, is associated with increased risk for age-related diseases as well as early all-cause and disease-specific mortality [3,4,5,6]. The present research study is to date the largest to demonstrate an association between elevated inflammatory activity and short LTL. Moreover, the present study

is the first to indicate that the combination of high IL-6 and TNF- α is associated with increased odds for short LTL, over and above the odds associated with having high levels of only one of these markers. Of note, having high levels of both IL-6 and TNF- α conferred almost twice the odds of being in the bottom tertile for LTL compared with having lower levels of both markers. In contrast, having high levels of CRP, as indexed by CRP levels either in the top tertile for the sample or above the established clinical cutoff, was not associated with increased odds for short LTL. Importantly, all of our results held when controlling for the contribution of numerous established risk factors for short LTL and potential confounds. Together with the observed causal relationships between inflammatory activity and LTL in animal

Table 2. Systemic inflammatory markers as predictors of short leukocyte telomere length.

	Unadjust	Unadjusted		Adjusted for age, gender and race		Adjusted for all covariates	
	OR	(95% CI)	OR	(95% CI)	OR	(95% CI)	
IL-6	1.3	(1.0–1.6)*	1.3	(1.0–1.6)*	1.3	(1.0–1.7)*	
TNF-α	1.6	(1.3-2.1)**	1.5	(1.2–1.91)**	1.5	(1.2–1.9)**	
CRP	0.9	(0.7-1.1)	1.1	(0.86-1.4)	1.1	(0.8-1.4)	
IL-6+TNF-α	1.9	(1.4–2.6)**	1.7	(1.3-2.4)**	1.8	(1.3-2.4)**	
IL-6+CRP	1.1	(0.9–1.5)	1.3	(0.9–1.7)	1.3	(0.9–1.7)	
TNF-α+CRP	1.6	(1.2-2.2)**	1.7	(1.2-2.4)**	1.7	(1.2-2.4)**	
IL-6+TNF-α+CRP	1.7	(1.2-2.5)**	1.7	(1.1–2.5)**	1.7	(1.1-2.6)**	

Notes.

*indicates p < .05 and

**indicates p < .01;

OR refers to odds ratio based on logistic regression; 95% CI is the 95% confidence interval; IL-6 = Interleukin-6; $TNF-\alpha$ = Tumor necrosis factor- α ; CRP = C-reactive protein. doi:10.1371/journal.pone.0019687.t002

and in vitro studies, these data provide preliminary evidence that adjunct anti-inflammatory therapies could potentially prevent accelerated leukocyte telomere shortening or protect against the negative effects of short LTL in older adults.

We included IL-6, TNF- α and CRP as three separate measures of systemic inflammatory activity in our study. Although all of these inflammatory markers were significantly correlated with one another, relationships between them were not strong (r's≤.47). IL-6 and CRP were the most strongly associated of the inflammatory markers, sharing 22% of variance. Given that IL-6 is a major inducer of CRP production by hepatocytes [34,35], the larger effect size for the relationship between these markers is not unexpected. However, high levels of IL-6, but not high levels of CRP, were associated with increased odds of short LTL in the sample. There is evidence that IL-6 is necessary but not sufficient to induce CRP synthesis [36], and also evidence that human coronary artery smooth muscle cells may produce CRP and that IL-6 is not a necessary inducer of CRP synthesis by these cells [37]. Thus, although IL-6 is an inducer of CRP and their concentrations are correlated, the physiological actions of IL-6 and CRP are not expected to be identical. The present findings, which need to be further investigated, suggest that in the case of LTL, these two factors do not have the same effects. Notably with regard to our overall findings, IL-6 and TNF- α shared only 7% of variance, supporting the formulation that circulating levels of these cytokines reflect different aspects of the inflammatory response. Thus, while each of these cytokines may contribute to leukocyte telomere shortening by promoting cell turnover and replicative senescence, inducing oxidative stress, and regulating telomerase activity [22,23,24,25,26], chronic exposure to the cumulative load of both high IL-6 and high TNF-α may have the greatest impact on the rate of leukocyte telomere shortening.

Equally plausible, however, is the emerging hypothesis that participants with short LTL in our study had the greatest proportion of accumulated senescent bodily cells overall, including fibroblasts and epithelial cells, which could plausibly contribute to the observed higher levels of IL-6 and TNF-α in this group [28,38]. Given that TNF- α and IL-6 may inhibit programmed cell death in specific cell types [39,40], high levels of these cytokines could even contribute to the maintenance of senescent cells in the system and hence the continued production of pro-inflammatory factors. However, the cross-sectional design of our study precludes drawing conclusions about the causal direction in the relationship between inflammatory activity and LTL.

In previous research, the cumulative load of high IL-6, high TNF-α and high CRP was found to confer the greatest risk for cardiovascular events in the Health ABC cohort [7]. However, CRP did not likewise contribute to the predictive value of the inflammatory markers with regard to LTL. Our finding of no association between CRP and LTL is in line with a previous finding of no associations between CRP and LTL in postmenopausal women, and in 'Cardiovascular Health Study' participants who were older than 73 years [7]. However, this finding does not support our hypothesis that higher levels of all measures of inflammatory activity would be associated with shorter LTL. Previous research has demonstrated that although CRP is reliably associated with cardiovascular disease (CVD) outcomes, CRP does not appear to be causally involved in the development of CVD [41]. Furthermore, CRP is less predictive of CVD in older compared with younger adults [42,43] and CRP levels were not predictive of cardiovascular events in a sample of high-risk Japanese adults in whom IL-6 was an independent predictor of such events [44]. However, it is possible that CRP would be associated with LTL in older populations with other diseases, such as inflammatory diseases characterized by very high levels of CRP. Our findings in relation to CRP may have particular relevance in clinical settings where CRP is the most commonly used index of inflammatory activity because they suggest that pro-inflammatory cytokine concentrations may serve as complementary indices of inflammation.

The present findings must be interpreted in the context of several limitations. First, the cross-sectional design of the present study precludes causal interpretations. Second, the narrow age range of our sample of older adults (70-79 years) is both a strength and weakness of our study. While this narrow age range allows us to maximize power to detect associations between inflammatory activity and LTL without many of the confounding factors associated with a sample of wider age range (e.g., cohort effects, hormonal effects), the findings of the present study may not be generalizable to other stages of the lifespan. Moreover, antiinflammatory medications were common in our sample of older adults. Given the wide range of medications that have antiinflammatory effects and the variability in dosage across individuals, our statistical control for anti-inflammatory medications is likely to be insufficient but is the best option currently

available. There has been debate about the relative merits of different methods used to assay LTL and the Q-PCR method has been subject to criticism [45,46]. However, strong associations between LTL as assessed by the Southern blot method and Q-PCR method have been documented [47], and the coefficient of variation of the LTL assay for this study was low at 4%. In addition, some findings related to LTL in Health ABC have been replicated in a cohort who had LTL measured with the traditional Southern blot technique [48]. Finally, it should be noted that effect sizes were small in this study and caution is therefore advised in the interpretation of the results pending future studies in independent cohorts.

Data from this large cohort of older adults indicates links between inflammatory activity and biological aging. In particular, the present data indicate that the cumulative load of high IL-6 and high TNF- α is accompanied by increased risk for short LTL. The present data also replicate a previous finding of no association between CRP and LTL in older adults, and suggest that systemic inflammatory cytokines may have more relevance to leukocyte telomere shortening than the more commonly used inflammatory marker CRP. In sum, older adults with high levels of inflammatory activity may be at increased risk for accelerated leukocyte telomere shortening, and those with short LTL may have increased risk for diseases with an inflammatory etiology.

Methods

Study Population

Study participants for this investigation included 1,962 wellfunctioning men and women aged 70-79 years who participated in Health ABC. Participants were identified from a random sample of white Medicare beneficiaries and all age-eligible black community residents in designated ZIP code areas surrounding Pittsburgh and Memphis. To be eligible for participation in Health ABC, participants had to report no difficulty in walking onequarter mile (0.5 km) or climbing 10 stairs without resting. Exclusion criteria included reported difficulty performing basic activities of daily living, obvious cognitive impairment, inability to communicate with the interviewer, intention of moving within 3 years, or participation in a trial involving a lifestyle intervention. Additionally, of the 3.075 participants who took part in Health ABC, our sample of 1,962 includes only participants who had complete data on LTL, inflammatory markers and all covariates included in our analysis. Our sample was not significantly different from excluded participants with missing data on age or site, but they were less likely to be female (50.1% of subsample versus 54% of non-sample, p = .04) and less likely to be Black (40% of subsample versus 44.7% of non-sample, p = .01). All participants gave written informed consent. The Institutional Review Boards at the University of Pittsburgh, the University of Tennessee and the University of California, San Francisco approved the protocol. Baseline data were collected from 1997 to 1998.

LTL Measurement

Quantitative polymerase chain reaction (Q-PCR) was used to measure LTL in the genomic DNA of peripheral leukocytes by determining the ratio of telomere repeat copy number to single-copy gene copy number (T/S ratio) in study samples relative to a reference sample [47,49]. The Q-PCR assay was conducted on three separate DNA samples per participant, and average LTL was calculated as the mean value of these triplicates. Each T/S value was later converted to number of base pairs (bp) by multiplying the T/S value by the known LTL of the reference DNA, which is a pooled sample of DNAs from several normal

Utah whites aged 65 years and older. The slope of the linear regression line through a plot of T/S ratio (the x axis) versus mean TRF length (the y axis) is the number of base pairs of telomeric DNA corresponding to a single T/S unit. All samples were measured in triplicate and the mean value of the triplicates was used in analyses. The coefficient of variation for this assay is 4% and results obtained with the Q-PCR method are strongly associated with the traditional terminal restriction fragment length index of LTL obtained by Southern blot technique [47]. DNA was available for 2,880 of the 3,075 individuals who participated in Health ABC, and LTL was successfully measured in 2,721 cases.

Inflammatory Markers

Blood samples for inflammatory markers were obtained in the morning (median time was 9:19 AM; interquartile range was from 8:49 AM to 9:52 AM). IL-6 and CRP levels were measured in serum and TNF-α levels were measured in plasma. Tubes for IL-6 and CRP were serum separator tubes containing no anticoagulant. Tubes for TNF-α were citrated tubes containing 0.5 mL of 3.8% sodium citrate. After processing, the specimens were aliquoted into cryovials, frozen at -70° C, and shipped to the Core Laboratory at the University of Vermont. Serum IL-6 levels and plasma TNF-α levels were measured in duplicate by enzyme-linked immunosorbent assay (ELISA) kits from R&D Systems (Minneapolis, MN). The detectable limit for IL-6 (by HS600 Quantikine kit) was .10 pg/mL, and for TNF-α (by HSTA50 kit) was .18 pg/mL. Plasma levels of CRP were also measured in duplicate by ELISA based on purified protein and polyclonal anti-CRP antibodies (Calbiochem, San Diego, CA). The CRP assay was standardized according to the World Health Organization First International Reference Standard, with a sensitivity of 0.08 µg/mL. Measures of IL-6, TNF-α and CRP were missing for 162, 203, and 38 participants respectively. Intra and inter-assay coefficients of variation for inflammatory markers were respectively: 10% and 15% for IL-6; 16% and 15% for TNF-α; and 8% and 5% for CRP

Covariates

Covariates were selected a priori based on previous research and included characteristics associated with inflammatory markers and LTL in previous research. Such a priori selection of covariates has been validated for logistic regression procedures [50,51]. The telomere batch number corresponds to a particular run of the O-PCR assay used to measure LTL. Because there is some variability between assays, we controlled for telomere batch in all LTL analyses, constructing a dummy variable for each batch to enter into regression models. Sociodemographic covariates included study site, age, gender, and race, as well as education (less than high school; high school; more than high school) and income (<\$10 k; \$10-25 k; \$25-50 k; >\$50 k). BMI was calculated as weight in kilograms divided by height in meters squared. Behavioral factors including smoking status (current, former, never), average alcohol use during the past year (0, <1, 1-7 or >7)drinks/week), exercise (total kilocalories burned per week) and hours of sleep per night on average were assessed in the baseline interview. Participants were asked whether they had experienced symptoms of respiratory infections within the last 2 weeks. The baseline presence of chronic illnesses including lung disease, heart disease (including myocardial infarction, angina pectoris, and congestive heart failure), stroke, diabetes mellitus, broken hip, and arthritis was adjudicated using standardized algorithms considering various sources of information: self-report of a medical diagnosis, medication use, and results of screening tests from a clinical examination where appropriate. All medications regularly taken in the past 2 weeks were recorded and coded according to the Iowa Drug Information System (IDIS) code [52]. Using this drug inventory, the daily use of antiinflammatory drugs (IDIS codes 2808 or 5208) and statins (IDIS code 2406) was assessed.

Our primary models were run with three different sets of covariates. First, adjusting only for telomere batch; second, adjusting for telomere batch, as well as potential confounds including study site, age, gender and race; and third, adjusting for telomere batch, as well as potential confounds and other potential covariates including study site, age, gender, race, income, education, BMI, smoking status, alcohol use, exercise, sleep, chronic disease, recent respiratory illness, and the use of anti-inflammatory drugs or statins.

Statistical Analysis

Logistic regression and analysis of covariance were used to test our primary hypothesis that high levels of inflammatory activity would be associated with increased odds for short LTL, as indexed by the odds of having LTL in the bottom tertile in the sample and by LTL in base pairs, respectively. For the purpose of these analyses, participants with values in the upper tertile of the sample were classified as "high" for inflammatory markers and those with values in the lower tertile of the sample were classified as "short" for LTL. We also examined if having CRP levels above the clinical cutoff of >3 mg/L was associated with increased odds for short LTL.

Logistic regression analyses were used to examine if participants with high levels of IL-6, TNF- α and CRP alone or in combination would be more likely than participants with lower levels of these inflammatory markers to have short LTL. In our first set of analyses, groups were not mutually exclusive, such that the same individual could be represented in multiple groups if they had high levels of IL-6, TNF- α or CRP. Based on the results of these analyses, we excluded CRP from our model and conducted follow-

References

- Willeit P, Willeit J, Mayr A, Weger S, Oberhollenzer F, et al. (2010) Telomere length and risk of incident cancer and cancer mortality. JAMA 304: 69-75
- Fitzpatrick AL, Kronmal RA, Gardner JP, Psaty BM, Jenny NS, et al. (2007) Leukocyte telomere length and cardiovascular disease in the Cardiovascular Health Study. Am J Epidemiol 165: 14–21.
- Farzaneh-Far R, Cawthon RM, Na B, Browner WS, Schiller NB, et al. (2008)
 Prognostic value of leukocyte telomere length in patients with stable coronary
 artery disease: data from the Heart and Soul Study. Arterioscler Thromb Vasc
 Biol 28: 1379–1384.
- Epel ES, Merkin SS, Cawthon R, Blackburn EH, Adler NE, et al. (2009) The rate of leukocyte telomere shortening predicts mortality from cardiovascular disease in elderly men. Aging 1: 81–88.
- Carrero JJ, Stenvinkel P, Fellstrom B, Qureshi AR, Lamb K, et al. (2008) Telomere attrition is associated with inflammation, low fetuin-A levels and high mortality in prevalent haemodialysis patients. J Int Med 263: 302–312.
- Cawthon RM, Smith KR, O'Brien E, Sivatchenko A, Kerber RA (2003) Association between telomere length in blood and mortality in people aged 60 years or older. Lancet 361: 393–395.
- Njajou OT, Hsueh WC, Blackburn EH, Newman AB, Wu SH, et al. (2009)
 Association between telomere length, specific causes of death, and years of healthy life in health, aging, and body composition, a population-based cohort study. J Gerontol A Biol Sci Med Sci 64: 860–864.
- 8. Blackburn EH (1991) Structure and function of telomeres. Nature 350: 569–573.
- Blackburn EH (2001) Switching and Signaling at the Telomere. Cell 106: 661–673.
- Sahin E, Colla S, Liesa M, Moslehi J, Muller FL, et al. (2011) Telomere dysfunction induces metabolic and mitochondrial compromise. Nature 470: 359–365.
- Jaskelioff M, Muller FL, Paik JH, Thomas E, Jiang S, et al. (2011) Telomerase reactivation reverses tissue degeneration in aged telomerase-deficient mice. Nature 469: 102–106.
- Chung HY, Kim HJ, Kim KW, Choi JS, Yu BP (2002) Molecular inflammation hypothesis of aging based on the anti-aging mechanism of calorie restriction. Microsc Res Tech 59: 264–272.

up analyses to examine if the odds for short LTL in participants with high levels of both IL-6 and TNF- α was significantly higher than that of participants with high levels of only IL-6, high levels of only TNF- α , or high levels of neither IL-6 nor TNF- α . Thus, this second group of logistic regression analyses were conducted on mutually exclusive groups. All models were run with the three different sets of covariates as described in the covariates section. Analysis of covariance was used to compare mean differences in base pairs of telomeres between those with and without high levels of inflammatory markers.

In order to compare characteristics of the sample by telomere length tertile we performed analysis of variance and Kruskal-Wallis rank tests for continuous variables, and chi-squared tests for categorical variables. In order to analyze differences in LTL and cytokines by gender, linear regressions were used. Due to unacceptable levels of skewness, the variables IL-6, TNF- α and CRP were log-transformed when used in correlations and linear regressions. All analyses were performed using Stata version 9.2 and SPSS version 18.0.

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Author Contributions

Conceived and designed the experiments: AOD MP EHB EE. Analyzed the data: AOD MP EP RC. Contributed reagents/materials/analysis tools: RC. Wrote the paper: AOD MP EP FD EHB KY RC PLO W-CH SS ABN HNA SMR TBH EE. Interpretation of data: AOD MP EP FD EHB KY PLO W-CH SS ABN TBH EE. Critical administrative and technical support: HNA SMR.

- Chung HY, Kim HJ, Kim JW, Yu BP (2001) The Inflammation Hypothesis of Aging: molecular modulation by calorie restriction. Ann NY Acad Sci 928: 327–335.
- Kinouchi Y, Hiwatashi N, Chida M, Nagashima F, Takagi S, et al. (1998)
 Telomere shortening in the colonic mucosa of patients with ulcerative colitis.
 I Gastroenterol 33: 343–348.
- Pradhan AD, Manson JE, Rifai N, Buring JE, Ridker PM (2001) C-reactive protein, interleukin 6, and risk of developing type-2 diabetes mellitus. JAMA 286: 327–334.
- 280: 327–334. 16. Libby P, Ridker PM, Maseri A (2002) Inflammation and atherosclerosis. Circulation 105: 1135–1143.
- Wyss-Coray T (2006) Inflammation in Alzheimer disease: driving force, bystander or beneficial response? Nat Med 12: 1005–1015.
- Moss SF, Blaser MJ (2005) Mechanisms of disease: inflammation and the origins of cancer. Nat Clin Prac Oncol 2: 90–97.
- Harris TB, Ferrucci L, Tracy RP, Corti MC, Wacholder S, et al. (1999)
 Associations of elevated interleukin-6 and C-reactive protein levels with mortality in the elderly. Am J Med 106: 506–512.
- Volpato S, Guralnik JM, Ferrucci L, Balfour J, Chaves P, et al. (2001) Cardiovascular disease, interleukin-6, and risk of mortality in older women: The Women's Health and Aging Study. Circulation 103: 947–953.
- Slattery ML, Curtin K, Baumgartner R, Sweeney C, Byers T, et al. (2007) IL6, aspirin, nonsteroidal anti-inflammatory drugs, and breast cancer risk in women living in the southwestern United States. Cancer Epidemiol Biomarkers Prev 16: 747-755
- Aviv A (2004) Telomeres and human aging: facts and fibs. Sci Aging Knowledge Environ 2004: pe43.
- Jaiswal M, LaRusso NF, Burgart LJ, Gores GJ (2000) Inflammatory cytokines induce DNA damage and inhibit DNA repair in cholangiocarcinoma cells by a nitric oxide-dependent mechanism. Cancer Res 60: 184–190.
- Xu D, Erickson S, Szeps M, Gruber A, Sangfelt O, et al. (2000) Interferon alpha down-regulates telomerase reverse transcriptase and telomerase activity in human malignant and nonmalignant hematopoietic cells. Blood 96: 4313–4318.
- Parish ST, Wu JE, Effros RB (2009) Modulation of T lymphocyte replicative senescence via TNF-{alpha} inhibition: role of caspase-3. J Immunol 182: 4237-4243.



- Akiyama M, Yamada O, Hideshima T, Yanagisawa T, Yokoi K, et al. (2004) TNF[alpha] induces rapid activation and nuclear translocation of telomerase in human lymphocytes. Biochem Biophys Res Commun 316: 528-532
- 27. Greider CW, Blackburn EH (1989) A telomeric sequence in the RNA of Tetrahymena telomerase required for telomere repeat synthesis. Nature 337:
- 28. Rodier F, Coppe JP, Patil CK, Hoeijmakers WA, Munoz DP, et al. (2009) Persistent DNA damage signalling triggers senescence-associated inflammatory cytokine secretion. Nat Cell Biol 11: 973-979.
- Krtolica A, Parrinello S, Lockett S, Desprez PY, Campisi J (2001) Senescent fibroblasts promote epithelial cell growth and tumorigenesis: a link between cancer and aging. Proc Natl Acad Sci U S A 98: 12072-12077.
- Aikata H, Takaishi H, Kawakami Y, Takahashi S, Kitamoto M, et al. (2000) Telomere reduction in human liver tissues with age and chronic inflammation. Exp Cell Res 256: 578-582.
- 31. Steer SE, Williams FM, Kato B, Gardner JP, Norman PJ, et al. (2007) Reduced telomere length in rheumatoid arthritis is independent of disease activity and duration. Ann Rheum Dis 66: 476-480.
- 32. Houben JM, Mercken EM, Ketelslegers HB, Bast A, Wouters EF, et al. (2009) Telomere shortening in chronic obstructive pulmonary disease. Respir Med 103: 230-236.
- 33. Pearson TA, Mensah GA, Alexander RW, Anderson JL, Cannon RO, 3rd, et al. (2003) Markers of inflammation and cardiovascular disease: application to clinical and public health practice: A statement for healthcare professionals from the Centers for Disease Control and Prevention and the American Heart Association, Circulation 107: 499-511.
- 34. Castell JV, Gomez-Lechon MJ, David M, Andus T, Geiger T, et al. (1989) Interleukin-6 is the major regulator of acute phase protein synthesis in adult human hepatocytes. FEBS Lett 242: 237-239.
- Castell JV, Gomez-Lechon MJ, David M, Hirano T, Kishimoto T, et al. (1988) Recombinant human interleukin-6 (IL-6/BSF-2/HSF) regulates the synthesis of acute phase proteins in human hepatocytes. FEBS Lett 232: 347-350.
- 36. Weinhold B, Bader A, Poli V, Ruther U (1997) Interleukin-6 is necessary, but not sufficient, for induction of the humanC-reactive protein gene in vivo. Biochem J 325(Pt 3): 617-621.
- 37. Calabro P, Willerson JT, Yeh ET (2003) Inflammatory cytokines stimulated Creactive protein production by human coronary artery smooth muscle cells. Circulation 108: 1930-1932
- 38. Coppe JP, Patil CK, Rodier F, Sun Y, Munoz DP, et al. (2008) Senescenceassociated secretory phenotypes reveal cell-nonautonomous functions of oncogenic RAS and the p53 tumor suppressor. PLoS Biol 6: 2853-2868.

- 39. Lin MT, Juan CY, Chang KJ, Chen WJ, Kuo ML (2001) IL-6 inhibits apoptosis and retains oxidative DNA lesions in human gastric cancer AGS cells through up-regulation of anti-apoptotic gene mcl-1. Carcinogenesis 22: 1947-1953.
- 40. Mangan DF, Wahl SM (1991) Differential regulation of human monocyte programmed cell death (apoptosis) by chemotactic factors and pro-inflammatory cytokines, I Immunol 147: 3408-3412.
- 41. Elliott P, Chambers JC, Zhang W, Clarke R, Hopewell JC, et al. (2009) Genetic Loci associated with C-reactive protein levels and risk of coronary heart disease. IAMA 302: 37-48.
- 42. Kritchevsky SB, Cesari M, Pahor M (2005) Inflammatory markers and cardiovascular health in older adults. Cardiovasc Res 66: 265-275.
- 43. Cesari M, Penninx BW, Newman AB, Kritchevsky SB, Nicklas BI, et al. (2003) Inflammatory markers and cardiovascular disease: The Health, Aging and Body Composition [Health ABC] Study. Am J Cardiol 92: 522-528.
- Nishida H, Horio T, Suzuki Y, Iwashima Y, Tokudome T, et al. (2011) Interleukin-6 as an independent predictor of future cardiovascular events in high-risk Japanese patients: comparison with C-reactive protein. Cytokine 53: 342 - 346.
- 45. Aviv A (2009) Commentary: Raising the bar on telomere epidemiology. Int J Epidemiol 38: 1735-1736.
- Aviv A (2008) The epidemiology of human telomeres: faults and promises. J Gerontol A Biol Sci Med Sci 63: 979-983.
- Cawthon RM (2002) Telomere measurement by quantitative PCR. Nucl Acids Res 30: e47
- Njajou OT, Blackburn EH, Pawlikowska L, Mangino M, Damcott CM, et al. (2010) A common variant in the telomerase RNA component is associated with short telomere length. PLoS ONE 5: e13048.
- 49. Cawthon RM (2009) Telomere length measurement by a novel monochrome multiplex quantitative PCR method. Nucleic Acids Res 37: e21.
- 50. Steyerberg EW, Eijkemans MJ, Harrell FE, Jr., Habbema JD (2001) Prognostic modeling with logistic regression analysis: in search of a sensible strategy in small data sets. Med Decis Making 21: 45-56.
- Sun GW, Shook TL, Kay GL (1996) Inappropriate use of bivariable analysis to screen risk factors for use in multivariable analysis. J Clin Epidemiol 49:
- 52. Pahor M. Chrischilles EA. Guralnik IM. Brown SL. Wallace RB, et al. (1994) Drug data coding and analysis in epidemiologic studies. Eur J Epidemiol 10: 405-411.