

COMMENTARY

Covid-19 vaccination in patients with multiple myeloma: Focus on immune response

Heinz Ludwig¹  | Jesús San-Miguel² | Nikhil Munshi³ | Pieter Sonneveld⁴  |
María-Victoria Mateos⁵ | Philippe Moreau⁶  | Evangelos Terpos⁷ 

¹First Department of Medicine, Clinic Ottakring, Wilhelminen Cancer Research Institute, Vienna, Austria

²CIMA, IDISNA, CIBERONC, Clínica Universidad de Navarra, Pamplona, Spain

³Jerome Lipper Multiple Myeloma Center, Dana-Farber Cancer Institute, Harvard Medical School, Boston, Massachusetts, USA

⁴Erasmus MC Cancer Institute, Erasmus University of Rotterdam, Rotterdam, The Netherlands

⁵IBSAL, Cancer Research Center, University Hospital of Salamanca, Salamanca, Spain

⁶Department of Hematology, University Hospital Nantes, Nantes, France

⁷Hematology & Medical Oncology, Department of Clinical Therapeutics, National and Kapodistrian University of Athens, Athens, Greece

Correspondence

Heinz Ludwig, Wilhelminen Cancer Research Institute, c/o First Department of Medicine, Clinic Ottakring, Montleartstrasse 37, 1160 Vienna, Austria.
Email: heinz.ludwig@extern.gesundheitsverbund.at

Funding information

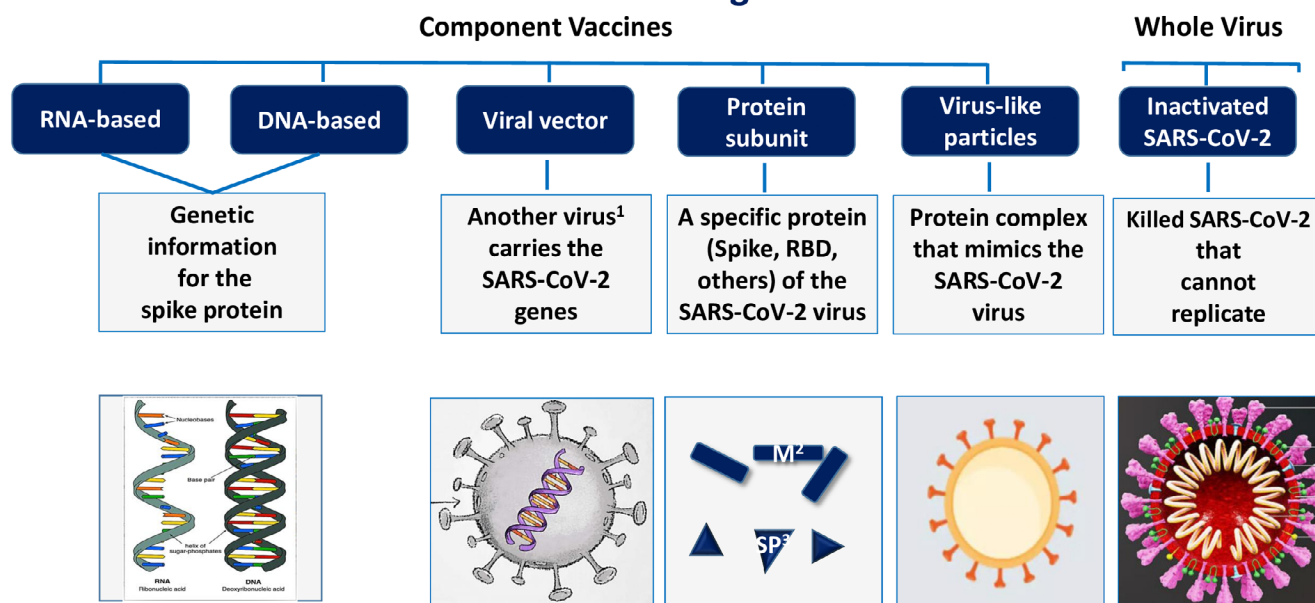
Austrian Forum against Cancer

Patients with multiple myeloma (MM) are at increased risk for severe clinical symptoms and mortality due to SARS-CoV-2 infection. To protect against these complications, the International Myeloma Society recommends vaccinating all patients with monoclonal gammopathy of undetermined significance (MGUS), monoclonal gammopathies of clinical significance, smoldering MM (SMM), and MM.¹ Protective immunity against SARS-CoV2 in those patients is partially relevant given their increased risk for infections, which may cause significant morbidity and represent the second most frequent cause of mortality.² The extent of the immune impairment in MM largely depends on the disease-inherent immune suppression exerted by the malignant clone that can affect all immune effector mechanism including B cells, T cells, NK cell, and dendritic cells, and the complement system.^{3,4} The myeloma-induced immune deficiency can resolve completely during periods of deep response, such as minimal residual disease negativity, but deep responses are achieved in a proportion of patients only, and several of those eventually relapse and become immune suppressed again and/or suffer from other causes of reduced immune competence. Many patients present at older age with comorbidities and immunosenescence⁵ with an impaired defense against infections and compromised development of long-term immune memory, which is aimed at by vaccination. Myeloma therapy including proteasome inhibitors, dexamethasone, high dose melphalan, monoclonal antibodies, antibody-drug conjugates, BiTEs

and cellular therapies, such as CAR-T cells, result in specific and cumulative immune suppression, and after extensive therapy in T cell exhaustion.⁶ Hence, myeloma and myeloma treatment-induced immunosuppression and other above mentioned factors frequently impose critical hurdles for an effective vaccination response in many patients and emerging data seem to underpin these concerns.⁷

A recent study in 48 elderly patients with MM (median age 83 years) reported a significantly lower proportion of MM patients with a neutralizing antibody response on day 22 after the first dose of the BNT162b2 vaccine compared to controls of the same age category (20.6% versus 32.5%, respectively). Virus neutralizing antibody titers of $\geq 50\%$ deemed to be clinically relevant, were noted in four (8.3%) of 48 patients and in 21 of 104 (20.2%) controls only; all these four patients were in remission without any therapy.⁸ Only one of the nine patients with SMM had neutralizing antibody titers. This poor response did not improve substantially with the second dose. Part of the observed low response rate could be due to the high age of the study participants as specific antibody production in response to vaccination declines with older age.^{9,10} Similar results were noted in 93 patients (median age 67 years) after one dose of the BNT162b2 or ChAdOx1-S vaccine.¹¹ After a median time from first dose to antibody testing of 33 days, SARS-CoV-2 IgG antibodies were noted in 55.9% of patients, with no difference between both vaccines.

Platforms used for manufacturing of SARS-CoV-2 vaccines



¹Adenovirus, Newcastle disease virus, Lentivirus, Vesicular stomatitis virus, Measles virus, ²Membrane proteins, ³Spike proteins

FIGURE 1 Platforms used for development of anti-SARS-CoV2 vaccines

These results compare with a positivity rate of 98.8% observed in hospital staff. When testing was expanded for IgA and IgM antibodies in 40 of the IgG antibody negative patients, a positive result was obtained in further 13 patients, adding up to a total antibody response in 65/93 patients (69.9%). Factors associated with antibody response were depth of response (CR-PR), absence of immunoparesis, and fewer lines of therapy, while being on therapy was associated with poor antibody response. Another presently unpublished study included seven patients with MM within a group of 38 patients with hematological cancers, which showed an IgG antibody response to the SARS-CoV-2 spike protein in only 13% of them, compared to 97% in healthy controls after one dose of the BNT162b2 vaccine.¹² Furthermore, half of the patients with haematologic malignancies tested had an impaired T cell response, and eight of the nine who showed a cellular immune response were serological non-responders. Similarly, a discordance between cellular and IgG antibody response was noted in previously Covid-19 infected health care workers; during follow up a T cell response, but no antibody formation was observed in seven out of 70 individuals, while in two subjects weak antibody titers, but no cellular responses were detected.¹³

These findings raise the question about the kind of humoral and cellular immune response which is required for protection against infection with SARS-CoV-2 virus, and which test systems provide most relevant information. Neutralizing antibodies have been shown to correlate with protection in rhesus macaques,¹⁴ and may be more informative than assessing levels of antibodies against spike proteins,

but data comparing their clinical relevance are not available as yet. For evaluation of cellular immunity, spike peptides are used for T-cell stimulation, but Covid-infected patients show reactivity against other peptides as well.¹⁵ This shows that available evidence is insufficient for clear guidance, therefore more laboratory data need to be generated and linked with clinical outcome including asymptomatic and symptomatic infection rate, hospitalization and mortality. Another issue of importance is the duration of the protection in patients with a documented immune response. Will antibodies and T-cell reactivity wane with time, even in patients in complete remission, and how fast will protection be lost in patients with progressive or recurrent disease? Will it be worthwhile to add additional doses of the same vaccine to non-responders or low-responders, or will a change of one vaccine to another elicit the desired immune response? For the latter strategy, several options are available, for instance mixing an RNA vaccine with an adenovirus vector, peptide, protein, inactivated virus, or adjuvanted vaccine, or a vaccine designed by a new molecular technology called NDV-HXP-S, which facilitates the manufacturing process. Figure 1 shows a schematic overview the various platforms used for anti-SARS-CoV2 vaccine production. More specifically new developments of interest are self-amplifying mRNA vaccines that produce more antigens, the design of vaccines for intranasal administration for induction of an additional secretory IgA response, or the production of bi- or trivalent vaccines, or of pan-respiratory virus vaccines including SARS-CoV-2, influenza, and other respiratory viruses.

The present vaccination program is endangered by the high mutation rate of SARS-CoV-2 virus leading to new mutants with relevant

TABLE 1 List of recent SARS-CoV-2 variants of concern, their clinical attributes and coverage by selected common vaccines

SARS-CoV-2 Variant	First detection		Evidence of clinical changes				Protected by
	Location	Date	Notable mutations	Transmissibility	Virulence	Antigenicity	
B.1.1.7	UK	Sep 2020	N501Y, 69-70del, P681H	Increased ~ 74% (NERVTAG)	32%–104% (~64%) more lethal (BMJ)	Reduced antigenic activity (ECDC)	BNT162b2 (Pfizer/BioNTech) mRNA-1273 (Moderna) AZD1222 (Oxford/AstraZeneca)
B.1.351	South Africa	Dec 2020	N501Y, K417N, E484K	Increased 50% (ECDC)	-	Reduced neutralization by antibodies (ECDC)	BNT162b2 (Pfizer/BioNTech) and mRNA-1273 (Moderna) may be 2/3 less effective (serum neutralizing antibodies), AZD1222 (Oxford/AstraZeneca) effective only in 10%
P.1	Brazil, Japan	Jan 2021	N501Y, E484K, K417T	Likely increased (CDC)	10%–80% (~45%) more lethal (CADDE)	Overall reduction in effective neutralization (ECDC)	Possible reduction of vaccine efficacy (ECDC)
B.1.525	Nigeria, UK	Dec 2020	E484K, F888L	Likely increased (CDC)	Likely increased (CDC)	„Modestly “reduced neutralization (COG-UK)	No data available as yet
B.1.427/B.1.429*	US	May 2020 July	L452R, D614G *plus S131, W152C	~20% increased (CDC)	Increased (CDC)	4.0–6.7 and two-fold decrease in neutralization titers, from convalescent patients and vaccine recipients, respectively (CDC)	No data available as yet
B.1.617	India	Oct 2020	E484Q, L452R, P681R	Increased (WHO)	Under investigation	Slight reduction in effective neutralization	No data available as yet

Abbreviations: CDC, Center for Disease Control; COG-UK, Covid-19 Genomics UK Consortium; ECDC, European Center for Disease Prevention and Control; NERVTAG, New and Emerging Respiratory Virus Threats Advisory Group; WHO, World health organization.

changes in the ACE-2 receptor-binding domain of the spike protein, which is one of the 26 virus proteins, and of other epitopes¹⁶ some of which resulting in higher transmissibility, virulence and lower antigenicity for all or some of the existing vaccines^{17–19} (Table 1). The neutralizing activity of the immune response elicited by the available mRNA vaccines was found to be significantly lower for the B.1.315 variant compared to the USA-WA1/2020 virus (identical to the original Wuhan SARS-CoV-2 virus).²⁰ Although neutralization titers usually are high in the normal population,¹⁰ lower neutralization efficacy can be critical in immunosuppressed patients like those with MM. Recent research indicates that certain mutations can also alter T-cell epitopes hiding mutated virus from recognition by T killer cells. Hence, adaptation of vaccines to the corresponding epitopes is necessary. A clinical trial has already been activated evaluating whether a vaccine targeting the B.1.315 variant (mRNA-1273.351) will provide better protection when given after a dose of the original mRNA-1273, or when administered concomitantly with the original vaccine.²¹

The above-mentioned considerations mandate close surveillance of the humoral and cellular vaccination response obtained in individual patients with MM to correlate findings with their level of protection. If improvement with further doses, other vaccines or vaccination strategies is unlikely in non-responders, new strategies such as employing monoclonal antibody cocktails against SARS-CoV-2 for protection against Covid-19 infection may be considered. These treatments are presently not approved for prophylactic use, but may turn out to protect all those with insufficient immune response to Covid-19 vaccines, particularly when preparations with extended half-life will be introduced. But until this option will become available for clinical practice, creation of “herd immunity” using a strategy of “ring vaccination” around affected patients will be required. Those patients will also be obliged to strictly follow general recommendations for infection risk reduction. They will desperately hope for new effective anti-COVID treatments, and finally for an end of the pandemic.

CONFLICT OF INTEREST

H.L.: Research Funding: Amgen, Takeda, Speaker's Bureau/Advisory Boards: Amgen, Takeda, Janssen, Celgene-BMS, Sanofi, Seattle Genetics; J.S.M.: Honoraria/Consultancy: Amgen, Celgene-BMS, GlaxoSmithKline, Janssen, Karyopharm, Novartis, MSD, Takeda, Sanofi, Roche; NM: Advisory Boards: Takeda, Celgene, Novartis; PS: Research Funding: Karyopharm, Amgen, Celgene, Janssen, Member BOD or advisory committee: SkylineDx, Honoraria: Amgen, Celgene, Janssen; M.V.M.: Honoraria: Janssen-Cilag, Celgene, Amgen, Takeda, GlaxoSmithKline, Abbvie/Genentech, Adaptive Biotechnologies, Consulting or Advisory Role: Takeda, Janssen-Cilag, Celgene, Amgen, Abbvie, GlaxoSmithKline, Pharmamar; P.M.: Honoraria/Advisory Boards: Abbvie, Janssen, Celgene-BMS, Amgen; ET: Research Funding: Amgen, Janssen, Celgene, Genesis Pharma, GlaxoSmithKline, Sanofi, Consultancy/Honoraria: Amgen, Celgene-BMS, Janssen, Takeda, Genesis Pharma, GlaxoSmithKline, Sanofi.

AUTHOR CONTRIBUTIONS

All authors contributed equally to concept and writing of the manuscript. All authors approved the final version.

DATA AVAILABILITY STATEMENT

Data sharing is not applicable to this article as no new data were created or analyzed in this study.

ORCID

Heinz Ludwig  <https://orcid.org/0000-0002-3302-8726>

Pieter Sonneveld  <https://orcid.org/0000-0002-0808-2237>

Philippe Moreau  <https://orcid.org/0000-0003-1780-8746>

Evangelos Terpos  <https://orcid.org/0000-0001-5133-1422>

REFERENCES

- <https://cms.cws.net/content/beta.myelomasociety.org/files/PM%20COVID%20vaccination%20in%20MM%20guidelines%20The%20Final.pdf>. Accessed March 24, 2021.
- Ludwig H, Boccadoro M, Moreau P, et al. Recommendations for vaccination in multiple myeloma: a consensus of the European myeloma network. *Leukemia*. 2021;35(1):31–44.
- Pratt G, Goodyear O, Moss P. Immunodeficiency and immunotherapy in multiple myeloma. *Br J Haematol*. 2007;138(5):563–579.
- Ratta M, Fagnoni F, Curti A, et al. Dendritic cells are functionally defective in multiple myeloma: the role of interleukin-6. *Blood*. 2002;100(1):230–237.
- Suen H, Brown R, Yang S, et al. Multiple myeloma causes clonal T-cell immunosenescence: identification of potential novel targets for promoting tumour immunity and implications for checkpoint blockade. *Leukemia*. 2016;30(8):1716–1724.
- Zelle-Rieser C, Thangavadivel S, Biedermann R, et al. T cells in multiple myeloma display features of exhaustion and senescence at the tumor site. *J Hematol Oncol*. 2016;9(1):116.
- Gavriatopoulou M, Ntanas-Stathopoulos I, Korompoki E, Terpos E, Dimopoulos MA. SARS-CoV-2 vaccines in patients with multiple myeloma. *HemaSphere*. 2021;5(3):e547.
- Terpos E, Trougakos I, Gavriatopoulou M, et al. Low neutralizing antibody responses against SARS-CoV-2 in elderly myeloma patients after the first BNT162b2 vaccine dose. *Blood*. 2021;137(26):3674–3676.
- Kim HJ, Moon BI, Lee JW, Kim SC, Kim HJ. Age-related reduction of antibody response against the human endogenous retrovirus K envelope in women. *Oncotarget*. 2016;7(14):17327–17337.
- Terpos E, Trougakos IP, Apostolou F, et al. Age-dependent and gender-dependent antibody responses against SARS-CoV-2 in health workers and octogenarians after vaccination with the BNT162b2 mRNA vaccine. *Am J Hematol*. 2021;96:E257–E259.
- Bird S, Panopoulou A, Shea RL, et al. Response to first vaccination against SARS-CoV-2 in patients with multiple myeloma. *Lancet Haematol*. 2021;8:e389–e392.
- Monin-Aldama L, Laing AG, Muñoz-Ruiz M, et al. Interim results of the safety and immune-efficacy of 1 versus 2 doses of COVID-19 vaccine BNT162b2 for cancer patients in the context of the UK vaccine priority guidelines. *medRxiv: The Preprint Server for Health Sciences*. 2021; 2021.03.17.21253131. <https://doi.org/10.1101/2021.03.17.21253131>
- Reynolds CJ, Swadling L, Gibbons JM, et al. Discordant neutralizing antibody and T cell responses in asymptomatic and mild SARS-CoV-2 infection. *Sci Immunol*. 2020;5(54):eabf3698.
- Yu J, Tostanoski LH, Peter L, et al. DNA vaccine protection against SARS-CoV-2 in rhesus macaques. *Science (New York, NY)*. 2020;369(6505):806–811.
- Predecki M, Clarke C, Brown J, et al. Effect of previous SARS-CoV-2 infection on humoral and T-cell responses to single-dose BNT162b2 vaccine. *Lancet (London, England)*. 2021;397(10280):1178–1181.

16. van Dorp L, Acman M, Richard D, et al. Emergence of genomic diversity and recurrent mutations in SARS-CoV-2. *Infect Genet Evol.* 2020;83: 104351.
17. Madhi SA, Baillie V, Cutland CL, et al. Efficacy of the ChAdOx1 nCoV-19 Covid-19 vaccine against the B.1.351 variant. *The New England Journal of Medicine.* 2021;384(20):1885–1898.
18. Supasa P, Zhou D, Dejnirattisai W, et al. Reduced neutralization of SARS-CoV-2 B.1.1.7 variant by convalescent and vaccine sera. *Cell.* 2021;184(8):2201–2211.e7.
19. Faria NR, Mellan TA, Whittaker C, et al. Genomics and epidemiology of a novel SARS-CoV-2 lineage in Manaus, Brazil. *medRxiv: The Preprint Server for Health Sciences.* 2021;2021.02.26.21252554. <https://doi.org/10.1101/2021.02.26.21252554>
20. Liu Y, Liu J, Xia H, et al. Neutralizing activity of BNT162b2-elicited serum. *N Engl J Med.* 2021;384:1466–1468.
21. <https://www.clinicaltrials.gov/ct2/show/NCT04785144?term=mRNA-1273351&cond=COVID&draw=2&rank=1>. Accessed April 21, 2021.

How to cite this article: Ludwig H, San-Miguel J, Munshi N, et al. Covid-19 vaccination in patients with multiple myeloma: Focus on immune response. *Am J Hematol.* 2021;96:896–900. <https://doi.org/10.1002/ajh.26263>