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Analysis of a Long Non-coding RNA associated Signature to Predict Survival in Patients with Bladder Cancer

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Abstract

Background: The study aimed to find a potential long non-coding RNA (lncRNA) model related to survival in bladder cancer by analyzing data in The Cancer Genome Atlas (TCGA).

Methods: We downloaded the gene expression data from the TCGA and analyzed the differentially expressed lncRNAs (DELs) between tumor and normal tissues. Patients were divided into training and testing groups, and a prognostic risk score model with lncRNAs was constructed by using data in the training group using multivariate Cox and lasso regression analysis. We divided patients into high-and low-risk groups according to the median value in the lncRNA signature model. Survival and receiver operating characteristic (ROC) curves were visualized in both groups. Further, we validated the model in the testing group.

Results: We screened 169 DELs for bladder cancer. The univariate Cox regression analysis showed that 13 lncRNAs were associated with prognosis with a *p*-value <0.01. We selected 12 of these lncRNAs to perform a multivariate Cox analysis to build the lncRNA signature. The formula with nine lncRNAs, namely, MIR497HG, LINC00968, NALCN-AS1, LINC02321, RNF144A-AS1, MNX1-AS1, FLJ22447, LINC01956, FLJ42969, was significantly related to prognosis. Patients in the high-risk group had a lower survival rate compared with the low-risk group in the training and testing sets (both p-values < 0.05) and the area of the ROC curve was 0.737 and 0.68, respectively.

Conclusions: The study illustrated a significant lncRNA signature and indicated the risk score Cox model could be an important biomarker to predict the prognosis of bladder cancer.

Categories: Genetics, Urology, Oncology

Keywords: predict, the cancer genome atlas, bladder cancer, survival, long non-coding rna

Introduction

Bladder cancer is one of the common types of malignant tumors, causing heavy social and economic burdens to the public. The incidence of bladder cancer ranks 10th among tumors and 6th among men, and it has the 9th highest mortality rate in the world [1]. With the aging of the population, changes in living habits and the environment and the popularization of cancer screening, the incidence and mortality rates of bladder cancer show a relatively stable status year by year in China [2]. Bladder cancer still exhibits the features of high aggressiveness and malignancy, and the majority of patients are usually diagnosed and treated at the advanced disease stage with unfavorable outcomes [3]. In the past decades, treatments for bladder cancer have rapidly improved. However, the overall survival (OS) time of bladder cancer patients has not been significantly extended, especially for patients with muscular invasive bladder cancers, with high malignancy and an extremely low 5-year survival rate of approximately 20-40% after radical surgery [4]. Recently, molecular biomarkers with prognostic and predictive values in bladder cancer have been illustrated due to the development of next-generation sequencing and gene expression profiling [5]. Additionally, long noncoding RNAs (lncRNAs) play a crucial role in predicting clinical outcomes [6]. However, a prognostic model based on the expression of lncRNAs has scarcely been studied. Therefore, it is critical to identify a novel IncRNA signature to better predict the prognosis of patients with bladder cancer and to provide potential targeted strategies.

LncRNAs, representing the largest and most diverse class of non-coding transcripts, are transcripts with lengths of more than 200 nucleotides with little or no protein-coding ability [7-8]. They were redundant segments of the genome but were considered useless transcripts in the past. However, in recent years, emerging studies have indicated the importance of lncRNAs in multiple cellular physiological and pathological processes [9-10]. Increasing evidence suggests that lncRNAs inhibit or activate gene expression by interacting with transcription factors or by binding to specific regions of mRNAs [11]. The roles and mechanisms of lncRNAs and their interactions with target genes have attracted the attention of scholars and represent a popular topic in cancer research. Researchers have identified lncRNA signatures in the prognosis of many cancers, including hepatocellular carcinoma and colorectal cancer. For bladder cancer,

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however, there are few published papers focusing on lncRNA signatures associated with prognosis.

In this study, we analyzed data from The Cancer Genome Atlas (TCGA) and aimed to find a potential long non-coding RNA (IncRNA) model related to survival in bladder cancer patients. RNA-seq data of bladder cancer were downloaded from The Cancer Genome Atlas (TCGA) official website to further analyze the characteristics of IncRNAs. We screened the differentially expressed lncRNAs (DELs) between tumor and normal tissues. Furthermore, we divided patients between the training and testing sets and performed Cox analysis to identify potential lncRNAs to construct the lncRNA risk score model to predict prognosis in bladder cancer. By dividing patients into high-and low-risk groups according to the median risk score, survival and receiver operating characteristic (ROC) curves as well as heatmaps were visualized. An identical analysis was repeated in the testing group to validate the Cox formula. In this work, we hope to determine the prognostic lncRNAs that may provide promising value to the prognosis and progression of bladder cancer in future research.

Materials And Methods

TCGA dataset

We downloaded a dataset containing the transcriptome profiles of RNA-seq data with gene expression quantification including 406 patients with information on survival time and vital status from the TCGA database on July 09, 2019. Furthermore, we screened the DELs between normal and tumor tissues in bladder cancer. In addition, we evaluated the correlation between lncRNA expression and OS rates in bladder cancer patients. Patients were divided into the training set (n=205) to construct the lncRNA signature and the testing set (n=201) to verify the lncRNA signature constructed with the multivariate Cox analysis. The flow chart of study is shown in Figure 1.



Identification of DELs

By using the "limma" package in R project (version 3.5.2; https://bioconductor.org/packages/limma), we performed differential expression analysis to identify DELs with the criteria of an adjusted p-value < 0.05 and a $|\log 2$ fold change $(\log 2FC)| > 2$ between tumor and adjacent normal tissues. Moreover, we used the "survival" package in R to further explore the association of lncRNAs with OS.

Selection of IncRNAs related to prognosis in bladder cancer

We divided patients into the training set (n=204) and the testing set (n=201) by using the R project. We performed a univariate Cox regression analysis to discover the DELs that were significantly related to OS with a p-value <0.01. Thirteen lncRNAs were selected as a result. Next, we applied lasso regression analysis to prevent overfitting of the model and to identify the most valuable lncRNAs to perform the multivariate Cox analysis. Finally, 12 lncRNAs were screened. A multivariate Cox analysis was conducted to construct the lncRNA signature by using the 12 lncRNAs found in the lasso regression model. Ultimately, a nine-lncRNA signature was identified.

Establishment of the risk score of the IncRNA signature

In the present study, the risk score model was constructed as follows: risk score = expression of DEL1 × β 1 DEL1 + expression of DEL2 × β 2 DEL2 +...expression of DELn × β n DELn. After the lncRNA prognostic model was found, we divided patients into high-and low-risk groups according to the median lncRNA expressionbased risk score. The survival rate in both groups was compared with the Kaplan-Meier survival analysis. Then, the receiver operating characteristic (ROC) curve was plotted. To reveal the sensitivity and specificity of the risk score model to predict the prognosis of patients with bladder cancer, the area under the curve (AUC) of the ROC curve was also calculated in the R project.

Validation of the IncRNA signature

To further determine the risk score model's independent impact on the prognosis of bladder cancer, risk score distribution, patient vital status, and a heatmap of these nine lncRNA profiles were further examined using the R project. The testing set data was applied to verify the prognostic value of the nine-lncRNA signature model.

Results

Screening of DELs

In this study, we included 406 samples with complete survival information. With the threshold definition of an adjusted p-value < 0.05 and a $|\log 2$ fold change $(\log 2FC)| > 2$, we eventually screened 169 DELs between the bladder cancer tumor and normal tissues. As a result, 116 lncRNAs were overexpressed in tumor tissues, while only approximately one-third of the lncRNAs were downregulated, as shown in the volcano plot (Figure 2).



FIGURE 2: Volcano plot of the differentially expressed IncRNAs between tumor and normal tissues in bladder cancer

The red dots indicate high-expression lncRNAs and the green ones indicate low- expression lncRNAs. The red ones demonstrate the lncRNA expression with both the log2FC > 2 and adjusted p-value < 0.05. FC: fold change; FDR: false discovery rate.

And the top nine lncRNAs most relevant to survival prognosis are visualized in Figure 3.



FIGURE 3: Survival curve of the top nine differentially expressed IncRNAs in bladder cancer

(A) The relationship of ADAMTS9.AS1 expression with overall survival (B) The relationship of ADAMTS9.AS2 expression with overall survival (C) The relationship of CARMN expression overall survival (D) The relationship of ELDR expression with overall survival (E) The relationship of GAS6.DT expression with overall survival (F) The relationship of LINC00702 expression with overall survival (G) The relationship of MIR99AHG expression with overall survival (H) The relationship of MIR90AHG expression with overall survival (H) The relationship of MIR100HG expression with overall survival (I) The relationship of PTPRD.AS1 expression with overall survival.

Construction of a IncRNA-related prognostic signature

Thirteen lncRNAs in the univariate Cox regression analysis were significantly related to OS, with a p-value <0.01 in bladder cancer patients, as shown in Table 1.

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Gene	HR	HR.95L	HR.95H	p value
FLJ42969	1.005	1.002	1.007	9.15E-05
RNF144A-AS1	1.003	1.002	1.005	0.000
LINC00968	1.007	1.004	1.011	0.000
FLJ22447	1.000	1.000	1.001	0.000
LINC01956	1.011	1.004	1.018	0.001
LINC02321	1.022	1.008	1.035	0.001
GAS1RR	1.009	1.003	1.014	0.001
MIR100HG	1.000	1.000	1.001	0.003
NALCN-AS1	1.014	1.004	1.023	0.004
MNX1-AS1	1.002	1.001	1.004	0.004
LINC02525	1.004	1.001	1.007	0.005
MIR497HG	1.006	1.002	1.010	0.007
LINC02544	1.001	1.000	1.002	0.007

TABLE 1: Univariate cox analysis of 13 IncRNAs with p-value <0.01

Lasso regression analysis was performed to identify the optimal lncRNAs to construct the lncRNA signature. Finally, 12 lncRNAs were screened in the lasso regression model (Figure *4A*). Then, a multivariable Cox analysis was applied to identify the prognostic lncRNAs among the 12 candidate lncRNAs. Importantly, nine-lncRNA signatures, including MIR497HG, LINC00968, NALCN-AS1, LINC02321, RNF144A-AS1, MNX1-AS1, FLJ22447, LINC01956, and FLJ42969, was identified to be significantly associated with prognosis based on the Cox model, as shown in Figure *4B*. The risk score model composed of the nine lncRNAs was constructed as follows: risk score = (0.0050 × expression level of MIR497HG) + (0.0051 × expression level of LINC00968) + (0.0129 × expression level of NALCN-AS1) + (0.0141 × expression level of LINC02321) + (0.0030 × expression level of RNF144A-AS1) + (0.0023 × expression level of MNX1-AS1) + (0.0005 × expression level of FLJ22447) + (0.0106 × expression level of LINC01956) + (0.0059 × expression level of FLJ42969). The nomogram (Figure *4C*) was drawn to show the risk score value along with other clinical parameters to predict the survival rate in bladder cancer.



FIGURE 4: The Cox analysis results and survival nomogram

(A) Lasso regression analysis of IncRNAs in univariate cox analysis. (B) The forest plot of the prognosis-related IncRNAs in the multivariate cox result. (C) Survival nomogram based on the risk score value of 369 patients with age, gender, stage, TNM stage parameters.

TNM is a standard for classifying the extent of the spread of cancer. T describes the size of the original (primary) tumor and if it has invaded nearby tissue; N is to describe nearby (regional) lymph nodes that are involved; M describes distant metastasis (spread of cancer from one part of the body to another).

IncRNA	coef	HR	HR.95L	HR.95H	p-value
MIR497HG	0.0050	1.0050	0.9987	1.0113	0.1194
LINC00968	0.0051	1.0051	0.9987	1.0115	0.1178
NALCN-AS1	0.0129	1.0129	1.0031	1.0229	0.0098
LINC02321	0.0141	1.0142	0.9996	1.0290	0.0558
RNF144A-AS1	0.0030	1.0030	1.0010	1.0050	0.0026
MNX1-AS1	0.0023	1.0023	1.0007	1.0040	0.0046
FLJ22447	0.0005	1.0005	1.0003	1.0007	1.48E-05
LINC01956	0.0106	1.0106	1.0036	1.0177	0.0032
FLJ42969	0.0059	1.0058	1.0032	1.0084	1.29E-05

As the coefficient of the lncRNA derived from the multivariable Cox model is demonstrated in Table 2,

TABLE 2: The multivariate cox analysis of nine long non-coding RNAs associated with overallsurvival of bladder cancer patients

Furthermore, the relative expression levels of these nine lncRNA between tumor and normal tissues were

compared, as shown in Figure 5.



FIGURE 5: The expression level between tumor and normal tissues of the nine IncRNAs in the IncRNA signature of bladder cancer

(A) The expression level of LINC00968 between tumor and normal tissues in bladder cancer (B) The expression level of MIR497HG between tumor and normal tissues in bladder cancer (C) The expression level of NALCN-AS1 between tumor and normal tissues in bladder cancer (D) The expression level of LINC02321 between tumor and normal tissues in bladder cancer (E) The expression level of RNF144A-AS1 between tumor and normal tissues in bladder cancer (G) The expression level of RNF144A-AS1 between tumor and normal tissues in bladder cancer (G) The expression level of RNS1-AS1 between tumor and normal tissues in bladder cancer (G) The expression level of FLJ22447 between tumor and normal tissues in bladder cancer (I) The expression level of LINC01956 between tumor and normal tissues in bladder cancer (I) The expression level of FLJ42969 between tumor and normal tissues in bladder cancer

As a result, the nine lncRNAs were differentially expressed, with a p-value < 0.05. By using the score model, we calculated the risk score for each patient in the training group and divided the training group into a high-risk group (n = 102) and a low-risk group (n = 102) according to the median risk score. In the training set, the high-risk group had a lower survival rate compared with the low-risk group, with approximate 3-year and 5-year survival rates of 34.7% and 22.6%, respectively, in the high-risk group, which were lower than those in the low-risk group (61.0% and 50.9%, respectively) (p-value = 9.79e-05, Figure *6A*). In addition, the area of the ROC curve for the nine-lncRNA signature in the training set was 0.737, which indicated a good performance (Figure *6B*). The patients' risk scores ranged considerably (Figure *6C*), and the survival status is described on the dot plot (Figure *6D*). From the information in the plots, we can tell that patients with a high-risk group. The heatmap of these nine lncRNAs shown in Figure *6E* revealed that MNX1-AS1, RNF144A-AS1, MIR497HG, and FLJ22447 were upregulated in the high-risk group with positive coefficients, while the other lncRNAs seemed to have similar but less obvious trends.



FIGURE 6: The overall survival prediction of the nine long non-coding RNA signature-based risk score in bladder cancer in the training set (n = 00.0

= 204)

(A): Kaplan-Meier analysis of patients' overall survival in the high-risk and low-risk subgroups; (B): The receiver operating characteristic (ROC) analysis of the risk score to predict the overall survival. (C): The risk score distribution based on the nine-IncRNA signature; (D): patient survival status and duration based on the nine-IncRNA signature; (D): patient survival status and duration based on the nine-IncRNA signature; (D): patient survival status and the risk and low-risk subgroups in the training set. The red color in figure 6D means dead status, the green one means alive status. The color trend from red to blue in figure 6E demonstrates a trend from low expression to high expression.

Verification of the IncRNA signature in the testing set

To verify the prognostic value of the nine-lncRNA signature, data in the testing set were used. First, we calculated the risk scores of patients in the test group according to the formula based on the expression of the nine lncRNAs and divided the test group into high-risk and low-risk groups using the same cutoff value as in the training group. As the results demonstrated, the survival rate in the low-risk group was much higher than that in the high-risk group (Figure 7*A*, p = 2.183e-02), similar to the result in the training set. The 3-year and 5-year survival rates were approximately 41.8% and 35.7%, respectively, in the high-risk group, which were lower than those in the low-risk group. The AUC for the nine-lncRNA-based risk score in the testing group was 0.68 (Figure 7*B*). Similar trend results were obtained in the risk score distribution (Figure 7*C*), patient vital status (Figure 7*D*), and heatmap of the nine lncRNA profiles (Figure 7*E*). The results in the testing set illustrated that the nine-lncRNA signature could reasonably reflect the prognosis of bladder cancer.



FIGURE 7: The overall survival prediction of the nine-long non-coding RNA signature-based risk score in bladder cancer in the testing set (n = 201)

(A): Kaplan-Meier analysis of patients' overall survival in the high-risk and low-risk subgroups; (B): The receiver operating characteristic (ROC) analysis of the risk score to predict the overall survival. (C): The risk score distribution based on the nine-IncRNA signature; (D): patient survival status and duration based on the nine-IncRNA signature; (D): patient survival status and duration based on the nine-IncRNA signature risk score; (E): Heatmap of the nine-IncRNA expression profiles in the high risk and low-risk subgroups in the training set. The red color in figure 7D means dead status, the green one means alive status. The color trend from red to blue in figure 7E demonstrates a trend from low expression to high expression.

Discussion

Bladder cancer is one of the most common and significant cancers with multiple potential risk factors, including smoking and chemical exposure [12]. Accumulating investigations have been carried out to explore the pathogenesis and aggravation of bladder cancer [13-15]. An increasing number of studies have revealed that long non-coding RNAs play a crucial role in the carcinogenesis and progression of bladder cancer through various mechanisms [16-20]. However, the specific regulation of lncRNAs and a related prognosis model in bladder cancer are still far from clear. The identification of a lncRNA-associated signature will help to predict the prognosis of bladder cancer and facilitate the development of novel therapeutic targets in further research.

In this study, we measured data in the TCGA to identify DELs and constructed a lncRNA-based signature related to prognosis in bladder cancer. Through an integrative analysis with bioinformatics methods, we identified a nine-lncRNA signature (including MIR497HG, LINC00968, NALCN-AS1, LINC02321, RNF144A-AS1, MNX1-AS1, FLJ22447, LINC01956, and FLJ42969) to predict the OS and prognosis of bladder cancer patients. In contrast to other diseases, there is limited research about the roles of these nine lncRNAs in

bladder cancer. Several studies have found that the long noncoding RNA LINC00968 promoted cell proliferation and migration in different cancers through different mechanisms [21-24] and accelerated the progression of coronary artery disease [25] and diabetic nephropathy [26]. These findings suggest that LINC00968 is probably a positive risk factor in several diseases, which contradicts our result showing that LINC00968 was downregulated in tumor tissues. MIR497HG was identified as a hub lncRNA in ischemic stroke [27]. The lncRNA RNF144A-AS1 was reported as a prognostic factor of chemoresistance in ovarian cancer, while its expression level was downregulated, contradicting its expression in bladder cancer [28]. Additionally, RNF144A-AS1 was illustrated as a positive gene related to Alzheimer's disease [29]. Ding et al. reported that FLJ22447 was remarkably upregulated in stromal carcinoma-related fibroblasts [30]. However, more studies need to be performed to explain the specific roles of these nine lncRNAs.

As the lncRNA signature was verified in the TCGA dataset, it will probably provide efficient values in practice. Nevertheless, there are limitations to the present work. First, it was a bioinformatic study with a limited sample size that may influence the lncRNA signature results. Second, experimental confirmation, including cell lines and animal studies, has been lacking thus far. There are limited publications exploring the unique mechanism of each lncRNA of the nine lncRNAs. We propose further research on the potential mechanism of each lncRNA in the model and the use of clinical samples to verify the accuracy of the lncRNA signature.

Conclusions

We analyzed the TCGA dataset with bioinformatics methods including differential analysis, lasso and Cox regression analysis. The present study illustrated a valuable lncRNA signature to predict the prognosis of bladder cancer and the risk score based on the signature could be an important biomarker for future studies. Meanwhile, further research on the potential mechanism of each lncRNA in the model is necessary and clinical samples would be used to verify the accuracy of lncRNA signatures next.

Additional Information

Disclosures

Human subjects: All authors have confirmed that this study did not involve human participants or tissue. Animal subjects: All authors have confirmed that this study did not involve animal subjects or tissue. Conflicts of interest: In compliance with the ICMJE uniform disclosure form, all authors declare the following: Payment/services info: All authors have declared that no financial support was received from any organization for the submitted work. Financial relationships: All authors have declared that they have no financial relationships at present or within the previous three years with any organizations that might have an interest in the submitted work. Other relationships: All authors have declared that there are no other relationships or activities that could appear to have influenced the submitted work.

References

- Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A: Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin. 2018, 68:394-424. 10.3322/caac.21492
- Chen W, Zheng R, Baade PD, et al.: Cancer statistics in China, 2015. CA Cancer J Clin. 2016, 66:115-32. 10.3322/caac.21338
- Fonteyne V, Ost P, Bellmunt J, et al.: Curative treatment for muscle invasive bladder cancer in elderly patients: a systematic review. Eur Urol. 2018, 73:40-50. 10.1016/j.eururo.2017.03.019
- 4. Witjes JA: Bladder cancer in 2015: improving indication, technique and outcome of radical cystectomy . Nat Rev Urol. 2016, 13:74-6. 10.1038/nrurol.2015.272
- Miyamoto DT, Mouw KW, Feng FY, Shipley WU, Efstathiou JA: Molecular biomarkers in bladder preservation therapy for muscle-invasive bladder cancer. Lancet Oncol. 2018, 19:e683-95. 10.1016/S1470-2045(18)30693-4
- Zhu W, Liu H, Wang X, Lu J, Yang W: Long noncoding RNAs in bladder cancer prognosis: a meta-analysis. Pathol Res Pract. 2019, 215:152429. 10.1016/j.prp.2019.04.021
- Schmitt AM, Chang HY: Long noncoding RNAs: at the intersection of cancer and chromatin biology. Cold Spring Harb Perspect Med. 2017, 7:10.1101/cshperspect.a026492
- Arun G, Diermeier SD, Spector DL: Therapeutic targeting of long non-coding RNAs in cancer . Trends Mol Med. 2018, 24:257-77. 10.1016/j.molmed.2018.01.001
- Mercer TR, Dinger ME, Mattick JS: Long non-coding RNAs: insights into functions. Nat Rev Genet. 2009, 10:155-9. 10.1038/nrg2521
- Moran VA, Perera RJ, Khalil AM: Emerging functional and mechanistic paradigms of mammalian long noncoding RNAs. Nucleic Acids Res. 2012, 40:6391-400. 10.1093/nar/gks296
- 11. Mercer TR, Mattick JS: Structure and function of long noncoding RNAs in epigenetic regulation . Nat Struct Mol Biol. 2013, 20:300-7. 10.1038/nsmb.2480
- 12. Cumberbatch MG, Jubber I, Black PC, et al.: Epidemiology of bladder cancer: a systematic review and contemporary update of risk factors in 2018. Eur Urol. 2018, 74:784-95. 10.1016/j.eururo.2018.09.001
- 13. Casadevall D, Kilian AY, Bellmunt J: The prognostic role of epigenetic dysregulation in bladder cancer: a systematic review. Cancer Treat Rev. 2017, 61:82-93. 10.1016/j.ctrv.2017.10.004
- 14. Cormio A, Sanguedolce F, Musicco C, et al.: Mitochondrial dysfunctions in bladder cancer: exploring their role as disease markers and potential therapeutic targets. Crit Rev Oncol Hematol. 2017, 117:67-72.

10.1016/j.critrevonc.2017.07.001

- Robertson AG, Kim J, Al-Ahmadie H, et al.: Comprehensive molecular characterization of muscle-invasive bladder cancer. Cell. 2017, 171:540-556.e25. 10.1016/j.cell.2017.09.007
- Heubach J, Monsior J, Deenen R, et al.: The long noncoding RNA HOTAIR has tissue and cell typedependent effects on HOX gene expression and phenotype of urothelial cancer cells. Mol Cancer. 2015, 14:108. 10.1186/s12943-015-0371-8
- 17. Hedegaard J, Lamy P, Nordentoft I, et al.: Comprehensive transcriptional analysis of early-stage urothelial carcinoma. Cancer Cell. 2016, 30:27-42. 10.1016/j.ccell.2016.05.004
- Avgeris M, Tsilimantou A, Levis PK, et al.: Loss of GAS5 tumour suppressor lncRNA: an independent molecular cancer biomarker for short-term relapse and progression in bladder cancer patients. Br J Cancer. 2018, 119:1477-86. 10.1038/s41416-018-0320-6
- 19. Gulia C, Baldassarra S, Signore F, et al.: Role of non-coding RNAs in the etiology of bladder cancer . Genes (Basel). 2017, 8:10.3390/genes8110339
- Li HJ, Gong X, Li ZK, et al.: Role of long non-coding RNAs on bladder cancer . Front Cell Dev Biol. 2021, 9:672679. 10.3389/fcell.2021.672679
- Sun X, Huang T, Zhang C, Zhang S, Wang Y, Zhang Q, Liu Z: Long non-coding RNA LINC00968 reduces cell proliferation and migration and angiogenesis in breast cancer through up-regulation of PROX1 by reducing hsa-miR-423-5p. Cell Cycle. 2019, 18:1908-24. 10.1080/15384101.2019.1632641
- Yao N, Sun JQ, Yu L, Ma L, Guo BQ: LINC00968 accelerates the progression of epithelial ovarian cancer via mediating the cell cycle progression. Eur Rev Med Pharmacol Sci. 2019, 23:4642-9. 10.26355/eurrev 201906 18043
- Liu G, Yuan D, Sun P, Liu W, Wu PF, Liu H, Yu GY: LINC00968 functions as an oncogene in osteosarcoma by activating the PI3K/AKT/mTOR signaling. J Cell Physiol. 2018, 233:8639-47. 10.1002/jcp.26624
- Wang Y, Zhou J, Xu YJ, Hu HB: Long non-coding RNA LINC00968 acts as oncogene in NSCLC by activating the Wnt signaling pathway. J Cell Physiol. 2018, 233:3397-406. 10.1002/jcp.26186
- Wang X, Zhao Z, Zhang W, Wang Y: Long noncoding RNA LINC00968 promotes endothelial cell proliferation and migration via regulating miR-9-3p expression. J Cell Biochem. 2018, 10.1002/jcb.28103
- Li Z, Yu Z, Meng X, Yu P: LncRNA LINC00968 accelerates the proliferation and fibrosis of diabetic nephropathy by epigenetically repressing p21 via recruiting EZH2. Biochem Biophys Res Commun. 2018, 504:499-504. 10.1016/j.bbrc.2018.08.048
- 27. Zheng Y, Sun S, Yu M, Fu X: Identification of potential hub-lncRNAs in ischemic stroke based on Subpathway-LNCE method. J Cell Biochem. 2019, 120:12832-42. 10.1002/jcb.28554
- Song J, Zhang W, Wang S, Liu K, Song F, Ran L: A panel of 7 prognosis-related long non-coding RNAs to improve platinum-based chemoresistance prediction in ovarian cancer. Int J Oncol. 2018, 53:866-76. 10.3892/ijo.2018.4403
- Cao M, Li H, Zhao J, Cui J, Hu G: Identification of age- and gender-associated long noncoding RNAs in the human brain with Alzheimer's disease. Neurobiol Aging. 2019, 81:116-26. 10.1016/j.neurobiolaging.2019.05.023
- Ding L, Ren J, Zhang D, et al.: A novel stromal lncRNA signature reprograms fibroblasts to promote the growth of oral squamous cell carcinoma via LncRNA-CAF/interleukin-33. Carcinogenesis. 2018, 39:397-406. 10.1093/carcin/bgy006