

# **Process Condition Optimization and Structural Feature Analysis of Humic Acid Extraction from Weathered Lignite**

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ABSTRACT: In this study, response surface methodology (RSM) was adopted to investigate the optimization of process conditions for extracting humic acid (HA) from coal, aiming to enhance the yield of humic acid. Additionally, UV−vis spectroscopy, FTIR, XRD, TG-DTG, CP/MAS13CNMR, XPS, and molecular fluorescence were utilized to examine the properties of HAs. The extraction time significantly influenced the yield from Lishi weathered lignite, while the liquid− solid ratio had a significant impact on the yield from Wuhai weathered lignite. The interactive effect between factors did not have a significant effect on the yield. The optimal extraction conditions for Lishi humic acid (LSHA) were determined to be an extraction time of 4.4 h, NaOH concentration of 0.30 mol/L, and liquid−solid ratio of 21 mL/g, while those for Wuhai humic acid (WHHA) were 3.1 h, 0.21 mol/L, and 12



mL/g, respectively. Under these optimal conditions, the true yield values closely matched the predicted value obtained from the model optimization. Comparative analysis of the HAs revealed similarities in their chemical properties, including the degree of aromaticity, molecular weight, and distribution of functional groups. The aromaticity of WHHA was higher compared to that of LSHA. The higher hydrophilic−hydrophobic index of LSHA contributed to its relatively high biological activity compared to that of WHHA. Both humic acids belong to terrestrial humic acids. The results of the study provide a reference for further application of humic acid.

# **1. INTRODUCTION**

Humic acid (HA) is a mixture of macromolecular organic compounds naturally formed through the decomposition and transformation of plants by microorganisms, followed by long-term geochemical processes.<sup>[1](#page-12-0)</sup> This compound possesses numerous reactive functional groups, such as carboxyl and phenolic hydroxyl groups, which possess excellent physiological activity and functions including absorption, complex-ation, and ion exchange.<sup>[2](#page-12-0),[3](#page-12-0)</sup> Additionally, these unique characteristics facilitate its widespread applications in agriculture, environmental protection, medicine, and industry. $4-7$  $4-7$ Low-grade coals, such as peat, lignite, and weathered coal, have low utilization value as fuels and are currently the main raw materials for the production of humic acid. Weathered lignite typically contains 30−80% humic acid, with the specific content depending on the original characteristics of the coal and the conditions of the weathering process.<sup>[8](#page-12-0)</sup> However, the extraction of HA from natural sources, particularly from coal, presents challenges due to low extraction rates, high energy consumption, and limited economic benefits. Therefore, it is crucial to optimize the extraction process of this compound to ensure the maximum yield and purity. Additionally, further exploration and enhancement of its potential applications in various fields are essential to harness its full potential.

In this context, response surface methodology (RSM) offers a promising approach for modeling and optimizing the extraction process. The RSM provides a systematic framework for studying the interaction between multiple factors and optimizing the extraction conditions, thus minimizing the number of experimental runs and enhancing the efficiency of the process.<sup>[9](#page-12-0),[10](#page-12-0)</sup> Ehsan Sarlaki et al. employed a three-level, three-factor central composite design to model and optimize the extraction process of humic acid from lignite waste, achieving the highest extraction efficiency of 54.2% for HA during their study. $11$ 

Currently, the alkali-dissolution and acid-precipitation methods are primarily used to extract humic acid from organic substances such as coal. This method leverages the fact that humic acid is soluble in alkali but insoluble in acid. The humic acid is initially dissolved in a NaOH/KOH solution and then precipitated out using a HCl solution.<sup>11-[14](#page-12-0)</sup> This study is based

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<span id="page-1-0"></span>on an alkaline extraction method and aims to apply RSM to investigate the extraction of HA from weathered lignite in Inner Mongolia Wuhai (WH) and Shanxi Lishi (LS), with a focus on understanding the influence of various factors on the yield of HAs and optimizing the extraction conditions. The composition and properties of extracted HA were thoroughly analyzed. Comprehensive characterization techniques including elemental analysis, UV−vis spectroscopy, FTIR, XRD, TG-DTG, CP/MAS<sup>13</sup>CNMR, XPS, and molecular fluorescence were employed for this purpose. This study will provide valuable insights into the industrial production and further application of HA.

# **2. EXPERIMENTAL SECTION**

**2.1. Materials.** Coal samples were acquired from leonardite in Wuhai (WH) of Inner Mongolia and Lishi (LS) of Shanxi Province, China. The raw coal samples were finely milled to a grain size of less than 0.2 mm. Sodium hydroxide (NaOH) was purchased from Tianjin Fengchuan Chemical Reagent Co., Ltd., and hydrochloric acid (HCl, 37%) was purchased from Chongqing Chuandong Chemical (Group) Co., Ltd. All reagents used were of analytical grade.

**2.2. Total Humic Acid in Coal.** The total humic acid content in the raw coal was determined by the volumetric method.<sup>15</sup> A sodium pyrophosphate alkaline solution was used to extract humic acid from the coal sample, and then, the carbon in humic acid was oxidized to carbon dioxide by potassium dichromate in a strongly acidic solution. The content of humic acid in the raw coal sample was calculated based on the consumption of potassium dichromate and the carbon content of humic acid.

**2.3. HA Extraction from the Coal Sample.** In this study, humic acids were extracted from coal using the alkalidissolution and acid-precipitation method. The coal samples and NaOH solution were added into a conical flask at a certain liquid−solid ratio (the ratio of alkaline solution volume to coal sample mass,  $mL/g$ ). The conical flask was reacted for a variable time, ranging from 1 to 6 h at room temperature (298 K, taking into account the issue of production costs in the actual industrial production process). After centrifugation (7000 rpm, 5 min) and suction filtration, the pH of the sample was modulated to less than 2 by HCl and left to stand for 24 h. The precipitate of HA was separated by centrifugation and then dialyzed. Finally, the HA sample was dried by using a freeze-dryer.

**2.4. RSM Optimization Experiment.** Based on the singlefactor experimental results, the effects of the time, NaOH solution concentration, and liquid−solid ratio on extraction efficiency of HA (%) were investigated using a Box−Behnk design. Design-Expert was for developing three-factor and three-level experiments. A total of 17 test points, 12 analysis points, and 5 zeros were designed to assess the error (Table 1 and Table 2). Three coded ranges—low  $(-1)$ , center  $(0)$ , and high  $(1)$ —were nominated from the subsequent preliminary experimentation [\(Table](https://pubs.acs.org/doi/suppl/10.1021/acsomega.4c01840/suppl_file/ao4c01840_si_001.pdf) S1). To assess the significance of each regression coefficient on HA extraction efficiency, an analysis of variance (ANOVA) was carried out. The optimum conditions to simulate the humic acid extraction process were used as the response surface model. A quadratic equation (eq 1) was employed for the approximation of resulting HA percentage extraction.

Table 1. RSM-BBD Design and Results of LSHA*<sup>a</sup>*

	LSHA							
		Factor		HA Extraction (%)				
No.	A	B	C	Actual Value	Predictive Value			
$\mathbf{1}$	$\overline{4}$	0.30	20	51.9	52.3			
$\overline{2}$	5	0.25	20	50.9	50.9			
3	3	0.30	16	48.2	47.9			
$\overline{4}$	$\overline{4}$	0.25	16	49.7	49.6			
5	5	0.30	24	51.1	51.3			
6	$\overline{4}$	0.25	24	50.9	50.7			
7	$\overline{4}$	0.30	20	52.7	52.3			
8	4	0.30	20	52.2	52.3			
9	5	0.30	16	50.7	50.8			
10	$\overline{4}$	0.35	16	49.9	50.1			
11	$\overline{4}$	0.35	24	50.5	50.6			
12	$\overline{4}$	0.30	20	52.3	52.3			
13	5	0.35	20	51.1	50.8			
14	3	0.30	24	49.1	48.9			
15	3	0.25	20	47.7	47.9			
16	3	0.35	20	48.4	48.4			
17	$\overline{4}$	0.30	20	52.7	52.3			

 ${}^a$ A is the time, h; B is the NaOH solution concentration, mol/L; C is the liquid−solid ratio, mL/g.

Table 2. RSM-BBD Design and Results of WHHA*<sup>a</sup>*

	<b>WHHA</b>								
		Factor		HA Extraction (%)					
No.	A	B	$\mathsf{C}$	Actual Value	Predictive value				
$\mathbf{1}$	3	0.25	5	60.8	60.4				
2	3	0.20	10	69.4	68.9				
3	2	0.20	15	63.0	63.5				
$\overline{4}$	3	0.25	15	67.3	66.8				
5	$\overline{4}$	0.15	10	64.1	63.9				
6	3	0.20	10	68.5	68.9				
7	4	0.20	15	65.1	64.8				
8	$\overline{4}$	0.25	10	64.1	64.9				
9	3	0.20	10	69.3	68.9				
10	$\overline{2}$	0.25	10	62.9	63.0				
11	3	0.15	5	56.9	57.5				
12	3	0.15	15	65.2	65.5				
13	3	0.20	10	68.7	68.9				
14	4	0.20	5	59.8	59.3				
15	3	0.20	10	68.9	68.9				
16	2	0.20	5	54.3	54.6				
17	2	0.15	10	60.7	59.9				

 ${}^a$ A is the time, h; B is the NaOH solution concentration, mol/L; C is the liquid−solid ratio, mL/g.

$$
Y = d_{o} + \sum_{i=1}^{k} d_{i}X_{i} + \sum_{i=1}^{k} d_{ii}X_{i}^{2} + \sum_{i=1}^{k} \sum_{j=i+1}^{k} d_{ij}X_{i}X_{j}
$$
(1)

where *Y* is response variable (HA extraction efficiency),  $d_0$  is the intercept value,  $d_i$  ( $i = 1, 2, ..., n$ ) is the first-order model coefficients for  $X_i$ ,  $d_{ij}$  is the interaction coefficients for  $X_i X_j$ , and  $d_{ii}$  represents the quadratic coefficients of  $X_i$ .

**2.5. Chemical and Spectral Features Analysis.** The proximate analysis was subjected to testing with reference to  $GB/T$  212-2008.<sup>[16](#page-12-0)</sup> The moisture content was measured by air drying in a blast drying oven. The ash and volatile yields of the samples were automatically determined by an intelligent muffle

<span id="page-2-0"></span>Table 3. ANOVA Analysis of Response Surface Model

			<b>LSHA</b>						<b>WHHA</b>			
Source	Sum of Squares	<b>D</b> fES	Mean Square F value		P value		Remark Sum of Squares	def.	Mean Square	F value	P value	Remark
Model	38.16	9	4.24	30.64	< 0.0001	с	317.68	9	35.30	71.49	< 0.0001	с
A	13.47	1	13.47	97.32	< 0.0001	с	18.24		18.24	36.94	0.0005	
B	0.063		0.063	0.46	0.5215	a	8.43		8.43	17.06	0.0044	
C	1.21		1.21	8.74	0.0212	b	103.18		103.18	208.97	< 0.0001	с
AB	0.078	1	0.078	0.57	0.4762	a	1.12		1.12	2.28	0.1752	a
AC	0.058	1	0.058	0.42	0.5394	a	2.89		2.89	5.85	0.0461	ь
BC	0.099		0.099	0.72	0.4251	a	0.68		0.68	1.38	0.2788	a
$A^2$	11.42		11.42	82.51	< 0.0001	с	67.16		67.16	136.02	< 0.0001	с
B <sup>2</sup>	5.86		5.86	42.31	0.0003	с	17.03		17.03	34.50	0.0006	с
$C^2$	3.64		3.64	26.27	0.0014	с	81.75		81.75	165.57	< 0.0001	с
Residual	0.97	7	0.14				3.46	7	0.49			
Lack of fit	0.45	3	0.15	1.14	0.4339	$\boldsymbol{a}$	2.85	3	0.95	6.27	0.0524	a
Total	39.13	16					321.14	16				
	<sup>a</sup> Not significant (P > 0.05). <sup>b</sup> Significant (P < 0.05). <sup>c</sup> Extremely significant (P < 0.01).											

furnace with model 5E-MF6100K (Changsha Kaiyuan Instrument Co., Ltd.), and the fixed carbon content could be obtained by using the subtraction method. The elemental compositions (C, H, N, and S) were determined with an elemental analyzer of the Vario macro cube model produced by Elementary, Germany. Oxygen content was calculated by the difference.

The contents of total acid groups and carboxyl groups were determined by the barium ion and calcium ion exchange methods, respectively.<sup>[17](#page-12-0)</sup> The phenolic hydroxyl groups were determined by the difference between the total acid groups and the carboxyl groups. Ultraviolet−visible spectroscopy (UV− vis) analysis was carried out by a UV-1800PC Series (Shanghai Mepoda Instrument Co., Ltd.). For absorbance measurements, the humic acids were dissolved in 100 mL of  $NaHCO<sub>3</sub>$ solution (0.05 mol/L), and the absorbance (Abs) was measured at 400, 465, 600, and 665 nm. The E4/E6 ratios (Abs465/Abs665) and ΔLog *K* coefficients (Log Abs400 − Log Abs<sub>600</sub>) were calculated for characterization of HAs.

FT-IR spectra were recorded on a Fourier transform infrared spectrometer (model Nicolet iS50) using the KBr pellet technique. The spectrum was recorded from 4000 to 500  $cm^{-1}$ at a resolution of 4  $\rm cm^{-1}$ . The thermal analyses (TG-DTG) of the HA samples were carried out in a thermal analyzer (model Netzsch STA 449F3) in an air atmosphere. A heating rate of 10 °C/min was chosen based on previous studies with continuous heating from 25 to 900 °C. Solid-state CP/MAS  $^{13}$ C NMR spectra of the HAs were collected on an AVWBIII600 nuclear magnetic resonance spectroscopy instrument produced by Bruker. The test speed was 14 kHz, and the resonance frequency was 150.9 MHz.

The X-ray diffraction (XRD) spectra were obtained with an X-ray diffractometer (X′Pert 3, Panalytical, Netherlands). Samples were scanned over a 2*θ* range from 5 to 90 with a scanning voltage of 40 kV and target Cu/K $\alpha$  ( $\lambda$  = 1.5406 Å).<sup>18</sup> XPS spectra were measured with a PHI5000 Versaprobe-II spectrometer (50 W, 15 kV). The Al target was selected as the X-ray light source. High-resolution spectra were obtained using 46.95 eV pass energy, and the spectra were corrected by C 1s with the binding energy of 284.8 eV. All fluorescence spectra were run on a Hitachi F-4600 fluorophotometer with a scan speed of 120 nm/min<sup>-1</sup>, using excitation and emission slit bandwidths of 5 mm. Xe was used as the measuring light

source. The scanning speed for three-dimensional fluorescence determination was 12 000 nm/min.

# **3. RESULTS AND DISCUSSION**

**3.1. Optimization of HA Extraction Conditions.** The initial experimental results are shown in [Figure](https://pubs.acs.org/doi/suppl/10.1021/acsomega.4c01840/suppl_file/ao4c01840_si_001.pdf) S1. For LSHA, as the extraction time increases from 2 h to 5 h, the yield shows an upward trend, rising from 32.12% to 40.96%; for WHHA, as the extraction time increases from 1 h to 5 h, the yield shows an upward trend, rising from 62.57% to 66.53% [\(Figure](https://pubs.acs.org/doi/suppl/10.1021/acsomega.4c01840/suppl_file/ao4c01840_si_001.pdf)  $S1(1,4)$ ). However, with the prolongation of extraction time, both the extraction amounts of LSHA and WHHA decrease. This phenomenon can be attributed to the ease of decomposition and degradation of HA under extended extraction conditions.<sup>[19,20](#page-12-0)</sup> Longer time periods correspond to increased energy usage and financial consequences. Thus, a time range of 3−5 h was chosen for additional optimization studies on LSHA extraction and 2−4 h for additional optimization on WHHA extraction. Once the optimal extraction time had been determined, HA extraction was studied using various NaOH concentrations. Figure  $S1(2,5)$ depicts that HA extraction increased significantly with an increase in NaOH concentration (LSHA: 0.15−0.30 mol/L; WHHA: 0.1−0.2 mol/L). Then, when the concentration of NaOH exceeds 0.3 mol/L for LSHA and 0.2 mol/L for WHHA, respectively, the extraction rates of both HA types decrease. Considering these data, NaOH concentrations in the range of 0.25−0.35 and 0.15−0.25 mol/L were carefully chosen for further optimization studies. This could prevent the need for using excessive solvent during the HA extraction process. The liquid−solid ratio presented the same trend as extraction time and NaOH concentration in extraction of two HAs.<sup>[21](#page-12-0)</sup> Within the ranges of 10 to 20 for LSHA and 5 to 10 mL/g for WHHA, respectively, the yields of LSHA and WHHA increase as the liquid−solid ratio increases. Beyond these ranges, the yields of HAs gradually decrease. Based on the current experiment, the liquid−solid ratio in the range of 16−24 mL/g for LSHA and 5−15 mL/g for WHHA was chosen for additional optimization studies (Figure  $S1(3,6)$ ).

After conducting initial screening experiments, the matrix design for the trials is presented in [Table](#page-1-0) 1 and [Table](#page-1-0) 2, using the Box−Behnken design. The analysis reveals a small deviation between the actual values of the two HAs of yields and the predicted values of the model. By comparing the fitting



Figure 1. (a) Normal plot of residuals and (b) residuals of LSHA yield.



Figure 2. (a) Normal plot of residuals and (b) residuals of the WHHA yield.

degrees of the various fitting equations, two quadratic multiple regression equations have been derived:

$$
Y_{LSHA} = -57.85 + 15.91A + 311.75B + 2.78C - 2.80AB
$$
  
- 0.03AC - 0.79BC - 1.65A<sup>2</sup> - 471.70B<sup>2</sup>  
- 0.06C<sup>2</sup>(R<sup>2</sup> = 0.9752; R<sup>2</sup><sub>adj</sub> = 0.9434) (2)

$$
Y_{WHHA} = -47.36 + 29.29A + 390.63B + 5.09C
$$
  
- 10.60AB - 0.17AC - 1.65BC - 3.99A<sup>2</sup> - 804.50B<sup>2</sup>  
- 0.18C<sup>2</sup>(R<sup>2</sup> = 0.9892; R<sup>2</sup><sub>adj</sub> = 0.97540) (3)

[Table](#page-2-0) 3 depicts the results of the analysis of variance, which showed that the *P*-values of the two models were both less than 0.0001, indicating that both models were significant. The *P*-value of the lack-of-fit was relatively low (*P* > 0.05), indicating that the lack-of-fit was not significant. This indicates that two models have a good fitting degree within the respective regression range, comply with the experimental requirement, and could be used for statistical analysis on experimental results.<sup>[22](#page-12-0)</sup> The correlation coefficient  $(R^2)$  and

adjusted correlation coefficient  $(R^2_{adj})$  were both higher than 0.90. The experimental results showed that the model was in good agreement with the predicted results, and the experimental error was low, which can describe the relationship between the factors and their response values well. According to the experimental data, the residual probability distribution diagram and the residual curve of yields are drawn as shown in Figure 1 and Figure 2. The residual distribution was on both sides of the prediction curve, and there was no systematical bias toward over- or underestimating humic acid yields. Therefore, the error could be considered to follow the normal distribution, and the residual curve of yields further verifies the assumption that the error follows this distribution. Based on this analysis, the results have shown that the two models can effectively predict and analyze the extraction process of humic acid from both types of weathered lignite. The statistical significance, excellent fit, high correlation coefficients, low experimental error, and normal distribution of errors all indicate that the two models are highly effective tools for understanding and optimizing the extraction of humic acid from weathered lignite. In practical terms, these results

<span id="page-4-0"></span>

Figure 3. Contour and response surface plots showing the effect of interaction between two factors on HA yield (LSHA: a, b, c; WHHA: d, e, f).

mean that researchers can use the RSM to establish models to predict optimal extraction conditions for maximizing the humic acid yield from weathered lignite.

Figure 3 reflects the impact of the interaction between AB, AC, and BC on the yields of HAs in [Table](#page-2-0) 3. The 3D response

surface can reflect the influence of various factors on the response value of the model. The contour diagram is the projection of three-dimensional graphics. The yield of HA increased and decreased with the increase of extraction time (A), sodium hydroxide solution concentration (B), and

		Proximate analyses (%)			Ultimate analyses (%, daf)					
Sample	Yield $(\%)$	$M_{\rm ad}$	$A_{\rm d}$	$V_{\text{daf}}$	$FC_{\text{daf}}$	C	H	N	S	O
WH	$\overline{\phantom{0}}$	3.6	54.62	45.5	54.5	62.41	4.55	1.21	4.55	27.27
LS	$\overline{\phantom{a}}$	7.79	44.37	51.8	48.2	64.29	3.41	1.46	1.6	29.24
WHHA	52.77	8.74	4.12	$\overline{\phantom{a}}$	$\overline{\phantom{a}}$	62.83	4.19	1.32	0.15	31.51
<b>LSHA</b>	69.52	10.64	5.09	$\overline{\phantom{a}}$	$\sim$	59.04	4.25	1.34	0.22	35.15
${}^aA_d$ is air-dried basis; d is dry basis; daf is dry ash free basis.										

<span id="page-5-0"></span>Table 4. Proximate Analyses and HA Yield*<sup>a</sup>*

liquid−solid ratio (C). As shown in [Figure](#page-4-0) 3(a, b, c), in the process of extracting HA from weathered lignite in Lishi, the results indicate that A exerts a paramount influence on HA yield, corroborating its pivotal role in facilitating the release of HAs from the lignite matrix. Meanwhile, C demonstrates a moderately significant impact, suggesting that optimizing this parameter can further enhance the yield. Conversely, B and the interaction terms AB, AC, and BC had a certain but not significant impact, implying that the studied range adjustments to this parameter may not yield substantial gains in HA yield. As described in [Figure](#page-4-0)  $3(d, e, f)$ , in the process of extracting HA from weathered lignite in Wuhai, C emerges as the most critical factor, emphasizing the importance of solvent quantity in enhancing mass transfer and, consequently, HA dissolution. Additionally, A, B, and interaction item AC significantly contribute to yield, indicating that a synergistic adjustment of these parameters is crucial for maximizing HA recovery. Notably, interaction items AB and BC are found to be insignificant, suggesting that their combined effect does not significantly alter the overall yield trend. The analysis results of the *F* value and *P* value in [Table](#page-2-0) 3 were consistent with this. The significant differences between Lishi and Wuhai in the process of HA extraction indicate that geographical factors have a crucial impact on the extraction efficiency. This may be attributed to the disparities in physicochemical properties such as mineral composition, organic matter content, and particle size distribution of the weathered lignite in the two regions. In summary, the extraction efficiency of humic acid is jointly influenced by various factors, including extraction time, sodium hydroxide solution concentration, and liquid-to-solid ratio, as well as the interactions among these factors. Furthermore, regional differences also constitute a significant factor that cannot be overlooked. To enhance the extraction efficiency of HA, it is recommended to implement customized production based on raw materials sourced from different regions. This can be achieved by optimizing the extraction conditions, particularly the extraction time and liquid-to-solid ratio, and taking into account the interactions among factors to realize efficient extraction.

**3.2. Determination and Verification of Optimum Process Conditions.** The optimization function of Design-Expert10.4.4 software was used to optimize the extraction conditions of LSHA and WHHA with 100% HA yield as the target value. Optimal extraction conditions of LSHA were as follows: the extraction time was 4.4 h, the NaOH concentration was 0.30 mol/L, the liquid/solid ratio was  $20.74$  mL/g, and the maximum yield of LSHA predicted by the model was 52.8%. Integer values for the liquid−solid ratio were taken based on the feasibility of the actual operation, and the optimal conditions were adjusted as follows: the extraction time was 4.4 h, the concentration of NaOH was 0.30 mol/L, and the liquid−solid ratio was 21 mL/g. The average yield of LSHA was 52.7%, and the relative error was 0.13% compared

to the predicted value. Optimal extraction conditions of WHHA were as follows: the extraction time was 3.14 h, the NaOH concentration was 0.21 mol/L, the liquid−solid ratio was 11.93 mL/g, and the maximum yield of WHHA predicted by the model was 69.8%. Considering the feasibility of the actual operation, the liquid−solid ratio is rounded to an integer value, and the optimal conditions were adjusted as follows: extraction time was 3.1 h, NaOH concentration was 0.21 mol/ L, liquid to solid ratio was 12 mL/g, and the average yield of WHHA was 69.5%, with the relative error of 0.5% compared with the predicted value. The results demonstrate that the model had good predictability, was feasible in operation, and had a practical application value. Optimal extraction conditions of LSHA were as follows: extraction time of 4.4 h, NaOH concentration of 0.30 mol/L, and liquid−solid ratio of 21 mL/ g. Optimal extraction conditions of WHHA were as follows: extraction time 3.1 h, NaOH concentration 0.21 mol/L, liquid−solid ratio of 12 mL/g.

**3.3. Characteristics of Two HAs.** The total humic acid contents of the two weathered lignites used in this study were 30.66% and 36.10%, respectively. The proximate and ultimate analyses of coal and HA samples are listed in Table 4 and Table 5. Compared with WH, LS contains a higher moisture





content. According to the classification of coal quality grades, $23,24$  WH belongs to the category of extremely high ash content coal  $(A_d > 40\%)$  and high volatile matter coal  $(37\% \lt V_{\text{daf}} \lt 50\%)$ , while LS belongs to the category of extremely high ash content coal  $(A_d > 40%)$  and extremely high volatile matter coal  $(V_{\text{daf}} > 50\%).$ 

The ash yields of HAs were 4.12% and 5.09%, and the moisture contents were 8.74% and 10.64%. The results reflected that the purity of the two HAs was high. The higher organic matter content in WHHA compared to that in LSHA suggests that the former may possess a more complex molecular structure or a higher concentration of bioactive compounds, which could potentially lead to enhanced functional properties. According to elemental analysis, although the sources of the two HAs were different, the elemental compositions of the HA samples were similar (Table 4). In all samples, the elemental composition of HA has shown a range similar to those described in the literature.<sup>[1,29](#page-12-0)</sup>

# <span id="page-6-0"></span>Table 6. Acidic Functional Groups, UV−vis, and FTIR Parameters of HAs



The H/C and O/C atomic ratios, which are considered to be important indicators of the aromaticity and maturity of HAs, have been used to study the structural changes of humus. $30,31$  $30,31$  The similarity in the H/C ratio between WHHA and WH coal samples indicates that they have comparable maturity levels, suggesting that WHHA has preserved the original chemical structural characteristics of WH coal samples to a good extent. The H/C ratio of LSHA is somewhat higher compared to that of the LS coal sample, which may be attributed to the possible destruction or alteration of some aromatic structures in the coal, resulting in a relative decrease in the carbon content. This phenomenon can be attributed to the difference in extraction conditions. The H/C ratios of the two types of HA were less than 1, indicating that the maturity of these two types of HAs was higher, and the maturity of weathered lignite HAs was higher than that of previously studied lignite HAs.<sup>[32](#page-12-0)</sup> The aromaticity of the samples under study decreases with the H/C ratio. Thus, LSHA had the lowest aromatic degree ([Table](#page-5-0) 5). Compared with the existing research, the aromatic degree of weathered lignite HA was higher than lignite HA.<sup>[32](#page-12-0)</sup> The O/C atomic ratio reflects the number of oxygen-containing groups (e.g., carboxylic acids) in HAs. The O/C ratio of coal HA generally ranges from 0.32 to 0.54, $30,33$  while the O/C ratios of the HAs in this paper were 0.38 and 0.45 ([Table](#page-5-0) 5). The O/C ratio of LSHA was much higher than that of WHHA, which may be due to the high ash content in LSHA. The N/C atomic ratio reflects the content of nitrogen in HA. A higher N/C ratio (usually around 0.05) is characteristic of HA in soil and peat. The N/C ratio of coal HA is generally less than  $0.05$ ,  $31$  and the N/C ratios of the two HAs are within this range. In terms of O/C and N/C, HAs are similar to coal samples.

The polarity index  $((O + N)/C)$  can be used to predict the adsorption behavior of HAs according to the characteristics of the organic phase.<sup>[25](#page-12-0)</sup> The  $(O + N)/C$  values of HAs used in this work are summarized in [Table](#page-5-0) 5. There is almost no difference from those reported in the literature.<sup>[25,26](#page-12-0)</sup> The greater the value of the  $(O + N)/C$  ratio, the greater the polarity. Obviously, compared with WHHA, LSHA is more polar, indicating that the content of the polar structure is higher. The polarity order indicated by the  $(O + N)/C$  value is the same as that of the O/C value, so it is speculated that the polarity of HAs is mainly determined by oxygen-containing functional groups. Internal oxidation degree (*ω*) indicates the degree of the HA oxidation process. The higher its value, the higher the degree of humification. The order of the *ω* value was as follows: LSHA < WHHA. This outcome may indicate that LSHA experienced fewer oxidative reactions or microbial transformation processes during its formation. The *ω* value of weathered lignite is positive, which is the characteristic value under aerobic conditions.<sup>34</sup>

Acid functional groups in HA play an important role in HA application. Table 6 shows the acid functional groups of the two HAs. The contents of total acid groups and carboxyl groups of the HAs are LSHA > WHHA, while the contents of phenolic hydroxyl groups are LSHA > WHHA. The carboxyl

group has a significant contribution to the solubility, buffer capacity, and cation exchange capacity of humic acid. Therefore, LSHA is more attractive in agricultural applications where nutrient retention and remediation are key issues. UV− vis spectroscopy is a nondestructive and simple method that has been used to determine the properties of HA for many years. There is a common understanding that the  $E_4/E_6$  ratio decreases with the increase of condensed ring, humification degree, molecular weight, or size. $35$  Data in Table 6 show that the  $E_4/E_6$  ratio is lower than 6 and increases in the following order: WHHA < LSHA. Thus, it can be inferred that the molecular weight of WHHA is greater than that of LSHA. The above order is almost determined by the ΔLog *K* values, which confirm that the humification degree of sample WHHA is higher than that of LSHA. In addition, the ΔLog *K* values of the two HAs are 0.61 and 0.69, indicating that the two HAs belong to type B humic acid.

In order to better understand the findings, FTIR spectroscopy was performed to confirm the chemical structure of extracted HA from coal samples (Figure 4). The main



Figure 4. Fourier transform infrared spectroscopy (FTIR) spectra of LSHA and WHHA.

absorption positions of the two HAs were roughly similar, but there were significant differences in the relative intensity of certain bands among different entities, which were related to the source and nature of the samples. The valence vibration of the O−H group in the aromatic and aliphatic regions of HA appeared at 3450−3400 cm<sup>−</sup><sup>1</sup> . This confirms the hydrophilicity of HA, which is crucial for its interaction with water and other polar solvents. The broader peak of the O−H stretching vibration may be attributed to the high  $O/C$  and  $H/C$  ratios.<sup>1</sup> The alcoholic and amide N−H nonbonded groups showed a deep absorption band at 3418 cm<sup>-1</sup>, confirming the presence of these groups.[36](#page-12-0) The aliphatic C−H stretching in HA appeared at  $2930-2850$  cm<sup>-1,[37](#page-13-0)</sup> The length and branching of these chains can significantly impact the solubility, reactivity, and adsorption properties of HA. In the range of 2700−2500

<span id="page-7-0"></span>cm<sup>−</sup><sup>1</sup> , O−H in the carboxyl group exhibited a stretching vibration.[38](#page-13-0) A small peak appeared in the range of 1700−1720 cm<sup>-1</sup> that was attributed to the stretching vibration of the C= O bond in the carboxyl group. $37$  It can be seen from [Figure](#page-6-0) 4 that at 1618 cm<sup>−</sup><sup>1</sup> the sharp edge loop band prominently indicates the presence of the  $C = C$  stretch (aromatic ring). The deformation vibrations of methylene and methyl groups occurred in the spectra at [1](#page-12-0)381  $\text{cm}^{-1}$ . In the range of 1200– 1260 cm<sup>−</sup><sup>1</sup> , the characteristic peaks are caused by the C−O vibration in aryl ethers and to the C−O stretching and O−H bending of carboxylic acids and/or phenolic units.<sup>[39](#page-13-0),[40](#page-13-0)</sup> At about 1046 cm<sup>−</sup><sup>1</sup> , the C−O stretching vibration in the polysaccharide and the Si−O stretching vibration in the silicate led to the appearance of this peak, which was often attributed to the mineral matter of the sample.<sup>[1](#page-12-0),[36,](#page-12-0)[41](#page-13-0)</sup> The bands occurring in the 900−700 cm<sup>−</sup><sup>1</sup> range is assigned to the out-ofplane bending vibration of aromatic C−H bonds.[18](#page-12-0),[29](#page-12-0) The bands observed in FTIR showed that it was evident that humic acids had an aromatic structure and contain aliphatic side chains. In addition, HA in coal samples has a complex heterogeneous chemical structure, which varies greatly with the source and properties of coal.

The range of  $1500-1800$  cm<sup>-1</sup> contains the absorption peak for oxygen-containing functional groups (Figure 5). The area



Figure 5. Oxygen functional group peaks of LSHA and WHHA

of each peak was integrated and normalized to obtain the relative percentage of each absorption peak. The results are shown in [Table](https://pubs.acs.org/doi/suppl/10.1021/acsomega.4c01840/suppl_file/ao4c01840_si_001.pdf) S3. It illustrated that the carboxyl content of the studied HAs was in the order of LSHA > WHHA, which was consistent with the determination results of acid functional groups in [Table](#page-6-0) 6. The aromaticity  $(f_a$  (FTIR)) of HAs can be calculated by the corresponding peak area in the FTIR spectrum.[42](#page-13-0) Its results are listed in [Table](#page-6-0) 6 and are consistent with the elemental analysis.

The XRD patterns of LSHA and WHHA are presented in Figure 6. The amorphous nature is predominant in the two humic acids. The XRD pattern of the isolated humic acids also indicated low impurities. Broad peaks are observed at  $2\theta = 25^{\circ}$ , indicating that HAs possess more aromatic carbon. This wide peak feature is usually associated with the complexity and amorphous properties of organic substances. The XRD approach can be used to analyze the aromatic degree of HA. It also allows for the calculation of structural parameters such as the average thickness of microcrystalline structure stacking  $(L_c)$ , the spacing between carbon planes  $(d_{002})$ , the average number of carbon plane stacking layers (*N*), and the purity of HA[.43](#page-13-0)<sup>−</sup>[46](#page-13-0) However, there are few studies on its application to coal HA. Schweitzer et al.<sup>[43](#page-13-0)</sup> divided the XRD patterns of HA into three zones, namely, *γ* band (distance between layers, 0.43 nm), G band (distance between layers, 0.35 nm), and 10 band (distance between layers, 0.23 nm). The *γ* band was a peak at



Figure 6. X-ray diffraction profiles of the studied HAs.

around  $2\theta = 20^\circ$  that was attributed primarily to aliphatic side chains, and the G (002) band was at  $2\theta = 25^{\circ}$ , which was due to aromatic carbon.

The XRD profiles of the HAs after waveform separation are shown in Figure 7 to obtain the aroma and corresponding



Figure 7. Fitted-peak curves of XRD patterns of four HAs.

Table 7. XRD and CP/MAS<sup>13</sup>C NMR Parameters of Four **HAs** 

Sample $d_{002}$ (Å) $L_c$ (Å) $N$ $f_{a-XRD}$ $f_{a-NMR}$ $f_{ar/al}$ $f_{h/h}$			
WHHA 3.45 14.73 4.27 0.48 0.65 2.95 0.29			
LSHA 3.43 16.73 4.88 0.46 0.64 3.05 0.36			

parameters of HA (Table 7). The peak-fitting results are shown in [Table](https://pubs.acs.org/doi/suppl/10.1021/acsomega.4c01840/suppl_file/ao4c01840_si_001.pdf) S4. The aromatic degree of the HAs was WHHA > LSHA, which was in agreement with the FITR and elemental analysis results. The layer space  $d_{002}$  of the studied HAs was not different, at around 3.4 Å. This may mean that the interaction forces between the two HA molecules are similar. The order of stacking thickness  $L_c$  and layer number  $N$  was WHHA > LSHA, which was related to the number of aromatic structures in HA. It indicates that WHHA molecular layers are more tightly stacked in the vertical direction, and the number of stacked layers is greater. This may be due to the fact that WHHA molecules contain more aromatic ring structures, which enhances the *π*−*π* interaction between molecules and promotes the stacking of molecular layers.

The CP/MAS<sup>13</sup>CNMR spectra of two kinds of HAs are shown in [Figure](#page-8-0) 8. All the spectra show weak signals in the range of 240−165 ppm. These signals may arise mainly from carbonyl carbon. The small but distinct signals between 190 and 165 ppm appear to arise mainly from carboxyl carbon in

<span id="page-8-0"></span>

Figure 8. CP/MAS<sup>13</sup>CNMR spectra of the HAs.

HA. Strong signals appear between 165 and 100 ppm in the range corresponding approximately to the aromatic carbons. The aliphatic carbons show signals in the range of 100−0 ppm.[42](#page-13-0) It can be seen from Figure 8 that the aromatic carbon content of the HAs is higher than that of the aliphatic carbon. Moreover, the content of aliphatic carbon in weathered lignite is lower than that in lignite.<sup>[32](#page-12-0)</sup>

The relative content of different types of carbon atoms in HA was obtained by CP/MAS<sup>13</sup>C NMR spectroscopy and peak fitting. Figure 9 is the fitting result of the two HAs' CP/



Figure 9. Peak fitting curves of CP/MAS<sup>13</sup>C NMR spectra of the two HAs.

MAS<sup>13</sup>CNMR spectra. After the classification of the functional groups containing carbon and the chemical displacement were obtained by peak separation processing on each level of the spectrogram, the relative content of each type of carbon was calculated according to the area of the fitted peak ([Table](https://pubs.acs.org/doi/suppl/10.1021/acsomega.4c01840/suppl_file/ao4c01840_si_001.pdf) S5). The relative content of aliphatic carbon (0−100 ppm) of the two HAs was WHHA (21.96%) > LSHA (20.67%). This indicates that WHHA may contain more aliphatic chains or branched chains, which may affect its solubility and biodegradability. The relative content of aromatic carbon (100−165 ppm) was WHHA (64.73%) > LSHA (63.93%), which was consistent with the XRD analysis. The relative contents of carboxyl carbon (165−190 ppm) of the two HAs were as follows: LSHA  $(5.97%) > WHHA (4.14%)$ , which was consistent with the trend of carboxyl carbon content measured by the calcium acetate method in [Table](#page-6-0) 6. The relative contents of the phenolic hydroxyl carbon (150−165 ppm) of the two HAs were LSHA  $(4.24%) > WHHA (2.91%)$ , which was consistent with the trend of phenolic hydroxyl size analysis in [Table](#page-6-0) 6. WHHA and LSHA have significant differences in structure, mainly in the contents of aromatic carbon and fatty carbon. These differences may result from differences in their original origin and subsequent weathering or treatment processes.

The related parameters in the  $CP/MAS^{13}CNMR$  of the HAs are summarized in [Table](#page-7-0) 7. The degree of aromaticity ( $f_{\text{a-NMR}}$ ) of the two HAs was WHHA > LSHA. This result was consistent with the XRD and FTIR analysis results. The hydrophilic−hydrophobic parameter ( $f_{\text{h/h}}$ ) was LSHA > WHHA, indicating that LSHA has the most hydrophilic groups. Thus, LSHA exhibited the highest biological activity in agricultural application.<sup> $4/$ </sup> This may be related to the extraction conditions. The extraction time and NaOH concentration for LSHA are both higher than those for WHHA, which facilitates a more effective disruption of the coal sample structure during the extraction process, releasing more bioactive components. Additionally, factors such as the type of coal sample, the degree of weathering, and environmental conditions can all potentially influence the biological activity of humic acid.

Figure 10 shows the TG and DTG curves for the HA samples studied, and data are given in Table 8. TG curves of

Table 8. Mass Changes of the Two HAs

600 °C Remnant
46.50% 4.12%
52.96% 5.52%

the studied samples showed gradual weight loss as a function of temperature increase. As observed, the thermal analysis of analyzed samples indicated the occurrence of three decomposition stages for LSHA and WHHA[.18](#page-12-0)

The first one, between 28 and 180 °C, corresponded to a mass loss of 11.6 9% and 13.57%; the second one, between 200 and 350 °C, corresponded to a mass loss of 10.86% and 7.03%; and the third one, between 350 and 600 °C, corresponded to a



Figure 10. TG-DTG curves of the two HAs.

mass loss of 69.93% and 75.28%. After that, weight loss remains steady, without further observable features, until the highest temperature was measured. The residual mass at 800 °C was around 5.52% and 4.12%. The first stage was attributed to the loss of free water (dehydration) from the HA sample. The second stage was associated with the degradation of polysaccharides, the decarboxylation of acidic groups, and the dehydration of aliphatic alcohols. The third stage (which showed a larger mass loss) was attributed to the decomposition of aromatic structures and cleavage of C−C bonds.[48](#page-13-0) Consequently, the strong exothermic peaks at approximately 457 and 417 °C showed a larger number of aromatic structures relative to the aliphatic structures in the HAs. Also, sample WHHA seems to be slightly more resistant to thermal degradation than LSHA, since the decomposition of the former samples occurs at relatively higher temperatures.<sup>[49](#page-13-0)</sup> This indicates that the aromatic structure in WHHA samples may be more complex or more cross-linked, thereby improving its thermal stability. In the total analyzed temperature range the weight loss was higher for WHHA than for LSHA, which could be connected with the lower humification degree (the lower values of  $N/C$ ).<sup>[50](#page-13-0)</sup> The differences exhibited by WHHA and LSHA samples during TGA may result from a combination of factors, such as their original sources, extraction methods, environmental conditions, and subsequent processing procedures. Together these factors determine the HA's chemical composition, structure, and properties of the sample.

XPS has been used to characterize the surface elemental compositions of LSHA and WHHA.<sup>1</sup> As Figure 11 displays,



Figure 11. Fitted-peak curves of C 1s spectra of the two HAs.

the XPS C 1s spectrum of HAs was fitted with 6 peaks at 284.4  $\pm$  0.2, 285.0  $\pm$  0.2, 286.0  $\pm$  0.2, 286.8  $\pm$  0.2, 288.1  $\pm$  0.2, and 288.8  $\pm$  0.2 eV, corresponding to aromatic, aliphatic, ether/ alcohol, ketonic, pyrrole/amide, and carboxyl carbons, respectively. The chemical assignments, binding energy (BE), and relative proportion (RP) of each functional group in the C 1s spectra of HA fractions are listed in Table 9. Aromatic carbon and aliphatic carbon were the main forms of carbon element present on the surface of the HAs, which was consistent with 13C NMR data. In addition, the relative proportion of carboxylic carbon in LSHA (8.50%) was higher than that of WHHA (6.06%), which indicates that LSHA

contains more carboxyl than WHHA. These results are in line with data from the FTIR and acid functional groups and CP/ MAS <sup>13</sup>CNMR of the HAs. For the XPS O 1s signals, four peaks were fitted as shown in [Figure](#page-10-0) 12. Chemical assignments and binding energy values of the peaks were C=O (531.7  $\pm$ 0.2 eV), C−O (532.7 ± 0.2 eV), COO (533.5 ± 0.2 eV), and adsorbed oxygen (534.6  $\pm$  0.2 eV), respectively.<sup>[51](#page-13-0)</sup> The experimental area percentages for each component of the O 1s spectra of HAs are listed in [Table](#page-10-0) 10. The data demonstrated that the main form of surface oxygen elements of the HAs was C�O, followed by C−O and COO, and the adsorbed oxygen content of the two samples was low. The above results show that the HAs from weathered lignite and previously studied HAs from lignite<sup>32</sup> have a very similar chemical composition.

The fluorescence index (FI) is a parameter used to distinguish organic matter derived from terrestrial and microbial sources. It is the ratio of the fluorescence intensity at 450/500 nm in the emission spectrum (with a fixed excitation wavelength of 370 nm). According to [Table](#page-10-0) 11, the FI values of the HAs were 1.15−1.36, indicating that the two HAs belonged to terrestrial  $HAs<sup>1</sup>$  $HAs<sup>1</sup>$  $HAs<sup>1</sup>$ . The biological index (BIX) values of the two HAs were less than 1, indicating that the content of primary organic matter of the two HAs was less, which was consistent with the results in the literature, $<sup>1</sup>$  $<sup>1</sup>$  $<sup>1</sup>$  and the</sup> content of primary organic matter of weathered lignite HA was less than that of lignite HA. This may be due to the fact that microbial activity and chemical decomposition during weathering further reduce the content of primary biomass.

The fluorescence excitation spectra of the studied HAs are shown in [Figure](#page-10-0)  $13(a)$ . The excitation spectra of LSHA and WHHA featured one intense peak at about 270 nm. This peak position is usually associated with the aromatic structure or conjugate system in the HA molecule, indicating that both LSHA and WHHA contain a certain number of aromatic compounds. The fluorescence emission spectra of LSHA and WHHA are shown in [Figure](#page-10-0)  $13(b)$ . The values of the fluorescence intensity (in arbitrary units, au) and fluorescence maxima are presented in [Table](#page-10-0) 11. The intensity of WHHA was significantly higher than LSHA, which was attributed to the fact that WHHA had more aromatic structures replaced by electron-donating groups or more aromatic structures were replaced by electron-withdrawing groups and/or an unsaturated conjugate structure because the HA molecules of electron-withdrawing groups (for example, carboxyl and carbonyl) can make the fluorescence intensity of HA decrease and electronic groups (such as amino, hydroxyl, methods, etc.) can make the fluorescence intensity of HA increase. The presence of these oxygen-containing and nitrogen-containing functional groups can transfer fluorescence to longer wavelengths by reducing the energy difference between the ground state and the first excited state.<sup>[36](#page-12-0)[,52](#page-13-0),[53](#page-13-0)</sup>

The impact of scattered light on the spectrum and spectral overlap can be reduced by using a synchronous fluorescence

Table 9. Fitted-Peak Results of <sup>C</sup> 1s Spectra of the Two HAs*<sup>a</sup>*

	Aromatic		Aliphatic		Ether/Alcohol		Ketonic		Pyrrole		Carboxylic	
Sample	BE	RP	<b>BE</b>	<b>RP</b>	BE	RP	BE	<b>RP</b>	BE	<b>RP</b>	BE	RP
<b>WHHA</b>	284.48	40.76	285.07	31.31	286.10	11.20	286.77	4.93	288.24	5.74	288.87	6.06
LSHA	284.50	38.77	285.04	30.44	285.90	12.52	286.90	7.95	288.06	1.82	288.81	8.50

*a* BE is binding energy, eV; RP is relative percentage, %.

<span id="page-10-0"></span>

Binding Energy (eV)

Figure 12. Fitted-peak curves of O 1s spectra of HAs.

Table 10. Fitted-Peak Results of <sup>O</sup> 1s Spectra of HAs*<sup>a</sup>*

	<b>LSHA</b>		WHHA				
Sample	<b>BE</b>	RP	<b>BE</b>	RP			
$C=0$	531.68	55.32	531.74	46.25			
$C - O$	532.82	21.65	532.67	13.86			
COO	533.59	18.20	533.38	38.64			
Adsorbed oxygen	534.64	4.82	534.66	1.25			
${}^a$ BE is binding energy, eV; RP is relative percentage, %.							

Table 11. Fluorescence Parameters and Maximum Emission Wavelength of HAs

			$E_{\rm m}$		
Sample	FI	<b>BIX</b>	$\lambda_{\text{max}}$ (nm)	$I$ (au)	
WHHA	1.36	0.49	414.2	130.6	
<b>LSHA</b>	1.15	0.39	437.0	125.2	



538 537 536 535 534 533 532 531 530 529 528 Binding Energy (eV)

spectrum. The most commonly used step length Δ*λ* (difference between the emission wavelength and excitation wavelength) is 18 nm.<sup>[53](#page-13-0)</sup> The synchronous fluorescence spectra of the HAs are shown in Figure 13(c). As can be observed, there were three obvious peaks in the synchronous fluorescence spectra of LSHA and WHHA, indicating that LSHA and WHHA had more complex structures and stronger dispersion. It may arise from the presence of different functional groups and structures in HA molecules, which contribute differently to fluorescence emission. In contrast, the previously studied lignite had the simplest structure and the least dispersion, which reflects the relatively unitary composition of its molecules. $32,53$  $32,53$  This reflects the relative uniformity of its molecular composition. In addition, when the excitation wavelength  $E_x$  of synchronous fluorescence spectrum ( $\Delta \lambda = 18$ nm) is in the range of 340−370 nm, the corresponding polycyclic aromatic hydrocarbons (PAHs) are composed of 3− 4 viscous benzene rings; when *Ex* is in the range of 370−420



Figure 13. Excitation spectra (a), emission spectra (b), and synchronous fluorescence (c) of the two HAs ( $\Delta \lambda = 18$  nm).



<span id="page-11-0"></span>Table 12. Location and Intensity of Ex/Em Peaks of HAs

nm, the corresponding PAHs are composed of 5 benzene rings; and when  $E_x$  is in the range of 438–487 nm, the corresponding PAHs have about 7 benzene rings or molecules containing lignin structures.<sup>[54](#page-13-0)</sup> Therefore, the PAHs in LSHA and WHHA were composed of three to five thickened benzene rings and were mainly composed of five thickened benzene rings. The HAs also contained a small amount of PAHs consisting of about seven fused benzene rings and lignin structures.

Table 12 shows that the EEM wavelength range is consistent with that in the literature, indicating that the two HAs both belong to coal  $HA_1^{\perp}$  $HA_1^{\perp}$  $HA_1^{\perp}$  and that the Ex/Em peak positions of the two HAs are very similar, indicating that the fluorophore in the two HA samples is the same or similar. The similarity of fluorophore may mean the two HA have common characteristics in structure and functional groups. When the excitation wavelength is 245−280 nm and the fluorescence emission wavelength is 340−400 nm, PAHs are composed of three condensed benzene rings. When the emission wavelength is 400−450 nm, PAHs are composed of four aromatic rings. When the  $\lambda_{\rm em}$  > 450 nm, PAHs may be composed of more than four benzene rings.<sup>[1](#page-12-0),[55](#page-13-0)</sup> As displayed in Table 12, PAHs composed of four thickened benzene rings exist in the two raw coal HAs. The results were in good agreement with those obtained by fluorescence synchronous spectroscopy.

## **4. CONCLUSIONS**

In this study, response surface methodology was employed to develop an efficient model aimed at optimizing the extraction of humic acids from two types of weathered lignite. The RSM outcomes for LSHA ( $R^2 = 0.9752$ ) and WHHA ( $R^2 = 0.9892$ ) indicated a high fitness of the models for predicting HA extraction efficiency. The extraction efficiency of HAs is governed by a confluence of factors, encompassing extraction time, NaOH concentration, the liquid-to-solid ratio, and their interactive effects. Additionally, regional differences emerged as a pivotal consideration that cannot be overlooked. The optimal extraction conditions were identified as 4.4 h for extraction time, 0.30 mol/L NaOH concentration, and a liquid-to-solid ratio of 21 mL/g for LSHA, whereas for WHHA, these were 3.1 h, 0.21 mol/L NaOH, and a ratio of 12 mL/g. The actual and predicted yields fell within the acceptable relative error limits.

UV−vis, FTIR, XPS, and fluorescence spectroscopy confirmed the presence of carboxyl, phenolic, ketonic, aromatic, and aliphatic functional groups in HAs extracted from weathered lignite. Notably, WHHA exhibited a higher aromaticity than LSHA, suggesting potentially greater chemical reactivity and adsorption capacity. The high aromatic carbon content and low aliphatic carbon content in WHHA likely contribute to its enhanced structural stability and reduced biodegradability. However, the abundant carboxyl and phenolic carbon in LSHA may confer heightened environmental reactivity and biological activity. This finding was

further corroborated by hydrophilic−hydrophobic parameters. WHHA samples demonstrated superior thermal stability compared to LSHA. Both belonged to terrestrial HAs with a relatively low native organic matter content. The polycyclic aromatic hydrocarbons in LSHA and WHHA were predominantly composed of three to five benzene rings, with trace amounts of higher-ring PAHs.

The subtle differences in chemical composition and structure between LSHA and WHHA can be traced back to their original coal samples, weathering degrees, and subsequent processing procedures. Weathering processes alter the chemical composition and structure of coal samples, thereby influencing the properties of the extracted HAs. Furthermore, variations in treatment conditions may lead to changes in the distribution and content of functional groups within the HAs. Environmental pollution and resource waste can be reduced by optimizing the extraction process. In the future, the RSM can be extended to predict the extraction efficiency of different coal sources with different properties. The results of this study provide strong support for the sustainable utilization of HA.

## ■ **ASSOCIATED CONTENT**

#### $\bullet$  Supporting Information

The Supporting Information is available free of charge at [https://pubs.acs.org/doi/10.1021/acsomega.4c01840.](https://pubs.acs.org/doi/10.1021/acsomega.4c01840?goto=supporting-info)

Effects of single factors on the yield of HAs; factors and levels of BBD-RSM design; fitted-peak assignment of FTIR; fitted-peak results of XRD; and fitted-peak results of  $CP/MAS$ <sup>13</sup>C NMR ([PDF\)](https://pubs.acs.org/doi/suppl/10.1021/acsomega.4c01840/suppl_file/ao4c01840_si_001.pdf)

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**Notes**

The authors declare no competing financial interest.

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# ■ **REFERENCES**

(1) Doskočil, L.; Burdíková-Szewieczková, J.; Enev, V.; Kalina, L.; Wasserbauer, J. Spectral [characterization](https://doi.org/10.1016/j.fuel.2017.10.114) and comparison of humic acids isolated from some [European](https://doi.org/10.1016/j.fuel.2017.10.114) lignites. *Fuel* 2018, *213*, 123−132. (2) de Melo, B. A.; Motta, F. L.; Santana, M. H. [Humic](https://doi.org/10.1016/j.msec.2015.12.001) acids: Structural properties and multiple [functionalities](https://doi.org/10.1016/j.msec.2015.12.001) for novel technological [developments.](https://doi.org/10.1016/j.msec.2015.12.001) *Mater. Sci. Eng. C Mater. Biol. Appl.* 2016, *62*, 967−74.

(3) Havelcova, M.; Mizera, J.; Sykorova, I.; Pekar, M. [Sorption](https://doi.org/10.1016/j.jhazmat.2008.03.136) of metal ions on lignite and the derived humic [substances.](https://doi.org/10.1016/j.jhazmat.2008.03.136) *J. Hazard Mater.* 2009, *161* (1), 559−64.

(4) Nardi, S.; Pizzeghello, D.; Muscolo, A.; Vianello, A. [Physiological](https://doi.org/10.1016/S0038-0717(02)00174-8) effects of humic [substances](https://doi.org/10.1016/S0038-0717(02)00174-8) on higher plants. *J. Soil Biology Biochemistry*2002, *34* (11), 1527.

(5) Sun, Z.; Tang, B.; Xie, H. [Treatment](https://doi.org/10.1021/ef502299k?urlappend=%3Fref%3DPDF&jav=VoR&rel=cite-as) of Waste Gases by Humic [Acid.](https://doi.org/10.1021/ef502299k?urlappend=%3Fref%3DPDF&jav=VoR&rel=cite-as) *Energy Fuels* 2015, *29* (3), 1269−1278.

(6) Peña-Méndez, E. M., H, J., Patocka, ̌ J., Humic [substances](https://doi.org/10.32725/jab.2005.002)compounds of still unknown structure: [applications](https://doi.org/10.32725/jab.2005.002) in agriculture, industry, [environment,](https://doi.org/10.32725/jab.2005.002) and biomedicine. *Journal of Applied Biomedicine* 2005, *3*, 13.

(7) Dong, L. H.; Yang, J. S.; Yuan, H. L.; Wang, E. T.; Chen, W. X. Chemical [characteristics](https://doi.org/10.1016/j.ejsobi.2007.07.002) and influences of two fractions of Chinese lignite humic acids on [urease.](https://doi.org/10.1016/j.ejsobi.2007.07.002) *European Journal of Soil Biology* 2008, *44* (2), 166−171.

(8) Qian, S.; Ding, W.; Li, Y.; Liu, G.; Sun, J.; Ding, Q. [Characterization](https://doi.org/10.1080/09542299.2015.1118361) of humic acids derived from Leonardite using a solid-state NMR [spectroscopy](https://doi.org/10.1080/09542299.2015.1118361) and effects of humic acids on growth and [nutrient](https://doi.org/10.1080/09542299.2015.1118361) uptake of snap bean. *Chemical Speciation & Bioavailability* 2015, *27* (4), 156−161.

(9) Trinh, T. K.; Kang, L. S. Response surface [methodological](https://doi.org/10.1016/j.cherd.2010.12.004) approach to optimize the coagulation−[flocculation](https://doi.org/10.1016/j.cherd.2010.12.004) process in drinking water [treatment.](https://doi.org/10.1016/j.cherd.2010.12.004) *Chem. Eng. Res. Des.* 2011, *89* (7), 1126− 1135.

(10) Betiku, E.; Omilakin, O. R.; Ajala, S. O.; Okeleye, A. A.; Taiwo, A. E.; Solomon, B. O. [Mathematical](https://doi.org/10.1016/j.energy.2014.05.033) modeling and process parameters [optimization](https://doi.org/10.1016/j.energy.2014.05.033) studies by artificial neural network and response surface [methodology:](https://doi.org/10.1016/j.energy.2014.05.033) A case of non-edible neem (Azadirachta indica) seed oil biodiesel [synthesis.](https://doi.org/10.1016/j.energy.2014.05.033) *Energy* 2014, *72*, 266−273.

(11) Sarlaki, E.; Sharif Paghaleh, A.; Kianmehr, M. H.; Asefpour Vakilian, K. [Valorization](https://doi.org/10.1016/j.renene.2020.08.096) of lignite wastes into humic acids: Process [optimization,](https://doi.org/10.1016/j.renene.2020.08.096) energy efficiency and structural features analysis. *Renewable Energy* 2021, *163*, 105−122.

(12) Wang, C.-F.; Fan, X.; Zhang, F.; Wang, S.-Z.; Zhao, Y.-P.; Zhao, X.-Y.; Zhao, W.; Zhu, T.-G.; Lu, J.-L.; Wei, X.-Y. [Characterization](https://doi.org/10.1039/C7RA01497J) of humic acids extracted from a lignite and [interpretation](https://doi.org/10.1039/C7RA01497J) for the mass [spectra.](https://doi.org/10.1039/C7RA01497J) *RSC Adv.* 2017, *7* (33), 20677−20684.

(13) Haider, R.; Ghauri, M. A.; Akhtar, K. [Isolation](https://doi.org/10.1080/01490451.2015.1039673) of Coal [Degrading](https://doi.org/10.1080/01490451.2015.1039673) Fungus from Drilled Core Coal Sample and Effect of Prior Fungal [Pretreatment](https://doi.org/10.1080/01490451.2015.1039673) on Chemical Attributes of Extracted Humic [Acid.](https://doi.org/10.1080/01490451.2015.1039673) *Geomicrobiology Journal* 2015, *32* (10), 944−953.

(14) Jia, J.; Zhang, Y.; Liu, Q.; Huang, G.; Xing, B.; Zhang, C.; Guo, H.; Pan, J.; Cao, Y. [Characterization](https://doi.org/10.1080/15567036.2020.1824034) of coal-based humic acids in relation to their [preparation](https://doi.org/10.1080/15567036.2020.1824034) methods. *Energy Sources, Part A: Recovery, Utilization, and Environmental Effects* 2020, 1−11.

(15) Li, S. *Humic Acid Product Analysis and Standards*; Chemical Industry Press, 2007.

(16) GB/T212−2008, Proximate Analysis of Coal. *National Standard of the People's Republic of China*; 2008.

(17) Klavins, M.; Purmalis, O. [Properties](https://doi.org/10.1016/j.molstruc.2013.07.021) and structure of raised bog peat [humic](https://doi.org/10.1016/j.molstruc.2013.07.021) acids. *J. Mol. Struct.* 2013, *1050*, 103−113.

(18) Das, T.; Saikia, B. K.; Baruah, B. P.; Das, D. [Characterizations](https://doi.org/10.1007/s12594-015-0334-0) of humic acid isolated from coals of two Nagaland [Coalfields](https://doi.org/10.1007/s12594-015-0334-0) of India in [relation](https://doi.org/10.1007/s12594-015-0334-0) to their origin. *Journal of the Geological Society of India* 2015, *86*, 468−474.

(19) Cheng, G.; Niu, Z.; Zhang, C.; Zhang, X.; Li, X. [Extraction](https://doi.org/10.3390/app9071356) of Humic Acid from Lignite by [KOH-Hydrothermal](https://doi.org/10.3390/app9071356) Method. *Applied Sciences* 2019, *9* (7), 1356.

(20) Li, Y.; Yuan, S. [Influence](https://doi.org/10.1007/s42452-020-04087-x) of addition of KOH on the yield and [characteristics](https://doi.org/10.1007/s42452-020-04087-x) of humic acids extracted from lignite using NaOH. *SN Applied Sciences* 2021, *3* (1), DOI: [10.1007/s42452-020-04087-x.](https://doi.org/10.1007/s42452-020-04087-x?urlappend=%3Fref%3DPDF&jav=VoR&rel=cite-as)

(21) Rashid, T.; Sher, F.; Jusoh, M.; Joya, T. A.; Zhang, S.; Rasheed, T.; Lima, E. C. Parametric [optimization](https://doi.org/10.1016/j.envres.2022.115160) and structural feature analysis of humic acid [extraction](https://doi.org/10.1016/j.envres.2022.115160) from lignite. *Environ. Res.* 2023, *220*, No. 115160.

(22) Tian, D. Z. W. Z. S. C. J. Optimization of Stir-fried with Saltwater Technology in Plantain Seed by BBD-RSM. *Medicinal Plant* 2014, *5* (03), 9−13.

(23) MT/T 849-2000, Classification of volatile matter yield of coal. *Coal Industry Standard of the People's Republic of China*, 2000.

(24) GB/T15224.1-2018, Classification for quality of coal-Part 1: Ash. *National Standard of the People's Republic of China*, 2018.

(25) Xing, B.; McGill, W. B.; Dudas, M. J. [Cross-correlation](https://doi.org/10.1021/es00060a025?urlappend=%3Fref%3DPDF&jav=VoR&rel=cite-as) of polarity curves to predict partition [coefficients](https://doi.org/10.1021/es00060a025?urlappend=%3Fref%3DPDF&jav=VoR&rel=cite-as) of nonionic organic [contaminants.](https://doi.org/10.1021/es00060a025?urlappend=%3Fref%3DPDF&jav=VoR&rel=cite-as) *Environ. Sci. Technol.* 1994, *28* (11), 1929−33.

(26) Ojwang, L. M.; Cook, R. L. [Environmental](https://doi.org/10.1021/es4004922?urlappend=%3Fref%3DPDF&jav=VoR&rel=cite-as) conditions that influence the ability of humic acids to induce [permeability](https://doi.org/10.1021/es4004922?urlappend=%3Fref%3DPDF&jav=VoR&rel=cite-as) in model [biomembranes.](https://doi.org/10.1021/es4004922?urlappend=%3Fref%3DPDF&jav=VoR&rel=cite-as) *Environ. Sci. Technol.* 2013, *47* (15), 8280−7.

(27) Sun, H.; Zhu, D.; Mao, J. Sorption of polar and [nonpolar](https://doi.org/10.1897/08-124.1) aromatic [compounds](https://doi.org/10.1897/08-124.1) to two humic acids with varied structural [heterogeneity.](https://doi.org/10.1897/08-124.1) *Environ. Toxicol. Chem.* 2008, *27* (12), 2449−56.

(28) Bozena ̇ Debska, M. D., Tobiasova, E. *Effect of Post-Harvest Residue of Maize, Rapeseed, and Sunflower on Humic Acids Properties in Various Soils*, 2012.

(29) Ibarra, J.; Juan, R. [Structural](https://doi.org/10.1016/0016-2361(85)90050-X) changes in humic acids during the [coalification](https://doi.org/10.1016/0016-2361(85)90050-X) process. *Fuel* 1985, *64*, 650−656.

(30) Francioso, O.; Montecchio, D.; Gioacchini, P.; Ciavatta, C. Thermal analysis (TG−DTA) and isotopic [characterization](https://doi.org/10.1016/j.apgeochem.2004.10.003) (13C− 15N) of humic acids from [different](https://doi.org/10.1016/j.apgeochem.2004.10.003) origins. *Appl. Geochem.* 2005, *20* (3), 537−544.

(31) Allard, B. A comparative study on the chemical [composition](https://doi.org/10.1016/j.geoderma.2005.01.010) of humic acids from forest soil, [agricultural](https://doi.org/10.1016/j.geoderma.2005.01.010) soil and lignite deposit. *Geoderma* 2006, *130* (1−2), 77−96.

(32) Wang, M.; Li, Y.; Zhang, Y.; Hu, X.; Li, Q.; Su, Y.; Zhao, W. Exploration of the  $H_2O_2$  Oxidation Process and [Characteristic](https://doi.org/10.1021/acsomega.1c03257?urlappend=%3Fref%3DPDF&jav=VoR&rel=cite-as) [Evaluation](https://doi.org/10.1021/acsomega.1c03257?urlappend=%3Fref%3DPDF&jav=VoR&rel=cite-as) of Humic Acids from Two Typical Lignites. *ACS Omega* 2021, *6* (37), 24051−24061.

(33) Nasir, S.; Sarfaraz, T. B.; Verheyen, T. V.; Chaffee, A. L. Structural [elucidation](https://doi.org/10.1016/j.fuproc.2010.12.020) of humic acids extracted from Pakistani lignite using [spectroscopic](https://doi.org/10.1016/j.fuproc.2010.12.020) and thermal degradative techniques. *Fuel Process. Technol.* 2011, *92* (5), 983−991.

(34) Cieslewicz, J.; Gonet, S. S. [Properties](https://doi.org/10.1007/s00027-004-0702-0) of humic acids as biomarkers of lake catchment [management.](https://doi.org/10.1007/s00027-004-0702-0) *Aquatic. Sci.* 2004, *66* (2), 178−184.

(35) Zalba, P.; Amiotti, N. M.; Galantini, J. A.; Pistola, S. Soil [Humic](https://doi.org/10.1080/00103624.2016.1206558) and Fulvic Acids from Different Land-Use Systems [Evaluated](https://doi.org/10.1080/00103624.2016.1206558) By E4/ E6 [Ratios.](https://doi.org/10.1080/00103624.2016.1206558) *Communications in soil science and plant analysis* 2016, *47* (13−14), 1675−1679.

(36) Enev, V.; Pospíšilová, L.; Klučáková, M.; Liptaj, T.; Doskočil, L. Spectral [characterization](https://doi.org/10.17221/39/2013-SWR) of selected humic substances. *Soil and Water Research* 2014, *9* (1), 9−17.

<span id="page-13-0"></span>(37) Zykova, M. V.; Brazovsky, K. S.; Veretennikova, E. E.; Danilets, M. G.; Logvinova, L. A.; Romanenko, S. V.; Trofimova, E. S.; Ligacheva, A. A.; Bratishko, K. A.; Yusubov, M. S.; Lyapkov, A. A.; Belousov, M. V. New artificial network model to estimate [biological](https://doi.org/10.1016/j.envres.2020.109999) [activity](https://doi.org/10.1016/j.envres.2020.109999) of peat humic acids. *Environ. Res.* 2020, *191*, No. 109999.

(38) Baigorri, R.; Fuentes, M.; Gonzalez-Gaitano, G.; Garcia-Mina, J. M.; Almendros, G.; Gonzalez-Vila, F. J. [Complementary](https://doi.org/10.1021/jf8035353?urlappend=%3Fref%3DPDF&jav=VoR&rel=cite-as) multianalytical approach to study the [distinctive](https://doi.org/10.1021/jf8035353?urlappend=%3Fref%3DPDF&jav=VoR&rel=cite-as) structural features of the main humic fractions in [solution:](https://doi.org/10.1021/jf8035353?urlappend=%3Fref%3DPDF&jav=VoR&rel=cite-as) gray humic acid, brown humic acid, and [fulvic](https://doi.org/10.1021/jf8035353?urlappend=%3Fref%3DPDF&jav=VoR&rel=cite-as) acid. *J. Journal of agricultural and food chemistry* 2009, *57* (8), 3266.

(39) Novák, F.; Š estauberová, M.; Hrabal, R. [Structural](https://doi.org/10.1016/j.molstruc.2015.03.054) features of [lignohumic](https://doi.org/10.1016/j.molstruc.2015.03.054) acids. *J. Mol. Struct.* 2015, *1093*, 179−185.

(40) Dick, D. P.; Mangrich, A. S.; Menezes, S. M. C.; Pereira, B. F. Chemical and Spectroscopical [Characterization](https://doi.org/10.1590/S0103-50532002000200008) of Humic Acids from two South Brazilian Coals of [Different](https://doi.org/10.1590/S0103-50532002000200008) Ranks. *J. Braz. Chem. Soc.*. 2002, *13* (2), 177.

(41) Liebezeit, U. F. G. *An IR study of humic acids isolated from sediments and soils*, 2003.

(42) Gong, G.-q.; Zhao, Y.-f.; Zhang, Y.-j.; Deng, B.; Liu, W.-x.; Wang, M.; Yuan, X.; Xu, L.-w. [Establishment](https://doi.org/10.1016/j.fuel.2020.117210) of a molecular structure model for classified products of [coal-based](https://doi.org/10.1016/j.fuel.2020.117210) fulvic acid. *Fuel* 2020, *267*, No. 117210.

(43) Schnitzer, M.; Kodama, H.; Ripmeester, J. A. [Determination](https://doi.org/10.2136/sssaj1991.03615995005500030018x) of the [Aromaticity](https://doi.org/10.2136/sssaj1991.03615995005500030018x) of Humic Substances by X-Ray Diffraction Analysis. *J. Soil Science Society of America Journal* 1991, *55* (3), 745.

(44) Katsumi, N.; Yonebayashi, K.; Okazaki, M. [Evaluation](https://doi.org/10.1080/00380768.2015.1034638) of stacking [nanostructure](https://doi.org/10.1080/00380768.2015.1034638) in soil humic acids by analysis of the 002 band of their X-ray [diffraction](https://doi.org/10.1080/00380768.2015.1034638) profiles. *Soil Science and Plant Nutrition* 2015, *61* (4), 603−612.

(45) Ikeya, K.; Hikage, T.; Arai, S.; Watanabe, A. Size [distribution](https://doi.org/10.1016/j.orggeochem.2010.10.006) of [condensed](https://doi.org/10.1016/j.orggeochem.2010.10.006) aromatic rings in various soil humic acids. *Org. Geochem.* 2011, *42* (1), 55−61.

(46) Naidja, A.; Huang, P. M.; Anderson, D. W.; Kessel, C. V. Fourier Transform Infrared, [UV-Visible,](https://doi.org/10.1366/0003702021954908) and X-Ray Diffraction [Analyses](https://doi.org/10.1366/0003702021954908) of Organic Matter in Humin, Humic Acid, and Fulvic Acid Fractions in Soil Exposed to Elevated CO2 and N [Fertilization.](https://doi.org/10.1366/0003702021954908) *J. Appl. Spectrosc.* 2002, *56* (3), 318.

(47) Zherebtsov, S. I.; Malyshenko, N. V.; Votolin, K. S.; Androkhanov, V. A.; Sokolov, D. A.; Dugarjav, J.; Ismagilov, Z. R. [Structural-Group](https://doi.org/10.3103/S0361521919030121) Composition and Biological Activity of Humic Acids Obtained from Brown Coals of Russia and [Mongolia.](https://doi.org/10.3103/S0361521919030121) *Solid Fuel Chemistry* 2019, *53* (3), 145−151.

(48) Jl, M.; Dm, X.; Vs, S.; Rp, S.; Mf, M.; Sn, C.; Vn, F.; Bs, C. [Characterization](https://doi.org/10.9735/0975-3710.4.5.238-242) of the Coal Humic Acids from the Candiota [Coalfield,](https://doi.org/10.9735/0975-3710.4.5.238-242) Brazil. *International Journal of Agriculture Sciences* 2012, *4* (5), 238−242.

(49) Giovanela, M.; Crespo, J. S.; Antunes, M.; Adamatti, D. S.; Fernandes, A. N.; Barison, A.; da Silva, C. W. P.; Guégan, R.; Motelica-Heino, M.; Sierra, M. M. D. Chemical and [spectroscopic](https://doi.org/10.1016/j.molstruc.2010.07.038) [characterization](https://doi.org/10.1016/j.molstruc.2010.07.038) of humic acids extracted from the bottom sediments of a Brazilian subtropical [microbasin.](https://doi.org/10.1016/j.molstruc.2010.07.038) *J. Mol. Struct.* 2010, *981* (1−3), 111−119.

(50) Boguta, P.; Sokołowska, Z.; Skic, K., Use of [thermal](https://doi.org/10.1371/journal.pone.0189653) analysis coupled with differential scanning [calorimetry,](https://doi.org/10.1371/journal.pone.0189653) quadrupole mass spectrometry and infrared spectroscopy [\(TG-DSC-QMS-FTIR\)](https://doi.org/10.1371/journal.pone.0189653) to monitor chemical [properties](https://doi.org/10.1371/journal.pone.0189653) and thermal stability of fulvic and humic [acids](https://doi.org/10.1371/journal.pone.0189653). *J. PLoS ONE* 2017, *12* (12), e0189653.

(51) Li, Z.-K.; Wei, X.-Y.; Yan, H.-L.; Zong, Z.-M. [Insight](https://doi.org/10.1016/j.fuel.2015.02.117) into the structural features of Zhaotong lignite using multiple [techniques.](https://doi.org/10.1016/j.fuel.2015.02.117) *Fuel* 2015, *153*, 176−182.

(52) Fuentes, M.; González-Gaitano, G.; García-Mina, J. M. [The](https://doi.org/10.1016/j.orggeochem.2006.07.024) usefulness of UV−visible and fluorescence [spectroscopies](https://doi.org/10.1016/j.orggeochem.2006.07.024) to study the chemical nature of humic [substances](https://doi.org/10.1016/j.orggeochem.2006.07.024) from soils and composts. *Org. Geochem.* 2006, *37* (12), 1949−1959.

(53) Rodríguez, F.; Schlenger, P.; García-Valverde, M. [A](https://doi.org/10.1016/j.scitotenv.2013.11.150) [comprehensive](https://doi.org/10.1016/j.scitotenv.2013.11.150) structural evaluation of humic substances using several [fluorescence](https://doi.org/10.1016/j.scitotenv.2013.11.150) techniques before and after ozonation. *Part I: Structural characterization of humic substances*. 2016, *476*−*477*, 718.

(54) Peuravuori, J.; Koivikko, R.; Pihlaja, K. [Characterization,](https://doi.org/10.1016/S0043-1354(02)00172-0) [differentiation](https://doi.org/10.1016/S0043-1354(02)00172-0) and classification of aquatic humic matter separated with different sorbents: [synchronous](https://doi.org/10.1016/S0043-1354(02)00172-0) scanning fluorescence spectros[copy](https://doi.org/10.1016/S0043-1354(02)00172-0) *J. Water Research* 2002, *36* (18), 4552.

(55) Doskočil, L.; Enev, V.; Pekař, M.; Wasserbauer, J. [The](https://doi.org/10.1016/j.orggeochem.2016.02.008) spectrometric [characterization](https://doi.org/10.1016/j.orggeochem.2016.02.008) of lipids extracted from lignite samples from [various](https://doi.org/10.1016/j.orggeochem.2016.02.008) coal basins. *Org. Geochem.* 2016, *95*, 34−40.