

Journal of International Medical Research 50(2) 1–7 © The Author(s) 2022 Article reuse guidelines: sagepub.com/journals-permissions DOI: 10.1177/03000605221079102 journals.sagepub.com/home/imr



Platelet phagocytosis by leukocytes in a patient with cerebral hemorrhage and thrombocytopenia caused by gram-negative bacterial infection

Xiuji Wu¹, Youjun Li² and Xiaoyang Yang³ (D)

Abstract

Bacteria-induced thrombocytopenia is a common clinical disease that is often ignored by clinical and scientific research. Thus, exploring the mechanism and principle of bacteria-induced thrombocytopenia could facilitate the development of new diagnostic, preventative, and treatment modalities for thrombocytopenia. This case report describes a case of platelet phagocytosis by neutrophils and monocytes in a patient with cerebral hemorrhage and thrombocytopenia caused by gram-negative bacterial infection. After the infection was eradicated, platelet phagocytosis was alleviated, and his platelet count normalized. Cellular immunity may be an important cause of bacteria-induced thrombocytopenia in patients with cerebral hemorrhage.

Keywords

Cerebral haemorrhage, thrombocytopenia, leukocyte, phagocytosis, case report, neutrophil, bacterial infection

Date received: 28 July 2021; accepted: 11 January 2022

¹Department of Laboratory Medicine, Affiliated Haikou Hospital of Xiangya Medical College, Central South University, Haikou, Hainan Province, China ²Department of Neurosurgery, Affiliated Haikou Hospital of Xiangya Medical College, Central South University, Haikou, Hainan Province, China ³Department of Hematology, Affiliated Haikou Hospital of Xiangya Medical College, Central South University, Haikou, Hainan Province, China

Corresponding author:

Xiaoyang Yang, Department of Hematology, Affiliated Haikou Hospital of Xiangya Medical College, Central South University, No. 43 Renmin Avenue, Haikou City, Hainan 410083, China. Email: y108108@126.com

Creative Commons Non Commercial CC BY-NC: This article is distributed under the terms of the Creative Commons Attribution-NonCommercial 4.0 License (https://creativecommons.org/licenses/by-nc/4.0/) which permits non-commercial use, reproduction and distribution of the work without further permission provided the original work is attributed as specified on the SAGE and Open Access pages (https://us.sagepub.com/en-us/nam/open-access-at-sage).

Introduction

Thrombocytopenia caused by infection is common in clinical practice. Its mechanism may be related to the direct damage of megakaryocytes, the direct destruction or consumption of platelets by pathogens, and platelet immune-mediated destruction. specific mechanism is However, the unknown, and little clinical or scientific research has addressed this issue. We found that the phagocytosis of neutrophils and macrophages played an important role in a patient with intracerebral hemorrhage and thrombocytopenia caused by gramnegative bacterial infection, indicating that cellular immunity might be an important cause of thrombocytopenia caused by infection.

Case report

Patient details

A 15-year-old boy was hospitalized with head injury and coma after riding in an electric car on January 17, 2020. At the time of admission, his temperature was 36.8°C, his pulse was 108 beats/minute, his respiratory rate was 19 breaths/minute, and his blood pressure was 110/60 mmHg. His abdomen was soft without tenderness. and his liver and spleen were not swollen. Subarachnoid hemorrhage, left frontotemporal extradural hematoma. left frontotemporal subdural hematoma. and left frontotemporal bone fracture were detected by computed tomography (CT). On the same day, hematoma removal and cranial flap decompression were performed under emergency general anesthesia. Postoperative CT revealed that the intracerebral hemorrhage had been cleared, but his consciousness was not restored. We suspected the existence of a brain hernia that could affect cerebral function. His temperature was 39.5°C on January 17 and 18, 2020, gradually falling to 38.2°C on January 19, 2020 and 37.5°C on January 30, 2020. His temperature returned to normal on February 1, 2020, and he gradually recovered consciousness on February 5, 2020. The patient was healthy, and he had no family history of thrombocytopenia.

This work was conducted in accordance with the Code of Ethics of the World Medical Association (Declaration of Helsinki) for experiments involving humans. The reporting of this study conforms to the CARE guidelines.¹ The patient provided written informed consent for the publication of this case report. Because all performed tests are routinely used in clinical diagnosis and treatment, the ethics committee of our hospital waived the requirement for ethics approval.

Laboratory examination

Routine blood testing was performed using an XN9000 automatic 5-part differential analyzer (Sysmex. hematology Kobe. Japan) and supporting reagents. Fibrinogen content was tested using a Ca7000 automatic coagulation analyzer (Sysmex) and supporting reagents. Blood biochemistry testing was performed using a Dxc800 automatic biochemical analyzer (Beckman-Coulter, Brea, CA, USA) and supporting reagents. The isolation, culture, and identification of sputum bacteria were performed in accordance with the national guide for clinical laboratory procedures (3rd edition) using the VITEK2 COMPACT automatic microbiology analvzer (BioMérieux. Marcy-l'Étoile, France).

Results of laboratory examination

The platelet count was normal at admission, but it decreased after infection before gradually returning to normal after the infection was cured (Tables 1–2). There was no obvious abnormality regarding bleeding and coagulation. After hematoma

Date	Platelets (10 ⁹ /L)	Hemoglobin (g/dL)	Erythrocyte (10 ¹² /L)	Leucocytes (10 ⁹ /L)	Lymphocyte ratio (%)	Neutrophil ratio (%)
1-17-2020	126	157	5.27	22.22	2.1	93.8
1-19-2020	89	95	3.4	15.9	4.5	91.1
2-5-2020	119	94	3.35	6.26	29.1	58.8

Table 1. Changes in the patient's blood routine test data.

Table 2. Changes in the patient's biochemistry test data.

Date	Blood glucose (mmol/L)	Triglyceride (mmol/L)	Lactate dehydrogenase (U/L)
1-17-2020	12.66	0.33	262
1-19-2020	5.5	0.36	312
2-50-2020	1	1.11	177

removal, the patient continued to have a fever. The patient had no history of infection outside the hospital, and his fever started after admission, strongly suggesting nosocomial infection. Haemophilus influenzae was detected in sputum culture, indicating the existence of pulmonary infection. On January 19, 2020, peripheral blood smear analysis revealed high levels of platelet phagocytosis by neutrophils and monocytes, toxic granules and vacuoles in the cytoplasm of the neutrophils, and a small amount of aggregation and mild satellite phenomena in platelets. After the infection was cured, platelet phagocytosis was no longer observed (Figures 1-2). After treatment, the patient recovered, after which he was discharged from the hospital (Figure 3).

Treatment

On January 17, 2020, intracranial hematoma removal surgery was performed under general anesthesia. One unit of platelets was transfused on January 19, 2020. Meropenem and phenobarbital sodium were injected intramuscularly on January 20, 2020, and cefoperazone sodium was injected on February 2, 2020.

Discussion

Thrombocytopenia caused by infection is a common clinical phenomenon, but it has not been widely investigated. In addition, the specific mechanism is unclear. We found that gram-negative bacteria played an important role in the development of thrombocytopenia in a patient with subarachnoid hemorrhage. The patient was admitted to our hospital with cerebral hemorrhage after a fall. On admission, his body temperature and platelet count were normal, but his leukocyte and neutrophil counts were elevated. After hematoma removal, the patient continued to have a fever. H. influenzae was detected in sputum culture, indicating the presence of pulmonary infection. On January 19, 2020, his leukocyte count (mainly neutrophils) remained elevated, and his platelet count was decreased. Other findings included high numbers neutrophilof and monocyte-phagocytized platelets, satellite phenomena, and aggregation of a small number of platelets, which were observed

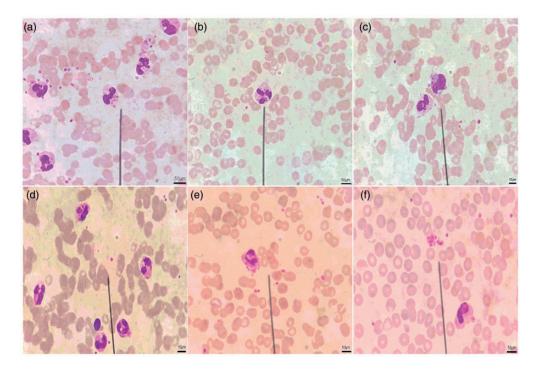


Figure 1. Platelet phagocytosis in patients with thrombocytopenia. (a) Platelets are phagocytized by neutrophils (Wright–Giemsa stain, $\times 1000$). (b) Platelets are phagocytized by monocytes (Wright–Giemsa stain, $\times 1000$). (c) Platelets are phagocytized by monocytes (Wright–Giemsa stain, $\times 1000$). (d) Platelets are phagocytized by monocytes, and toxic particles are present (Wright–Giemsa stain, $\times 1000$). (e) Platelets adhere to neutrophils, followed by platelet phagocytosis (Wright–Giemsa stain, $\times 1000$). (f) Platelet aggregation (Wright–Giemsa stain, $\times 1000$).

after the peripheral blood smear was stained with Wright's staining.

Intracerebral hemorrhage can cause endothelial cell damage and activate platelets.² After activation, platelets adhere to the vascular endothelium and react with neutrophils and monocytes.^{3,4} The main physiological functions of platelets include hemostasis, coagulation, inflammation, and immune reactions. Thrombin-sensitive protein (TSP) can mediate the interaction between activated platelets and monocytes because both leukocytes and activated platelets have receptors for TSP on their surfaces. Relevant studies have illustrated that platelet membrane protein-140 can also mediate the interaction between activated platelets and neutrophils and cause

thrombocytopenia by participating in the binding of activated platelets and leukocytes.⁵ In addition, vascular adhesion protein-1 can mediate the interaction between platelets and neutrophils. Static platelets cannot react with neutrophils, but thrombin-activated platelets react with neutrophils in a calcium-dependent manner.⁶

In this case, the patient had a stress reaction caused by an accident, elevated blood glucose levels, and marked leukocytosis. Acute stress and infection mainly trigger innate immune responses. Cells involved in innate immunity do not target any specific pathogen; rather, they widely attack a variety of different pathogens quickly (minutes to hours) when challenged. Neutrophils and macrophages play major

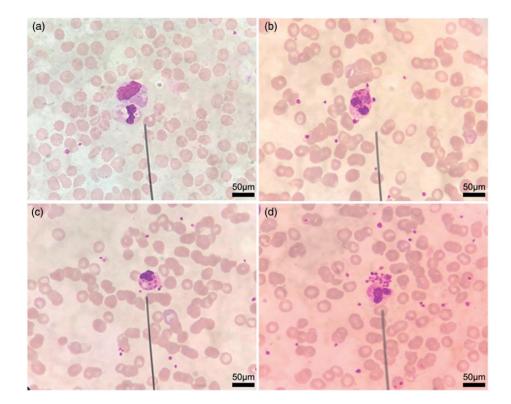


Figure 2. Sodium citrate anticoagulant. (a) Platelets are phagocytized by neutrophils and monocytes (Wright–Giemsa stain, $\times 1000$). (b) Platelets are phagocytized by neutrophils (Wright–Giemsa stain, $\times 1000$). (c) Platelets are phagocytized by neutrophils (Wright–Giemsa stain, $\times 1000$). (d) Platelet aggregation (Wright–Giemsa stain, $\times 1000$).

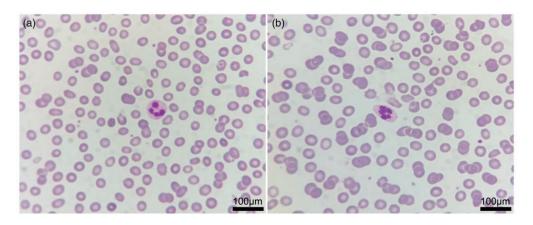


Figure 3. Platelet phagocytosis was alleviated after thrombocytopenia was resolved. (a–b) Patient's blood smear after recovery (Wright–Giemsa stain, $\times 1000$).

roles in innate immunity, and activation of these cells can lead to inflammation. Neutrophils and macrophages concentrate at the injured or infected site, releasing toxic substances such as oxygen-free radicals to damage the invading pathogens and simultaneously phagocytize pathogens and damaged tissues.⁷ Furthermore, it has been reported that the thrombocytopenia associated with increased lactate dehydrogenase (LDH) levels is often caused by infection.8 In this case, the observations of increased LDH levels and gram-negative bacterial infection were consistent with changes in platelet counts (Table 2). In summary, it was likely that cerebral hemorrhage and gram-negative bacterial infection led to these overreactions that sensitized the platelets, resulting in their phagocytosis by leukocytes.

To confirm thrombocytopenia in this case, the possibility of reduced platelet counts caused by the use of the anticoagulant EDTA-K2 had to be discounted. Therefore, in this study, sodium citrate and heparin were used as anticoagulants to detect platelets. The results showed that platelet counts were still depressed, and leukocyte-phagocytized platelets were detected.

The pathogenic mechanism of human granulocvtic anaplasmosis (HGA) is mainly that the function of neutrophils is damaged following infection by pathogens, resulting in abnormal adhesion, migration, neutrophil degranulation, and phagocytosis, which in turn cause a series of immunopathological changes.9 In this case, the patient had fever and thrombocytopenia, and the possibility of Anaplasma phagocytophilum infection existed. However, considering that the cause of the disease was clear in this case, HGA could be excluded.

The etiology of hemophagocytic lymphohistiocytosis (HLH) is diverse, and infection is the most common cause.¹⁰ Because leukocyte-phagocytized platelets and consistent fever were detected in this case, the possibility of HLH had to be considered. This patient had normal triglyceride levels, his spleen could not be touched under the rib margin, his hemoglobin level exceeded 90 g/L, and his prognosis was good. Therefore, a HLH diagnosis was excluded.

Viral infection can cause platelet aggregation and destruction, resulting in thrombocytopenia. Acute thrombocytopenia is more common in children.¹¹ In this case, cerebral hemorrhage and gram-negative bacterial infection were clearly present, and thus, virus-induced thrombocytopenia was not considered.

Infection following cerebral hemorrhage has rarely been reported as a cause of thrombocytopenia associated with the phagocytosis of platelets by leukocytes in peripheral blood. We cannot clarify whether the stress of intracerebral hemorrhage or infection alone leads to platelet phagocytosis. It is unclear why thrombocytopenia caused by infection is common in such patients even though platelet phagocytosis is rare. However, few studies have examined thrombocytopenia caused by craniocerebral hemorrhage alone. It remains unknown whether platelet phagocytosis is caused by craniocerebral trauma and intracerebral hemorrhage alone. We hope this will be clarified in the future.

Declaration of conflicting interests

No conflict of interest exists in the submission of this manuscript, and the manuscript was approved for publication by all authors. Signed consent to publish from the patient has been obtained. We have de-identified all patient details.

Funding

This research received no specific grant from any funding agency in the public, commercial, or not-for-profit sectors.

ORCID iD

Xiaoyang Yang D https://orcid.org/0000-0002-4386-1535

Supplemental material

Supplemental material for this article is available online.

References

- Gagnier JJ, Kienle G, Altman DG, et al. The CARE guidelines: consensus-based clinical case reporting guideline development. *Headache* 2013; 53: 1541–1547. 2013/11/26. DOI: 10.1111/head.12246.
- Lad SP, Hegen H, Gupta G, et al. Proteomic biomarker discovery in cerebrospinal fluid for cerebral vasospasm following subarachnoid hemorrhage. J Stroke Cerebrovasc Dis 2012; 21: 30–41. 2010/09/21. DOI: 10.1016/j. jstrokecerebrovasdis.2010.04.004.
- Zhang X, Zhu M, Jiang XL, et al. P-selectin glycoprotein ligand 1 deficiency prevents development of acute pancreatitis by attenuating leukocyte infiltration. *World J Gastroenterol* 2020; 26: 6361–6377. 2020/11/ 28. DOI: 10.3748/wjg.v26.i41.6361.
- Cheng B, Li J, Peng L, et al. Efficacy and safety of restarting antiplatelet therapy for patients with spontaneous intracranial haemorrhage: A systematic review and metaanalysis. J Clin Pharm Ther 2021: 46: 957–965. 2021/02/05. DOI: 10.1111/jcpt.13377.
- 5. Gremmel T, Ay C, Riedl J, et al. Plateletspecific markers are associated with monocyte-platelet aggregate formation and thrombin generation potential in advanced atherosclerosis. *Thromb Haemost*

2016; 115: 615–621. 2015/10/16. DOI: 10.1160/th15-07-0598.

- Ollivier V, Roques C, Receveur N, et al. Bioreactivity of stent material: Activation of platelets, coagulation, leukocytes and endothelial cell dysfunction in vitro. *Platelets* 2017; 28: 529–539. 2016/12/30. DOI: 10.1080/09537104.2016.1252836.
- Navegantes KC, de Souza Gomes R, Pereira PAT, et al. Immune modulation of some autoimmune diseases: the critical role of macrophages and neutrophils in the innate and adaptive immunity. *J Transl Med* 2017; 15: 36. 2017/02/17. DOI: 10.1186/s12967-017-1141-8.
- Liu J, Fu H, Sun D, et al. Analysis of the laboratory indexes and risk factors in 189 cases of severe fever with thrombocytopenia syndrome. *Medicine (Baltimore)* 2020; 99: e18727. 2020/01/09. DOI: 10.1097/md.000 0000000018727.
- Lee SH, Shin NR, Kim CM, et al. First identification of Anaplasma phagocytophilum in both a biting tick Ixodes nipponensis and a patient in Korea: a case report. *BMC Infect Dis* 2020; 20: 826. 2020/11/13. DOI: 10.1186/ s12879-020-05522-5.
- Yang X, Zhang S, Huang D, et al. Treatment of refractory secondary hemophagocytic lymphohistiocytosis with umbilical cord mesenchymal stem cells. *J Int Med Res* 2019; 47: 2135–2144. 2019/04/10. DOI: 10.1177/0300060519836533.
- Alexandropoulou O, Kossiva L, Giannaki M, et al. The epidemiology, clinical course and outcome of febrile cytopenia in children. *Acta Paediatr* 2015; 104: e112–118. 2014/12/ 19. DOI: 10.1111/apa.12903.