

DNA Lesions, Inducible DNA Repair, and Cell Division: Three Key Factors in Mutagenesis and Carcinogenesis

by Bruce N. Ames,¹ Mark K. Shigenaga,¹ and Lois Swirsky Gold²

DNA lesions that escape repair have a certain probability of giving rise to mutations when the cell divides. Endogenous DNA damage is high: 10^6 oxidative lesions are present per rat cell. An exogenous mutagen produces an increment in lesions over the background rate of endogenous lesions. The effectiveness of a particular lesion depends on whether it is excised by a DNA repair system and the probability that it gives rise to a mutation when the cell divides. When the cell divides, an unrepaired DNA lesion has a certain probability of giving rise to a mutation. Thus, an important factor in the mutagenic effect of an exogenous agent whether it is genotoxic or non-genotoxic, is the increment it causes over the background cell division rate (mitogenesis) in cells that appear to matter most in cancer, the stem cells, which are not on their way to being discarded. Increasing their cell division rate increases mutation and therefore cancer. There is little cancer from nondividing cells. Endogenous cell division rates can be influenced by hormone levels, decreased by calorie restriction, or increased by high doses of chemicals. If both the rate of DNA lesions and cell division are increased, then there will be a multiplicative effect on mutagenesis (and carcinogenesis), for example, by high doses of a mutagen that also increases mitogenesis through cell killing. The defense system against reactive electrophilic mutagens, such as the glutathione transferases, are also almost all inducible and buffer cells against increments in active forms of chemicals that can cause DNA lesions. A variety of DNA repair defense systems, almost all inducible, buffer the cell against any increment in DNA lesions. Therefore, the effect of a particular chemical insult depends on the level of each defense, which in turn depends on the past history of exposure. Exogenous agents can influence the induction and effectiveness of these defenses. Defenses can be partially disabled by lack of particular micronutrients in the diet (e.g., antioxidants).

Endogenous DNA Damage and Mutagenesis

Endogenous rates of DNA damage are enormous. Mutagens are often thought to be only exogenous agents, but endogenous mutagens cause extensive DNA damage (oxidative and other lesions), some of which is converted to mutations during cell division.

Four endogenous processes leading to significant DNA damage are oxidation (1-3), methylation, deamination, and depurination (2). The importance of these processes is supported by the existence of specific DNA repair glycosylases for oxidative, methylated, and deaminated adducts and a repair system for apurinic sites that are produced by spontaneous depurination (3).

DNA damage produced by oxidation appears to be the most significant endogenous damage (4). We estimate that the DNA hits per cell per day from endogenous oxidants are normally 10^5 in the rat and 10^4 in the human (5-7). These oxidative lesions are effectively but not perfectly repaired; the normal steady-state level of oxidative DNA lesions is about 10^6 per cell in the young rat and about twice this in the old rat (6,8). Oxidants are produced as byproducts of mitochondrial electron transport, various oxygen-utilizing enzyme system, peroxisomes, and other processes associated with normal aerobic metabolism, as well as by lipid

¹Division of Biochemistry and Molecular Biology, 401 Barker Hall, University of California, Berkeley, CA 94720.

²Life Sciences Division, Lawrence Berkeley Laboratory, Berkeley, CA 94720.

Address reprint requests to B.N. Ames, Division of Biochemistry and Molecular Biology, 401 Barker Hall, University of California, Berkeley, CA 94720.

This paper was presented at the Symposium on Cell Proliferation and Chemical Carcinogenesis that was held January 14-16, 1992, in Research Triangle Park, NC.

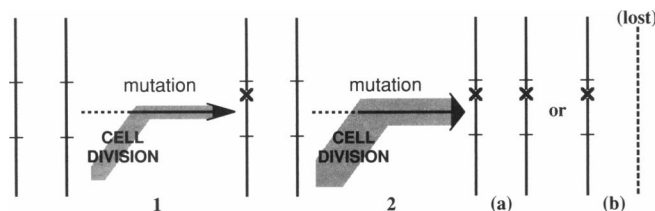


FIGURE 1. Mitogenesis (induced cell division) is a major multiplier of endogenous (or exogenous) DNA damage leading to mutation. The pathway to inactivating (x) both copies of a recessive tumor-suppressor gene is shown (two vertical lines represent the pair of chromosomes carrying the genes). Cell division increases mutagenesis due to DNA adducts converted to mutations before they are repaired (1 and 2a); mutations due to DNA replication (1 and 2a); replicating DNA is more vulnerable to damage (1 and 2a). Mitotic recombination (2a), gene conversion (2a), and nondisjunction (2b) are more frequent, and the first two give rise to the same mutations on both chromosomes. This diagram does not attempt to deal with the complex mutational pathway to tumors (164,165).

peroxidation. Oxidants that escape the numerous antioxidant defenses can damage cellular macromolecules, including DNA, and such damage can lead to mutations and cancer.

We have argued that this oxidative DNA damage is a major contributor to aging and the degenerative diseases associated with aging (4,9). Because of the high background of endogenous DNA lesions, any agent causing chronic mitogenesis can be indirectly mutagenic, and consequently carcinogenic, by increasing the probability of these endogenous DNA lesions being converted to mutations (Fig. 1). Furthermore, endogenous rates of DNA damage are so high that at low doses that do not increase mitogenesis, it may be difficult for exogenous mutagens to make a significant increment in the total DNA damage. Dietary micronutrients, e.g., antioxidants, are necessary for cellular defense systems; deficiency of micronutrients can alter the balance between damage and defense and thus can increase DNA lesion rates. Oxidants are also important in mitogenesis (wound healing) and progression (see below).

Mitogenesis Increases Mutagenesis

When the cell divides, an unrepaired DNA lesion has a certain probability of giving rise to a mutation. Thus, an important factor in the mutagenic effect of an exogenous agent, whether it is genotoxic or nongenotoxic, is the increment it causes over the background cell division rate (mitogenesis) (10). There is little cancer from nondividing cells. The time interval for DNA repair during cell division is short, and lesions can be converted to point mutations or gaps during replication. In support of higher "spontaneous" mutation rates in dividing cells is the observation that background hypoxanthine phosphoribosyltransferase mutations that arise *in vivo* in human T-lymphocytes arise preferentially in dividing T-cells (11–14).

During cell division, single-stranded DNA is without base-pairing, nucleosomes, or histones and is thus more sensitive to damage than double-stranded DNA. Cell division triggers mitotic recombination, gene conversion, and nondisjunction, which together seem more effective than an independent second mutation (15–19) in converting a heterozygous recessive gene (e.g., p53, a tumor-suppressor gene) to homo- or hemizygosity. Thus, the second mutational step toward cancer is more dependent on mitogenesis than the first. Heterozygotes at the human HLA-A gene are spontaneously converted to homozygotes during cell division (20). The above mechanisms could account for gross chromosomal alterations that occur frequently in human tumors (21–27). Cell division allows gene duplication, which can increase expression of oncogenes that are otherwise weakly expressed (28).

Cell division (i.e., through mitotic recombination) and DNA damage (29) would be expected to increase the rate of loss of 5-methylcytosine. Epigenetic changes in DNA, such as 5-methylcytosine levels, appear to be important in turning off genes in differentiation and could play a role in both cancer (30–34) and aging (31,35). It has been observed that the 5-methylcytosine level decreases with age (36), and it is known that cells dedifferentiate with age (37,38).

Those cells that appear to be most important for cancer are the stem cells, which are not on their way to being discarded. Increasing the division rate of stem cells increases mutation and therefore cancer. In dividing cells such as in epithelial tissues, one important unknown factor is to what extent there is a queue of reserve stem cells to replace stem cells that are programmed to die (apoptosis) after a certain number of divisions. A recent study on transformation of ovarian cells examined various factors and emphasized the role of cell division (39). Thus, understanding cell division rates, apoptosis, cell lineages, and differentiation are all relevant to mutagenesis and carcinogenesis (40). Endogenous cell division rates can be influenced by hormone levels, decreased by calorie restriction, or increased by high doses of chemicals. If the rates of both DNA lesions and cell division are increased, there will be a multiplicative effect on mutagenesis (and carcinogenesis), for example, by high doses of a mutagen that also increase mitogenesis through cell killing.

Mitogenesis and Carcinogenesis

Epidemiological studies, a large body of experimental evidence, and theoretical work on the mechanisms of carcinogenesis point to mitogenesis as a major contributor to cancer.

Suppression of Intercellular Communication Causes Mitogenesis. At near-toxic doses, some chemicals interfere with cell-cell communication in quiescent tissues (e.g., the liver, the major target site for carcinogenesis in rodents), thereby causing mito-

genesis and carcinogenesis (41–43). Trosko and his associates (41,42) have shown that suppression of gap junctional intercellular communication, which is associated with an increase in Ha-ras expression in contact-inhibited cells, leads to mitogenesis (44). It is of great interest to identify chemicals causing mitogenesis at doses much below the maximum tolerated dose [e.g., certain peroxisome proliferators (45)].

Mitogenesis from Exogenous and Endogenous Factors. Chronic toxicity can cause injury to tissues, resulting in replacement cell division (46–50). In an experimental cancer model, the surgical removal of part of the liver causes neighboring cells to proliferate (46–48). The incidence of liver cancer is low in humans (but not in some strains of mice) unless the liver is chronically damaged. Viruses or alcohol excess, for example, cause damage to the liver, which is a risk factor for cancer. Salt excess is a risk factor in human stomach cancer because it causes mitogenesis (51–58). Chronic toxicity can also cause an inflammatory reaction because phagocytic cells unleash a barrage of oxidants in destroying dead cells at a wound. The oxidants produced are in part the same as those produced by ionizing radiation; therefore, chronic inflammation may be equivalent to irradiating the tissue (59). Nitrogen oxides from inflammation are also mutagens. The oxidants produced as a result of inflammation can stimulate proto-oncogenes and cell division (60–63). Chronic irritation and inflammation cause cancer in animals (64). Chronic inflammation is, as expected, a risk factor for human cancer (65–67); the carcinogenic effect of asbestos (68) and cigarette smoke, for example, may be due primarily to inflammation, which increases both mitogenesis and mutagenesis.

Chronic infection from viruses, bacteria, schistosomes, and other organisms that cause cell killing and consequent mitogenesis can be risk factors for cancer. Two examples are the human virus hepatitis B, a major cause of liver cancer in the world (69,70), and human papilloma virus 16, a risk factor for cervical cancer and one of whose major effects is to increase mitogenesis (71). In transgenic mice, overproduction of one protein of the hepatitis B virus, a surface antigen, results in cell turnover that causes all of the mice to develop hepatocellular carcinomas (72). Human T-cell lymphotropic virus type 1 causes constitutive expression of the T-cell interleukin-2 receptor. This may commit the cell to unremitting *in vivo* cell division with an increased likelihood for the occurrence of critical mutations leading to T-cell leukemia/lymphoma (73,74). Chronic *Helicobacter* (*Campylobacter*) infection is a risk factor for stomach cancer (75,78). Chronic schistosome infection is a risk factor for bladder and colorectal cancer (79). Dietary antioxidants would be expected to decrease both the mutagenic and proliferative effects of oxidants and therefore to lower tumor incidence.

Hormones can also cause mitogenesis, and hormone imbalances are major risk factors for a number of human cancers [e.g., breast cancer (80,82)]. Thus,

agents causing chronic mitogenesis can be proper carcinogens and are important in human cancer (80,82).

A calorie-restricted diet, compared to an *ad libitum* diet, significantly increases the life span of rats and mice and markedly decreases the cancer rate. It is striking that in calorie-restricted animals, mitogenesis rates are markedly lowered in a variety of tissues (83,84); this could in principle account for much of the decrease in the cancer rate (see below).

Thinking of chemicals as “initiators” or “promoters” confuses mechanistic issues of carcinogenesis (85). The idea that promoters are not in themselves carcinogens is not credible on mechanistic grounds and is not correct on experimental grounds (85–87). Every classical promoter that has been tested in a high-dose cancer test is a carcinogen (e.g., phenobarbital, catechol, TPA (88,89)). A promoter has been viewed as an agent that facilitates the development of tumors by selected growth of an initiated cell; however, a promoter can be viewed more accurately as inducing cell division and thus stimulating the rate of accumulation of key mutations that are needed to acquire a transformed phenotype. Thus, the very word promoter is confusing because mitogenesis is caused by one dose of a chemical and not by a lower dose. Nongenotoxic agents such as saccharin can be carcinogens at high doses just by causing cell killing and inducing chronic mitogenesis and inflammation, and the dose response would be expected to show a threshold (86,90–92).

Chronic mitogenesis alone can be a risk factor for cancer: theory predicts this and a large literature supports it (79,86). Work on radiation has also supported the idea that both mutagenesis and mitogenesis are important in tumor induction (93–96).

DNA Repair and Other Inducible Defenses

A variety of DNA repair defense systems, almost all inducible, protect the cell against any increment in DNA lesions by both exogenous and endogenous mutagens (97). The defense systems against reactive electrophilic mutagens, such as the glutathione transferases, are also almost all inducible and buffer cells against increases in active forms of chemicals that can cause DNA lesions (98,99). Therefore, the effect of a particular chemical insult is dependent on the level of each defense, which in turn is dependent on the past history of exposure. Thus, a small dose of a mutagen can protect against a subsequent challenge from a large dose, as has been shown for radiation (100) and a variety of other mutagens.

Different adducts are repaired with different effectiveness. Oxidative lesions, being so common in all aerobic creatures, may be particularly well repaired. However, even though radiation is an oxidative mutagen, it still can add to the background of DNA lesions to give increased mutation and cancer. Exogenous agents can influence the induction and effectiveness of

these defenses. Defenses can be partially disabled by lack of particular micronutrients in the diet [e.g., antioxidants (101)].

Mitogenesis and Animal Cancer Tests

Animal cancer tests are conducted at the maximum tolerated dose (MTD) and 1/2 the MTD of the test chemical for long periods of time—both high doses that can cause chronic mitogenesis (86,90,102). Future experimental work measuring mitogenesis will show how often chronic dosing at the MTD is like chronic wounding, which is known to increase tumor yields in rodent tests and to be a risk factor for cancer in humans (103). Our theoretical arguments (87,104,105) for taking mitogenesis into account in animal cancer tests help to explain many of the results that have been found in analyzing these tests, as discussed below.

To the extent that increases in tumor incidence in rodent studies are due to the secondary effects of administering high doses, then any chemical that increases mitogenesis (e.g., by chronic cell killing and cell replacement or by suppression of intercellular communication) may be a rodent carcinogen; thus, one would expect that a high proportion of chemicals tested at the MTD would be positive, and this is indeed what is found. In our large Carcinogenic Potency Database (CPDB), approximately half of the chemicals are positive in at least one test, and this proportion is similar for a variety of subsets of the CPDB (106–110). It is unlikely that the high proportion of carcinogens in rodent studies is due simply to selection of suspicious chemical structures because most chemicals were selected because of their use as industrial compounds, pesticides, drugs, or food additives. Moreover, historically, our knowledge to predict carcinogenicity has been inadequate (104).

Because mitogenesis indirectly increases mutation, one would expect that in animal tests at the MTD, nongenotoxic carcinogens are likely to be acting by this mechanism (86). Indeed, among chemicals in the CPDB that have been tested adequately in rats and mice, we find that half of the nonmutagens in *Salmonella* are carcinogens, and approximately 45% of the carcinogens are not mutagenic (110).

Mitogenesis at the MTD is an important, and possibly the dominant, factor in carcinogenesis for mutagens. Mutagenic chemicals, because they directly damage DNA, are generally more effective at killing cells at high doses (with consequent cell replacement) than nonmutagens and thus are more effective at causing mitogenesis. Mutagens, unlike nonmutagens, can damage DNA as well as increase mitogenesis at high doses, giving a multiplicative interaction for carcinogenesis. This would lead one to expect three results that are, in fact, found in analyses of animal cancer tests: Mutagens, compared to nonmutagens, are *a*) more likely to be carcinogenic (104,111,112), *b*) more likely to be

positive in both rats and mice (104,111,112), and *c*) more likely to induce tumors in multiple organs (112).

One would not expect the mutagenicity of a chemical in *Salmonella* to always indicate the mechanism in a rodent. In the CPDB, of 384 chemicals tested for both carcinogenicity in rats and mice and mutagenicity in *Salmonella*, 26% of the noncarcinogens are mutagens in *Salmonella*; these presumably are not acting as significant mutagens in the rodents (110). Additionally, some nonmutagens in *Salmonella* may be indirectly mutagenic in higher organisms, for example, peroxisome proliferators. Even those mutagens that are carcinogens may not all be acting as genotoxins in animals because of detoxification and other processes. The importance of mitogenesis, even for mutagens, has been shown in experiments with pairs of mutagenic isomers (1 versus 2-nitropropane and 2,4, versus 2,6-diaminotoluene). In each pair only one chemical was a carcinogen, and only the carcinogen was mitogenic (113,114).

Several recent analyses of dose response in animal tests are consistent with the idea that mitogenesis from cell killing and cell replacement at the high doses tested is important. In the usual experimental design of dosing at the MTD and 1/2 MTD, both dose levels are high and may result in mitogenesis. Even at these two high doses, we have found that 44% of the positive sites in National Toxicology Program bioassays are statistically significant at the MTD but not at 1/2 MTD (among 365 positive sites). Moreover, the proportion positive only at the high dose is similar for mutagens and nonmutagens (Gold et al., unpublished data). Another analysis of the shape of dose response curves indicates that a quadratic dose response is compatible with more of the data than a linear one for both mutagens and nonmutagens (115). That mitogenesis at near-toxic doses is important in the carcinogenic response at the MTD is also suggested by the lack of chemicals that are highly carcinogenic relative to their MTD. Such chemicals would be expected to produce very high tumor incidence rates at the MTD; however, in animal cancer tests at the MTD it is uncommon to find 100% of the animals developing tumors (102). Moreover, if chemicals were highly carcinogenic relative to their MTD, then the dose-response curve might plateau well below the MTD. However, we found that only 10% of the dose-response functions indicate a possible plateau (a leveling off of the dose response). Moreover, even for these 10% there was a lack of consistency: for the compounds in which an apparent plateau was observed in one site, the result was generally not replicated in other target sites in the same experiment, in other sex of the same species, or in other species (102).

In this paper, we have mainly discussed theoretical issues. The pioneering work on this subject by experimentalists is discussed in detail in other pages in this issue.

It is clear that the mechanisms of action for all rodent carcinogens are not the same. For some chemicals there is evidence to support mitogenesis effects

unique to high doses, for example, formaldehyde, melamine, and saccharin. For others (e.g., butadiene), carcinogenic effects have been found considerably below the MTD. Further studies of mechanism in rodent bioassays should help to clarify such differences. Adding routine measurements of mitogenesis to the 13-week toxicology study and the 2-year bioassay would provide information that would improve dose setting, interpretation of experimental results, and risk assessment.

As currently conducted, standard rodent bioassays do not provide sufficient information to assess carcinogenic risk to humans at doses thousands of times below the MTD. If mitogenesis is a dominant factor in carcinogenesis at the MTD, then at low doses where mitogenesis is not generally induced, the hazards to humans of rodent carcinogens may be much lower than commonly assumed. Defenses are inducible at low doses, and even for mutagens the increment in DNA damage over the enormous rate of endogenous background damage may not be significant.

Toxicological examination of synthetic chemicals such as pesticides and industrial pollutants, without similar examination of chemicals that occur naturally, has resulted in an imbalance in both data and perception about chemical carcinogens. About 80% of the chemicals tested in both rats and mice are synthetic. Yet, there is an enormous background of natural chemicals in the diet such as plant pesticides and the products of cooking that have not been a focus of carcinogenicity testing (105). Regulatory policy to prevent human cancer has primarily addressed synthetic chemicals, yet similar proportions of natural chemicals and synthetic chemicals test positive in rodent studies, as expected from an understanding of toxicological defenses (116). The vast proportion of human exposures are to natural chemicals [e.g., 99.99% by weight of pesticides humans ingest are natural (105)]. Thus, rodent carcinogens that humans are exposed to are ubiquitous (105,117). Natural chemicals are the experimental control for evaluating cancer-regulatory strategies for synthetic pollutants. Possible hazards from residues of synthetic chemicals should be routinely compared to the possible hazards from natural chemicals. We have found that when the same index is used for natural and synthetic chemicals, possible carcinogenic hazards from current levels of pesticide residues or water pollution are likely to be of minimal concern relative to the background levels of natural substances (117,118). Extrapolation to low dose human exposure from results of animal cancer tests done at high dose should be based on knowledge about mechanisms of carcinogenesis for each chemical, particularly mitogenic effects that are present at high but not at low doses.

Progression and Clonal Instability

One of the hallmarks of latter stages of cancer is a genetic instability that leads through a succession of

stages ("progression") to a metastasizing, aggressive tumor (119). Numerous authors have discussed progression in terms of a possible clonal instability (119-121). Because there is a balance between a high endogenous oxidant production and extensive oxidative defenses, a plausible way to get a clone with a high mutation rate is to upset this balance. Because oxidants are both mitogens [the wound healing response (122-124)] and mutagens, they furnish the selective growth and instability needed for progression to a malignant tumor. Recent work supports this idea: tumor cell lines from various organs produce excess H_2O_2 (125); breast tumors show high levels of oxidative DNA damage (126), and a number of tumors have altered antioxidant defenses (127-133). There is a large literature on antioxidants as anticarcinogens (134,135). Antioxidants have been shown to be effective in suppressing all stages of carcinogenesis (135,136).

Factors Decreasing Mitogenesis, Mutagenesis, Carcinogenesis

Dietary Imbalances and Degenerative Diseases.

The high endogenous level of oxidative DNA lesions reinforces evidence from epidemiology that deficiency of antioxidants (137-139) is likely to be an important risk factor for cancer. Epidemiologists have been accumulating evidence that unbalanced diets are major contributors to heart disease and cancer and are likely to be as important as smoking. It has been estimated that approximately 30% of all cancers are related to diet (140). The main culprit appears to be a dietary imbalance of too few fruits and vegetables and too much fat (137-139). Particular micronutrients in fruits and vegetables that appear to be important in disease prevention are antioxidants (carotenoids, tocopherols, ascorbate) and folic acid, but many more vitamins and essential minerals may also be of interest (137-139, 141). Micronutrients are components of defenses against oxidants and other endogenous mutagens that contribute to the degenerative diseases associated with aging, such as cancer, heart disease, and cataracts. Because endogenous oxidative DNA damage is enormous, there are good theoretical reasons for thinking that antioxidants should be as important as they are being found to be. Surveys have indicated that 91% of the U.S. population is not eating sufficient fruits and vegetables; for example, almost half the population had eaten neither fruits nor vegetables on the day of the survey (142).

A study of sperm DNA (101) indicates that levels of oxidative DNA damage are inversely correlated with ascorbate concentrations in seminal fluid which, in turn, are related to the amount of dietary ascorbate consumed. In 10 individuals whose dietary intake of ascorbate was started at 250 mg/day for 2 weeks and then reduced to 10 or 20 mg/day, the level of ascorbate in seminal fluid fell to one-fourth, and the levels of oxidative lesions in sperm DNA were increased 2.5

times; repletion of dietary ascorbate led to a decrease in lesion levels closer to those before ascorbate deprivation. In a separate group consisting of 24 free-living subjects, the steady-state levels of lesions were also found to correlate inversely with content of ascorbate in seminal plasma. These results demonstrate the protective effect of dietary ascorbate against oxidative damage to human sperm DNA. This protective effect may be especially relevant to about one third of the male population who have low ascorbate levels because of poor diets or smoking (139,143,144). Their progeny may be at a higher risk for birth defects and childhood cancer. It seems likely that the men's somatic cells are being mutated as well as their sperm.

Work showing that folate deficiency in mice results in chromosome breakage (145) reinforces studies indicating that folate deficiency is an important cause of chromosome breaks, cancer, and birth defects (137,138). Again, a sizeable proportion of the population (30% or more) may not be ingesting sufficient folate. Hypomethylation of DNA, which is associated with folate deficiency, might also contribute to epigenetic effects, such as dedifferentiation of cells, which occurs during tumorigenesis (146,147).

In the quest to delay aging and prevent cancer and heart disease, it is important to understand what level of each micronutrient is optimal for long-term effects. The RDA (U.S. Recommended Daily Allowance) for micronutrients is based on the level necessary to prevent an immediate pathological effect, but the optimal levels that maybe needed to suppress tumor formation are likely to be much higher. The great genetic variability of the human species makes it likely that the optimal RDA for particular micronutrients will be higher than average for many people. Techniques for noninvasive measurement of DNA damage in humans are clearly relevant (148,149).

Calorie Restriction Lowers Mitogenesis Rates.

In rodents, a high-calorie diet appears to be carcinogenic (150–152). A calorie-restricted diet, compared to an *ad libitum* diet, significantly increases the life span of rats and mice and dramatically decreases the cancer rate. Though a causative mechanism to account for the beneficial effects of calorie restriction has not been established, several physiological indexes are altered in rodents that are fed restricted diets. Calorie restriction activates the pituitary adrenocorticotrophic axis, resulting in a decrease in the release of reproductive and mitogenic hormones. It has been suggested that Darwinian fitness will be increased if reproductive function is delayed during periods of low food availability (153) and that the saved resources will be invested in maintaining the body until food resources are available for successful reproduction (154). If one accepts the concept of a trade-off between reproduction and maintenance as predicted by evolutionary biology (155), it becomes evident why calorie restriction is so effective in reducing cancer. Decreases in mitogenic hormones such as insulin, thyroid-stimulating hor-

mone, growth hormone, estrogen, and prolactin decrease the likelihood of hormone-induced cancers, as has been shown in various animal studies (156). The markedly lowered mitogenesis rates observed in a variety of tissues of calorie-restricted relative to *ad libitum* rodents (83,84) is consistent with suppression of mitogenic hormones (157) and decreased protooncogene expression (158). The lowered incidence of mammary tumors observed in calorie-restricted rats has been attributed to reduced circulating levels of the mammatropic hormones estrogen and prolactin (159). Thus, the decrease in mitogenesis rates in calorie-restricted rats is likely to account for much of the decrease in tumor incidence.

The suggestion that maintenance functions are enhanced in calorie-restricted rats is supported by the findings of more efficient DNA repair (160), better coupled mitochondrial respiration (161), and a delay in the age-dependent decline of antioxidant defenses (162). The overall effect of these enhanced maintenance activities would be a reduction in oxidative damage to cellular macromolecules, and a decrease in somatic mutations that are induced by endogenous oxidative mutagens. The higher levels of antioxidant defenses in calorie-restricted rodents could account for an enhanced immune response (163) that sometimes suppresses tumor formation.

This work was supported by the National Cancer Institute Outstanding Investigator Grant CA 39910 and by the National Institute of Environmental Health Sciences Center Grant ES01896 to B.N.A. and by the Lawrence Berkeley Laboratory through U.S. Department of Energy contract DE-AC-03-76SF00098 and U.S. Environmental Protection Agency agreement R-815619-01-0 to L.S.G. M.K.S. was supported by an NIA Postdoctoral Fellowship and by the University of California Toxic Substances Program. This paper was adapted in part from papers of Ames and Shigenaga (4), and Ames and Gold (9,86).

REFERENCES

1. Ames, B. N. Dietary carcinogens and anti-carcinogens (oxygen radicals and degenerative diseases). *Science* 221: 1256–1264 (1983).
2. Saul, R. L., and Ames, B. N. Background levels of DNA damage in the population. In: *Mechanisms of DNA Damage and Repair: Implications for Carcinogenesis and Risk Assessment* (M. Simic, L. Grossman, and A. Upton, Eds.), Plenum Press, New York, 1986, pp. 529–536.
3. Lindahl, T. DNA-repair enzymes. *Annu. Rev. Biochem.* 51: 61–87 (1982).
4. Ames, B. N., and Shigenaga, M. K. Oxidants are a major contributor to aging. In: *Annals of the N.Y. Academy of Sciences*. Vol. 663 (C. Franceschi, G. Crepaldi, V. J. Cristafalo, L. Masotti, and J. Vijg, Eds.), N.Y. Acad. Sci., New York, 1992, pp. 85–96.
5. Ames, B. N. Endogenous oxidative DNA damage, aging, and cancer. *Free Rad. Res. Commun.* 7:121–128 (1989).
6. Fraga, C. G., Shigenaga, M. K., Park, J.-W., Degan, P., and Ames, B. N. Oxidative damage to DNA during aging: 8-hydroxy-2'-deoxyguanosine in rat organ DNA and urine. *Proc. Natl. Acad. Sci. U.S.A.* 87: 4533–4537 (1990).
7. Shigenaga, M. K., Gimeno, C. J., and Ames, B. N. Urinary 8-hydroxy-2'-deoxyguanosine as a biological marker of *in vivo*

- oxidative DNA damage. *Proc. Natl. Acad. Sci. U.S.A.* 86: 9697-9701 (1989).
8. Richter, C., Park, J.-W., and Ames, B. N. Normal oxidative damage to mitochondrial and nuclear DNA is extensive. *Proc. Natl. Acad. Sci. U.S.A.* 85: 6465-6467 (1988).
9. Ames, B. N., and Gold, L. S. Endogenous mutagens and the causes of aging and cancer. *Mutat. Res.* 250: 3-16 (1991).
10. Tong, C., Fazio, M., and Williams, G. M. Cell cycle-specific mutagenesis at the hypoxanthine phosphoribosyltransferase locus in adult rat liver epithelial cells. *Proc. Natl. Acad. Sci. U.S.A.* 77: 7377-7389 (1980).
11. Nicklas, J. A., O'Neill, J. P., Sullivan, L. M., Hunter, T. C., Allegretta, M., Chastenay, B. V., Libbus, B. L., and Albertini, R. J. Molecular analyses of *in vivo* hypoxanthine-guanine phosphoribosyltransferase mutations in human T-lymphocytes II. Demonstration of a clonal amplification of *hprt* mutant T-lymphocytes *in vivo*. *Environ. Mol. Mutagen.* 12: 271-284 (1988).
12. Nicklas, J. A., Hunter, T. C., O'Neill, J. P., and Albertini, R. J. Molecular analyses of *in vivo* *hprt* mutations in human T-lymphocytes III. Longitudinal study of *hprt* gene structural alterations and T-cell clonal origins. *Mutat. Res.* 215: 147-160 (1989).
13. Albertini, R. J., O'Neill, J. P., Nicklas, J. A., Allegretta, M., Recio, L., and Skopek, T. R. Molecular and clonal analysis of *in vivo* *hprt* mutations in human cells. In: *Human Carcinogen Exposure: Biomonitoring and Risk Assessment* (R. C. Garner, P. D. Farmer, G. T. Steel, and A. S. Wright, Eds.), Oxford University Press, Oxford, England, 1991, pp. 103-125.
14. Allegretta, M., Nicklas, J. A., Sriram, S., and Albertini, R. J. T cells responsive to myelin basic protein in patients with multiple sclerosis. *Science* 247: 718-721 (1990).
15. Schiestl, R. H., Gietz, R. D., Mehta, R. D., and Hastings, P. J. Carcinogens induce intrachromosomal recombination in yeast. *Carcinogenesis* 10: 1445-1455 (1989).
16. Liskay, R. M., and Stachelek, J. L. Evidence of intrachromosomal gene conversion in cultured mouse cells. *Cell* 35: 157-165 (1983).
17. Fahrig, R. The effect of dose and time on the induction of genetic alterations in *Saccharomyces cerevisiae* by aminoacridines in the presence and absence of visible light irradiation in comparison with the dose-effect-curves of mutagens with other types of action. *Mol. Gen. Genet.* 144: 131-140 (1976).
18. Ramel, C. Short-term testing—are we looking at wrong endpoints? *Mutat. Res.* 205: 13-24 (1988).
19. Grodon, J., Nakamura, Y., and German, J. Molecular evidence that homologous recombination occurs in proliferating human somatic cells. *Proc. Natl. Acad. Sci. U.S.A.* 87: 4315-4319 (1990).
20. Cavenee, W. K., Dryja, T. P., Phillips, R. A., Benedict, W. F., Godbout, R., Gallie, B. L., Murphree, A. L., Strong, L. C., and White, R. L. Expression of recessive alleles by chromosomal mechanisms in retinoblastoma. *Nature* 305: 779-784 (1983).
21. Cavenee, W. K., Hansen, M. F., Nordenskjold, M., Kock, E., Maumenee, I., Squire, J. A., Phillips, R. A., and Gallie, B. L. Genetic origin of mutations predisposing to retinoblastoma. *Science* 228: 501-503 (1985).
22. Hansen, M. F., and Cavenee, W. K. Genetics of cancer predisposition. *Cancer Res.* 47: 5518-5527 (1987).
23. Sasaki, M., Okamoto, M., Sato, C., Sugio, K., Soejima, J., Iwama, T., Ikeuchi, T., Tonomura, A., Miyaki, M., and Sasazuki, T. Loss of constitutional heterozygosity in colorectal tumors from patients with familial polyposis coli and those with nonpolyposis colorectal carcinoma. *Cancer Res.* 49: 4402-4406 (1989).
24. Erisman, M. D., Scott, J. K., and Astrin, S. M. Evidence that the familial adenomatous polyposis gene is involved in a subset of colon cancers with a completable defect in *c-myc* regulation. *Proc. Natl. Acad. Sci. U.S.A.* 86: 4264-4268 (1989).
25. Vogelstein, B., Fearon, E. R., Hamilton, S. R., Kern, S. E., Preisinger, A. C., Leppert, M., Nakamura, Y., White, R., Smits, A. M. M., and Bos, J. L. Genetic alterations during colorectal-tumor development. *N. Engl. J. Med.* 319: 525-532 (1988).
26. Vogelstein, B., Fearon, E. R., Kern, S. E., Hamilton, S. R., Preisinger, A. C., Nakamura, Y., and White, R. Allelotype of colorectal carcinomas. *Science* 244: 207-211 (1989).
27. Turner, D. R., Grist, S. A., Janatipour, M., and Morley, A. A. Mutations in human lymphocytes commonly involve gene duplication and resemble those seen in cancer cells. *Proc. Natl. Acad. Sci. U.S.A.* 85: 3189-3192 (1988).
28. Orr-Weaver, T. L., and Spradling, A. C. *Drosophila* chorion gene amplification requires an upstream region regulating *s18* transcription. *Mol. Cell. Biol.* 6: 4624-4633 (1986).
29. Cannon, S. V., Cummings, A., and Teebor, G. W. 5-Hydroxymethylcytosine DNA glycosylases activity in mammalian tissue. *Biochem. Biophys. Res. Commun.* 151: 1173-1179 (1988).
30. Fearon, E. F., Cho, K. R., Nigro, J. M., Kern, S. E., Simons, J. W., Ruppert, J. M., Hamilton, S. R., Preisinger, A. C., Thomas, G., Kinzler, K. W., and Vogelstein, B. Identification of a chromosome 18q gene that is altered in colo-rectal cancer. *Science* 247: 49-56 (1990).
31. Holliday, R. DNA methylation and epigenetic defects in carcinogenesis. *Mutat. Res.* 181: 215-217 (1987).
32. Vorce, R. L., and Goodman, J. I. Hypomethylation of *ras* oncogenes in chemically induced and spontaneous B6C3F₁ mouse liver tumors. *J. Toxicol. Environ. Health* 34: 367-384 (1991).
33. Vorce, R. L. and Goodman, J. I. Altered methylation of *ras* oncogenes in benzdine-induced B6C3F₁ mouse liver tumors. *Toxicol. Appl. Pharmacol.* 100: 398-410 (1989).
34. Goodman, J. I., Ward, J. M., Popp, J. A., Klaunig, J. E., and Fox, T. R. Mouse liver carcinogenesis: mechanisms and relevance. *Fundam. Appl. Toxicol.* 17: 651-665 (1991).
35. Holliday, R. The inheritance of epigenetic defects. *Science* 238: 163-169 (1987).
36. Wilson, V. L., Smith, R. A., Ma, S., and Cutler, R. G. Genomic 5-methyldeoxycytidine decreases with age. *J. Biol. Chem.* 262: 9948-9951 (1987).
37. Harman, D. The aging process. *Proc. Natl. Acad. Sci. U.S.A.* 78: 7124-7128 (1981).
38. Hartman, P. E. Mutagens: Some possible health impacts beyond carcinogenesis. *Environ. Mutagen.* 5: 139-152 (1983).
39. Godwin, A. K., Testa, J. R., Handel, L. M., Liu, Z., Vanderveer, L. A., Tracey, P. A., and Hamilton, T. C. Spontaneous transformation of rat ovarian surface epithelial cells—association with cytogenetic changes and implications of repeated ovulation in the etiology of ovarian cancer. *J. Natl. Cancer Inst.* 84: 592-601 (1992).
40. Irons, R. D., Stillman, W. S., Colagiovanni, D. B., and Henry, V. A. Synergistic action of the benzene metabolite hydroquinone on myelopoietic stimulating activity of granulocyte/macrophage colony-stimulating factor *in vitro*. *Proc. Natl. Acad. Sci. U.S.A.* 89: 3691-3695 (1992).
41. Trosko, J. E. Towards understanding carcinogenic hazards: a crisis in paradigms. *J. Am. Coll. Toxicol.* 89: 1121-1132 (1989).
42. Trosko, J. E., Chang, C. C., and Madhukar, B. V. *In vitro* analysis of modulators of intercellular communications: implications for biologically based risk assessment models for chemical exposure. *Toxicol. In Vitro* 4: 635-643 (1990).
43. Yamasaki, H., Enomoto, K., Fitzgerald, D. J., Mesnil, M., Katoh, F., and Hollstein, M. Role of intercellular communication in the control of critical gene expression during multistage carcinogenesis. In: *Cell Differentiation, Genes and Cancer* (T. Kakunaga, T. Sugimura, L. Tomatis, and H. Yamasaki, Eds.), IARC Scientific Publication No. 92, International Agency for Research on Cancer, Lyon, France, 1988, pp. 57-75.
44. de Feijter, A. W., Trosko, J. E., Krizman, D. B., Lebovitz, R. M., and Lieberman, M. W. Correlation of increased levels of Ha-*ras* T24 protein with extent of loss of gap junction function in rat liver epithelial cells. *Mol. Carcinog.* 5: 205-212 (1992).
45. Wada, N., Marsman, D. S., and Popp, J. A. Dose-related effects of the hepatocarcinogen, WY-14,643, on peroxisomes and cell replication. *Fundam. Appl. Toxicol.* 18: 149-154 (1992).
46. Farber, E. Clonal adaptation during carcinogenesis. *Biochem. Pharmacol.* 39: 1837-1846 (1990).
47. Farber, E., Parker, S., and Gruenstein, M. The resistance of putative premalignant liver cell populations, hyperplastic nodules, to the acute cytotoxic effects of some hepatocarcinogens. *Cancer Res.* 36: 3879-3887 (1976).

48. Farber, E. Cellular biochemistry of the stepwise development of cancer with chemicals: G.H.A. Clowes Memorial Lecture. *Cancer Res.* 44: 5463-5474 (1984).
49. Mirsalis, J. C., and Steinmetz, K. L. The role of hyperplasia in liver carcinogenesis. In: *Mouse Liver Carcinogenesis: Mechanisms and Species Comparisons* (D. Stevenson, Ed.), Wiley-Liss, New York, 1990, pp. 149-161.
50. Mirsalis, J. C., Tyson, C. K., Steinmetz, K. L., Loh, E. K., Hamilton, C. M., Bakke, J. P., and Spalding, J. W. Measurement of unscheduled DNA synthesis and S-phase synthesis in rodent hepatocytes following *in vivo* treatment: testing of 24 compounds. *Environ. Mol. Mutagen.* 14: 155-164 (1989).
51. Joossens, J. V. and Geboers, J. Nutrition and gastric cancer. *Nutr. Cancer* 2: 250-261 (1981).
52. Furihata, C., Sato, Y., Hosaka, M., Matsushima, T., Furukawa, F., and Takahashi, M. NaCl induced ornithine decarboxylase and DNA synthesis in rat stomach mucosa. *Biochem. Biophys. Res. Commun.* 121: 1027-1032 (1984).
53. Tuyns, A. J. Salt and gastrointestinal cancer. *Nutr. Cancer* 11: 229-232 (1988).
54. Lu, J.-B. and Qin, Y.-M. Correlation between high salt intake and mortality rates for oesophageal and gastric cancers in Henan Province, China. *Int. J. Epidemiol.* 16: 171-176 (1987).
55. Furihata, C., Sudo, K., and Matsushima, T. Calcium chloride inhibits stimulation of replicative DNA synthesis by sodium chloride in the pyloric mucosa of rat stomach. *Carcinogenesis* 10: 2135-2137 (1990).
56. Coggon, D., Barker, D. J. P., Cole, R. B., and Nelson, M. Stomach cancer and food storage. *J. Natl. Cancer Inst.* 81: 1178-1182 (1989).
57. Charnley, G., and Tannenbaum, S. R. Flow cytometric analysis of the effect of sodium chloride on gastric cancer risk in the rat. *Cancer Res.* 45: 5608-5616 (1985).
58. Karube, T., Katayama, H., Takemoto, K., and Watanabe, S. Induction of squamous metaplasia, dysplasia and carcinoma *in situ* of the mouse tracheal mucosa by inhalation of sodium chloride mist following subcutaneous injection of 4-nitroquinoline 1-oxide. *Jpn. J. Cancer Res.* 80: 698-701 (1989).
59. Ward, J. F., Limoli, C. L., Calabro-Jones, P., and Evans, J. W. Radiation vs. chemical damage to DNA. In: *Anticarcinogenesis and Radiation Protection* (P. A. Cerutti, O. F. Nygaard, and M. G. Simic, Eds.) Plenum Press, New York, 1987, pp. 321-327.
60. Crawford, D., and Cerutti, P. Expression of oxidant stress-related genes in tumor promotion of mouse epidermal cells JB6. In: *Anticarcinogenesis and Radiation Protection* (O. Nygaard, M. Simic and P. Cerutti, Eds.), Plenum Press, New York, 1989, pp. 183-190.
61. Chan, T. M., Chen, E., Tatoyan, A., Shargill, N. S., Pleta, M., and Hochstein, P. Stimulation of tyrosine-specific protein phosphorylation in the rat liver plasma membrane by oxygen radicals. *Biochem. Biophys. Res. Commun.* 139: 439-445 (1986).
62. Craven, P. A., Pfanstiel, J., and DeRubertis, F. R. Role of activation of protein kinase C in the stimulation of colonic epithelial proliferation and reactive oxygen formation by bile acids. *J. Clin. Invest.* 79: 532-541 (1987).
63. Sieweke, M. H., Stoker, A. W., and Bissell, M. J. Evaluation of the cocarcinogenic effect of wounding in Rous Sarcoma virus tumorigenesis. *Cancer Res.* 49: 6419-6424 (1989).
64. Madsen, C. Squamous-cell carcinoma and oral, pharyngeal and nasal lesions caused by foreign bodies in food. Cases from a long-term study in rats. *Lab. Anim.* 23: 241-247 (1989).
65. Demopoulos, H. B., Pietronigro, D. D., Flamm, E. S., and Seligman, M. L. The possible role of free radical reactions in carcinogenesis. *J. Environ. Pathol. Toxicol.* 3: 273-303 (1980).
66. Templeton, A. Pre-existing, non-malignant disorders associated with increased cancer risk. *J. Environ. Pathol. Toxicol.* 3: 387-397 (1980).
67. Lewis, J. G., and Adams, D. O. Inflammation, oxidative DNA damage, and carcinogenesis. *Environ. Health Perspect.* 76: 19-27 (1987).
68. Petruska, J., Marsh, J. P., Kagan, E., and Mossman, B. T. Release of superoxide (O_2^-) by cells obtained from bronchoalveolar lavage (BAL) after exposure of rats to either crocidolite or chrysotile asbestos. *Am. Rev. Respir. Dis.* 137: 403 (1988).
69. Yeh, F.-S., Mo, C.-C., Luo, S., Henderson, B. E., Tong, M. J., and Yu, M. C. A serological case-control study of primary hepatocellular carcinoma in Guangxi, China. *Cancer Res.* 45: 872-873 (1985).
70. Wu, T. C., Tong, M. J., Hwang, B., Lee, S.-D., and Hu, M. M. Primary hepatocellular carcinoma and hepatitis B infection during childhood. *Hepatology* 7: 46-48 (1987).
71. Peto, R., and zur Hausen, H., Eds. *Viral Etiology of Cervical Cancer* Banbury Report 21, Cold Spring Harbor Laboratory, Cold Spring Harbor, NY, 1986.
72. Dunsford, H. A., Sell, S., and Chisari, F. V. Hepatocarcinogenesis due to chronic liver cell injury in hepatitis B virus transgenic mice. *Cancer Res.* 50: 3400-3407 (1990).
73. Inoue, J., Seiki, M., Taniguchi, T., Tsuru, S., and Yoshida, M. Induction of interleukin 2 receptor gene expression by p40x encoded by human T-cell leukemia virus type 1. *EMBO J.* 5: 2883-2888 (1986).
74. Taniguchi, T., Yamada, G., Shibuya, H., Maruyama, M., Harada, H., Hatakeyama, M., and Fujita, T. Regulation of the interleukin-2 system and T cell neoplasm. In: *Cell Differentiation, Genes and Cancer* (T. Kakunaga, T. Sugimura, L. Tomatis, and H. Yamasaki, Eds.), IARC Publication, International Agency for Research on Cancer, Lyon, France, 1988, pp. 181-184.
75. Rathbone, B. J., and Heatley, R. V., Eds. *Campylobacter pylori and Gastrointestinal Disease*. Blackwell, Oxford, 1989.
76. Blaser, M. J., Ed. *Campylobacter pylori in Gastritis and Peptic Ulcer Disease*. Igaku-Shoin, New York, 1989.
77. Wotherspoon, A. C., Ortiz, H. C., Falzon, M. R., and Isaacson, P. G. *Helicobacter pylori*-associated gastritis and primary B-cell gastric lymphoma. *Lancet* 338: 1175-1176 (1991).
78. Parsonnet, J., Friedman, G. D., Vandersteen, D. P., Chang, Y., Vogelmann, J. H., Orentreich, N., and Sibley, R. K. *Helicobacter pylori* infection and risk of gastric carcinoma. *N. Engl. J. Med.* 325: 1127-1131 (1991).
79. Preston-Martin, S., Pike, M. C., Ross, R. K., Jones, P. A., and Henderson, B. E. Increased cell division as a cause of human cancer. *Cancer Res.* 50: 7415-7421 (1990).
80. Preston-Martin, S., Pike, M. C., Ross, R. K., and Henderson, B. E. Epidemiologic evidence for the increased cell proliferation model of carcinogenesis. In: *Chemically Induced Cell Proliferation: Implications for Risk Assessment* (B. Butterworth, T. Slaga, W. Farland, and M. McClain, Eds.), Wiley-Liss, New York, 1991, pp. 21-34.
81. Henderson, B. E., Ross, R., and Bernstein, L. Estrogens as a cause of human cancer: The Richard and Hinda Rosenthal Foundation Award Lecture. *Cancer Res.* 48: 246-253 (1988).
82. Henderson, B. E., Ross, R. K., and Pike, M. C. Toward the primary prevention of cancer. *Science* 254: 1131-1138 (1991).
83. Lok, E., Scott, F. W., Mongeau, R., Nera, E. A., Malcolm, S., and Clayton, D. B. Calorie restriction and cellular proliferation in various tissues of the female Swiss Webster mouse. *Cancer Lett.* 51: 67-73 (1990).
84. Heller, T. D., Holt, P. R., and Richardson, A. Food restriction retards age-related histological changes in rat small intestine. *Gastroenterology* 98: 387-391 (1990).
85. Iversen, O. H., Ed. *Theories of Carcinogenesis*. Hemisphere Publishing, Washington, DC, 1988.
86. Ames, B. N., and Gold, L. S. Chemical carcinogenesis: too many rodent carcinogens. *Proc. Natl. Acad. Sci. U.S.A.* 87: 7772-7776 (1990).
87. Ames, B. N., and Gold, L. S. Too many rodent carcinogens: mitogenesis increases mutagenesis. *Science* 249: 970-971 (1990).
88. IARC. Phenobarbital and phenobarbital sodium. In: *IARC Monographs on the Evaluation of Carcinogenic Risk of Chemicals to Man. Some Miscellaneous Pharmaceutical Substances*. International Agency for Research on Cancer, Lyon, France, 1977, pp. 157-181.
89. Ingram, A. J., and Grasso, P. Evidence for and possible mechanisms of non-genotoxic carcinogenesis in mouse skin. *Mutat. Res.* 248: 333-340 (1991).
90. Butterworth, B. E. Chemically induced cell proliferation as a predictive assay for potential carcinogenicity. In: *Chemically*

- Induced Cell Proliferation: Implications for Risk Assessment, B. E. Butterworth, and T. J. Slaga, Eds.), Wiley-Liss, New York, 1991, pp. 457-467.
91. Cohen, S. M., and Ellwein, L. B. Cell proliferation in carcinogenesis. *Science* 249: 1007-1011 (1990).
 92. Cohen, S. M., and Ellwein, L. B. Genetic errors, cell proliferation, and carcinogenesis. *Cancer Res.* 51: 6493-6505 (1991).
 93. Jones, T. D., Griffin, G. D., and Walsh, P. J. A unifying concept for carcinogenic risk assessments. *J. Theor. Biol.* 105: 35-61 (1983).
 94. Jones, T. D. A unifying concept for carcinogenic risk assessments: comparison with radiation-induced leukemia and mice and men. *Health Phys.* 4: 533-558 (1984).
 95. Little, J. B., Kennedy, A. R., and McGandy, R. B. Effect of dose rate on the induction of experimental lung cancer in hamsters by radiation. *Radiat. Res.* 103: 293-299 (1985).
 96. Ootsuyama, A., and Tanooka, H. Threshold-like dose of local irradiation repeated throughout the life span of mice for induction of skin and bone tumors. *Radiat. Res.* 125: 98-101 (1991).
 97. Luethy, J. D., and Holbrook, N. J. Activation of *gadd153* promoter by genotoxic agents: rapid and specific response to DNA damage. *Cancer Res.* 52: 5-10 (1992).
 98. Friling, R. S., Bergelson, S., and Daniel, V. Two adjacent AP-1-like binding sites form the electrophile responsive element of the murine glutathione S-transferase Ya subunit gene. *Proc. Natl. Acad. Sci. U.S.A.* 89: 668-672 (1992).
 99. Talalay, P. Mechanisms of induction of enzymes that protect against chemical carcinogenesis. In: *Advances in Enzyme Regulation* (G. Weber, Ed.), Pergamon Press, Oxford, 1989, pp. 237-250.
 100. Kelsey, K. T., Memisoglu, A., Frenkel, D., and Liber, H. L. Human lymphocytes exposed to low doses of X-rays are less susceptible to radiation-induced mutagenesis. *Mutat. Res.* 263: 197-201 (1991).
 101. Fraga, C. G., Motchnik, P. A., Shigenaga, M. K., Helbock, H. J., Jacob, R. A., and Ames, B. N. Ascorbic acid protects against endogenous oxidative damage in human sperm. *Proc. Natl. Acad. Sci. U.S.A.* 88: 11003-11006 (1991).
 102. Bernstein, L., Gold, L. S., Ames, B. N., Pike, M. C., and Hoel, D. G. Some tautologous aspects of the comparison of carcinogenic potency in rats and mice. *Fundam. Appl. Toxicol.* 5: 79-86 (1985).
 103. Weitzman, S. A., and Gordon, L. I. Inflammation and cancer: role of phagocyte-generated oxidants in carcinogenesis. *Blood* 76: 655-663 (1990).
 104. Gold, L. S., Bernstein, L., Magaw, R., and Slone, T. H. Interspecies extrapolation in carcinogenesis: prediction between rats and mice. *Environ. Health Perspect.* 81: 211-219 (1989).
 105. Ames, B. N., Profet, M., and Gold, L. S. Dietary pesticides (99.99% all natural). *Proc. Natl. Acad. Sci. U.S.A.* 87: 7777-7781 (1990).
 106. Gold, L. S., Sawyer, C. B., Magaw, R., Backman, G. M., de Veciana, M., Levinson, R., Hooper, N. K., Havender, W. R., Bernstein, L., Peto, R., Pike, M. C. and Ames, B. N. A carcinogenic potency database of the standardized results of animal bioassays. *Environ. Health Perspect.* 58: 9-319 (1984).
 107. Gold, L. S., de Veciana, M., Backman, G., Magaw, R., Lopipero, P., Smith, M., Blumenthal, M., Levinson, R., Gerson, J., and Ames, B. N. Chronological supplement to the Carcinogenic Potency Database: standardized results of animal bioassays published through December 1982. *Environ. Health Perspect.* 67: 161-200 (1986).
 108. Gold, L. S., Slone, T. H., Backman, G. M., Magaw, R., Costa, M. D., Lopipero, P., Blumenthal, M., and Ames, B. N. Second chronological supplement to the Carcinogenic Potency Database: standardized results of animal bioassays published through December 1984 and by the National Toxicology Program through May 1986. *Environ. Health Perspect.* 74: 237-329 (1987).
 109. Gold, L. S., Slone, T. H., Backman, G. M., Eisenberg, S., Costa, M. D., Wong, M., Manley, N. B., Rohrbach, L., and Ames, B. N. Third chronological supplement to the Carcinogenic Potency Database: standardized results of animal bioassays published through December 1986 and by the National Toxicology Program through June 1987. *Environ. Health Perspect.* 84: 215-286 (1990).
 110. Gold, L. S., Manley, M. B., Rohrbach, L., Slone, T. H., Garfinkel, G. B., and Ames, B. N. The fifth plot of the Carcinogenic Potency Database: results of animal bioassays published in the general literature through 1988 and by the National Toxicology Program through 1989. *Environ. Health Perspect.* 100: 65-135 (1993).
 111. Gold, L. S., Slone, T. H., Stern, B. R., and Bernstein, L. Comparison of target organs of carcinogenicity for mutagenic and non-mutagenic chemicals. *Mutat. Res.* 286: 75-100 (1993).
 112. Ashby, J. and Tennant, R. W. Chemical structure, *Salmonella* mutagenicity and extent of carcinogenicity as indicators of genotoxic carcinogenesis among 222 chemicals tested in rodents by the U.S. NCI/NTP. *Mutat. Res.* 204: 17-115 (1988).
 113. Cunningham, M. L., and Matthews, H. B. Relationship of hepatocarcinogenicity and hepatocellular proliferation induced by mutagenic noncarcinogens vs. carcinogens. II. 1- vs. 2-nitropropane. *Toxicol. Appl. Pharmacol.* 110: 505-513 (1991).
 114. Cunningham, M. L., Foley, J., Maronpot, R., and Matthews, H. B. Correlation of hepatocellular proliferation with hepatocarcinogenicity induced by the mutagenic noncarcinogen: carcinogen pair-2,6 and 2,4-diaminotoluene. *Toxicol. Appl. Pharmacol.* 107: 562-567 (1991).
 115. Hoel, D. G., and Portier, C. J. Nonlinearity of dose-response functions for carcinogenicity. *Environ. Health Perspect.*, in press.
 116. Ames, B. N., Profet, M., and Gold, L. S. Nature's chemicals and synthetic chemicals: comparative toxicology. *Proc. Natl. Acad. Sci. U.S.A.* 87: 7782-7786 (1990).
 117. Gold, L. S., Slone, T. H., Stern, B. R., Manley, N. B., and Ames, B. N. Rodent carcinogens: setting priorities. *Science* 258: 261-265 (1992).
 118. Ames, B. N., Magaw, R., and Gold, L. S. Ranking possible carcinogenic hazards. *Science* 236: 271-280 (1987).
 119. Loeb, L. A. Mutator phenotype may be required for multistage carcinogenesis. *Cancer Res.* 51: 3075-3079 (1991).
 120. Strauss, B. S. The origin of point mutations in human tumor cells. *Cancer Res.* 52: 249-253 (1992).
 121. Battle, A., and Riley, P. A. Abnormality of heme synthesis as the initiating lesion in carcinogenesis. *Cancer J.* 4: 326-331 (1991).
 122. Amstad, P., Peskin, A., Shah, G., Mirault, M. E., Moret, R., Zbinden, I., and Cerutti, P. The balance between Cu,Zn-superoxide dismutase and catalase affects the sensitivity of mouse epidermal cells to oxidative stress. *Biochemistry* 30: 9305-9313 (1991).
 123. Cerutti, P. A. Oxidant stress and carcinogenesis. *Eur. J. Clin. Invest.* 21: 1-5 (1991).
 124. Cerutti, P. A., and Trump, B. F. Inflammation and oxidative stress in carcinogenesis. *Cancer Cells* 3: 1-7 (1991).
 125. Szatrowski, T. P., and Nathan, C. F. Production of large amounts of hydrogen peroxide by human tumor cells. *Cancer Res.* 51: 794-798 (1991).
 126. Malins, D. C., and Haimanot, R. Major alterations in the nucleotide structure of DNA in cancer of the female breast. *Cancer Res.* 51: 5430-5432 (1991).
 127. Borunov, E. V., Smirnova, L. P., Schepetkin, I. A., Lankin, V. Z., and Vasil'ev, N. V. High activity of antioxidant enzymes in a tumor as a factor of "avoidance of control" in the immune system. *Biull. Eksp. Biol. Med.* 107: 467-469 (1989).
 128. McCormick, M. L., Oberley, T. D., Elwell, J. H., Oberley, L. W., Sun, Y., and Li, J. J. Superoxide dismutase and catalase levels during estrogen-induced renal tumorigenesis, in renal tumors and their autonomous variants in the Syrian hamster. *Carcinogenesis* 12: 977-983 (1991).
 129. St. Clair, D. K., and Oberley, L. W. Manganese superoxide dismutase expression in human cancer cells: a possible role of mRNA processing. *Free Rad. Res. Commun.* 12-13: 771-778 (1991).
 130. Galeotti, T., Masotti, L., Borrello, S., and Casali, E. Oxy-radical metabolism and control of tumour growth. *Xenobiotica* 21: 1041-1051 (1991).
 131. Cheeseman, K. H., Emery, S., Maddix, S. P., Slater, T. F., Burton, G. W., and Ingold, K. U. Studies on lipid peroxidation in

- normal and tumor tissues. The Yoshida rat liver tumour. *Biochem. J.* 250: 247-252 (1988).
132. Bartoli, G. M., Bartoli, S., Galeotti, R., and Bertoli, E. Superoxide dismutase content and microsomal lipid composition of tumors with different growth rates. *Biochem. Biophys. Acta* 620: 205-211 (1980).
 133. de Vries, E. G., Meijer, C., Timmer-Bosscha, H., Berendsen, H. H., de Leij, L., Scheper, R. J., and Mulder, N. H. Resistance mechanisms in three human small cell lung cancer cell lines established from one patient during clinical follow-up. *Cancer Res.* 49: 4175-4178 (1989).
 134. Hasegawa, R., Tiwawech, D., Hirose, M., Takaba, K., Hoshiya, T., Shirai, T., and Ito, N. Suppression of diethylnitrosamine-initiated preneoplastic foci development in the rat liver by combined administration of four antioxidants at low doses. *Jpn. J. Cancer Res.* 83: 431-437 (1992).
 135. Perchellet, J. P., and Perchellet, E. M. Antioxidants and multi-stage carcinogenesis in mouse skin. *Free Rad. Biol. Med.* 7: 377-408 (1989).
 136. Kuroda, Y., Shankel, D. M., and Waters, M. D., Eds. *Antimutagenesis and Anticarcinogenesis Mechanisms II*. Plenum Press, New York, 1990.
 137. Bendich, A., and Butterworth, C. E., Eds. *Micronutrients in Health and in Disease Prevention*. Marcel Dekker, New York, 1991.
 138. National Research Council, *Diet and Health: Implications for Reducing Chronic Disease Risk*. National Academy Press, Washington, DC 1989.
 139. Block, G. Epidemiologic evidence regarding vitamin C and cancer. *Am. J. Clin. Nutr.* 54: 1310S-1314S (1991).
 140. Doll, R., and Peto, R. The causes of cancer. Quantitative estimates of avoidable risks of cancer in the United States today. *J. Natl. Cancer Inst.* 66: 1191-1308 (1981).
 141. Reddy, B. S., and Cohen, L. A., Eds. *Diet, Nutrition, and Cancer: A Critical Evaluation*. CRC Press, Boca Raton, FL, 1986.
 142. Patterson, B. H., and Block, G. Fruit and vegetable consumption national survey data. In: *Micronutrients in Health and in Disease Prevention* (A. Bendich and C. E. Butterworth, Eds.), Marcel Dekker, New York, 1991, pp. 409-436.
 143. Block, G., Patterson, B., and Subar, A. Fruit, vegetables and cancer prevention: a review of the epidemiologic evidence. *Nutr. Cancer* 18: 1-29 (1992).
 144. Schectman, G., Byrd, J. C., and Hoffmann, R. Ascorbic acid requirements for smokers: analysis of a population survey. *Am. J. Clin. Nutr.* 53: 1466-1470 (1991).
 145. MacGregor, J. T., Schlegel, R., Wehr, C. M., Alperin, P., and Ames, B. N. Cytogenetic damage induced by folate deficiency in mice is enhanced by caffeine. *Proc. Natl. Acad. Sci. U.S.A.* 87: 9962-9965 (1990).
 146. Bhawe, M. R., Wilson, M. J., and Poirier, L. A. c-H-ras and c-K-ras gene hypomethylation in the livers and hepatomas of rats fed methyl-deficient, amino acid-defined diets. *Carcinogenesis* 9: 343-348 (1988).
 147. Cravo, M., Mason, J., Salomon, R. N., Ordovas, J., Osada, J., Selhub, J., and Rosenberg, J. H. Folate deficiency in rats causes hypomethylation of DNA. *FASEB J.* 5: A914 (1991).
 148. Degan, P., Shigenaga, M. K., Park, E.-M., Alperin, P. E., and Ames, B. N. Immunoaffinity isolation of urinary 8-hydroxy-2'-deoxyguanosine and 8-hydroxyguanine and quantitation of 8-hydroxy-2'-deoxyguanosine in DNA by polyclonal antibodies. *Carcinogenesis* 12: 865-871 (1991).
 149. Park, E. M., Shigenaga, M. K., Degan, P., Korn, T. S., Kitzler, J. W., Wehr, C. M., Kolachana, P., and Ames, B. N. The assay of excised oxidative DNA lesions: isolation of 8-oxoguanine and its nucleoside derivatives from biological fluids with a monoclonal antibody column. *Proc. Natl. Acad. Sci. U.S.A.* 89: 3375-3379 (1992).
 150. Boutwell, R. K., and Pariza, M. W. Historical perspective: calories and energy expenditure in carcinogenesis. *Am. J. Clin. Nutr.* 45 (Suppl 1): 151-156 (1987).
 151. Roe, F. J. C. Non-genotoxic carcinogenesis: Implications for testing extrapolation to man. *Mutagenesis* 4: 407-411 (1989).
 152. Roe, F. J. C., Lee, P. N., Conybeare, G., Tobin, G., Kelly, D., Prentice, D., and Matter, B. Risks of premature death and cancer predicted by body weight in early adult life. *Hum. Exp. Toxicol.* 10: 285-288 (1991).
 153. Holehan, A. M., and Merry, B. J. Modification of the oestrous cycle hormonal profile by dietary restriction. *Mech. Ageing Dev.* 32: 63-76 (1985).
 154. Holliday, R. Food, reproduction and longevity: is the extended lifespan of calorie-restricted animals an evolutionary adaptation? *Bioassays* 10: 125-127 (1989).
 155. Williams, G. C. Pleiotropy, natural selection and the evolution of senescence. *Evolution* 11: 398-411 (1957).
 156. Ruggeri, B. The effects of calorie restriction on neoplasia and age-related degenerative processes. In: *Cancer and Nutrition. Human Nutrition: A Comprehensive Treatise* (R. B. Alfin-Slater and D. Dritzchevsky, Eds.), Plenum Press, New York, 1991, pp. 187-210.
 157. Armario, A., Montero, J. L., and Jolin, T. Chronic food restriction and circadian rhythms of pituitary-adrenal hormones, growth hormone and thyroid-stimulating hormone. *Ann. Nutr. Metab.* 31: 81-87 (1987).
 158. Nakamura, K. D., Duffy, P. H., Lu, M. H., Turturro, A., and Hart, R. W. The effect of dietary restriction on *myc* protooncogene expression in mice; a preliminary study. *Mech. Ageing Dev.* 48: 199-205 (1989).
 159. Sylvester, P. W., Aylsworth, C. F., Van Vugt, D. A., and Meites, J. Influence of underfeeding during the "critical period" or thereafter on carcinogen-induced mammary tumors in rats. *Cancer Res.* 42: 4943-4947 (1982).
 160. Weraarchakul, N., Strong, R., Wood, W. G., and Richardson, A. The effect of aging and dietary restriction of DNA repair. *Exp. Cell Res.* 181: 197-204 (1989).
 161. Weindruch, R. H., Cheung, M. K., Verity, M. A., and Walford, R. L. Modification of mitochondrial respiration by aging and dietary restriction. *Mech. Ageing Dev.* 12: 375-392 (1980).
 162. Koizumi, A., Weindruch, R., and Walford, R. L. Influences of dietary restriction and age in liver enzyme activities and lipid peroxidation in mice. *J. Nutr.* 117: 361-367 (1987).
 163. Kubo, C., Johnson, B. C., Day, N. K., and Good, R. A. Calorie source, calorie restriction, immunity and aging of (NZB/NZW) F1 mice. *J. Nutr.* 76: 1987 (1984).
 164. Kinzler, K. W., Nilbert, M. C., Vogelstein, B., Bryan, T. M., Levy, D. B., Smith, K. J., Preisinger, A. C., Hamilton, S. R., Hedge, P., Markham, A., Carlson, M., Joslyn, G., Groden, J., White, R., Miki, Y., Miyoshi, Y., Nishio, I., and Nakamura, Y. Identification of a gene located at chromosome 5q21 that is mutated in colorectal cancers. *Science* 251: 1366-1370 (1991).
 165. Weinberg, R. A. Oncogenes, antioncogenes, and the molecular bases of multistep carcinogenesis. *Cancer Res.* 49: 3713-3721 (1989).