



# Genome Sequence of a Potentially New *Buttiauxella* Species, Strain B2, Isolated from Rhizosphere of Olivillo Trees (*Aextoxicon punctatum*)

Romina Almasia,<sup>a</sup> Marlene Henríquez,<sup>a</sup> Arturo Levican,<sup>b</sup> Matías Poblete-Morales<sup>c</sup>

<sup>a</sup>Departamento I+D+i, Microagro SpA, Santiago, Chile

<sup>b</sup>Carrera Tecnología Médica, Facultad de Ciencias, Pontificia Universidad Católica de Valparaíso, Valparaíso, Chile

<sup>c</sup>Facultad de Ciencias de la Salud, Instituto de Ciencias Biomédicas, Universidad Autónoma de Chile, Santiago, Chile

Romina Almasia and Marlene Henríquez contributed equally to this article. Author order was determined alphabetically.

**ABSTRACT** We announce the draft genome sequence of strain B2, which belongs to a potentially new *Buttiauxella* species, isolated from soil associated with rhizosphere of olivillo trees (*Aextoxicon punctatum*). Its size is 4,967,099 bp, and its G+C content is 49.1%. The genome of strain B2 carries genes related to rhizobacteria that promote the growth of plants.

The genus *Buttiauxella* belongs to the family *Enterobacteriaceae* and includes Gram-negative motile rods which are facultative, anaerobic, and negative for oxidase (1). It was named in 1982 by Ferragut et al. (1) and currently comprises seven validly accepted species, *Buttiauxella agrestis*, *Buttiauxella brennerae*, *Buttiauxella ferragutiae*, *Buttiauxella gaviniae*, *Buttiauxella izardii*, *Buttiauxella noackiae*, and *Buttiauxella warmboldiae* (2). They have been isolated from mollusks and snails, as well as from water, food, soil, and human samples (2). The main characteristics that allow differentiating of the species of this genus are DNA-DNA hybridization characteristics and the guanine-plus-cytosine (G+C) ratio, which ranges from 48% to 50% (3). Recently, two potentially new species have been isolated from plants, one of them from the rhizosphere of a chrysanthemum (*Chrysanthemum* sp.) plantation in Brazil (*Buttiauxella chrysanthemi* sp. nov.) and the other from sedum (*Sedum alfredii*), a hyperaccumulator plant used for phytoremediation of heavy metal contamination in soil (strain SaSR13) (4, 5). The latter strain significantly improves plant growth, the development of the root, and the accumulation of cadmium in *Sedum alfredii* (5). Furthermore, among isolates from the genus *Buttiauxella*, there have been described phytases that allow the use of inorganic phosphate applicable as a nutritional supplement for animals (6). In fact, Ruangsanka (7), through the analysis of 16S rRNA gene sequences of bacteria associated with bamboo rhizosphere, observed that bacteria with the highest production of soluble phosphate belong to the genus *Buttiauxella*.

*Buttiauxella* sp. strain B2 was isolated from the rhizosphere of olivillo trees (*Aextoxicon punctatum*) in Cerro Santa Ines, Coquimbo Region, Chile (−32.1632, −71.4949; altitude, 663 m), in August 2015. For isolation, 20 g of roots with adhering soil was resuspended in 180 ml of sterile saline (0.85% NaCl). This mixture was homogenized in a previously sterilized glass blender three times for 1 min each with intermittent steps of placing the glass container in ice for 1 min. An aliquot of 1 ml of the supernatant was submitted to a serial 10-fold dilution in sterile saline. Then, 100 μl of each dilution was inoculated onto solid plates of LB medium (Beckton Dickinson, USA). The agar plates were incubated at 28°C for 72 hours, and single colonies were restreaked onto a new LB plate. The isolate was classified within the *Enterobacteriaceae* family through

**Citation** Almasia R, Henríquez M, Levican A, Poblete-Morales M. 2020. Genome sequence of a potentially new *Buttiauxella* species, strain B2, isolated from rhizosphere of olivillo trees (*Aextoxicon punctatum*). *Microbiol Resour Announc* 9:e01351-19. <https://doi.org/10.1128/MRA.01351-19>.

**Editor** Irene L. G. Newton, Indiana University, Bloomington

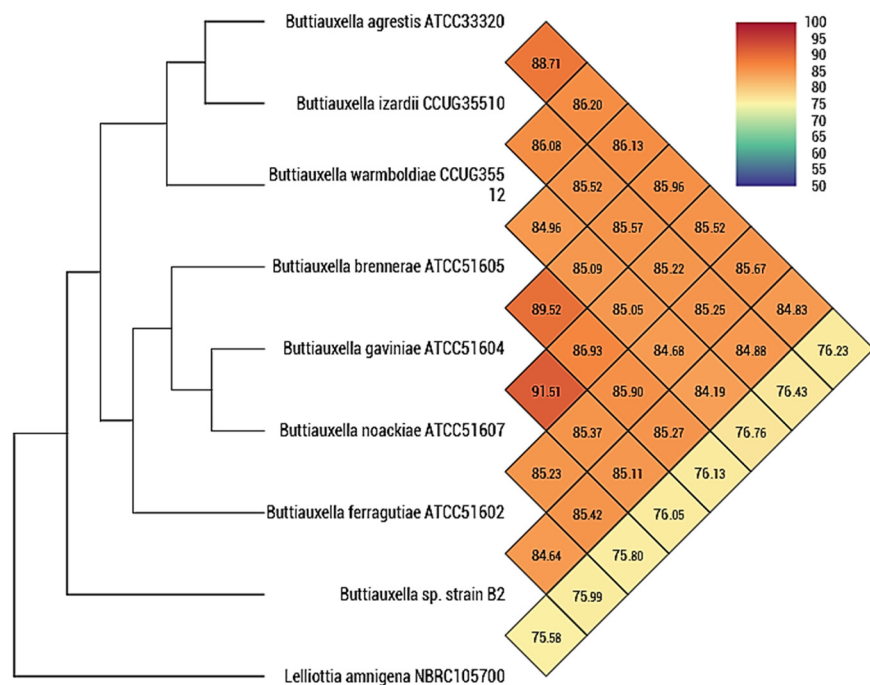
**Copyright** © 2020 Almasia et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Arturo Levican, [arturo.levican@pucv.cl](mailto:arturo.levican@pucv.cl), or Matías Poblete-Morales, [matias.poblete@uautonoma.cl](mailto:matias.poblete@uautonoma.cl).

**Received** 31 October 2019

**Accepted** 29 January 2020

**Published** 27 February 2020



**FIG 1** Heatmap showing the average nucleotide identity by orthology (OrthoANI) between strain B2 and all the type strains of the genus *Buttiauxella* calculated with OAT software with *Lelliottia amnigena* used as the outgroup.

phenotypic characterization, i.e., Gram-negative, motile rods which were negative for oxidase and positive for oxidation and fermentation of glucose in oxidation-fermentative (OF) medium. Furthermore, the 16S rRNA gene was amplified and sequenced as previously described (8). The sequence obtained (GenBank accession number [MN759732](https://www.ncbi.nlm.nih.gov/nuccore/MN759732)) showed an identity of 99.25% with bacteria of the genus *Buttiauxella* according to the online tool EzBioCloud (<https://www.ezbiocloud.net>).

For genomic DNA extraction, bacteria were grown on LB medium at 28°C for 24 hours under aerobic conditions, and then a single colony was picked and subcultured in 5 ml of LB broth medium, which was incubated overnight at 28°C in a shaker at 200 rpm. DNA was extracted using the Wizard genomic DNA purification kit (Promega Corp., USA) following the manufacturer's instructions for Gram-negative bacterial cells. The library was prepared with the TruSeq Nano DNA kit, and genomic sequencing was carried out with the Illumina MiSeq platform with 2 × 150-bp paired-end (PE) reads by Macrogen, Inc. (Seoul, South Korea). A total of 11,149,234 reads were obtained, and they were quality checked and filtered with the FastQC v0.11.5 application using the default parameters. K-mer analysis was performed in order to estimate coverage, heterozygosity, and genome size with Jellyfish v2.2.10 and GenomeScope (<http://qb.cshl.edu/genomescope/>). Then, *de novo* assembly was performed by using the filtered reads with SOAPdenovo2 v2.04. All of these basic bioinformatic analyses were performed by Macrogen, Inc. A total of 75 contigs were obtained ( $L_{50}$ , 8;  $N_{50}$ , 189,923 bp) with an average depth of 126.74×, and the estimated length of the chromosome was 4,967,099 bp with a G+C content of 49.1%.

Functional annotation of the assembled genome was performed with the Rapid Annotations using Subsystems Technology (RAST) server v2.0 (9) (<http://rast.nmpdr.org/rast.cgi>) via RASTtk (10). A total of 4,676 genes were identified, i.e., 4,610 coding sequences (CDS), 63 RNAs, and 3 rRNAs. PGAP annotation was also performed by default when genomes were deposited in NCBI (11). However, the annotations in RAST and PGAP were not compared, because that is beyond the scope of this study.

In order to determine the genomic identity of strain B2, the orthologous average nucleotide identity was performed to measure the overall similarity between genome

**TABLE 1** Genes associated with plant growth and phosphate-solubilizing traits (antiSMASH tool) present in the genome of *Buttiauxella* sp. strain B2

| Trait and category       | Subcategory                     | Subsystem                           | Role (EC no.)   | RAST features or no. of contigs/<br>start-stop position/strand<br>(antiSMASH tool)   |
|--------------------------|---------------------------------|-------------------------------------|---|--|
| Plant growth promotion   |                                 |                                     |   |  |
| Secondary metabolism     | Plant hormones                  | Auxin biosynthesis                  | Tryptophan synthase alpha chain (4.2.1.20)<br>Anthranilate phosphoribosyltransferase (2.4.2.18)<br>Tryptophan synthase beta chain (4.2.1.20)<br>Monoamine oxidase (1.4.3.4)<br>Phosphoribosylanthranilate isomerase (5.3.1.24)  | fig 6666666.374956.peg.200<br>fig 6666666.374956.peg.197<br>fig 6666666.374956.peg.199<br>fig 6666666.374956.peg.75<br>fig 6666666.374956.peg.198                                    |
| Miscellaneous            | Plant-prokaryote<br>DOE project | Single-rhodanese-domain<br>proteins | Thiosulfate sulfurtransferase GlpE (2.8.1.1)<br><br>Rhodanese domain protein, enterobacterial<br>subgroup, YceA homolog   | fig 6666666.374956.peg.4358<br><br>fig 6666666.374956.peg.1859   |
| Phosphate solubilization |                                 |                                     | Oligopeptide-binding protein (AppA)<br>Major phosphate-irrepressible acid phosphatase<br>Periplasmic nitrate reductase<br>Periplasmic nitrate reductase, electron<br>transfer subunit<br>Quinoprotein glucose dehydrogenase<br>Alpha-D-ribose 1-methylphosphonate<br>5-triphosphate synthase<br>subunit (PhnG)<br>Alpha-D-ribose 1-methylphosphonate<br>5-triphosphate synthase<br>subunit (PhnH)<br>Alpha-D-ribose 1-methylphosphonate<br>5-triphosphate synthase<br>subunit (PhnI)<br>Alpha-D-ribose 1-methylphosphonate<br>5-phosphate C-P lyase | 1/501295-502887/-<br>3/162750-164021/-<br>8/129512-131999/+<br>8/132010-132462/+<br>13/74571-76952/-<br>22/21340-21801/+<br>22/21798-22382/+<br>22/22382-23446/+<br>22/23439-24287/+ |

sequences using the OrthoANI v0.93.1 online tool available at EzBioCloud (<https://www.ezbiocloud.net/tools/orthoani>) (12). Low identity values were obtained between strain B2 and the type strain of the genus *Buttiauxella* (Fig. 1).

The genome of *Buttiauxella* sp. strain B2 encoded two genes classified in the category of plant-prokaryote by the U.S. Department of Energy (DOE) project. That project aims to determine the pangenomes of 100 to 200 species of soil- or plant-associated prokaryotes (<https://jgi.doe.gov/core-pangenomes-soil-plant-associated-prokaryotes/>). Furthermore, it encoded five genes associated with the production of plant hormones (Table 1). On the other hand, a strong capacity of phosphate solubilization was observed when culturing the bacteria in solid National Botanical Research Institute's phosphate growth medium (NBRIP) as previously described (13). Moreover, gene clusters associated with solubilization of phosphate were found in the genome by using the bacterial antiSMASH tool v3.0 with the default parameters (Table 1) (14, 15). Therefore, the presence of plant-associated genes, as well as the observed capacity of phosphate solubilization, warrants further studies to determine whether this bacterium possesses symbiotic interactions with the host species *Aextoxicon punctatum* and a biotechnological potential.

The genome sequence of strain B2 was compared with all available sequences of *Buttiauxella* spp. by using the OrthoANI standalone software tool that is freely available at <http://www.ezbiocloud.net/sw/oat> (12). OrthoANI is a robust and fast means of calculating average nucleotide identity for species delineation; it does not require gene-finding or functional annotation processes, allowing simple, reproducible, and standardized procedures (12). Although Lee et al. (12) recommended a 95% to 96% OrthoANI value threshold for species demarcation, the interspecies values between the accepted *Buttiauxella* spp. are even lower, i.e., 84.68% to 91.51% (Fig. 1). Along these lines, OrthoANI values between strain B2 and accepted species ranged from 84.19% to 85.42% (Fig. 1). Therefore, these results support the finding that strain B2 actually belongs to a potentially new *Buttiauxella* species, even though a future polyphasic

study is necessary to demonstrate this taxonomic position. Moreover, the presence of genes related to rhizobacteria that promote the growth of plants warrants future studies to determine their *in vivo* effects on the host species.

**Data availability.** This whole-genome sequence project has been deposited in GenBank under the accession number [VEXQ00000000](https://doi.org/10.1016/S0721-9571(81)80016-6). The version described in this paper is version VEXQ01000000. The BioProject number is [PRJNA548017](https://doi.org/10.1016/S0721-9571(81)80016-6), and the BioSample number is [SAMN11997788](https://doi.org/10.1016/S0721-9571(81)80016-6).

## ACKNOWLEDGMENTS

This work was supported by grants FIA PYT-2016-0486, Proyectos de Emprendimiento Innovador, Fondecyt Iniciación 11181262 by the Comisión Nacional de Ciencia y Tecnología (CONICYT), and a doctoral fellowship from Universidad Autónoma de Chile, Programa de Doctorado en Ciencias Biomédicas.

## REFERENCES

- Ferragut C, Izard D, Gavini F, Lefebvre B, Leclerc H. 1981. *Buttiauxella*, a new genus of the family Enterobacteraceae. Zentralbl Bakteriell Mikrobiol Hyg A 2:33–44. [https://doi.org/10.1016/S0721-9571\(81\)80016-6](https://doi.org/10.1016/S0721-9571(81)80016-6).
- Müller HE, Brenner DJ, Fanning GR, Grimont PAD, Kämpfer P. 1996. Emended description of *Buttiauxella agrestis* with recognition of six new species of *Buttiauxella* and two new species of *Kluyvera*: *Buttiauxella ferragutiae* sp. nov., *Buttiauxella gavinae* sp. nov., *Buttiauxella brennerae* sp. nov., *Buttiauxella izardii* sp. nov., *Buttiauxella noackiae* sp. nov., *Buttiauxella warmboldiae* sp. nov., *Kluyvera cochleae* sp. nov., and *Kluyvera georgiana* sp. nov. Int J Syst Bacteriol 46:50–63. <https://doi.org/10.1099/00207713-46-1-50>.
- Gavini F, Izard D, Ferragut C, Farmer JJ, Leclerc H. 1983. Separation of *Kluyvera* and *Buttiauxella* by biochemical and nucleic acid methods. Int J Syst Bacteriol 33:880–882. <https://doi.org/10.1099/00207713-33-4-880>.
- Furlan JPR, Braz VS, Paschoal JAR, Stehling EG. 2018. *Buttiauxella chrysanthemi* sp. nov., isolated from a chrysanthemum plantation in Brazil. Arch Microbiol 200:1365–1369. <https://doi.org/10.1007/s00203-018-1548-5>.
- Wu K, Luo J, Li J, An Q, Yang X, Liang Y, Li T. 2018. Endophytic bacterium *Buttiauxella* sp. SaSR13 improves plant growth and cadmium accumulation of hyperaccumulator *Sedum alfredii*. Environ Sci Pollut Res Int 25:21844–21854. <https://doi.org/10.1007/s11356-018-2322-6>.
- Dersjant-Li Y, Plumstead P, Awati A, Remus J. 2018. Productive performance of commercial growing and finishing pigs supplemented with a *Buttiauxella* phytase as a total replacement of inorganic phosphate. Anim Nutr 4:351–357. <https://doi.org/10.1016/j.aninu.2018.02.002>.
- Ruangsanka S. 2014. Identification of phosphate-solubilizing bacteria from the bamboo rhizosphere. ScienceAsia 40:204–211. <https://doi.org/10.2306/scienceasia1513-1874.2014.40.204>.
- Lane DJ. 1991. 16S/23S rRNA sequencing, p 115–175. In Stackebrandt E, Goodfellow M (ed), Nucleic acid techniques in bacterial systematics. John Wiley and Sons, New York, NY.
- Overbeek R, Olson R, Pusch GD, Olsen GJ, Davis JJ, Disz T, Edwards RA, Gerdes S, Parrello B, Shukla M, Vonstein V, Wattam AR, Xia F, Stevens R. 2014. The SEED and the Rapid Annotation of microbial genomes using Subsystems Technology (RAST). Nucleic Acids Res 42:D206–D214. <https://doi.org/10.1093/nar/gkt1226>.
- Brettin T, Davis JJ, Disz T, Edwards RA, Gerdes S, Olsen GJ, Olson R, Overbeek R, Parrello B, Pusch GD, Shukla M, Thomason JA, Stevens R, Vonstein V, Wattam AR, Xia F. 2015. RASTtk: a modular and extensible implementation of the RAST algorithm for building custom annotation pipelines and annotating batches of genomes. Sci Rep 5:8365.
- Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI Prokaryotic Genome Annotation Pipeline. Nucleic Acids Res 44:6614–6624. <https://doi.org/10.1093/nar/gkw569>.
- Lee I, Kim YO, Park SC, Chun J. 2016. OrthoANI: an improved algorithm and software for calculating average nucleotide identity. Int J Syst Evol Microbiol 66:1100–1103. <https://doi.org/10.1099/ijsem.0.000760>.
- Nautiyal CS. 1999. An efficient microbiological growth medium for screening phosphate solubilizing microorganisms. FEMS Microbiol Lett 170:265–270. <https://doi.org/10.1111/j.1574-6968.1999.tb13383.x>.
- Rodríguez H, Fraga R, Gonzalez T, Bashan Y. 2006. Genetics of phosphate solubilization and its potential for improving plant growth-promoting bacteria. Plant Soil 287:15–21. <https://doi.org/10.1007/s11104-006-9056-9>.
- Weber T, Blin K, Duddela S, Krug D, Kim HU, Bruccoleri R, Lee SY, Fischbach MA, Müller R, Wohlleben W, Breitling R, Takano E, Medema MH. 2015. antiSMASH 3.0: a comprehensive resource for the genome mining of biosynthetic gene clusters. Nucleic Acids Res 43:W237–W243. <https://doi.org/10.1093/nar/gkv437>.