



Draft Genome Sequence of Bioactive Strain *Streptomyces* sp. SMS_SU21, Isolated from Soil Sediment of the Sundarbans Mangrove Ecosystem

Sohan Sengupta,^a Arnab Pramanik,^a Pijush Basak,^a Maitree Bhattacharyya^{a,b}

^aJagadis Bose National Science Talent Search, Kolkata, West Bengal, India

^bDepartment of Biochemistry, University of Calcutta, Kolkata, West Bengal, India

ABSTRACT *Streptomyces* sp. SMS_SU21 possesses strong antimicrobial activity and antioxidant potential. This strain was isolated from the Sundarbans mangrove ecosystem, and its draft genome comprises 7,449,420 bp with 6,680 open reading frames. Genome analysis of strain SMS_SU21 provides insight into its secondary metabolite arsenal and reveals the gene clusters putatively responsible for its bioactive potential.

Mangrove streptomycetes are rich sources of natural products with significant biological activities and novel structures (1). Genome mining of mangrove streptomycetes accelerates the rapid discovery of useful products originating from them. In this study, *Streptomyces* sp. SMS_SU21 was isolated from the soil sediment of the Sundarbans mangrove ecosystem in India. This strain possesses potent antimicrobial activity against a broad spectrum of microorganisms, including multidrug-resistant strains and various phytopathogens (2). Interspecies competition within the residing microbial population is an obvious phenomenon in the Sundarbans, due to the rich index of species diversity and limited consumable nutrient sources (3) found there. Thus, in-depth information regarding the genomic edifice of this strain is required to understand its survival strategies in a competitive environment like the Sundarbans mangrove ecosystem.

Genomic DNA was extracted with a HiPurA streptomycetes DNA isolation and purification kit (Himedia, India). Shotgun sequencing was performed with a high-throughput HiSeq platform (Illumina) at AgriGenome Labs Private Limited in Kerala, India. Prior to whole-genome analysis, Cutadapt version 1.8 (4) was used to remove adapter sequences, and all low-quality data ($Q < 30$) were filtered out using Sickle version 1.33 (5). The cleaned reads were subjected to analysis with Kmergenie (6) to predict the optimal k -value and assembly size, which were found to be 31 and 7,449,420 bp, respectively. *De novo* assembly was performed using SPAdes version 3.9.0 (7), Velvet (8), and QUASt (9). The genome sequence of *Streptomyces* sp. SMS_SU21 was annotated using the Rapid Annotations using Subsystems Technology (RAST) server (10) and the NCBI Prokaryotic Genome Annotation Pipeline (PGAP) version 4.3 (11). The draft genome sequence of *Streptomyces* sp. SMS_SU21 constituted a total of 93 contigs ($>1,000$ bp), with a total size of 7,449,420 bp and a G+C content of 72.3%. The RAST server predicted 6,680 coding sequences, of which 2,208 (34%) were annotated as SEED subsystem features and 4,472 (66%) were annotated as outside the SEED subsystem; 3 rRNAs and 67 tRNAs were also predicted. The closest related type strains based on the 16S rRNA gene sequence are *S. griseorubens* NBRC 12780 (GenBank accession number AB184139), *S. althioticus* NRRL B-3981 (GenBank accession number AY999791), and *S. griseoincarnatus* LMG 19316 (GenBank accession number AJ781321), all with 99% sequence identity.

Received 31 May 2018 Accepted 1 June 2018 Published 5 July 2018

Citation Sengupta S, Pramanik A, Basak P, Bhattacharyya M. 2018. Draft genome sequence of bioactive strain *Streptomyces* sp. SMS_SU21, isolated from soil sediment of the Sundarbans mangrove ecosystem. Genome Announc 6:e00614-18. <https://doi.org/10.1128/genomeA.00614-18>.

Copyright © 2018 Sengupta et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Pijush Basak, pbasakjbnsts@gmail.com, or Maitree Bhattacharyya, bmaitree@gmail.com.

Secondary metabolite biosynthetic gene clusters (BGCs) were predicted using antiSMASH version 4.0 (12), which identified 24 putative BGCs in the genome. This includes nonribosomal peptide synthetase (NRPS) gene clusters, polyketide synthase (PKS), novel hybrid PKS-NRPS gene clusters, and other BGCs for producing siderophores, lantipeptides, lassopeptide, and bacteriocin. Numerous genes responsible for resistance to toxic compounds, including arsenic, mercury, cobalt, tellurium, and cadmium, were additionally detected. Hence, *Streptomyces* sp. SMS_SU21 may have great potential to produce exclusive bioactive natural compounds for clinical, industrial, and environmental applications.

Accession number(s). This whole-genome shotgun project has been deposited at DDBJ/ENA/GenBank under the accession number [PNRA00000000](https://www.ncbi.nlm.nih.gov/nuclink/PNRA00000000). The version described in this paper is the second version, PNRA02000000.

ACKNOWLEDGMENTS

We thank the World Bank, the ICZM project (54-ICZMP/3P), and DST-SERB for providing financial and instrumental support. Sohan Sengupta acknowledges UGC for providing his fellowship. Arnab Pramanik is supported by a research associateship from the ICZM project. Pijush Basak thanks the Science and Engineering Research Board (SERB), New Delhi, India, for providing the financial assistance in the form of grants and fellowships (project file number YSS/2015/001123/LS).

We are also grateful for the use of the instrument facility provided by UGC-CAS, DST-FIST, DBT-IPLS at the Department of Biochemistry, University of Calcutta, India, and to the local people of the Sundarbans for their moral support and active cooperation.

REFERENCES

- Xiao J, Wang Y, Luo Y, Xie S-J, Ruan J-S, Xu J. 2009. *Streptomyces avicenniae* sp. nov., a novel actinomycete isolated from the rhizosphere of the mangrove plant *Avicennia mariana*. *Int J Syst Evol Microbiol* 59:2624–2628. <https://doi.org/10.1099/ijs.0.009357-0>.
- Sengupta S, Pramanik A, Ghosh A, Bhattacharyya M. 2015. Antimicrobial activities of actinomycetes isolated from unexplored regions of Sundarbans mangrove ecosystem. *BMC Microbiol* 15:170. <https://doi.org/10.1186/s12866-015-0495-4>.
- Basak P, Majumder NS, Nag S, Bhattacharyya A, Roy D, Chakraborty A, SenGupta S, Roy A, Mukherjee A, Pattanayak A, Ghosh A, Chattopadhyay D, Bhattacharyya M. 2015. Spatiotemporal analysis of bacterial diversity in sediments of Sundarbans using parallel 16S rRNA gene tag sequencing. *Microb Ecol* 69:500–511. <https://doi.org/10.1007/s00248-014-0498-y>.
- Martin M. 2011. Cutadapt removes adapter sequences from high throughput sequencing reads. *EMBnet J* 17:10–12. <https://doi.org/10.14806/ej.17.1.200>.
- Joshi NA, Fass JN. 2011. Sickle: a sliding-window, adaptive, quality-based trimming tool for FastQ files (version 1.33). <https://github.com/najoshi/sickle>.
- Chikhi R, Medvedev P. 2014. Informed and automated *k*-mer size selection for genome assembly. *Bioinformatics* 30:31–37. <https://doi.org/10.1093/bioinformatics/btt310>.
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Pribelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J Comput Biol* 19:455–477. <https://doi.org/10.1089/cmb.2012.0021>.
- Zerbino DR. 2010. Using the velvet *de novo* assembler for short-read sequencing technologies. *Curr Protoc Bioinformatics* 11:1–13. <https://doi.org/10.1002/0471250953.b11105s31>.
- Gurevich A, Saveliev V, Vyahhi N, Tesler G. 2013. QUASt: quality assessment tool for genome assemblies. *Bioinformatics* 29:1072–1075. <https://doi.org/10.1093/bioinformatics/btt086>.
- Aziz RK, Bartels D, Best A, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: Rapid Annotations using Subsystems Technology. *BMC Genomics* 9:75. <https://doi.org/10.1186/1471-2164-9-75>.
- Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI Prokaryotic Genome Annotation Pipeline. *Nucleic Acids Res* 44:6614–6624. <https://doi.org/10.1093/nar/gkw569>.
- Blin K, Wolf T, Chevrette MG, Lu X, Schwalen CJ, Kautsar SA, Suarez Duran HG, de los Santos ELC, Kim HU, Nave M, Dickschat JS, Mitchell DA, Shelest E, Breitling R, Takano E, Lee SY, Weber T, Medema MH. 2017. antiSMASH 4.0—improvements in chemistry prediction and gene cluster boundary identification. *Nucleic Acids Res* 45:W36–W41. <https://doi.org/10.1093/nar/gkx319>.