

How Geometric Constraints Control the Hydride Position and Activity in [NiFe]-Hydrogenases and Their Biomimetic Complexes

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Cite This: *Inorg. Chem.* 2025, 64, 10078–10086



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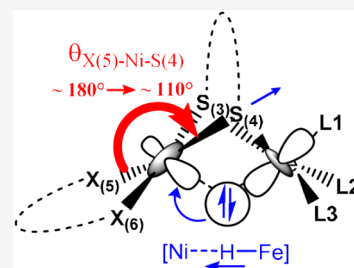


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ABSTRACT: The Ni-R active site in [NiFe]-hydrogenase features a bridging hydride between the Ni and Fe, displaced toward the Ni. However, all synthetic Ni-R models reported to date exhibit a hydride displaced toward Fe and display low turnover frequencies for H₂ evolution. Understanding the factors governing the hydride position and activity of Ni-R and biomimetic complexes is crucial for developing efficient hydrogen-evolving catalysts. By utilizing the CCSD theory, DFT, NBO, and QTAIM analysis, we investigated these factors in a Ni-R active-site model (1), and two representative biomimetic complexes, 2* and 3. Our results reveal that the Ni site of 1 inherently prefers a square-planar [S₂NiSH] configuration with an apically positioned thiolate and that hydride positioning is governed by the strength of [Ni–H–Fe] three-center two-electron bonding, which is modulated by the geometric torsion between the Ni terminal ligands and the bridging thiolates. By modifying the linkers between the Ni terminal ligands and bridging thiolate ligands of 2* and 3, we designed virtual biomimetic complexes (4–10). These complexes exhibit improved hydride nucleophilicity and increased potential for H₂ formation, providing valuable insights into how geometric and electronic factors influence hydride activity and informing the design of more effective biomimetic hydrogenase models.



INTRODUCTION

[NiFe]-hydrogenases ([NiFe]-H₂ase) are enzymes that facilitate the reversible conversion between H₂ and protons/electrons under mild conditions,^{1,2} making them ideal models for studying how nature developed hydrogen consuming and evolving catalysts (HECs) and for bioengineering and biomimetic approaches to hydrogen production, storage, and utilization.^{3,4} Extensive theoretical^{5–20} and experimental^{2,21–30} investigations of [NiFe]-H₂ase have led to the general acceptance of Ni-SI_a, Ni-C, and Ni-R species as intermediates in the catalytic cycle of hydrogen evolution (the top circle of Scheme 1). The Ni-R active site, [(CO)(CN)₂Fe(μ-S_{cys})₂(μ-H)Ni(HS_{cys})(S_{cys})]^{2–} (S_{cys} = cysteine), which is characterized by a bridging hydride between the Ni and Fe centers but displaced toward the Ni site,^{21,31} plays a critical role from hydrogen evolution to utilization.¹¹ However, synthetic biomimetic complexes of the Ni-R model (1) reported thus far, such as [(dppe)Ni(μ-pdt)(μ-H)Fe(CO)₃]⁺ (2*, pdt = 1,3-propanedithiolate; dppe = 1,2-C₂H₄(PPh₂)₂) and [Ni(N₂S₂)FeCp]⁺ (3, N₂S₂ = bismercaptoethane diazacycloheptane, Cp = η⁵-C₅H₅[–]) (Scheme 1, diamond at the bottom), exhibit the hydride shifted toward or bound mainly to Fe (Table S1),^{4,13,32–47} deviating from the Ni-R active site, and lower turnover frequencies (TOF) for H₂ evolution.^{13,32,38–40} Unraveling how variations in the geometric and electronic properties of the Ni and Fe ligands in the Ni-R active site and the biomimetic complexes influence hydride positioning and activity may aid synthetic efforts in accelerating the development of biomimetic catalysts for hydrogen evolution.

The high-resolution X-ray crystal structure of [NiFe]-H₂ase as Ni-R from *D. vulgaris* Miyazaki F protein revealed that the [S₂NiS₂] (S = sulfur of cysteines) framework and the bridging hydride form a nearly square pyramidal structure about the Ni with the hydride, Cys81, Cys84, and Cys546 in basal positions, and Cys549 in an apical position.²¹ The Fe site coordinates with one CO and two CN ligands and bridges to the hydride and the two cysteines (Cys84 and Cys549 bridging to Ni), which results in a nearly octahedral Fe geometry (Scheme 1). Overall, this arrangement leads to a longer Ni–Cys549 distance of 2.54 Å compared to the average of 2.21 Å for the other Ni–Cys bond lengths. Notably, the Ni–H bond length in Ni-R measures 1.58 Å, which is 0.20 Å shorter than the Fe–H bond length of 1.78 Å.

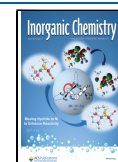
Theoretical studies have suggested both heterolytic and homolytic mechanisms for the reversible conversion of molecular hydrogen in [NiFe]-H₂ase.^{6,11,48} In the heterolytic mechanism, the H–H bond is formed via a nucleophilic addition pathway, where the bridging nucleophilic hydride (H[–]) and the electrophilic proton (H⁺) on Cys546 approach each other to form H₂, and the H–H bond is cleaved by the reverse reaction. In the homolytic mechanism, the H–H bond is formed/cleaved via a reductive-elimination/oxidation-addition pathway. Here,

Received: February 12, 2025

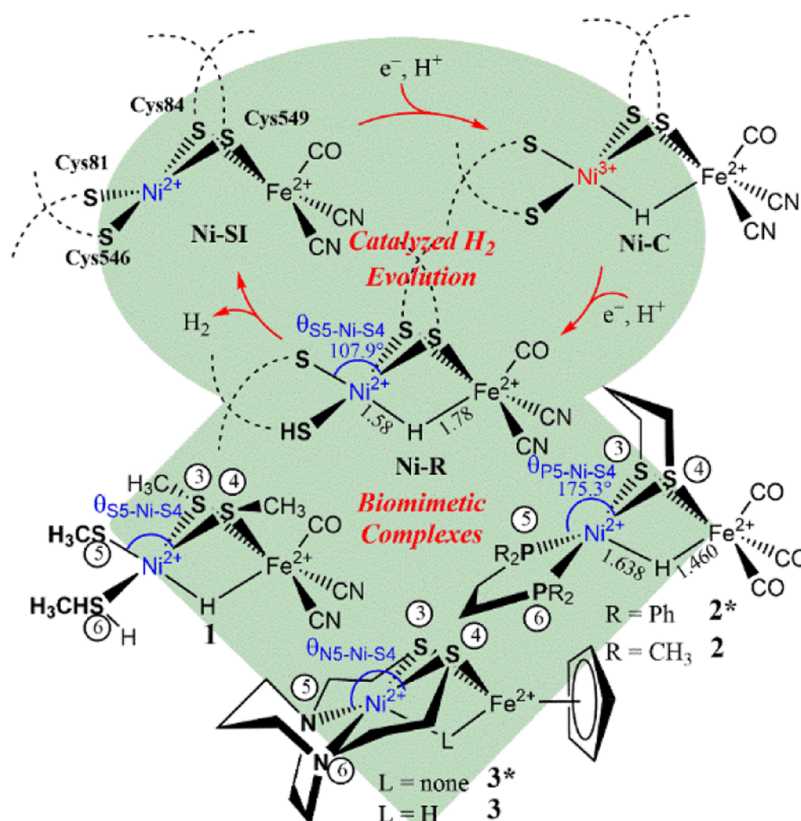
Revised: April 10, 2025

Accepted: April 24, 2025

Published: May 9, 2025



Scheme 1



the proton on Cys546 is transferred to the Ni(II) site, forming a dihydride Ni(IV) intermediate, which then releases H₂ by coupling the two H• radicals. Therefore, the displacement of the bridging hydride toward the Ni in Ni-R is likely conducive to the H₂ formation by either reaction mechanism.

Recent advancements have been made in the development of synthetic models of the [NiFe]-H₂ase active site, specifically mimicking the [Ni(μ-S)₂(μ-H)Fe] core found in the enzymes.^{4,13,29,32–47,49–51} These biomimetic complexes reported in the literature have shown similar structural characteristics, with the hydride positioned apically to the square-planar [X₂NiS₂] (X = N, P) moiety and in closer proximity to the Fe atom. In summary, the Ni–H and Fe–H bond lengths in these complexes range from 1.64 to 2.16 Å and 1.46 to 1.65 Å, respectively (see Table S1),^{32–37} and their TOF for H₂ evolution are low.^{32,40} In particular, the biomimetic complex, 2* (Scheme 1),³² exhibits a square pyramidal structure on the Ni site with hydride in the apical site with a Ni–H bond length of 1.64 Å and a short Fe–H bond of 1.46 Å. Additionally, several biomimetic complexes containing metallodithiolate [N₂NiS₂] moiety bound to CpFe or Fe(NO)₂, including 3* (Scheme 1), have been recently reported and have demonstrated modest electrocatalysis and H₂ production capabilities.^{43,44,46,51} Experimental and theoretical studies supported the existence of a hydride intermediate like 3 (Scheme 1), again with a long Ni–H distance, in their hydrogen evolution reaction.^{36–38,45,46}

To investigate how variations in the geometric and electronic properties of the Ni terminal ligands influence the hydride position and activity of the Ni-R and the biomimetic complexes 2* and 3*, we performed comprehensive density functional theory (DFT)⁵² and coupled-cluster singles-and-doubles (CCSD)⁵³ calculations, along with natural bond orbital

(NBO)⁵⁴ and quantum theory of atoms in molecules (QTAIM)⁵⁵ analyses. We begin by examining the factors that contribute to stability of the [S₂Ni(μ-S)₂(μ-H)Fe] core in Ni-R. Next, we identify the factors controlling the hydride position and activity relevant to H₂ evolution. Finally, based on these key factors (descriptors), we computationally generate a series of analog complexes of the biomimetic complexes 2* and 3* with the desired structural and electronic properties optimized to resemble those of Ni-R.

COMPUTATIONAL METHODS

The major model complexes studied are [(HSMc)(SMc)Ni(μ-SMe)₂(μ-H)Fe(CO)(CN)₂]²⁻ (1, where Me = CH₃), (dmpe)Ni(μ-pdt)(μ-H)Fe(CO)₃⁺ [2, where dmpe = 1,2-bis(dimethylphosphino)ethane], and [Ni(N₂S₂)(μ-H)FeCp]⁺ (3) (Scheme 1). Full and constraint geometry optimizations of complex 1, a model of the Ni-R active site, were performed using CCSD and DFT methods. In the CCSD calculations, the 6-31G** basis sets were employed for the Ni, Fe, S, and hydrogens involved in the H₂ formation/cleavage, while the STO-3G basis sets were utilized for methyl, CO, and CN groups. The frozen-core approximation was applied, in which all valence electrons were correlated, including the Ni and Fe 3s and 3p electrons. In the DFT calculations, spin-unrestricted DFT approach was performed with extra fine grids for numerical integration (target accuracy of 10⁻⁸ au in energy calculations). Five functionals,^{52,56} BP86, TPSS, B3LYP, M06, and LC-ωPBEh, with two basis sets, DZVP2 and Def2-TZVP, were assessed for their accuracy in determining geometric and energetic properties of Ni-R active site model complex 1 (Tables S2 and S3).

Previous studies have demonstrated the effectiveness of the M06/DZVP2 approach in accurately describing geometries, inner-sphere reduction energies, thermodynamic and kinetic properties of redox and catalytic sites in various first-row transition-metal systems like bimetallic hydrogenases,⁵⁷ iron–sulfur electron transfer proteins,⁵⁸ blue-copper proteins,⁵⁹ and [M(mnt)₂]ⁿ⁻ (M = Ni, Fe; mnt = 1,2-

$\text{S}_2\text{C}_2(\text{CN})_2^{2-}$; $n = 2, 1$) (see Figure S1). Overall, the M06/DZVP2 (or M06/DGDZVP2) offered a reliable balance of computational cost and accuracy for modeling geometries and energies of first-row transition-metal systems complexes. Therefore, M06/DZVP2 was primarily utilized for geometry optimizations and energy calculations in this study.

To account for the solvation free energy of the protein environments ($\epsilon \approx 4.0$) on the structure of the Ni-R active site, the full geometry optimizations were also carried out on **1** in the diethylamine solution phase ($\epsilon = 3.6$) utilizing the SMD continuum solvation model.⁶⁰ The relative energies (ΔE_e) and the relative free energy (ΔG) were determined using the total electronic energy (E_e) and Gibbs free energy (G) of molecules, calculated at either fully optimized or constrained optimized geometries. Gibbs free energies ($G = H - TS$) at temperature of 298.15 K were obtained by summing the calculated E_e and thermal energy correction to G . The thermal energy correction to enthalpy ($H_{\text{corr}} = \text{ZPE} + E_{\text{vib}} + E_{\text{rot}} + E_{\text{trans}} + RT$) and the total entropy (S) were derived from a frequency calculation. Additionally, the hydride bonding energies (ΔG_{H^-}) of the complexes (MH^-) in the MeCN solution were calculated at the M06/DZVP2 level and are defined as

$$\Delta G_{\text{H}^-} = G(\text{MH}^-) - [G(\text{M}) + G(\text{H}^-)]$$

The QTAIM⁵⁵ and NBO⁵⁴ analyses were employed to evaluate the atomic charges on the atom A (Q_A), the electron delocalization indexes of an A–B bond ($\delta_{\text{A-B}}$), the orbital occupancy $Q_{\text{Ni-H-Fe}}$, orbital energy $E_{\text{H-Ni-H-Fe}}$, and NBO stabilization energies E^2 of donor–acceptor interactions (using second-order perturbation theory).

All DFT calculations were conducted using the NWChem software packages.⁶¹ The G09 computational chemistry software package⁶² was utilized for CCSD and NBO calculations while the QTAIM calculations were performed with the AIMALL software package.⁶³ Structural and electronic structural visualizations and analysis were carried out using the GaussView,⁶² ChemOffice,⁶⁴ Molden,⁶⁵ and Chimrea.⁶⁶ The simulated reaction pathways were visualized and converted into videos using GaussView, with the IRC (intrinsic reaction coordinate) was generated by our mLST-IRC program. The mLST-IRC program generates reaction trajectories by linking key points, such as reactants, intermediates, transition states, and products, using multilinear synchronous transit (mLST) calculations (see Supporting Information).

RESULTS AND DISCUSSION

To understand the unusual structural characteristics of the Ni-R active site, we conducted detailed calculations on the model complex **1** at various levels of theory. Initially, DFT calculations of the model complex **1** in the gas phase utilizing various functionals and basis sets, ranging from the pure-GGA through hybrid/meta GGA to long-range corrected functionals, revealed a square-planar $[\text{S}_2\text{NiSH}]$ site with the apical fourth thiolate with a longer Ni–S distance bridging to the Fe. Specifically, for all the methods, the fully optimized structure **1a** of the model complex **1**, exhibited similar bond lengths except that of the apical Ni–S(4), which displayed a wider range of distances (Table S2). The DFT results are also consistent with those optimized at the CCSD/6-31G** level of theory and, aside from this longer Ni–S(4) $[\text{Ni}-\text{S}_{(\text{Cys549})}]$ distance, all the computed distances are in good agreement with the experimental values for Ni-R (Table S2).

Next, to assess the electrostatic effects of protein environments on the Ni– $\text{S}_{(\text{Cys549})}$ distance, we performed the full geometry optimization for **1a** in solution (diethylamine with $\epsilon = 3.6$, like protein environments of ~ 4). The slight decrease of 0.008 Å in the apical Ni–S(4) distance between the gas phase and solution phase suggests that the shorter Ni– $\text{S}_{(\text{Cys549})}$ bond in Ni-R compared to **1a** is likely due to the structural influence of protein backbone. Finally, the constraint-optimized geometric parameters of the model complex **1a1**, obtained through the

DFT calculations with a fixed bridging Ni–S(4) distance of 2.54 Å, like that in Ni-R, show excellent agreement with the active site of Ni-R (Table S3). For instance, the RMSD (Root Mean Square Deviation) of the calculated geometry at the M06/DZVP2 level of theory for **1a1** compared to the active site of Ni-R is 0.250 Å (Figure S2a). In particular, the calculated $\theta_{\text{S(5)-Ni-S(4)}}$ angle of 114.9°, the Ni–H, Fe–H, and Ni–S(3) bond lengths of 1.59 Å, 1.67 Å, and 2.21 Å for **1a1** at the M06/DZVP2 level closely match the experimental values of 107.9°, 1.58 Å, 1.78 Å, and 2.21 Å for the active site of Ni-R (Table S3), respectively. Notably, the differences in energy (ΔE_e) between **1a** and **1a1** at the CCSD/6-31G** and the M06/DZVP2 levels are only 0.71 and 3.87 kcal/mol, respectively (Table S3); suggesting that the structural changes arising from variation of the Ni–Cys549 are intrinsically low energy.

Furthermore, to understand the intrinsic structural properties of biomimetic complexes and their relationship the structure of Ni-R, we performed full geometry optimization on a conformer **1b** of the model complex **1**. The initial geometry for **1b** featured a $[\text{Ni}(\mu\text{-S})_2(\mu\text{-H})\text{Fe}]$ core with a square-planar $[\text{S}_2\text{NiS}_2]$ site, similar to the $[\text{X}_2\text{NiS}_2]$ site in **2*** and **3** (where X = P or N), with an apically positioned hydride. While attempts to optimize **1b** in the gas phase lead back to **1a** (Video S1), the geometry optimization of **1b** in the solution phase at the M06/DZVP2 level yielded a structural profile similar to those of **2*** and **3** with the $[\text{S}_2\text{NiS}_2]$ nearly square planar and the H in the apical site, where the Ni–S, Ni–H, and Fe–H bond distances of **1b** are 2.20 Å, 1.960 Å and 1.662 Å, respectively. The relative energy (ΔE_e) of 8.28 kcal/mol between **1a** and **1b** in the solution phase confirms that **1a** is a global minimum (Table S4).

Since **1b** has a structure like the biomimetic complexes, while **1a** (and especially **1a1** with the constrained Ni–S(4) distance) have structures that are nearly identical to that of Ni-R, we examined the influence of the Ni terminal ligand's angular geometry on the hydride position and properties of the model complex **1** through a potential energy surface (PES) scan and QTAIM topological analysis. With the model **1a1**, the geometry optimized PES scan at the M06/DZVP2 level of theory involved increasing the angle $\theta_{\text{S(5)-Ni-S(4)}}$ from 115° (**1a1**) to 140° (**1a2**), 160° (**1a3**) and finally 180° (**1a4**, which is like **1b**), while maintaining the bridging Ni–S(4) and Ni–S(3) distances of **1a2** to **1a4** at 2.54 and 2.21 Å, respectively, as observed in Ni-R and **1a1**. As the $\theta_{\text{S(5)-Ni-S(4)}}$ angle increased from **1a1** to **1a4**, a significant displacement of the hydride away from the Ni toward Fe was observed, an increase of 0.32 Å in the Ni–H bond length accompanied by an increase of 0.17 Å in the Ni–Fe distance (Figure 1a, Tables 1, S4, and Video S2). The total electronic energy (E_e) increased smoothly from **1a1** to **1a4** without encountering a barrier. QTAIM topological analysis (Tables 1 and S4) revealed a Ni–H electron delocalization index ($\delta_{\text{Ni-H}}$), which measures the degree of electron sharing or delocalization between the Ni and hydride in real space, of 0.54 au in **1a1**, larger than the $\delta_{\text{Fe-H}}$ of 0.41 au. As the $\theta_{\text{S(5)-Ni-S(4)}}$ angle increased from **1a1** to **1a4**, we observed a significant decrease in $\delta_{\text{Ni-H}}$ to 0.26 au and a corresponding increase in $\delta_{\text{Fe-H}}$ to 0.61 au. Additionally, the negative charge on the hydride (Q_{H}) experienced a slight increase, which was unexpected (*vide infra*), as one might have expected the Ni-R-like structure, **1a1**, to be more hydridic compared to the biomimetic-like structure, **1a4**.

Further molecular orbital (MO) and NBO analyses provide deeper insight into the exothermic nature and the decrease in hydridic character from **1a4** to **1a1**. The MO interaction diagram between an octahedral $[\text{FeH}(\text{SMe})_2(\text{CN})_2(\text{CO})]^{3-}$

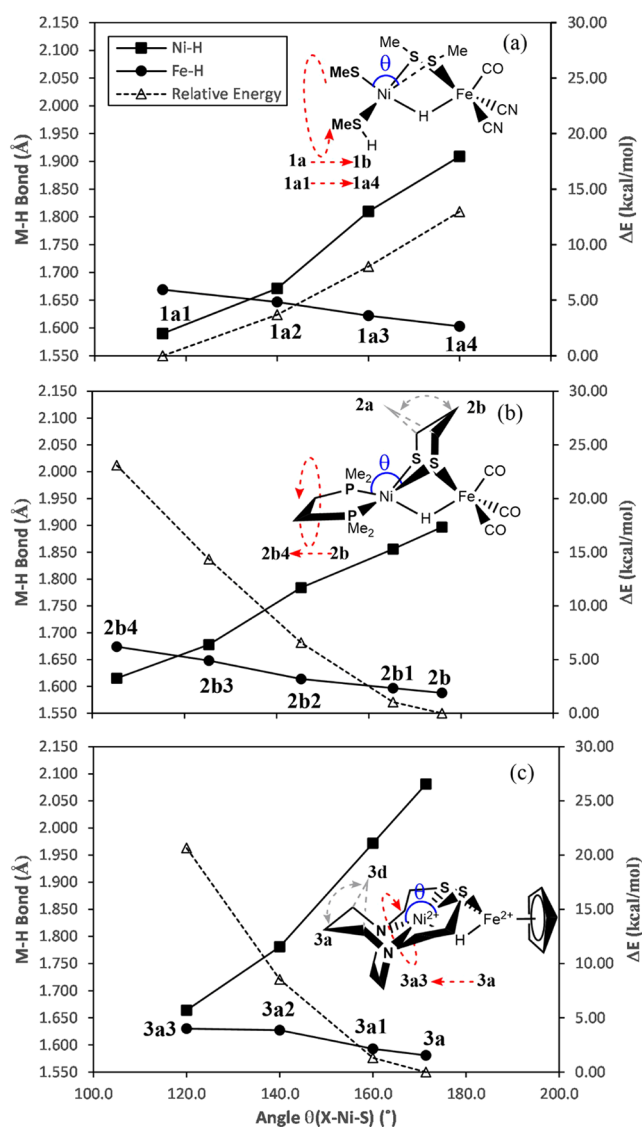


Figure 1. Plot of variations of the M–H bond lengths (M = Ni: solid square; M = Fe: solid circle), relative total electronic energy (ΔE_e ; open triangle) with the increase in the $\theta_{X(S)-Ni-S(4)}$ angle (X = S for 1, P for 2, N for 3): (a) from 1a1 to 1a4, (b) from 2b to 2b4, and (c) from 3a to 3a3, respectively. The changes in the conformational structure between 2a and 2b and between 3a and 3d are depicted by gray dashed lines.

and a right triangle $[(SMe)(SMeH)Ni]^+$ fragments for 1a4 reveals that a $[Ni-H-Fe]$ three-center-two-electron (3c-2e) bond ($\psi_{Ni-H-Fe}$) can be formed via interactions between the hydride orbital, which is the σ_{Fe-H} bond ($H_{1s} + Fe_{4p_z}$), and the unoccupied $4p_z$ and $4s$ orbitals of the Ni site (Figure S3). This 3c-2e bond has a longer Ni–H distance as it is destructively interacting with the doubly occupied Ni d_{z^2} . During the transition from 1a4 to 1a1, the overlap of the bonding orbitals between the hydride and the Ni site in this 3c-2e bond increases as the occupied Ni d_{z^2} moves away. This enhanced bonding interaction leads to greater stabilization energy for the $\psi_{Ni-H-Fe}$ orbital in 1a1 compared to 1a4 (Figure S4). As a result of the hydride shifting toward Ni, the frontier orbitals of 1a1 show notable changes, particularly in the Ni–H antibonding characteristics, as the doubly occupied Ni d_{z^2} was antibonding with the Fe–H bond in 1a4 and becomes antibonding with the Ni–S in bond in 1a1 (Figure S4). Simultaneously, electron

density on the Ni increases as observed by QTAIM (Table 1). NBO analysis reveals that, compared to 1a4, the Ni orbital coefficient in the $\psi_{Ni-H-Fe}$ orbital of 1a1 shows significant increase of 0.16, with a particularly notable rise in the Ni 3d orbital hybridization coefficient (Figure 2). This indicates that as 1a4 transits to 1a1, the Ni 3d orbital contribution to the $\psi_{Ni-H-Fe}$ orbital substantially increases. Meanwhile the contributions from the Fe and hydride orbital decrease (Figure 2). Compared to 1a4, the NBO $\psi_{Ni-H-Fe}$ orbital of 1a1 is more stable by 2.41 eV (Figures 2, S5 and Table 1). The MO and NBO analyses show that the exothermicity and the reduction in hydridic character from 1a4 to 1a1 can be attributed to variations in the stability of the $\psi_{Ni-H-Fe}$ orbital.

By introducing carbon linkers either between two bridging sulfurs as in 2* or between the terminal ligands and bridging sulfurs on the Ni site as in 3, the biomimetic complexes, 2* and 3, contain a $[Ni(\mu-S_2)(\mu-H)Fe]$ core that resembles Ni–R and 1a1, except that these linkers produce an active site geometry where the hydride is far from the Ni and apical with respect to the square-planar $[X_2NiS_2]$ Ni site (2* X = P and 3 X = N). Notably, the structural characteristics of the $[Ni(\mu-S_2)(\mu-H)Fe]$ core in these biomimetic complexes closely resemble those of the metastable 1b (or 1a4), where the hydride is predominantly bound to the Fe site. To guide synthetic efforts aimed at enhancing the catalytic properties of 2* and 3 for the hydrogen evolution, we examined how the angular geometry of the terminal ligands affects the hydride position and properties of these model complexes, 2 and 3 (Scheme 1). We will present the results for both 2 and 3 first and then compare them to those for 1.

As anticipated, the M06/DZVP2 approach provided a satisfactory representation of the geometric properties of 2* (Table S5), with a RMSD of 0.299 Å from the experimental structure (Figure S2b). Given that the M06/DZVP2 relative energy (ΔE_e) between 2a and 2b (alternative pdt geometries, chair/boat, Figure 1) is only 3.92 kcal/mol (Table S5) and that the energy barrier for the chair/boat flip of the pdt ligand is 9.63 kcal/mol,⁶⁷ 2b was used for the PES scan as it would better accommodate variations in the angular geometry between the pdt and the bulky Ni terminal ligands. Thus, the PES scan was conducted by decreasing the $\theta_{X(S)-Ni-S(4)}$ angle from 175.9° in 2b through 165.3° (2b1), 145.3° (2b2), 125.3° (2b3), to 105.3° (2b4) (X = P), with the bridging Ni–S(4) and Ni–S(3) bond lengths fixed as observed in 2b (Figure 1b, Table S4, and Video S3). Among all the conformers of 3, 3a–3c and 3d–3f with alternative linkage geometries, the species 3a is the most stable species based on the calculated relative energies (Figure S7 and Table S6). Thus, 3a was used for the PES scan by decreasing the $\theta_{X(S)-Ni-S(4)}$ angle from 171.4° (3a) through 160.0° (3a1), 140.0° (3a2), to 120.0° (3a3) (X = N) (Figure 1c and Table S6). All on the scans in Figure 1 proceed without an energy barrier; notably, unlike the transition from 1a4 back to 1a1, the transitions of 2b to 2b4 and 3a to 3a3 are endothermic (Figure 1b,c and Table 1). However, like the scan from 1a4 to 1a1, the NBO $[Ni-H-Fe]$ 3c-2e orbital energies ($E\psi_{Ni-H-Fe}$) for the scans 2b to 2b4 and 3a to 3a3 decrease by 2.22 and 2.44 eV, respectively. These results suggest that the hydride displacement toward the Ni inherently stabilizes the $[Ni(\mu-S_2)(\mu-H)Fe]$ core. The contrasting thermodynamic behaviors observed in the transitions from 2b to 2b4 and from 3a to 3a3, compared to the transition from 1a4 to 1a1, can be attributed to geometric constraints imposed by the carbon linkers. This highlights the critical role of the carbon linkers in stabilizing the $[Ni(\mu-S_2)(\mu-$

Table 1. M06/DZVP2 Geometric Parameters (In Deg and Å; X = S, P, N), NBO Properties (3c-2e Orbital Occupancy $Q_{\text{Ni-H-Fe}}$ and Orbital Energy $E\psi_{\text{Ni-H-Fe}}$ in Atomic Unit), QAIM Topological Properties (Charge Q and Electron Delocalization Index δ in Atomic Unit), Relative Total Electron Energy ΔE_e , Relative Free Energy ΔG , and Hydride Bonding Free Energy ΔG_{H^-} (kcal/mol) for the Ni-R Active Site Model 1, Biomimetic Models 2 and 3

	1a1 ^a	1a4 ^{a,b}	2b4 ^b	2b	3a3 ^b	3a
$\theta_{\text{X(S)}-\text{Ni-S(4)}}$	114.9	180.0	105.3	175.9	120.0	171.4
$r_{\text{Ni-Fe}}$	2.557	2.724	2.421	2.665	2.366	2.590
$r_{\text{Ni-H}}$	1.590	1.909	1.615	1.897	1.664	2.081
$r_{\text{Fe-H}}$	1.669	1.603	1.674	1.588	1.630	1.581
$Q_{\text{Ni-H-Fe}}$	1.74	1.77	1.72	1.73	1.83	1.88
$E\psi_{\text{Ni-H-Fe}}$	−0.12297	−0.03438	−0.52572	−0.44423	−0.36280	−0.27307
Q_{Ni}	0.392	0.401	0.166	0.190	0.515	0.544
Q_{Fe}	0.834	0.821	0.799	0.795	0.733	0.680
Q_{H}	−0.215	−0.298	−0.210	−0.219	−0.294	−0.373
$\delta_{\text{Ni-H}}$	0.544	0.261	0.480	0.251	0.510	0.182
$\delta_{\text{Fe-H}}$	0.408	0.609	0.465	0.625	0.542	0.731
ΔE_e^c	0.00	12.97	0.00	−23.11	0.00	−20.65
ΔG^d	0.00	9.08	0.00	−21.39	0.00	−24.81
$\Delta G_{\text{H}^-}^d$	−50.72	−30.66	−83.83	−73.03	−49.89	−28.47

^aConstrained $r_{\text{Ni-S(3)}}$ and $r_{\text{Ni-S(4)}}$ at 2.21 Å and 2.54 Å, respectively. ^bConstrained $\theta_{\text{S(S)}-\text{Ni-S(4)}}$ (in bold) in the geometry optimizations. ^cIn the gas phase. ^dIn the acetonitrile solution.

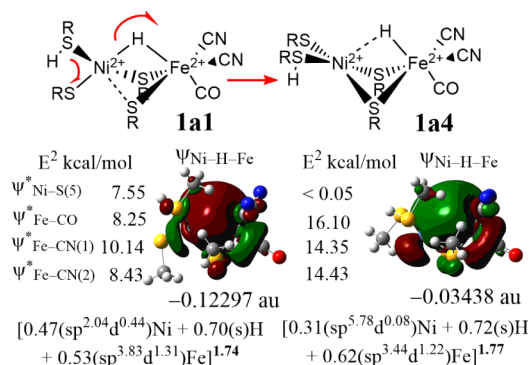


Figure 2. NBO MO ($\psi_{\text{Ni-H-Fe}}$), coefficients, hybrids, and occupancy (in bold), and orbital energy (in au) of the [Ni-H-Fe] 3c-2e bond of **1a1** and **1a4**. The stabilization energies E^2 (in kcal/mol) of the $\psi_{\text{Ni-H-Fe}}$ and $\psi_{\text{Fe-CN(1)}}$ with the NBOs of the Fe terminal ligands by second-order perturbation theory are shown in the left side of $\psi_{\text{Ni-H-Fe}}$.

H)Fe] core of biomimetic complexes in a geometry different from that observed for Ni-R.

In addition to the above trends in the energies, the charges Q_{X} (X = H, Ni, Fe), bond lengths, and binding energy of the hydride show related trends (Table 1 and Figure 3). As the hydride shifts closer the Ni (decreasing the $\theta_{\text{X(S)}-\text{Ni-S(4)}}$ angle) from **1a4** to **1a1**, **3a** to **3a3**, and **2b** to **2b4**, the hydride charge (Q_{H}) progressively increases (Figure 3a), while the Ni-H and Ni-Fe bond lengths ($r_{\text{Ni-H}}$ and $r_{\text{Ni-Fe}}$) decrease (Figure 3b). These shifts are accompanied by an increase in stability of the [Ni-H-Fe] 3c-2e bond, which enhances the hydride binding (Figure 3c). These trends lend further evidence that decreasing the $\theta_{\text{X(S)}-\text{Ni-S(4)}}$ angle diminishes the hydridic character while increasing Ni's nucleophilicity. Thus, in a charge-controlled reaction mechanism, which is primarily influenced by the distribution of charges (electrostatic interactions) on the reactants, the bridging hydride of **1a4**, **3a**, and **2b** possesses higher hydridic character, more favorable for proton bonding via a heterolytic reaction mechanism to generate H_2 . On the other hand, **1a1**, **3a3**, and **2b4** exhibits a greater nucleophilicity on the

Ni site, which would favor a homolytic reaction mechanism through a dihydride Ni(IV) intermediate to generate H_2 .

Furthermore, as shown in Figure 2, the significant changes in the NBO second-order perturbation energy E^2 of **1a1**, compared to **1a4**, suggest that the terminal ligands on the Fe and Ni sites can significantly impact the stability of the $\psi_{\text{Ni-H-Fe}}$ orbital by the interactions between the unoccupied $\psi_{\text{Ni-S(5)}}$ and $\psi_{\text{Fe-CO}}^*/\psi_{\text{Fe-CN}}^*$ orbitals and the $\psi_{\text{Ni-H-Fe}}$ orbital. Notably, as the [Ni-H-Fe] 3c-2e bonding strengthens from **1a4** to **1a1**, the E^2 originating from $\psi_{\text{Fe-CO}}^*$ with $\sigma_{\text{Fe-CO}}^*$ antibonding character decreases by 7.85 kcal/mol, leading to a shortening of both the Fe-CO and C≡O bond lengths (Table S4). The significant increase of about 20 cm^{-1} in the calculated C≡O vibrational frequency from **1a4** to **1a1** appears to align well with the substantial shifts in the C≡O vibrational frequency of Ni-R observed experimentally.² To evaluate the effects of ligands on the hydridic properties of Ni-R and biomimetic complex models, we examined the hydride binding free energies (ΔG_{H^-}) in the acetonitrile solution (Table S7). Notably, the calculated ΔG_{H^-} of −73 kcal/mol for **2b** closely matches that for **2a** (−73 kcal/mol) and the experimental value for **2a*** (−79 kcal/mol).³³ Overall, the ΔG_{H^-} of these models appear to correlate with the $E\psi_{\text{Ni-H-Fe}}$ as a function of the $\theta_{\text{X(S)}-\text{Ni-S(4)}}$ angle, both decreasing in the order of **1a4** > **1a1**, **3a** > **3a3**, and **2b** > **2b4** (Table 1 and Figure 3c). Interestingly, like the $\theta_{\text{X(S)}-\text{Ni-S(4)}}$ angle, the $E\psi_{\text{Ni-H-Fe}}$ values decrease in the order: **1a4** > **3a** > **2b**, and **1a1** > **3a3** > **2b4**, but the ΔG_{H^-} shows: **1a4** ≈ **3a** > **2b**, and **1a1** ≈ **3a3** > **2b4**. Thus, **1** (or Ni-R) and **3** are predicted to have similar but higher hydride activity than **2** (or **2***). Our results strongly suggest that the hydride activity decreases as the hydride shifts toward the Ni across the transitions from **1a4**, **2b**, and **3a** to **1a1**, **3a3**, and **2b4**, respectively (Figures 3 and S8). As mentioned above, both experimental and theoretical studies support that displacing hydride toward Ni facilitates H_2 formation via either heterolytic or homolytic reaction pathways.^{6,11} However, the higher ΔG_{H^-} and lower Q_{H} (*vide supra*) observed for **1a1**, **2b4**, and **3a3** compared to their precursors presents a challenge to fully understanding the activity of [Ni($\mu\text{-S}_2$)($\mu\text{-H}$)Fe] core in Ni-R versus biomimetic complexes. Thus, additional factors may help

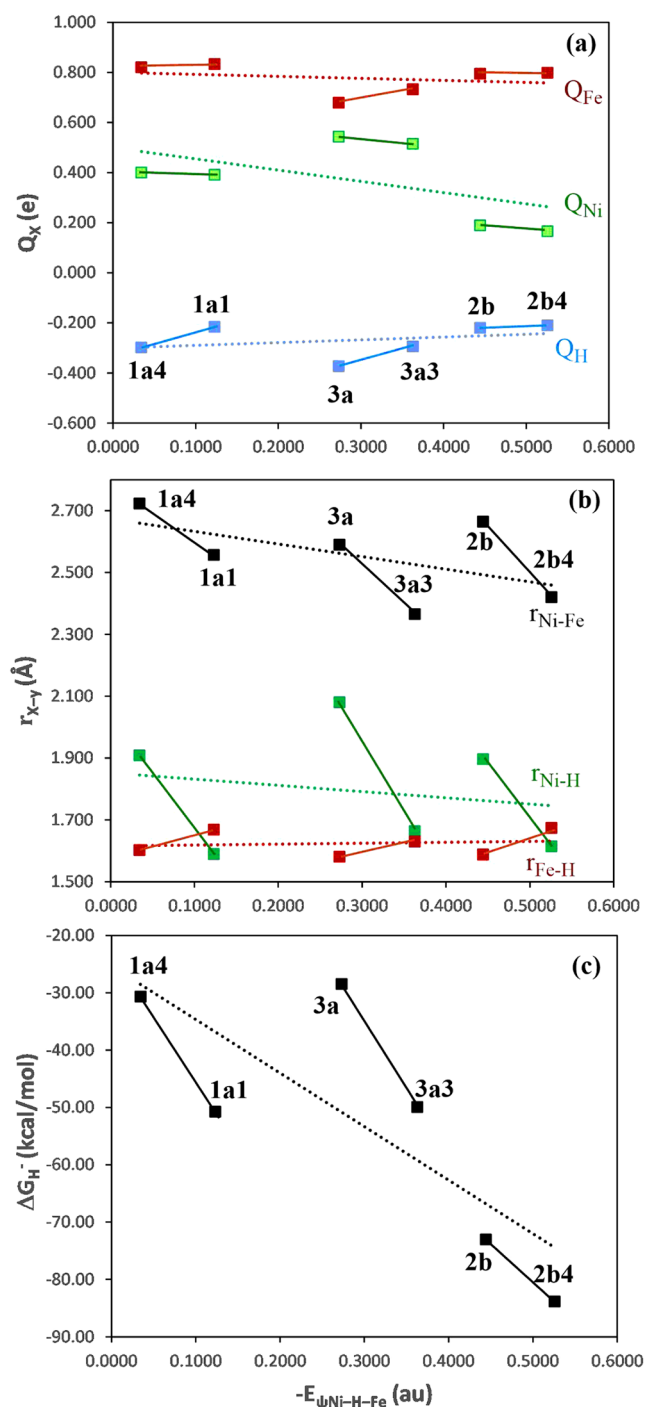


Figure 3. Plot of variations in the electronic and bonding properties as the $[\text{Ni-H-Fe}]$ 3c-2e orbital energies ($\epsilon_{\psi_{\text{Ni-H-Fe}}}$) decrease for the Ni-R and biomimetic complex models: (a) Charge (Q_x) on hydride (blue), Ni (green), and Fe (red); (b) Ni-H (green) and Fe-H (red) bond lengths and Ni-Fe distance (black); (c) Hydride (H^-) binding free energies (ΔG_{H^-}) in solution (CH_3CN). The dashed lines illustrate the overall correlation trends between each property and $\epsilon_{\psi_{\text{Ni-H-Fe}}}$.

facilitate H_2 formation in Ni-R. It is worth noting that as the strength of $[\text{Ni-H-Fe}]$ 3c-2e bonding increases, the frontier orbitals, specifically the doubly occupied $\text{Ni } d_{z^2}$ and the unoccupied $\text{Ni } d_{x^2-y^2}$, move closer in energy, leading to substantial changes in the hydride character within these orbitals (Figures S3 and S4). This increased hydride contribution in the frontier orbitals may significantly enhance H_2 formation via an

orbital-controlled reaction mechanism that lies intermediate between a homolytic and heterolytic pathways. Mimicking the $[\text{Ni}(\mu\text{-S}_2)(\mu\text{-H})\text{Fe}]$ core structure is a crucial step for the rational design of biomimetic catalyst, as this will provide a full understanding of how the structure of the $[\text{Ni}(\mu\text{-S}_2)(\mu\text{-H})\text{Fe}]$ core contributes to the H_2 formation.

Our findings demonstrated that the carbon linkers in reported biomimetic complexes are critical in maintaining the $[\text{Ni}(\mu\text{-S}_2)(\mu\text{-H})\text{Fe}]$ core structure, likewise the protein backbones stabilize the Ni-R active site. This insight suggests a strategy for the rational design of biomimetic complexes with higher activity might involve adjusting the $\theta_{\text{X}(\text{S})-\text{Ni}-\text{S}(4)}$ angle ($\text{X} = \text{P}, \text{N}$) of **2a*** and **3** by modifying the carbon linkers between the Ni terminal ligands and bridging thiolate ligands. Changing this angle might lead the complexes to more closely mimic the Ni-R active site in enzymes. Among a series of the “designer” complexes **4–10** (Scheme S1), **4–8** and **9a** exhibited stronger resemblance to the model complex **1a1** of the Ni-R active site in terms of hydride position and the electronic properties of the $[\text{Ni}(\mu\text{-H})(\mu\text{-S})_2\text{Fe}]$ core (Figures 4 and S9).

The calculated Ni-Fe distance, Ni-H, and Fe-H bond lengths in these complexes ranged from 2.488 to 2.519 Å, 1.615 to 1.685 Å, and 1.610 to 1.692 Å, respectively (Figures 4, S9 and Table S8). Especially, the biomimetic complex **2a** appears to exhibit a higher degree of adaptability in achieving species **4–8** that feature a $[\text{Ni}(\mu\text{-H})(\mu\text{-S})_2\text{Fe}]$ structural motif like that in Ni-R, as there appear to be more alternative options for modifying the carbon linker. The virtual complexes **4–8** exhibited total electron delocalization indices for the Ni-H and Fe-H bonds ($\delta_{\text{Ni-H-Fe}}$) ranging from 0.925 to 1.016 au, which are comparable to the value of 0.952 au calculated for **1a1**. Notably, compared to **1a1**, the smaller positive charge on the Ni (Q_{Ni}) and the larger negative charge on the hydride (Q_{H}) in **4–8** suggest that these virtual complexes might possess not only a higher nucleophilic Ni site, enabling an oxidative addition reaction for H_2 evolution via a homolytic pathway, but also exhibit a more nucleophilic hydride, facilitating the H_2 evolution via a heterolytic pathway. Additionally, the virtual complexes **9a** and **9b**, characterized by a more nucleophilic hydride (Q_{H}) and a longer bridging Ni-S(4) distance of 2.64–2.80 Å compared to **1a1** may offer a more favorable proton transfer channel through the bridging S(4) for H_2 formation via a heterolytic reaction pathway compared to their native complex **3**. It is noteworthy that a recent comprehensive DFT study on H_2 evolution catalyzed by a $[\text{Ni}(\text{N}2\text{S}2)\text{Fe}(\text{CO})\text{Cp}]^+$ complex revealed a significantly lower energy pathway through an E[ECEC] electrochemical reaction mechanism (E = electron transfer, C = chemical reaction, here corresponding to a proton transfer).^{45,46} In this mechanism, the $[(\text{N}2\text{S}2)\text{Ni}(\mu\text{-H})\text{Fe}(\text{CO})\text{Cp}]^+$ intermediate undergoes a reduction to form a Ni(I) site, followed by protonation on a bridging sulfur, leading to H_2 formation via a heterolytic reaction pathway. Overall, from the perspective of charge-controlled reactions, in comparison to their native complexes, **2a*** and **3**, the virtual biomimetic complexes, **4–9**, should demonstrate significant improvement in their reactivity for H_2 formation.

CONCLUSIONS

Our CCSD and DFT calculations indicate that the equilibrium geometry of the $[\text{X}_2\text{Ni}(\mu\text{-S})_2(\mu\text{-H})\text{Fe}]$ core ($\text{X} = \text{S}, \text{P}, \text{N}$) differs in Ni-R and the synthetic biomimetic complexes (**2*** and **3**). While Ni-R inherently favors a square-planar $[\text{X}_2\text{NiSH}]$ geometry with an apically positioned thiolate, the biomimetic

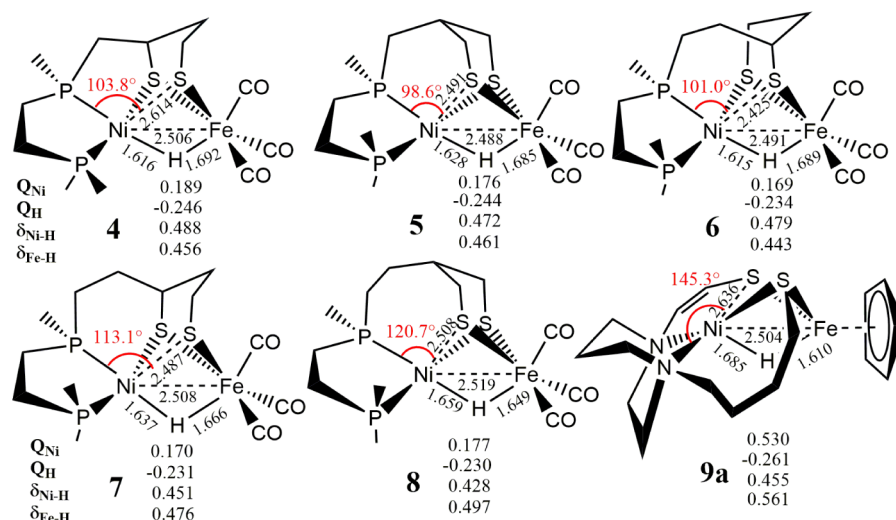


Figure 4. Optimized geometric parameters ($\theta_{\text{X(S)}-\text{Ni}-\text{S(4)}}$ in deg and $r_{\text{M-H}}$ in Å) and QTAIM topological properties (charge Q and electron delocalization index δ in au) of the designed biomimetic complexes 4–8 ($X = \text{P}$) and 9a ($X = \text{N}$).

complexes favor a square-planar $[\text{X}_2\text{NiS}_2]$ geometry with an apically positioned hydride that is bonded more strongly to Fe. Moving a bridging thiolate to the apical site, as in Ni-R, shifts the hydride from the Fe to a basal bonding site on Ni. This displacement of the bridging hydride from the Fe site to the Ni site is facilitated by $[\text{Ni-H-Fe}]$ 3c-2e bonding, which is only possible when the hydride is in the basal Ni site. Model complexes mimicking the geometry of Ni-R can be constructed by manipulating the geometric torsion of the Ni terminal ligands relative to the bridging ligands. Specifically, adjusting the $\theta_{\text{X(S)}-\text{Ni}-\text{S(4)}}$ angle ($X = \text{P}, \text{N}$) through modifications of the carbon linkers between the Ni terminal ligands and bridging thiolate ligands of **2*** and **3**, generates a series of virtual complexes that achieve a similar $[\text{Ni}(\mu\text{-S}_2)(\mu\text{-H})\text{Fe}]$ core with structural and electronic properties similar to those found in Ni-R. Finally, compared to the Ni-R model complex **1a1**, the modified biomimetic complexes 4–8 based on **2** exhibit improved protonation capabilities at either the Ni or the bridging hydride, while the complex **9** by modifying **3** demonstrates enhanced nucleophilicity on the bridging hydride, facilitating H_2 formation. These findings provide valuable guidance for the rational design and modifications of biomimetic complexes that may more closely mimic the biological functions of Ni-R. Further systematic studies are underway to elucidate the reasonable enzymatic mechanisms of Ni-R and identify key factors that activate hydride. Leveraging high-quality descriptors derived from the rate-determining step of a well-defined enzymatic process, we aim to design and optimize biomimetic complexes with improved catalytic efficiency for H_2 formation.

■ ASSOCIATED CONTENT

SI Supporting Information

The Supporting Information is available free of charge at <https://pubs.acs.org/doi/10.1021/acs.inorgchem.5c00670>.

Computational details; Scheme S1, Figures S1–S9, Tables S1–S8 (PDF)

Cartesian coordinates of calculated models at the M06/DZVP2 level (PDF)

Visualizing structural transitions of **1b** → **1a** → **1b** (MP4)

Visualizing structural transitions of **1b** → **1a1** → **1a** → **1a1** → **1b** (MP4)

Visualizing structural transitions of **2a** → **2b** → **2b4** → **2b** → **2a** (MP4)

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Notes

The authors declare no competing financial interest.

■ ACKNOWLEDGMENTS

This work was supported by the National Science Foundation (CHE-1300787) and the Welch Foundation (A-0648). We acknowledge Texas A&M University's High Performance Research Computing Facility and the Laboratory for Molecular Simulations for providing computing resources. The NWChem calculations were partially conducted at the Environmental Molecular Sciences Laboratory (EMSL), a national user facility sponsored by the U.S. DOE's Office of Biological and Environmental Research and located at Pacific Northwest National Laboratory, operated for the DOE by Battelle, under the grant EMSL 48430. We thank Wally L. Niu for his contribution to the mLST-IRC program, which greatly facilitated the visualization of the simulated reaction pathways for this work.

■ REFERENCES

- Fontecilla-Camps, J. C.; Volbeda, A.; Cavazza, C.; Nicolet, Y. Structure/Function Relationships of $[\text{NiFe}]$ - and $[\text{FeFe}]$ -Hydrogenases. *Chem. Rev.* **2007**, *107*, 4273–4303.
- Lubitz, W.; Ogata, H.; Ruediger, O.; Reijerse, E. Hydrogenases. *Chem. Rev.* **2014**, *114*, 4081–4148.

- (3) Esswein, A. J.; Nocera, D. G. Hydrogen production by molecular photocatalysis. *Chem. Rev.* **2007**, *107*, 4022–4047.
- (4) Artero, V.; Berggren, G.; Atta, M.; Caserta, G.; Roy, S.; Pecqueur, L.; Fontecave, M. From Enzyme Maturation to Synthetic Chemistry: The Case of Hydrogenases. *Acc. Chem. Res.* **2015**, *48*, 2380–2387.
- (5) Niu, S. Q.; Thomson, L. M.; Hall, M. B. Theoretical characterization, of the reaction intermediates in a model of the nickel-iron hydrogenase of *Desulfovibrio gigas*. *J. Am. Chem. Soc.* **1999**, *121*, 4000–4007.
- (6) Niu, S. Q.; Hall, M. B. Modeling the active sites in metalloenzymes 5. The heterolytic bond cleavage of H₂ in the [NiFe] hydrogenase of *Desulfovibrio gigas* by a nucleophilic addition mechanism. *Inorg. Chem.* **2001**, *40*, 6201–6203.
- (7) Bruschi, M.; Tiberti, M.; Guerra, A.; De Gioia, L. Disclosure of key stereoelectronic factors for efficient H₂ binding and cleavage in the active site of [NiFe]-hydrogenases. *J. Am. Chem. Soc.* **2014**, *136*, 1803–1814.
- (8) Wu, H.; Hall, M. B. Density functional theory on the larger active site models for [NiFe] hydrogenases: Two-state reactivity? *C. R. Chim.* **2008**, *11*, 790–804.
- (9) Dong, G.; Phung, Q. M.; Hallaert, S. D.; Pierloot, K.; Ryde, U. H₂ binding to the active site of [NiFe] hydrogenase studied by multiconfigurational and coupled-cluster methods. *Phys. Chem. Chem. Phys.* **2017**, *19*, 10590–10601.
- (10) Kramer, T.; Kamp, M.; Lubitz, W.; van Gestel, M.; Neese, F. Theoretical Spectroscopy of the Ni^{II} Intermediate States in the Catalytic Cycle and the Activation of [NiFe] Hydrogenases. *ChemBioChem* **2013**, *14*, 1898–1905.
- (11) Siegbahn, P. E.; Tye, J. W.; Hall, M. B. Computational Studies of [NiFe] and [FeFe] Hydrogenases. *Chem. Rev.* **2007**, *107*, 4414–4435.
- (12) Das, R.; Neese, F.; van Gestel, M. Hydrogen evolution in [NiFe] hydrogenases and related biomimetic systems: Similarities and differences. *Phys. Chem. Chem. Phys.* **2016**, *18*, 24681–24692.
- (13) Hugenbruch, S.; Shafaat, H. S.; Kramer, T.; Delgado-Jaime, M. U.; Weber, K.; Neese, F.; Lubitz, W.; DeBeer, S. In search of metal hydrides: An X-ray absorption and emission study of [NiFe] hydrogenase model complexes. *Phys. Chem. Chem. Phys.* **2016**, *18*, 10688–10699.
- (14) Chambers, G. M.; Huynh, M. T.; Li, Y.; Hammes-Schiffer, S.; Rauchfuss, T. B.; Reijerse, E.; Lubitz, W. Models of the Ni-L and Ni-SI₃ States of the [NiFe]-Hydrogenase Active Site. *Inorg. Chem.* **2016**, *55*, 419–431.
- (15) Dong, G.; Phung, Q. M.; Pierloot, K.; Ryde, U. Reaction Mechanism of [NiFe] Hydrogenase Studied by Computational Methods. *Inorg. Chem.* **2018**, *57*, 15289–15298.
- (16) Siegbahn, P. E. M.; Chen, S.-L.; Liao, R.-Z. Theoretical Studies of Nickel-Dependent Enzymes. *Inorganics* **2019**, *7*, 95.
- (17) Siegbahn, P. E. M.; Liao, R.-Z. The Energetics of Hydrogen Molecule Oxidation in NiFe-hydrogenase. *ACS Catal.* **2020**, *10*, 5603–5613.
- (18) Qiu, S.; Li, Q.; Xu, Y.; Shen, S.; Sun, C. Learning from nature: Understanding hydrogenase enzyme using computational approach. *Wiley Interdiscip. Rev.: Comput. Mol. Sci.* **2020**, *10*, No. e1422.
- (19) Isegawa, M.; Matsumoto, T.; Ogo, S. H₂ activation by hydrogenase-inspired NiFe catalyst using frustrated Lewis pair: Effect of buffer and halide ion in the heterolytic H–H bond cleavage. *RSC Adv.* **2021**, *11*, 28420–28432.
- (20) Isegawa, M.; Matsumoto, T.; Ogo, S. Hydrogen evolution, electron-transfer, and hydride-transfer reactions in a nickel–iron hydrogenase model complex: A theoretical study of the distinctive reactivities for the conformational isomers of nickel–iron hydride. *Dalton Trans.* **2021**, *51*, 312–323.
- (21) Ogata, H.; Nishikawa, K.; Lubitz, W. Hydrogens detected by subatomic resolution protein crystallography in a [NiFe] hydrogenase. *Nature* **2015**, *520*, 571–574.
- (22) Evans, R. M.; Krahn, N.; Weiss, J.; Vincent, K. A.; Söll, D.; Armstrong, F. A. Replacing a Cysteine Ligand by Selenocysteine in a [NiFe]-Hydrogenase Unlocks Hydrogen Production Activity and Addresses the Role of Concerted Proton-Coupled Electron Transfer in Electrocatalytic Reversibility. *J. Am. Chem. Soc.* **2024**, *146*, 16971–16976.
- (23) Pandelia, M.-E.; Ogata, H.; Lubitz, W. Intermediates in the Catalytic Cycle of [NiFe] Hydrogenase: Functional Spectroscopy of the Active Site. *ChemPhysChem* **2010**, *11*, 1127–1140.
- (24) Wang, H. X.; Ralston, C. Y.; Patil, D. S.; Jones, R. M.; Gu, W.; Verhagen, M.; Adams, M.; Ge, P.; Riordan, C.; Marganian, C. A.; Mascharak, P.; Kovacs, J.; Miller, C. G.; Collins, T. J.; Brooker, S.; Croucher, P. D.; Wang, K.; Stiefel, E. I.; Cramer, S. P. Nickel L-edge soft X-ray spectroscopy of nickel-iron hydrogenases and model compounds - Evidence for high-spin nickel(II) in the active enzyme. *J. Am. Chem. Soc.* **2000**, *122*, 10544–10552.
- (25) Wang, H. X.; Patil, D. S.; Gu, W.; Jacquamet, L.; Friedrich, S.; Funk, T.; Cramer, S. P. L-edge X-ray absorption spectroscopy of some Ni enzymes: Probe of Ni electronic structure. *J. Electron Spectrosc. Relat. Phenom.* **2001**, *114–116*, 855–863.
- (26) Basu, D.; Gray, D. L.; Woods, T. J.; Rauchfuss, T. B.; Arrigoni, F.; Zampella, G. Challenges in the Synthesis of Active Site Mimics for [NiFe]-Hydrogenases. *Organometallics* **2021**, *40*, 3306–3312.
- (27) Song, L.-C.; Chen, S.; Han, X.-F.; Zhang, Z.-Q.; Wang, Y.-P.; Dong, Y.-X. Synthesis, Structures and Chemical Reactivity of Dithiolato-Bridged Ni-Fe Complexes as Biomimetics for the Active Site of [NiFe]-Hydrogenases. *Inorganics* **2022**, *10*, 90.
- (28) Song, L. C.; Wang, Y. P.; Dong, Y. X.; Yang, X. Y. Functionalized nickel(II)-iron(II) dithiolates as biomimetic models of [NiFe]-H₂ases. *Dalton Trans.* **2023**, *52*, 3755–3768.
- (29) Zhang, F.; Woods, T. J.; Rauchfuss, T. B. Hybrids of [FeFe]- and [NiFe]-H₂ase Active Site Models. *Organometallics* **2023**, *42*, 1607–1614.
- (30) Lubitz, W.; Reijerse, E.; van Gestel, M. [NiFe] and [FeFe] Hydrogenases Studied by Advanced Magnetic Resonance Techniques. *Chem. Rev.* **2007**, *107*, 4331–4365.
- (31) Ogata, H.; Lubitz, W.; Higuchi, Y. [NiFe] hydrogenases: Structural and spectroscopic studies of the reaction mechanism. *Dalton Trans.* **2009**, 7577–7587.
- (32) Barton, B. E.; Whaley, C. M.; Rauchfuss, T. B.; Gray, D. L. Nickel-iron dithiolato hydrides relevant to the [NiFe]-hydrogenase active site. *J. Am. Chem. Soc.* **2009**, *131*, 6942–6943.
- (33) Barton, B. E.; Rauchfuss, T. B. Hydride-Containing Models for the Active Site of the Nickel-Iron Hydrogenases. *J. Am. Chem. Soc.* **2010**, *132*, 14877–14885.
- (34) Carroll, M. E.; Barton, B. E.; Gray, D. L.; Mack, A. E.; Rauchfuss, T. B. Active-Site Models for the Nickel-Iron Hydrogenases: Effects of Ligands on Reactivity and Catalytic Properties. *Inorg. Chem.* **2011**, *50*, 9554–9563.
- (35) Ogo, S.; Ichikawa, K.; Kishima, T.; Matsumoto, T.; Nakai, H.; Kusaka, K.; Ohhara, T. A functional [NiFe]hydrogenase mimic that catalyzes electron and hydride transfer from H₂. *Science* **2013**, *339*, 682–684.
- (36) Ogo, S.; Kishima, T.; Yatabe, T.; Miyazawa, K.; Yamasaki, R.; Matsumoto, T.; Ando, T.; Kikkawa, M.; Isegawa, M.; Yoon, K. S.; Hayami, S. [NiFe], [FeFe], and [Fe] hydrogenase models from isomers. *Sci. Adv.* **2020**, *6*, No. eaaz8181.
- (37) Basu, D.; Bailey, T. S.; Lalaoui, N.; Richers, C. P.; Woods, T. J.; Rauchfuss, T. B.; Arrigoni, F.; Zampella, G. Synthetic Designs and Structural Investigations of Biomimetic Ni–Fe Thiolates. *Inorg. Chem.* **2019**, *58*, 2430–2443.
- (38) Huynh, M. T.; Schilter, D.; Hammes-Schiffer, S.; Rauchfuss, T. B. Protonation of Nickel-Iron Hydrogenase Models Proceeds after Isomerization at Nickel. *J. Am. Chem. Soc.* **2014**, *136*, 12385–12395.
- (39) Ulloa, O. A.; Huynh, M. T.; Richers, C. P.; Bertke, J. A.; Nilges, M. J.; Hammes-Schiffer, S.; Rauchfuss, T. B. Mechanism of H₂ Production by Models for the NiFe -Hydrogenases: Role of Reduced Hydrides. *J. Am. Chem. Soc.* **2016**, *138*, 9234–9245.
- (40) Schilter, D.; Camara, J. M.; Huynh, M. T.; Hammes-Schiffer, S.; Rauchfuss, T. B. Hydrogenase Enzymes and Their Synthetic Models: The Role of Metal Hydrides. *Chem. Rev.* **2016**, *116*, 8693–8749.

- (41) Simmons, T. R.; Berggren, G.; Bacchi, M.; Fontecave, M.; Artero, V. Mimicking hydrogenases: From biomimetics to artificial enzymes. *Coord. Chem. Rev.* **2014**, *270*, 127–150.
- (42) Kaur-Ghumaan, S.; Stein, M. [NiFe] hydrogenases: How close do structural and functional mimics approach the active site? *Dalton Trans.* **2014**, *43*, 9392–9405.
- (43) Ding, S.; Ghosh, P.; Darensbourg, M. Y.; Hall, M. B. Interplay of hemilability and redox activity in models of hydrogenase active sites. *Proc. Natl. Acad. Sci. U. S. A.* **2017**, *114*, No. E9775–E9782.
- (44) Ding, S.; Ghosh, P.; Lunsford, A. M.; Wang, N.; Bhuvanesh, N.; Hall, M. B.; Darensbourg, M. Y. Hemilabile Bridging Thiolates as Proton Shuttles in Bioinspired H₂ Production Electrocatalysts. *J. Am. Chem. Soc.* **2016**, *138*, 12920–12927.
- (45) Tang, H.; Hall, M. B. Biomimetics of [NiFe]-Hydrogenase: Nickel- or Iron-Centered Proton Reduction Catalysis? *J. Am. Chem. Soc.* **2017**, *139*, 18065–18070.
- (46) Brazzolotto, D.; Wang, L.; Tang, H.; Gennari, M.; Queyriaux, N.; Philouze, C.; Demeshko, S.; Meyer, F.; Orio, M.; Artero, V.; Hall, M. B.; Duboc, C. Tuning Reactivity of Bioinspired [NiFe]-Hydrogenase Models by Ligand Design and Modeling the CO Inhibition Process. *ACS Catal.* **2018**, *8*, 10658–10667.
- (47) Wang, L.; Gennari, M.; Barrozo, A.; Fize, J.; Philouze, C.; Demeshko, S.; Meyer, F.; Orio, M.; Artero, V.; Duboc, C. Role of the Metal Ion in Bio-Inspired Hydrogenase Models: Investigation of a Homodinuclear FeFe Complex vs Its Heterodinuclear NiFe Analogue. *ACS Catal.* **2020**, *10*, 177–186.
- (48) Niu, S. Q.; Hall, M. B. Theoretical studies on reactions of transition-metal complexes. *Chem. Rev.* **2000**, *100*, 353–405.
- (49) Yang, X.; Elrod, L. C.; Le, T.; Vega, V. S.; Naumann, H.; Rezenom, Y.; Reibenspies, J. H.; Hall, M. B.; Darensbourg, M. Y. Controlling O₂ Reactivity in Synthetic Analogues of [NiFeS]- and [NiFeSe]-Hydrogenase Active Sites. *J. Am. Chem. Soc.* **2019**, *141*, 15338–15347.
- (50) Yang, X.; Elrod, L. C.; Reibenspies, J. H.; Hall, M. B.; Darensbourg, M. Y. Oxygen uptake in complexes related to [NiFeS]- and [NiFeSe]-hydrogenase active sites. *Chem. Sci.* **2019**, *10*, 1368–1373.
- (51) Brazzolotto, D.; Gennari, M.; Queyriaux, N.; Simmons, T. R.; Pecaut, J.; Demeshko, S.; Meyer, F.; Orio, M.; Artero, V.; Duboc, C. Nickel-centred proton reduction catalysis in a model of [NiFe] hydrogenase. *Nat. Chem.* **2016**, *8*, 1054–1060.
- (52) Cramer, C. J. *Essentials of computational chemistry: Theories and models*, 2nd ed.; John Wiley & Sons Inc., 2004.
- (53) Lewars, E. *Computational chemistry introduction to the theory and applications of molecular and quantum mechanics*, 2nd ed.; Springer: Dordrecht, 2011.
- (54) Weinhold, F.; Landis, C. R. *Discovering Chemistry With Natural Bond Orbitals*; John Wiley & Sons Inc.: Hoboken, NJ, 2012.
- (55) Cortesguzman, F.; Bader, R. F. W. Complementarity of QTAIM and MO theory in the study of bonding in donor-acceptor complexes. *Coord. Chem. Rev.* **2005**, *249*, 633–662.
- (56) Cramer, C. J.; Truhlar, D. G. Density functional theory for transition metals and transition metal chemistry. *Phys. Chem. Chem. Phys.* **2009**, *11*, 10757–10816.
- (57) Niu, S. Q.; Nelson, A. E.; De La Torre, P.; Li, H.; Works, C. F.; Hall, M. B. Photoinduced Terminal Hydride of [FeFe]-Hydrogenase Biomimetic Complexes. *Inorg. Chem.* **2019**, *58*, 13737–13741.
- (58) Niu, S. Q.; Ichiye, T. Density functional theory calculations of redox properties of iron-sulphur protein analogues. *Mol. Simul.* **2011**, *37*, 572–590.
- (59) Niu, S. Q.; Huang, D. L.; Dau, P. D.; Liu, H. T.; Wang, L. S.; Ichiye, T. Assessment of Quantum Mechanical Methods for Copper and Iron Complexes by Photoelectron Spectroscopy. *J. Chem. Theory Comput.* **2014**, *10*, 1283–1291.
- (60) Marenich, A. V.; Cramer, C. J.; Truhlar, D. G. Universal solvation model based on solute electron density and on a continuum model of the solvent defined by the bulk dielectric constant and atomic surface tensions. *J. Phys. Chem. B* **2009**, *113*, 6378–6396.
- (61) Aprà, E.; Bylaska, E. J.; Jong, W. A. D.; Govind, N.; Kowalski, K.; Straatsma, T. P.; Valiev, M.; Dam, H. J. J. V.; Alexeev, Y.; Anchell, J.; et al. NWChem: Past, present, and future. *J. Chem. Phys.* **2020**, *152* (18), 184102.
- (62) Frisch, M. J.; Trucks, G.; Schlegel, H. B.; Scuseria, G. E.; Robb, M. A.; Cheeseman, J. R.; Scalmani, G.; Barone, V.; Petersson, G. A.; Nakatsuji, H.; et al. *Gaussian 09, Revision A.01*; Gaussian, Inc.: Wallingford CT, 2009.
- (63) Keith, T. A. *AIMAll*; TK Gristmill Software: Overland Park KS, USA, 2019.
- (64) Buntrock, R. E. ChemOffice Ultra 7.0. *J. Chem. Inf. Comput. Sci.* **2002**, *42*, 1505–1506.
- (65) Schaftenaar, G.; Noordik, J. H. Molden: A pre- and post-processing program for molecular and electronic structures. *J. Comput.-Aided Mol. Des.* **2000**, *14*, 123–134.
- (66) Pettersen, E. F.; Goddard, T. D.; Huang, C. C.; Couch, G. S.; Greenblatt, D. M.; Meng, E. C.; Ferrin, T. E. UCSF Chimera—A visualization system for exploratory research and analysis. *J. Comput. Chem.* **2004**, *25*, 1605–1612.
- (67) Crouthers, D. J.; Denny, J. A.; Bethel, R. D.; Munoz, D. G.; Darensbourg, M. Y. Conformational Mobility and Pendent Base Effects on Electrochemistry of Synthetic Analogues of the [FeFe]-Hydrogenase Active Site. *Organometallics* **2014**, *33*, 4747–4755.