





Complete Genome Analysis of Flavobacterium psychrophilum Strain FPRT1, Isolated from Diseased Rainbow Trout (Oncorhynchus mykiss) in South Korea

Jiyeon Park, a Hyeong Jin Roh, a Yoonhang Lee, a Junewoo Park, a Do-Hyung Kima

^aDepartment of Aquatic Life Medicine, Pukyong National University, Busan, Republic of Korea

ABSTRACT Here, we report the complete genome sequence of Flavobacterium psychrophilum FPRT1, isolated from the spleen and kidney of diseased rainbow trout (Oncorhynchus mykiss). Whole-genome sequencing was performed using the PacBio RS II platform, which yielded a circular chromosome of 2,795,347 bp harboring 2,895 protein-coding genes.

lavobacterium psychrophilum is a Gram-negative bacterial pathogen that causes bacterial cold-water disease and rainbow trout fry syndrome (1, 2). The clinical signs of diseased rainbow trout in this study included ulcerative lesions in the mandible, pale gills (anemia), exophthalmia, and increased mucus secretion. Fifty rainbow trout (body weight, \sim 70 g) were purchased from a fish farm in South Korea. Fish were acclimated in a 500-liter tank and maintained at 15°C. Freshly dead fish were aseptically dissected, and swabbed materials derived from the kidney and spleen were placed onto tryptone-glucose-yeast extract (TGYE) agar. Bacterial cultures isolated as a pure culture were used to extract DNA using the AccuPrep genomic DNA extraction kit (Bioneer, South Korea). PCR was performed using the primers 27F (5'-AGAGTTTGATCMTGGCTCAG-3') and 1492R (5'-TACGGYTACCTTGTTACGACTT-3') for 16S rRNA gene sequencing. The organism was identified as F. psychrophilum (identity, 100%). F. psychrophilum FPRT1 growth required at least 72 h of incubation on TGYE agar at 15°C under aerobic conditions and was stored in TGYE broth supplemented with 20% glycerol at -80°C. Antibiotic susceptibility testing according to the guidelines of the Clinical and Laboratory Standards Institute (3) showed inhibition zones were not formed for oxolinic acid, trimethoprim, and sulfamethoxazole, indicating that this strain was resistant to sulfa drugs and the quinolone.

A single colony of F. psychrophilum FPRT1 was inoculated into TGYE broth and incubated at 15°C for 36 h for DNA extraction. Genomic DNA of FPRT1 was extracted using a Wizard genomic DNA purification kit (Promega, USA) according to the manufacturer's protocol. The whole-genome sequencing was performed using the PacBio RS II (Pacific Biosciences, USA) platform. Library preparation was performed using single-molecule real-time (SMRT) cell 8Pac V3 and DNA polymerase binding kit P6 (Pacific Biosciences). Genomic DNA was sheared with g-TUBE (Covaris, USA) into a 20-kb size and purified using AMPure PB magnetic beads (Beckman Coulter, USA). A bioanalyzer 2100 instrument (Agilent, USA) was used to determine the actual size distribution. After subread filtering of the PacBio raw data (minimum polymerase read quality, 0.80), 132,029 PacBio subreads (average subread length, 10,730 bp; subread N_{50} , 15,690 bp) of FPRT1 were generated. Raw sequences were assembled de novo using Hierarchical Genome Assembly Process 3 (HGAP3), and error correction was conducted using Quiver (4). When both ends of the contig overlapped, contigs were connected to form a circular shape, and overlapped regions were manually trimmed. The coding DNA sequence (CDS), tRNA, and rRNA genes on each contig were predicted and annotated using Rapid Annotations using Subsystems Technology (RAST) and SEED viewer (5, 6). Antibiotic resistance-related genes

Citation Park J, Roh H, Lee Y, Park J, Kim D-H. 2021. Complete genome analysis of Flavobacterium psychrophilum strain FPRT1. isolated from diseased rainbow trout (Oncorhynchus mykiss) in South Korea. Microbiol Resour Announc 10:e00151-21. https://doi.org/10.1128/MRA.00151-21.

Editor Frank J. Stewart, Georgia Institute of

Copyright © 2021 Park et al. This is an openaccess article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Do-Hyung Kim, dhkim@pknu.ac.kr.

Received 9 February 2021 Accepted 9 March 2021 Published 25 March 2021

Park et al.

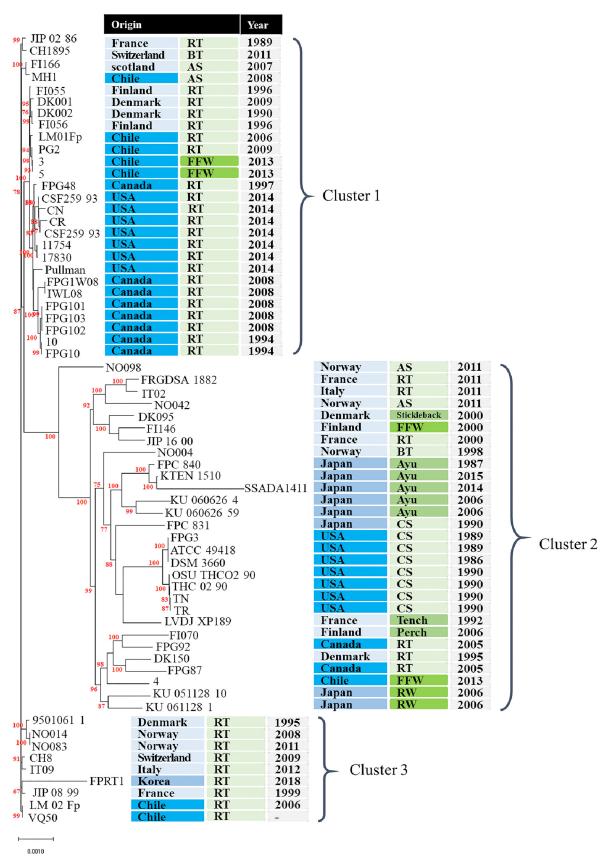


FIG 1 Neighbor-joining phylogenetic tree of 65 strains of *Flavobacterium psychrophilum* based on 831 orthologous genes. Bootstrap values (only >50%) from 500 replicates are indicated at each node. Abbreviations: RT, rainbow trout; BT, brown trout; AS, Atlantic salmon; CS, Coho salmon; FFW, freshwater fish farm water; RW, river water.

Volume 10 Issue 12 e00151-21 mra.asm.org **2**



were searched in the Comprehensive Antibiotic Resistance Database (CARD) (7). Default settings were used for all software.

The genome consisted of 2,795,347 bp in a single circular chromosome with an overall G+C content of 32.6%, containing 2,895 CDS, 49 tRNA, and 14 rRNA genes. Additionally, phylogenetic analysis (8) based on 831 orthologous genes retrieved from 65 publicly available *F. psychrophilum* genome sequences showed that they were phylogenetically divided into 3 clusters (Fig. 1). Our strain was most closely related to isolates originating from some European countries and Chile within cluster 3. In addition, strain FPRT1 harbors several antibiotic resistance-related genes, such as *emrA* and *emrB*, that encode multidrug efflux pumps in the MFS family (9). This is the first report of genomic sequencing of *F. psychrophilum* from infected fish in South Korea. In the era of whole-genome sequencing, this study might provide important epidemiological information on *F. psychrophilum*.

Data availability. The whole-genome sequence of *F. psychrophilum* FPRT1 has been deposited in the GenBank database with the accession number CP059061.1. The raw sequencing data are available under accession number SRR11814644. The BioProject and BioSample numbers are PRJNA633452 and SAMN14943929, respectively.

ACKNOWLEDGMENT

This research was supported by the Basic Science Research Program through the National Research Foundation of Korea (NRF) funded by the Ministry of Education, Science and Technology (2018R1D1A1A09083861).

REFERENCES

- Cipriano RC, Holt RA. 2005. Flavobacterium psychrophilum, cause of bacterial cold-water disease and rainbow trout fry syndrome. Fish disease leaflet no. 86. U.S. Department of the Interior. U.S. Geological Service, National Fish Health Research Laboratory, Kearneysville, WV.
- Lorenzen E, Dalsgaard I, Bernardet JF. 1997. Characterization of isolates of Flavobacterium psychrophilum associated with cold water disease or rainbow trout fry syndrome I: phenotypic and genomic studies. Dis Aquat Org 31:197–208. https://doi.org/10.3354/dao031197.
- Clinical and Laboratory Standards Institute. 2006. Methods for antimicrobial disk susceptibility testing of bacteria isolated from aquatic animals, approved guideline M42-A. Clinical and Laboratory Standards Institute, Wayne, PA.
- Chin CS, Alexander DH, Marks P, Klammer AA, Drake J, Heiner C, Clum A, Copeland A, Huddleston J, Eichler EE, Turner SW, Korlach J. 2013. Nonhybrid, finished microbial genome assemblies from long-read SMRT sequencing data. Nat Methods 10:563–569. https://doi.org/10.1038/nmeth.2474.
- Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST

- server: Rapid Annotations using Subsystems Technology. BMC Genomics 9:75. https://doi.org/10.1186/1471-2164-9-75.
- Overbeek R, Olson R, Pusch GD, Olsen GJ, Davis JJ, Disz T, Edwards RA, Gerdes S, Parrello B, Shukla M, Vonstein V, Wattam AR, Xia F, Stevens R. 2014. The SEED and the Rapid Annotation of microbial genomes using Subsystems Technology (RAST). Nucleic Acids Res 42:D206–D214. https://doi.org/10.1093/nar/qkt1226.
- McArthur AG, Waglechner N, Nizam F, Yan A, Azad MA, Baylay AJ, Bhullar K, Canova MJ, De Pascale G, Ejim L, Kalan L, King AM, Koteva K, Morar M, Mulvey MR, O'Brien JS, Pawlowski AC, Piddock LJV, Spanogiannopoulos P, Sutherland AD, Tang I, Taylor PL, Thaker M, Wang W, Yan M, Yu T, Wright GD. 2013. The comprehensive antibiotic resistance database. Antimicrob Agents Chemother 57:3348–3357. https://doi.org/10.1128/AAC.00419-13.
- Blom J, Kreis J, Spänig S, Juhre T, Bertelli C, Ernst C, Goesmann A. 2016. EDGAR 2.0: an enhanced software platform for comparative gene content analyses. Nucleic Acids Res 44:W22–W28. https://doi.org/10.1093/nar/gkw255.
- Lomovskaya O, Lewis K. 1992. Emr, an Escherichia coli locus for multidrug resistance. Proc Natl Acad Sci U S A 89:8938–8942. https://doi.org/10.1073/ pnas.89.19.8938.

Volume 10 lssue 12 e00151-21 mra.asm.org **3**