Microdamage Caused by Fatigue Loading in Human Cancellous Bone: Relationship to Reductions in Bone Biomechanical Performance

Floor M. Lambers¹, Amanda R. Bouman¹, Clare M. Rimnac³, Christopher J. Hernandez^{1,2,4}*

1 Sibley School of Mechanical and Aerospace Engineering, Cornell University, Ithaca, New York, United States of America, 2 Department of Biomedical Engineering, Cornell University, Ithaca, New York, United States of America, 3 Department of Mechanical and Aerospace Engineering, Case Western Reserve University, Cleveland, Ohio, United States of America, 4 Hospital for Special Surgery, New York, United States of America

Abstract

Vertebral fractures associated with osteoporosis are often the result of tissue damage accumulated over time. Microscopic tissue damage (microdamage) generated in vivo is believed to be a mechanically relevant aspect of bone quality that may contribute to fracture risk. Although the presence of microdamage in bone tissue has been documented, the relationship between loading, microdamage accumulation and mechanical failure is not well understood. The aim of the current study was to determine how microdamage accumulates in human vertebral cancellous bone subjected to cyclic fatigue loading. Cancellous bone cores (n = 32) from the third lumbar vertebra of 16 donors (10 male, 6 female, age 76 ± 8.8 , mean \pm SD) were subjected to compressive cyclic loading at $\sigma/E_0 = 0.0035$ (where σ is stress and E_0 is the initial Young's modulus). Cyclic loading was suspended before failure at one of seven different amounts of loading and specimens were stained for microdamage using lead uranyl acetate. Damage volume fraction (DV/BV) varied from 0.8±0.5% (no loading) to 3.4±2.1% (fatigue-loaded to complete failure) and was linearly related to the reductions in Young's modulus caused by fatigue loading ($r^2 = 0.60$, p<0.01). The relationship between reductions in Young's modulus and proportion of fatigue life was nonlinear and suggests that most microdamage generation occurs late in fatigue loading, during the tertiary phase. Our results indicate that human vertebral cancellous bone tissue with a DV/BV of 1.5% is expected to have, on average, a Young's modulus 31% lower than the same tissue without microdamage and is able to withstand 92% fewer cycles before failure than the same tissue without microdamage. Hence, even small amounts of microscopic tissue damage in human vertebral cancellous bone may have large effects on subsequent biomechanical performance.

Citation: Lambers FM, Bouman AR, Rimnac CM, Hernandez CJ (2013) Microdamage Caused by Fatigue Loading in Human Cancellous Bone: Relationship to Reductions in Bone Biomechanical Performance. PLoS ONE 8(12): e83662. doi:10.1371/journal.pone.0083662

Editor: Ryan K. Roeder, University of Notre Dame, United States of America

Received August 28, 2013; Accepted November 13, 2013; Published December 30, 2013

Copyright: © 2013 Lambers et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Funding: This publication was made possible by Grant Number R01 AR057362 from NIAMS/NIH (PI CJH). The authors acknowledge use of human vertebral bodies provided by the National Disease Research Interchange (NDRI, Philadelphia, PA), with support from NIH grant 8U42OD011158-22. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.

* E-mail: cjh275@cornell.edu

Introduction

Vertebral fractures are the most common form of osteoporosisrelated fractures [1,2]. Only 51% of all vertebral fractures are associated with a discrete loading event, suggesting that many vertebral fractures are the result of tissue damage caused by multiple loading events over time [3,4]. Cancellous bone is the primary load-carrying component in human vertebral bodies, suggesting that damage accumulation and related degradation in biomechanical performance of vertebral cancellous bone is part of the development of vertebral fractures [5].

Microscopic tissue damage (microdamage) in the form of linear microcracks, diffuse damage, or trabecular microfracture occurs *in vivo* [6,7], shows a higher prevalence in older individuals [8,9,10], and is believed to be a biomechanically relevant aspect of bone quality [11]. Microdamage in bone tissue can be visualized through bulk staining with basic fuchsin [12] or fluorochromes [13,14] followed by sectioning and histomorphometry. Alternatively microdamage may be stained using radio-opaque contrast agents combined with x-ray computed tomography (micro-CT,

etc) [15,16,17]. Previous studies have demonstrated that greater amounts of cyclic loading applied to cortical bone tissue are related to greater amounts of microdamage [18,19,20,21,22,23]. In cancellous bone, relations between a single loading event and the amount of resulting microdamage have been reported [16,24,25,26], but less is known about microdamage accumulation during cyclic fatigue loading.

Cancellous bone tissue submitted to cyclic loading displays creep-fatigue behavior in three phases: a primary phase in which stiffness reduces relatively rapidly, a secondary phase in which stiffness declines slowly and a tertiary phase in which stiffness rapidly declines before failure (**Fig. 1**) [27,28,29]. Microdamage generated during fatigue loading has been examined in bovine cancellous bone [13,30,31] and in human femoral cancellous bone after an initial overload [32], but quantitative relationships between the amount of fatigue loading, microdamage and alterations in biomechanical performance in less dense cancellous bone such as that in the vertebral body have not yet been presented. Human vertebral cancellous bone is more rod-like and



Figure 1. Fatigue loading of cancellous bone. (A) A creep-fatigue curve is shown. Lines represent minimum and maximum strain per cycle, as indicated in the inset. The creep-fatigue curve consists of three phases: the primary phase where the strain accumulation per cycle decreases, the secondary phase where the strain accumulation per cycle is constant, and the tertiary phase where the strain accumulation per cycle increases. (B) Stress-strain plots for the first load cycle (N_1) and a later load cycle (N_i) are shown. Mechanical properties measured within the loading cycle include the Young's modulus (E, shown in red), energy dissipation (U, measured as the area labeled in blue), maximum strain_i = creep strain_i+cyclic strain_i, doi:10.1371/journal.pone.0083662.g001

porous than cancellous bone tissue from the femoral neck and is therefore expected to show different failure processes [33].

The overall goal of the current line of investigation is to understand the contributions of microdamage to vertebral fractures. Specifically, the aim of the current study is to determine the relationship between the amount of fatigue loading, the accumulation of microdamage, and reductions in the Young's modulus during cyclic loading of human vertebral cancellous bone.

Methods

Sample Preparation

The third lumbar vertebral bodies of 16 donors (10 male, 6 female, mean age 76±8.8 years, range 62-92 years) were examined (tissue from NDRI, Philadelphia, PA). The work examined human tissue acquired from a licensed tissue bank (http://ndriresource.org/) and was therefore exempt from Institutional Review Board evaluation. Donors had no medical history of metabolic bone disease or cancer. Vertebrae were dissected free of soft tissue and cylindrical cores of cancellous bone (nominally 8 mm diameter, 19 mm in length) aligned in the superior-inferior direction (as described previously [34]) were collected using a diamond tipped coring tool (Starlite Industries, Bryn Mawr, PA, USA). Trabecular alignment was ensured by aligning the vertebrae visually during specimen collection. Two specimens were collected from each vertebra resulting in a total of 32 specimens. Specimens from the anterior, left, or right sites in the vertebra were included in the study. Little variation in mechanical properties has been observed among anterior, left and right sites within lumbar vertebrae [35,36,37]. Samples from the posterior side were avoided because of bone volume fraction inhomogeneity due to the vascular supply. Specimens were wrapped in saline soaked gauze and stored in airtight containers at $-20^{\circ}C$ until mechanical testing. Prior to mechanical testing, bone marrow was removed from the specimens with a low-pressure water jet. The specimens were glued (Loctite 401, Newington, CT, USA) into brass cylindrical end-caps as previously described [34]. The cyanoacrylate glue was allowed to cure overnight at 4°C, with specimens wrapped in saline soaked gauze to maintain hydration.

Mechanical Loading

Fatigue loading was applied to the cancellous bone cores to induce microdamage. Mechanical loading was conducted at room temperature (23°C) using a servohydraulic testing machine (858 Mini Bionix, MTS, Eden Prairie, MN, USA) with a 1 kN load cell (Model 661.18E-01, MTS, Eden Prairie, MN, USA, USA). Mechanical properties degrade faster at higher temperatures [28,38,39], thus testing at body temperature would have accelerated mechanical degradation compared to testing at room temperature. By testing at room temperature in the current study, microdamage propagated more slowly and we were better able to stop fatigue loading at prescribed amounts of fatigue loading. Strain was measured with a 25 mm gage length extensometer attached to the end-caps. The effective gage length was defined as the exposed plus half the embedded length of the specimen [34]. During testing, specimens were hydrated with physiologically buffered saline (pH of 7.4) containing 10 µM protease inhibitors (E-64, Sigma Aldrich, St. Louis, MO, USA). A rubber membrane placed around the specimen was used to contain the solution. Mechanical loading consisted of an initial set of ten preconditioning cycles between 0 and 0.1% strain at a rate of 0.5%/sec. After preconditioning, the specimens were submitted to cyclic loading between 0 N and a load corresponding to a value of σ/E_0 of 0.0035 mm/mm, where σ is stress and E₀ is the initial Young's modulus. Fatigue loading was performed at 4 Hz in load control using a haversine waveform.

Six specimens were loaded to 5% strain, which we defined as failure (group 7). The value of 5% strain was defined as failure, because from preliminary tests it was found that loading to 5% strain would capture the primary, secondary, and tertiary phase. Our definition is conservative in that others have defined failure at lower strain magnitudes (2.5% [30] or even "the cycle before which a specimen could no longer sustain the applied stress" which was at a strain of $3.36\% \pm 2.13\%$ [29]). Five specimens were not subjected to fatigue loading (group 1). Fatigue loading in five other groups was stopped before failure based on monitoring the creep-fatigue curve, secant modulus, and hysteresis loops to achieve specimens loaded to a specified region of the creep-fatigue curve. Fatigue loading was stopped at the start of the secondary phase (group 2), in the middle of the secondary phase (group 3), and at three different points within the tertiary phase (groups 4-6). Specimen group assignment was performed to evenly distribute male and female donors and ensure that multiple specimens from

the same donor were not in the same group. After loading, the stress-strain curves were examined to confirm group classification based on reduction in Young's modulus $(1 - E/E_0)$, and specimens were reassigned group number if necessary. Eleven specimens were reassigned group based on the Young's modulus reduction. Reassignment did not alter the relatively even distribution of male/female donors and specimens among donors. The distribution in Young's modulus reduction is shown in Figure 2B (inset): For group 1 (n=5) no fatigue loading was applied, for the other groups the Young's modulus reduction was 6-9% (group 2, n=4), 12-17% (group 3, n=3), 32-35% (group 4, n=4), 37-43% (group 5, n=5), 44-51% (group 6, n=4) or 57-82% (group 7, n=6).

Mechanical Properties

Force and strain data were collected every cycle at 102.4 Hz, and at 1024 Hz every tenth cycle and during the last 10 cycles (MTS TestSuite Multipurpose Elite/FlexTest 40 digital controller, MTS, Eden Prairie, MN, USA). Young's modulus was calculated from the data sampled at 1024 Hz. Young's modulus (E) was determined with a polynomial fit of the first 0.2% portion of the loading segment [40]. Force data was converted to stress using the cross-sectional area of the specimen gage region. Creep strain, cyclic strain, and maximum strain were determined per cycle (Fig. 1B). Creep strain was defined as the minimum strain and cyclic strain as the difference between the maximum and minimum strain per cycle [41,42]. Maximum strain was the sum of the creep and cyclic strain [28,29]. Energy dissipation (U), which describes the amount of energy lost per cycle, was calculated as the area within the hysteresis loop of each cycle. The number of cycles to failure (Nf) was determined for specimens loaded to failure (defined as achieving a maximum strain of 5%).

Microdamage and Bone Microarchitecture

After mechanical testing, specimens were removed from the end-caps using a low-speed diamond saw (Isomet, Buehler Ltd., Lake Bluff, IL) and stained with lead uranyl acetate. Lead uranyl acetate binds to regions of bone tissue in which microdamage has formed [43] and has high x-ray absorption, allowing visualization of microdamage with micro-computed tomography [16,17]. Lead uranyl acetate staining was performed as follows (derived from [16]): cylindrical cores were dehydrated in 70% acetone for 24 hours. Next, the specimens were stained for one week in a solution of equal parts 8% uranyl acetate (Ted Pella, Inc., Redding, CA, USA) in 70% acetone and 20% lead (II) acetate (Sigma-Aldrich, St. Louis, MO, USA) in 70% acetone, while on a shaker covered to limit exposure to light. Thereafter, the samples were placed in a 1% ammonium sulfide (Sigma-Aldrich, St. Louis, MO, USA) in 70% acetone solution for one week with one change of solution at 3 days. Samples were rinsed with 70% acetone, submersed in 70% acetone for one week, rinsed again and submitted to ultrasonic vibration for 10 minutes in 70% acetone to remove the solution from the pores. Specimens were glued on metal nails. Images were collected with X-ray microscopy (VersaXRM-500, Xradia, Pleasanton, CA, USA) in air at an energy of 100 kVp, a current of 90 µA, an integration time of 1000 ms, and an isotropic voxel size of 10 µm. The region of interest was 10.2 mm in length and contained the complete cross-sections of the specimen. The volume analyzed was 7.5 mm in diameter (at least 250 µm away from machined edges) and 8.2 mm in length (central slices), unless the specimen was shorter, in which case the volume analyzed started 500 µm below the machined edge. To segment bone, a Gaussian filter (sigma = 1.2, support = 1) was applied to the images, followed by a global threshold, determined with the Otsu method. To segment microdamage from bone, manual thresholds were chosen by an observer blinded to the experimental group of each specimen. Bone microarchitecture was quantified using a three-dimensional approach (IPL, Scanco Medical, Brüttisellen, Switzerland), including bone volume fraction (BV/TV), trabecular thickness, trabecular number, bone surface, structure model index, connectivity density, and degree of anisotropy. Damage volume fraction was calculated as the damage volume divided by the bone volume (DV/BV).

Statistical Analysis

Regression was performed to determine the relationship between loading group (as an ordinal) and mechanical properties or DV/BV. Regression models were also performed to determine relations between microdamage, mechanical properties and measures of cancellous microarchitecture. Donor was included as a random effect to take into account the use of multiple specimens from each donor. Statistical tests were conducted using JMP 9 (SAS Institute Inc., Cary, NC, USA) or MATLAB (7.12.1, R2011a, The MathWorks Inc., Natick, MA, USA).



Figure 2. Reduction in Young's modulus with fatigue loading. (A) Reduction in Young's modulus for 6 samples loaded to failure is shown. Each color represents a different specimen. (B) The average reduction in Young's modulus of all samples loaded to failure is shown relative to the proportion of fatigue life used. The numbers and colored ranges on the curve indicate the points where fatigue loading was stopped for each group (the range shown in the inset). doi:10.1371/journal.pone.0083662.q002

PLOS ONE | www.plosone.org

Results

Damage Volume Fraction

Damage volume fraction was increased in specimens receiving more fatigue loading ($r^2 = 0.71$, p < 0.01, **Fig. 3B**). DV/BV was 0.8% $\pm 0.6\%$ (mean \pm SD) in samples with no fatigue loading and 3.4% $\pm 2.1\%$ in the group loaded to failure. Although there was considerable variability within each loading group, within a donor, DV/BV was greater in specimens with greater amounts of fatigue loading (visualization of specimens from the same donor in **Fig. 4**).

Mechanical Properties

Young's modulus was reduced in specimens receiving more fatigue loading ($r^2 = 0.96$, p<0.01, **Fig. 3A**). Maximum strain was increased in specimens receiving more fatigue loading, but did not exceed 2% in the groups where cyclic loading was stopped before failure (Groups 1-6, Fig. 3C). Maximum energy dissipation ranged between 0.1 µJ/mm³ and 1.3 µJ/mm³ for groups 2–6, and increased to an average of 4.6 μ J/mm³ for group 7 (r² = 0.65, p<0.01, Fig. 3D). Early during fatigue loading the maximum strain was similar to the cyclic strain, but with continued loading, creep strain increased (Fig. 5A), indicating that changes in maximum strain were mostly an effect of accumulation of creep strain. Maximum strain was related to both creep strain ($r^2 = 0.95$, p < 0.01) and cyclic strain ($r^2 = 0.64$, p < 0.01, **Table 1**). In contrast to the continuous reduction in Young's modulus throughout fatigue loading, energy dissipation remained relatively constant and only increased in the tertiary phase (Fig. 5B).

Relationship between Damage Volume Fraction and Mechanical Properties

Greater amounts of microdamage were associated with reductions in mechanical properties (Fig. 6). DV/BV was

positively related to modulus reduction ($r^2 = 0.60$, p < 0.01) and maximum strain ($r^2 = 0.61$, p < 0.01). Creep strain ($r^2 = 0.57$, p < 0.01), cyclic strain ($R^2 = 0.54$, p < 0.01), and energy dissipation ($r^2 = 0.53$, p < 0.01) were positively correlated with DV/BV (**Table 1, Table 2, Fig. S1**).

Relationship between Damage Volume Fraction and Fatigue Life

The relationship between reductions in Young's modulus and proportion of fatigue life was consistent among specimens loaded to failure (n = 6, **Fig. 2A**) allowing quantitative estimates of the relationship between DV/BV and proportion of fatigue life by combining the relationship between the reduction in Young's modulus and proportion of fatigue life (**Fig. 2B**) with the relationship between the reduction in Young's modulus and DV/BV (**Fig. 6A**). The relationship between DV/BV and the proportion of fatigue life was nonlinear, with most damage accumulation late in the fatigue life (**Fig. 7**). A DV/BV of 1.5% (95% CI: 1.1–1.9%) corresponded, on average, to a 31% reduction in Young's modulus and to 92% (95% CI: 88–96%) of the fatigue life. A DV/BV of 2% was associated, on average, with 98% of the fatigue life.

Effect of Cancellous Microarchitecture on Microdamage Accumulation

No differences in bone microarchitectural parameters (**Table 3**) were detected among groups. No correlations between DV/BV and microarchitecture were observed (**Table 2, Fig. S1**). Including microarchitectural parameters as a covariate in the regression between DV/BV and mechanical properties did not improve the correlation coefficients.



Figure 3. The distribution of DV/BV and mechanical properties for each of the groups. Colors represent different donors. Female donors are shown as circles, male donors as squares. Lines connect specimens from the same donor. (A) Reduction in Young's modulus ($r^2 = 0.96$, p < 0.01), (B) Damage volume fraction ($r^2 = 0.71$, p < 0.01), (C) Maximum strain ($r^2 = 0.93$, p < 0.01), and (D) Maximum energy dissipation ($r^2 = 0.65$, p < 0.01) were increased in groups experiencing more fatigue loading. doi:10.1371/journal.pone.0083662.q003



Figure 4. Visualization of microdamage in cancellous bone. Red represents microdamage and transparent white represents bone. Shown are two specimens from the same donor subjected to (A) No loading (Group 1) and (B) Fatigue loading in group 5 (Young's modulus was reduced by 42%). (C) An enlarged view of a damage site in the cancellous bone is shown. doi:10.1371/journal.pone.0083662.q004

Discussion

The current study provides the first quantitative measures of stained microdamage accumulation during cyclic loading in human vertebral cancellous bone and shows that microdamage generated by cyclic loading is linearly related to reductions in specimen stiffness and non-linearly related to the proportion of fatigue life. Additionally, our results suggest that microdamage has a greater effect on fatigue life of cancellous bone tissue than it does on Young's modulus.

There are a number of strengths to the current study. First, our study describes microdamage accumulation with cyclic loading both in terms of microscopically observed microdamage as well as reductions in specimen stiffness. Second, cyclic loading was applied to vertebral cancellous bone from elderly humans with low bone volume fraction (BV/TV range 5–15%), thereby characterizing the population in which vertebral fractures are more prevalent. Third, microdamage was quantified in three-dimensions. Examination of microdamage in three-dimensions reduces variance in microdamage measures by measuring the entire specimen [44].

There are some limitations that must be considered in interpreting our results. First, the current study measured microdamage based on lead uranyl acetate staining. Lead uranyl acetate binds to exposed phosphate on the bone surface (e.g. hydroxyapatite crystals) [45]. Recent studies have demonstrated that fluorescent dyes used to stain microdamage do not stain all regions of permanent deformation within bone tissue [46,47,48]. Hence, it is possible that there are small forms of microdamage that are not stained by lead uranyl acetate. Nevertheless, in a study where high resolution imaging (order of 10 nm voxel size) was performed, lead uranyl acetate staining was typically localized to areas of maximum stress [17], suggesting lead uranyl acetate binding occurred in the regions where microdamage would be expected. Second, our staining protocol is expected to stain microdamage caused by fatigue loading as well as any pre-existing microdamage that had been caused in vivo. The use of multiple specimens from each donor mediated the effect of pre-existing damage on our results, as can be seen by the fact that in all but one donor increasing amounts of fatigue loading were associated with increased amounts of stained microdamage. Third, a limitation of



Figure 5. Changes in mechanical properties with fatigue loading. (A) The proportion of creep strain to maximum strain is shown for the six specimens fatigue loaded to failure. Each color represents a different specimen. (B) The alterations in modulus and energy dissipation in one specimen fatigue loaded to failure is shown. Modulus reduction increased progressively throughout fatigue loading, with a rapid loss in stiffness close to failure. Energy dissipation remained relatively constant until in the tertiary phase, where it increased rapidly. doi:10.1371/journal.pone.0083662.g005

Table 1. Regression models relating measured mechanical properties and DV/BV are shown.

Independent variable	Dependent variable	r²	Slope (95% CI)	Intercept (95% CI)
Reduction in Young's Modulus (%)	DVBV (%)	0.60	0.034 (0.018,0.050)	0.434 (-0.265,1.134)
Maximum strain (%)	DVBV (%)	0.61	0.491 (0.283,0.699)	0.690 (0.125,1.255)
Creep strain (%)	DVBV (%)	0.57	0.596 (0.327,0.864)	0.905 (0.377,1.433)
Cyclic strain (%)	DVBV (%)	0.54	2.029 (1.094,2.964)	0.200 (-0.550,0.950)
Energy dissipation (µJ/mm ³)	DVBV (%)	0.53	0.413 (0.221,0.605)	1.014 (0.512,1.516)
Maximum strain (%)	Reduction in Young's Modulus (%)	0.60	11.18 (8.54,13.82)	13.27 (7.07,19.48)
Creep strain (%)	Reduction in Young's Modulus (%)	0.35	12.86 (9.10,16.62)	18.93 (12.80,25.05)
Cyclic strain (%)	Reduction in Young's Modulus (%)	0.83	53.87 (45.58,62.15)	-3.06 (-9.37,3.26)
Energy dissipation (µJ/mm³)	Reduction in Young's Modulus (%)	0.46	8.05 (4.71,11.39)	22.44 (14.34,30.55)
Creep strain (%)	Maximum strain (%)	0.95	1.22 (1.17,1.28)	0.42 (0.33,0.52)
Cyclic strain (%)	Maximum strain (%)	0.64	4.01 (3.22,4.80)	-0.91 (-1.51, -0.33)
Energy dissipation (µJ/mm ³)	Maximum strain (%)	0.76	0.75 (0.57,0.93)	0.77 (0.32,1.24)

Donor was included as a random effect in all of the regression models to account for the use of multiple specimens from each donor.

doi:10.1371/journal.pone.0083662.t001

our study was that the maximum applied strain (3500 $\mu\epsilon$) was greater than is common during activities *in vivo*. Lifting an 8 kg weight has been estimated to induce an *in vivo* apparent strain of 2300 $\mu\epsilon$ in vertebral cancellous bone [49]. Thus, an apparent strain of 3500 $\mu\epsilon$ is representative of an overload, but is still considerably lower than required for failure [50]. The maximum strain level was selected to balance the goals of attaining observable microdamage while limiting the amount of degradation of the material from environmental conditions (temperature, etc.) during loading [19,51,52].

The current study provides a detailed description of the accumulation of microdamage during fatigue loading in human vertebral cancellous bone, making it possible to estimate the biomechanical importance of microdamage, including that presumably generated *in vivo* in donor tissue [7,53]. Determination of the relation between microdamage accumulation and reductions in Young's modulus is not entirely new in itself; others have described such relations in bovine cancellous bone or cortical bone, but the current study is the first to determine the relation for human cancellous bone. Consistent with the linear relation between damage volume fraction and reduction in Young's modulus reported here, Moore and colleagues found that the number of damaged trabeculae per total section area increased linearly with increasing modulus reduction in bovine cancellous bone [30].

Our measures of microdamage are of similar magnitude as previous studies. In the current study, the damage volume fraction ranged between 0.3% and 7.4%. In bovine cancellous bone subjected to an overload the damage volume fraction was $4.54\pm2.94\%$ [15] and the damaged area fraction was $3\%\pm1.9\%$ [24]. Similar ranges in microdamage are achieved in our laboratory using serial milling or two-dimensional sections. Nevertheless, our measures of microdamage are much smaller than *in vivo* microdamage reported from analysis of basic fuchsin,



Figure 6. The relationships between DV/BV and mechanical properties. A positive relation between DV/BV and (A) reduction in Young's modulus ($r^2 = 0.60$, p < 0.01); and (B) maximum strain applied during cyclic loading ($r^2 = 0.61$, p < 0.01) was observed. Dotted lines indicate the 95% confidence interval of the regression line. doi:10.1371/journal.pone.0083662.q006

Table 2. Correlations between mechanical properties, DV/BV and parameters of bone microarchitecture are shown.

	Reduction in Young's modulus (%)	Maximum strain (%)	Creep strain (%)	Cyclic strain (%)	Energy dissipation (µJ/mm³)	DV/BV (%)	Trabecular Thickness (um)	BV/TV (%)
Reduction in Young's modulus (%)	-	(0.74,0.93)	(0.66,0.91)	(0.87,0.97)	(0.46,0.84)	(0.30,0.78)	(-0.34,0.35)	(-0.50,0.17)
Maximum strain (%)	0.87 *	-	(0.98,1.0)	(0.79,0.95)	(0.71,0.92)	(0.39,0.82)	(-0.25,0.44)	(-0.40,0.29)
Creep strain (%)	0.82 *	0.99 *	-	(0.68,0.91)	(0.67,0.91)	(0.37,0.80)	(-0.25,0.44)	(-0.37,0.33)
Cyclic strain (%)	0.94 *	0.9 *	0.83 *	-	(0.66,0.91)	(0.36,0.80)	(-0.28,0.42)	(-0.52,0.15)
Energy (dissipation (µJ/mm³)	0.70 *	0.85 *	0.83 *	0.82 *	-	(0.36,0.80)	(-0.20,0.48)	(-0.33,0.36)
DV/BV (%)	0.59 *	0.65 *	0.63 *	0.63 *	0.63 *	-	(-0.25,0.44)	(-0.37,0.33)
Trabecular Thickness (um)	-0.01	0.11	0.11	0.08	0.16	0.1	-	(-0.46,0.22)
BV/TV (%)	-0.19	-0.06	-0.02	-0.21	0.02	-0.02	-0.14	-

The lower triangle displays the correlation coefficients and the 95% confidence interval of the correlation coefficient is shown in the upper triangle. Correlation scatterplots are included in Fig. S1.

*p<0.05.

doi:10.1371/journal.pone.0083662.t002

where the percent bone area with diffuse damage reached values up to 50% [53]. Differences in damage volume fraction could arise from the fact that in the current study microdamage was induced by mechanical loading, which may differ from microdamage sustained in vivo. In the current study, microdamage was induced by cyclic compressive loading, while in vivo there is more likely a mixed loading mode including bending and torsion that might cause differences in the amount of microdamage. Also, differences in the population of the subjects investigated with the current study only containing older donors (range 62-92 years) and the previous study including younger donors (range 23-96 years) could lead to differences in damage volume fraction. Furthermore, staining methodology (lead uranyl acetate v. basic fuchsin), sectioning (three-dimensional v. two-dimensional sections) or measurement (thresholding v. point counts) could have contributed to the differences in the magnitudes of microdamage. The damage volume fraction in the current study was also lower than that generated in vitro and visualized using lead uranyl acetate [16,54], where DV/BV was as high as 25%. Differences in skeletal region examined could contribute to the observed differences in



Figure 7. The estimated relationship between the amount of microdamage (DV/BV) and the proportion of fatigue life used (N/N_f). The relationship is derived from the average reduction in Young's modulus in Fig. 2B and regression model in Fig. 6A. Error bars in blue indicate the 95% confidence intervals at specified points. doi:10.1371/journal.pone.0083662.g007

microdamage. Vertebral cancellous bone is more rod-like and porous than bone from the femoral neck or tibial plateau (used by the Vashishth laboratory). Failure mechanisms of cancellous bone have been shown to differ between plate-like and rod-like bone [33], with tissue predominantly yielding in plates [55]. Therefore, more microdamage might accumulate in tibial or femoral cancellous bone before failure than in vertebral cancellous bone. There are also possible differences between damage generated from uniaxial loading (previous studies) and from fatigue loading (the current study). In addition, other factors including the lead uranyl acetate staining protocol (immersion time, rinsing methodology), image resolution, micro-CT settings (energy and integration time) and thresholding (manual v. automated thresholding with Otsu) may contribute to the differences in DV/BV between our study and studies of the Vashishth laboratory. To examine the effect of thresholding we analyzed the specimens with manually selected thresholds and found a similar range in DV/BV to that determined with the Otsu method (0.3-7.4% manually v. 0.9-8.8% with Otsu) and found that global thresholds necessary to achieve DV/BV measures as high as 25% did not match user observations, suggesting that global threshold selection alone is less likely to explain the differences in magnitude of DV/BV between our study and prior work. Further study is necessary to identify the true reason(s) for the differences.

Table 3. The mean and standard deviation of the bone microarchitectural parameters are shown.

	Mean±SD	Range
Bone volume fraction (%)	8.47±2.05	5.53-15.04
Trabecular thickness (μm)	128±13	106–149
Trabecular number (1/mm)	0.88±0.15	0.60-1.28
Bone surface (mm ²)	631±154	371-1104
Structure model index (–)	1.51 ± 0.32	0.64-2.07
Degree of anisotropy (-)	1.55 ± 0.21	1.28–2.16
Connectivity density (1/mm ³)	3.06±1.23	1.00–6.68

doi:10.1371/journal.pone.0083662.t003

In agreement with previous studies that have clearly shown that creep contributes to fatigue behavior in cancellous bone [28,29,56], creep strain dominated the accumulation of strain. Whether creep strain contributed to microdamage accumulation was, however, less clear since DV/BV and creep strain were correlated (Fig. S1) and in vivo loading models in animals have shown that creep loading causes microdamage [57,58]. Nevertheless, others have proposed that the contribution of creep strain on damage accumulation is negligible [29,52,59], because most of the creep strain is recoverable upon unloading and can therefore only cause minor permanent deformation to cancellous bone [56]. Maximum strain was the single best predictor of DV/BV (tested using multivariate linear regressions), which is consistent with a strain-based failure criterion for cancellous bone [30,42,60], however, because creep strain and maximum strain were highly correlated it is not possible to conclude which was causative.

Consistent with previous studies that found no effect of gender on fatigue properties in bone [32,61], the current study did not detect differences in fatigue properties between male and female samples.

In contrast to others that have reported correlations between the bone microstructure and bone microdamage [54,62,63,64], in the current study DV/BV did not correlate with the bone microarchitecture, and the bone microarchitecture did not appear to influence the relationship between DV/BV and modulus reduction. In the current study BV/TV ranges from 5.5% to 15.0% while prior examinations of microdamage in cancellous bone have ranged from 17.6% to 38.4% [64]. Thus the current study may not have had a sufficient range in microarchitecture to detect a correlation between microdamage and microarchitecture.

Although the current study did not examine the bone ultrastructure, it is useful to consider the mechanisms of energy dissipation that can explain changes in mechanical properties with fatigue loading. Bone is a hierarchical structure with energy dissipation mechanisms at different length scales of bone [65]. To resist fracture, bone tissue can deform through fibrillar sliding or through the formation of microcracks [66,67], where larger microcracks dissipate greater amounts of energy. Our observation of a correlation between DV/BV and energy dissipation per cycle are consistent with the idea that microdamage contributes to energy dissipation in cancellous bone. We speculate that in the secondary phase of fatigue loading crack-limiting boundaries (such as cement lines) limited the size of microdamage and therefore the amount of energy dissipation, while in the tertiary phase, microcracks extended through boundaries and result in greater microcracks with greater levels of energy dissipation.

References

- Cooper C, Melton IJ, 3rd (1992) Epidemiology of osteoporosis. Trends Endocrinol Metab 3: 224–229.
- Riggs BL, Melton LJ, 3rd (1995) The worldwide problem of osteoporosis: insights afforded by epidemiology. Bone 17: 505S–511S.
- Cooper C, Atkinson FJ, O'Fallon WM, Melton LJ, 3rd (1992) Incidence of clinically diagnosed vertebral fractures: a population-based study in Rochester, Minnesota, 1985–1989. J Bone Miner Res 7: 221–227.
- Keaveny TM, Wachtel EF, Kopperdahl DL (1999) Mechanical behavior of human trabecular bone after overloading. J Orthop Res 17: 346–353.
- Kopperdahl DL, Pearlman JL, Keaveny TM (2000) Biomechanical consequences of an isolated overload on the human vertebral body. J Orthop Res 18: 685–690.
- Burr DB, Forwood MR, Fyhrie DP, Martin RB, Schaffler MB, et al. (1997) Bone microdamage and skeletal fragility in osteoporotic and stress fractures. J Bone Miner Res 12: 6–15.
- Wenzel TE, Schaffler MB, Fyhrie DP (1996) In vivo trabecular microcracks in human vertebral bone. Bone 19: 89–95.
- Schaffler MB, Choi K, Milgrom C (1995) Aging and matrix microdamage accumulation in human compact bone. Bone 17: 521–525.

Our study provides a quantitative estimation of the relationship between DV/BV and proportion of fatigue life. Our findings suggest that small amounts of microdamage in human vertebral cancellous bone cause reductions in fatigue life that are much greater than reductions in Young's modulus. Therefore, even small amounts of stained microdamage may indicate large reductions in the number of loading cycles that the tissue can sustain before failure even when alterations in cancellous bone stiffness are small. Although the current study examined only the effects of microdamage generated by cyclic compressive loading, we expect our estimations of the effects of microdamage on subsequent biomechanical performance to be conservative because alterations in loading mode and mixed mode loading have been shown to be more effective at propagating microdamage [26,64]. Recognizing the limitations of our study mentioned above, the current study provides a means of estimating the biomechanical importance of in vivo microdamage observed within cancellous bone tissue.

In conclusion, we have demonstrated the relation between microscopically observed microdamage, reductions in specimen stiffness and reduced fatigue life during cyclic loading of human vertebral cancellous bone. Our findings suggest that microdamage has a much greater effect on fatigue life than on Young's modulus and provide a means to estimate the biomechanical repercussions of *in vivo* microdamage observed in tissue from patients.

Supporting Information

Figure S1 Scatterplots. Scatterplots of correlations between DV/BV, mechanical properties, BV/TV and trabecular thickness are shown.

(TIF)

Acknowledgments

For technical support, we would like to thank Garry Brock for explaining the lead uranyl acetate staining protocol, Katherine Ehlert and Daniel Brooks for help with the mechanical testing protocol, Christopher Chapa for contributing to specimen preparation, Kristin Regan for helping with calculation of the bone microarchitectural parameters, and Mark Riccio for X-ray CT scanning of the samples.

Author Contributions

Conceived and designed the experiments: FML CMR CJH. Performed the experiments: FML. Analyzed the data: FML ARB. Wrote the paper: FML CJH. Critical revision and final approval of the manuscript: FML ARB CMR CJH.

- Mori S, Harruff R, Ambrosius W, Burr DB (1997) Trabecular bone volume and microdamage accumulation in the femoral heads of women with and without femoral neck fractures. Bone 21: 521–526.
- Fazzalari NL, Forwood MR, Smith K, Manthey BA, Herreen P (1998) Assessment of cancellous bone quality in severe osteoarthrosis: bone mineral density, mechanics, and microdamage. Bone 22: 381–388.
- Hernandez CJ, Keaveny TM (2006) A biomechanical perspective on bone quality. Bone 39: 1173–1181.
- Burr DB, Hooser M (1995) Alterations to the En-Bloc Basic Fuchsin Staining Protocol for the Demonstration of Microdamage Produced in-Vivo. Bone 17: 431–433.
- Lee TC, Arthur TL, Gibson IJ, Hayes WC (2000) Sequential labelling of microdamage in bone using chelating agents. J Orthop Res 18: 322–325.
- Lee TC, Mohsin S, Taylor D, Parkesh R, Gunnlaugsson T, et al. (2003) Detecting microdamage in bone. J Anat 203: 161–172.
- Wang X, Masse DB, Leng H, Hess KP, Ross RD, et al. (2007) Detection of trabecular bone microdamage by micro-computed tomography. J Biomech 40: 3397–3403.

- Tang SY, Vashishth D (2007) A non-invasive in vitro technique for the threedimensional quantification of microdamage in trabecular bone. Bone 40: 1259– 1264.
- Brock GR, Kim G, Ingraffea AR, Andrews JC, Pianetta P, et al. (2013) Nanoscale examination of microdamage in sheep cortical bone using synchrotron radiation transmission x-ray microscopy. PLoS One 8: e57942.
- Forwood MR, Parker AW (1989) Microdamage in response to repetitive torsional loading in the rat tibia. Calcif Tissue Int 45: 47–53.
- Schaffler MB, Radin EL, Burr DB (1989) Mechanical and morphological effects of strain rate on fatigue of compact bone. Bone 10: 207–214.
- Burr DB, Turner CH, Naick P, Forwood MR, Ambrosius W, et al. (1998) Does microdamage accumulation affect the mechanical properties of bone? J Biomech 31: 337–345.
- O'Brien FJ, Taylor D, Lee TC (2003) Microcrack accumulation at different intervals during fatigue testing of compact bone. J Biomech 36: 973–980.
- Landrigan MD, Li J, Turnbull TL, Burr DB, Niebur GL, et al. (2011) Contrastenhanced micro-computed tomography of fatigue microdamage accumulation in human cortical bone. Bone 48: 443–450.
- Diab T, Vashishth D (2005) Effects of damage morphology on cortical bone fragility. Bone 37: 96–102.
- Moore TLA, Gibson LJ (2002) Microdamage accumulation in bovine trabecular bone in uniaxial compression. J Biomech Eng 124: 63–71.
- Wachtel EF, Keaveny TM (1997) Dependence of trabecular damage on mechanical strain. J Orthop Res 15: 781–787.
- Wu Z, Laneve AJ, Niebur GL (2013) In vivo microdamage is an indicator of susceptibility to initiation and propagation of microdamage in human femoral trabecular bone. Bone 55: 208–215.
- Michel MC, Guo XD, Gibson LJ, McMahon TA, Hayes WC (1993) Compressive fatigue behavior of bovine trabecular bone. J Biomech 26: 453– 463.
- Bowman SM, Guo XE, Cheng DW, Keaveny TM, Gibson LJ, et al. (1998) Creep contributes to the fatigue behavior of bovine trabecular bone. J Biomech Eng 120: 647–654.
- Haddock SM, Yeh OC, Mummaneni PV, Rosenberg WS, Keaveny TM (2004) Similarity in the fatigue behavior of trabecular bone across site and species. J Biomech 37: 181–187.
- Moore TL, Gibson LJ (2003) Fatigue microdamage in bovine trabecular bone. J Biomech Eng 125: 769–776.
- Thurner PJ, Wyss P, Voide R, Stauber M, Stampanoni M, et al. (2006) Timelapsed investigation of three-dimensional failure and damage accumulation in trabecular bone using synchrotron light. Bone 39: 289–299.
- Green JO, Wang J, Diab T, Vidakovic B, Guldberg RE (2011) Age-related differences in the morphology of microdamage propagation in trabecular bone. J Biomech 44: 2659–2666.
- Bevill G, Eswaran SK, Gupta A, Papadopoulos P, Keaveny TM (2006) Influence of bone volume fraction and architecture on computed large-deformation failure mechanisms in human trabecular bone. Bone 39: 1218–1225.
- Keaveny TM, Pinilla TP, Crawford RP, Kopperdahl DL, Lou A (1997) Systematic and random errors in compression testing of trabecular bone. J Orthop Res 15: 101–110.
- Giambini H, Khosla S, Nassr A, Zhao C, An KN (2013) Longitudinal changes in lumbar bone mineral density distribution may increase the risk of wedge fractures. Clin Biomech (Bristol, Avon) 28: 10–14.
- Gong H, Zhang M, Qin L, Hou Y (2007) Regional variations in the apparent and tissue-level mechanical parameters of vertebral trabecular bone with aging using micro-finite element analysis. Ann Biomed Eng 35: 1622–1631.
- Morgan EF, Keaveny TM (2001) Dependence of yield strain of human trabecular bone on anatomic site. J Biomech 34: 569–577.
- Carter DR, Hayes WC (1976) Fatigue life of compact bone–I. Effects of stress amplitude, temperature and density. J Biomech 9: 27–34.
- Rimnac CM, Petko AA, Santner TJ, Wright TM (1993) The effect of temperature, stress and microstructure on the creep of compact bovine bone. J Biomech 26: 219–228.
- Morgan EF, Yeh OC, Chang WC, Keaveny TM (2001) Nonlinear behavior of trabecular bone at small strains. J Biomech Eng 123: 1–9.
- Kummari SR, Davis AJ, Vega LA, Ahn N, Cassinelli EH, et al. (2009) Trabecular microfracture precedes cortical shell failure in the rat caudal vertebra under cyclic overloading. Calcif Tissue Int 85: 127–133.

- Uthgenannt BA, Silva MJ (2007) Use of the rat forelimb compression model to create discrete levels of bone damage in vivo. J Biomech 40: 317–324.
- Schaffler MB, Pitchford WC, Choi K, Riddle JM (1994) Examination of compact bone microdamage using back-scattered electron microscopy. Bone 15: 483–488.
- Ehlert KM, O'Brien RG, Hernandez CJ (2011) Statistical power in measures of microscopic tissue damage in cancellous bone. American Society for Bone and Mineral Research. San Diego, CA, USA. pp. MO0052.
- Hayat MA (2000) Principles and Techniques of Electron Microscopy: Biological Applications. Cambridge: Cambridge University Press.
- Sun X, Hoon Jeon J, Blendell J, Akkus O (2010) Visualization of a phantom post-yield deformation process in cortical bone. J Biomech 43: 1989–1996.
- Xu Z, Sun X, Liu J, Song Q, Muckley M, et al. (2010) Spectroscopic visualization of nanoscale deformation in bone: interaction of light with partially disordered nanostructure. J Biomed Opt 15: 060503.
- Sun X, Kishore V, Fites K, Akkus O (2012) Osteoblasts detect pericellular calcium concentration increase via neomycin-sensitive voltage gated calcium channels. Bone 51: 860–867.
- Kopperdahl DL, Keaveny TM (1998) Yield strain behavior of trabecular bone. J Biomech 31: 601–608.
- Brinckmann P, Frobin W, Hierholzer E, Horst M (1983) Deformation of the vertebral end-plate under axial loading of the spine. Spine (Phila Pa 1976) 8: 851–856.
- Pattin CA, Caler WE, Carter DR (1996) Cyclic mechanical property degradation during fatigue loading of cortical bone. J Biomech 29: 69–79.
- Moore TL, Gibson LJ (2003) Fatigue of bovine trabecular bone. J Biomech Eng 125: 761–768.
- Vashishth D, Koontz J, Qiu SJ, Lundin-Cannon D, Yeni YN, et al. (2000) In vivo diffuse damage in human vertebral trabecular bone. Bone 26: 147–152.
- Karim L, Vashishth D (2011) Role of trabecular microarchitecture in the formation, accumulation, and morphology of microdamage in human cancellous bone. J Orthop Res 29: 1739–1744.
- Shi X, Liu XS, Wang X, Guo XE, Niebur GL (2010) Type and orientation of yielded trabeculae during overloading of trabecular bone along orthogonal directions. J Biomech 43: 2460–2466.
- Yamamoto E, Paul Crawford R, Chan DD, Keaveny TM (2006) Development of residual strains in human vertebral trabecular bone after prolonged static and cyclic loading at low load levels. J Biomech 39: 1812–1818.
- Lynch JA, Silva MJ (2008) In vivo static creep loading of the rat forelimb reduces ulnar structural properties at time-zero and induces damage-dependent woven bone formation. Bone 42: 942–949.
- Seref-Ferlengez Z, Kennedy OD, Schaffler MB (2012) Diffuse Microdamage Induced in Cortical Bone in vivo Decreases with Time in the Absence of Bone Remodeling. Orthopaedic Research Society (ORS). San Francisco, CA, USA. 1348.
- Moore TL, O'Brien FJ, Gibson LJ (2004) Creep does not contribute to fatigue in bovine trabecular bone. J Biomech Eng 126: 321–329.
- Nalla RK, Kinney JH, Ritchie RO (2003) Mechanistic fracture criteria for the failure of human cortical bone. Nat Mater 2: 164–168.
- Sobelman OS, Gibeling JC, Stover SM, Hazelwood SJ, Yeh OC, et al. (2004) Do microcracks decrease or increase fatigue resistance in cortical bone? J Biomech 37: 1295–1303.
- Arlot ME, Burt-Pichat B, Roux JP, Vashishth D, Bouxsein ML, et al. (2008) Microarchitecture influences microdamage accumulation in human vertebral trabecular bone. Journal of Bone and Mineral Research 23: 1613–1618.
- Karim L, Vashishth D (2012) Heterogeneous glycation of cancellous bone and its association with bone quality and fragility. PLoS One 7: e35047.
- Wang X, Niebur GL (2006) Microdamage propagation in trabecular bone due to changes in loading mode. J Biomech 39: 781–790.
- Vashishth D (2007) Hierarchy of Bone Microdamage at Multiple Length Scales. Int J Fatigue 29: 1024–1033.
- Akkus O, Rimnac CM (2001) Cortical bone tissue resists fatigue fracture by deceleration and arrest of microcrack growth. J Biomech 34: 757–764.
- Zimmermann EA, Schaible E, Bale H, Barth HD, Tang SY, et al. (2011) Agerelated changes in the plasticity and toughness of human cortical bone at multiple length scales. Proc Natl Acad Sci U S A 108: 14416–14421.