




Draft Genome Sequences of *Salmonella enterica* Isolates Containing Incompatibility Group I1 Plasmids from Swine, Poultry, and Human Sources

Pravin R. Kaldhone,^{a,b}  Bijay K. Khajanchi,^a Jing Han,^a Rajesh Nayak,^a Steven C. Ricke,^b Steven L. Foley^a

U.S. Food and Drug Administration, National Center for Toxicological Research, Jefferson, Arkansas, USA^a;
Center for Food Safety, Department of Food Science, University of Arkansas, Fayetteville, Arkansas, USA^b

ABSTRACT The draft genome sequences of eight *Salmonella enterica* isolates from various sources were evaluated for the influence of incompatibility group I1 (Incl1) plasmids on virulence. Strains SE142, SE143, SE144, and SE146 originated from swine, SE36N and SE89N from poultry-related sources, and SE991 and SE1148 from human patients.

Salmonella enterica is one of the top five bacterial pathogens contributing to foodborne illnesses. *Salmonella* is also a leading foodborne pathogen associated with hospitalizations and deaths in the United States (1, 2). Food products originating from diverse sources like poultry, swine, and cattle are commonly associated with disease outbreaks caused by *Salmonella enterica* (2). Some serotypes of *Salmonella*, including Enteritidis, Typhimurium, and Heidelberg, are more prevalent as foodborne pathogens than other serotypes, such as Kentucky (3). Isolates containing certain mobile genetic elements, such as plasmids, have been associated with clinical manifestations of *Salmonella* infection (4). Plasmids encode genes responsible for antimicrobial resistance and virulence, which may have clinical significance associated with severe manifestations of diseases (5). Incompatibility group I1 (Incl1) plasmids have been reported to carry genes related to antimicrobial resistance and virulence (6).

Eight strains of *Salmonella enterica* containing Incl1 plasmids were sequenced (Table 1). Four of these strains were isolated from swine, two from poultry-related sources, and two from human patients. Previous studies showed that SE1148 and SE146 carried Incl1 plasmids and antimicrobial resistance genes (5). In addition to resistance genes, SE146 also contained an IncX4 plasmid which encodes a VirB/D4 type 4 secretion system that is likely involved in the increased virulence potential of this strain (7). Strains SE142, SE143, SE144, and SE146 were found to contain one or more plasmids and were resistant to commonly used antimicrobial agents (8). Overall analysis of the whole-genome sequences of these strains will improve our current understanding of the potential role of Incl1 plasmids in the pathogenicity of *Salmonella enterica* isolated from various foods and hosts.

To conduct the sequencing, total DNA was extracted using a DNeasy blood and tissue kit (Qiagen, Valencia, CA, USA). Nextera XT DNA sample kits (Illumina, San Diego, CA, USA) were used to construct a DNA library. Sequencing reactions were carried out at the DNA Sequencing Core at the University of Arkansas for Medical Sciences (UAMS) (Little Rock, AR, USA) and the Division of Microbiology, National Center for Toxicological Research (NCTR) (Jefferson, AR, USA) on an Illumina MiSeq instrument to generate 2 × 250 (UAMS) or 2 × 300 (NCTR) paired-end reads (9). Trimming and *de novo* assembly of the paired-end reads was performed using CLC

Received 24 August 2017 Accepted 25 August 2017 Published 28 September 2017

Citation Kaldhone PR, Khajanchi BK, Han J, Nayak R, Ricke SC, Foley SL. 2017. Draft genome sequences of *Salmonella enterica* isolates containing incompatibility group I1 plasmids from swine, poultry, and human sources. *Genome Announc* 5:e01056-17. <https://doi.org/10.1128/genomeA.01056-17>.

This is a work of the U.S. Government and is not subject to copyright protection in the United States. Foreign copyrights may apply.

Address correspondence to Jing Han, jing.han1@fda.hhs.gov, or Steven L. Foley, steven.foley@fda.hhs.gov.

TABLE 1 Summary of the genome sequence analysis of *Salmonella enterica* strains containing IncI1 plasmids

Strain	Serovar	Source	Location, yr of isolation	No. of contigs	Assembly size (bp)	G+C content (%)	No. of CDS ^a	No. of functional proteins	Accession no.
SE142	Heidelberg	Swine	Indiana, 2002	205	5,197,369	51.85	5,450	4,683	NPFC00000000
SE143	Heidelberg	Swine	Minnesota, 2002	306	5,361,922	51.69	5,718	4,779	NPEL00000000
SE144	Heidelberg	Swine	Minnesota, 2002	111	5,279,737	51.82	5,488	4,673	NPEQ00000000
SE146	Heidelberg	Swine	Minnesota, 2002	221	5,356,597	51.59	5,704	4,801	NPEM00000000
SE36N	Typhimurium	Chicken	West Virginia, 2000	158	5,160,965	51.90	5,358	4,705	NPER00000000
SE89N	Kentucky	Poultry house water	West Virginia, 2000	120	5,146,652	51.69	5,799	4,811	NPE50000000
SE991	Heidelberg	Human feces	Arkansas, 2009	98	5,053,493	51.82	5,161	4,493	NPEP00000000
SE1148	Heidelberg	Human blood	Wisconsin, 2007	166	4,867,737	52.17	4,995	4,435	NPEO00000000

^aCDS, coding sequences.

Genomics Workbench versions 8.5.1 and 9 (Qiagen, Germantown, MD, USA). The Rapid Annotation using Subsystem Technology (RAST) server (10), the Pathosystems Resource Integration Center (PATRIC) (11), and the NCBI Prokaryotic Genome Automatic Annotation Pipeline (PGAAP) (12) were employed to annotate the draft genomes of these strains (Table 1). The average G+C content of these strains is estimated to be 51.81%, as determined by PATRIC. Table 1 lists individual G+C content (%) and numbers of contigs, coding sequences, and functional proteins for respective strains.

Accession number(s). This whole-genome shotgun project has been deposited at DDBJ/ENA/GenBank under the accession numbers listed in Table 1.

ACKNOWLEDGMENTS

We acknowledge Allen Gies for his assistance with the DNA sequencing run and the UAMS Sequencing Facility, which is supported in part by grants from NIH to UAMS's Translational Research Institute (UL1TR000039) and the Center for Microbiological Pathogenesis and Host Inflammatory Response (P20GM103625).

The opinions expressed in this report are solely the responsibility of the authors and do not necessarily represent the official views and policy of the U.S. Food and Drug Administration or Department of Health and Human Services. Reference to any commercial material, equipment, or process does not in any way constitute approval, endorsement, or recommendation by the Food and Drug Administration.

This work was funded by the U.S. Food and Drug Administration (FDA) (E0759601) and the Center for Food Safety, Department of Food Science, University of Arkansas, Fayetteville, AR. Pravin R. Kaldhone's graduate assistantship was provided by the Center for Advanced Surface Engineering, under National Science Foundation Grant IIA-1457888, and the Arkansas EPSCoR Program, ASSET III.

REFERENCES

- Centers for Disease Control and Prevention. 2012. National *Salmonella* surveillance overview. Centers for Disease Control and Prevention, Atlanta, GA. <https://www.cdc.gov/national-surveillance/salmonella-surveillance.html>.
- Scallan E, Hoekstra RM, Angulo FJ, Tauxe RV, Widdowson MA, Roy SL, Jones JL, Griffin PM. 2011. Foodborne illnesses acquired in the United States—major pathogens. *Emerg Infect Dis* 17:7–15. <https://doi.org/10.3201/eid1701.P11101>.
- Foley SL, Johnson TJ, Ricke SC, Nayak R, Danzeisen J. 2013. *Salmonella* pathogenicity and host adaptation in chicken-associated serovars. *Microbiol Mol Biol Rev* 77:582–607. <https://doi.org/10.1128/MMBR.00015-13>.
- Han J, David DE, Deck J, Lynne AM, Kaldhone P, Nayak R, Stefanova R, Foley SL. 2011. Comparison of *Salmonella enterica* serovar Heidelberg isolates from human patients with those from animal and food sources. *J Clin Microbiol* 49:1130–1133. <https://doi.org/10.1128/JCM.01931-10>.
- Han J, Lynne AM, David DE, Tang H, Xu J, Nayak R, Kaldhone P, Logue CM, Foley SL. 2012. DNA sequence analysis of plasmids from multidrug-resistant *Salmonella enterica* serotype Heidelberg isolates. *PLoS One* 7:e51160. <https://doi.org/10.1371/journal.pone.0051160>.
- Smith H, Bossers A, Harders F, Wu G, Woodford N, Schwarz S, Guerra B, Rodríguez I, van Essen-Zandbergen AV, Brouwer M, Mevius D. 2015. Characterization of epidemic IncI1- γ plasmids harboring ambler class A and C genes in *Escherichia coli* and *Salmonella enterica* from animals and humans. *Antimicrob Agents Chemother* 59:5357–5365. <https://doi.org/10.1128/AAC.05006-14>.
- Gokulan K, Khare S, Rooney AW, Han J, Lynne AM, Foley SL. 2013. Impact of plasmids, including those encoding VirB4/D4 type IV secretion systems, on *Salmonella enterica* serovar Heidelberg virulence in macrophages and epithelial cells. *PLoS One* 8:e77866. <https://doi.org/10.1371/journal.pone.0077866>.
- Lynne AM, Kaldhone P, David D, White DG, Foley SL. 2009. Characterization of antimicrobial resistance in *Salmonella enterica* serotype Heidelberg isolated from food animals. *Foodborne Pathog Dis* 6:207–215. <https://doi.org/10.1089/fpd.2008.0172>.
- Khajanchi BK, Han J, Gokulan K, Zhao S, Gies A, Foley SL. 2016. Draft genome sequences of four *Salmonella enterica* strains isolated from turkey-associated sources. *Genome Announc* 4:e01122-16. <https://doi.org/10.1128/genomeA.01122-16>.
- Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL,

- Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: Rapid Annotations using Subsystems Technology. *BMC Genomics* 9:75. <https://doi.org/10.1186/1471-2164-9-75>.
11. Wattam AR, Abraham D, Dalay O, Disz TL, Driscoll T, Gabbard JL, Gillespie JJ, Gough R, Hix D, Kenyon R, Machi D, Mao C, Nordberg EK, Olson R, Overbeek R, Pusch GD, Shukla M, Schulman J, Stevens RL, Sullivan DE, Vonstein V, Warren A, Will R, Wilson MJ, Yoo HS, Zhang C, Zhang Y, Sobral BW. 2014. PATRIC, the bacterial bioinformatics database and analysis resource. *Nucleic Acids Res* 42:D581–D591. <https://doi.org/10.1093/nar/gkt1099>.
12. Angiuoli SV, Gussman A, Klimke W, Cochrane G, Field D, Garrity G, Kodira CD, Kyrpides N, Madupu R, Markowitz V, Tatusova T, Thomson N, White O. 2008. Toward an online repository of standard operating procedures (SOPs) for (meta)genomic annotation. *Omic* 12:137–141. <https://doi.org/10.1089/omi.2008.0017>.