



## Draft Genome Sequence of Serratia grimesii Strain A2

Ayslu M. Mardanova,<sup>a</sup> Anna A. Toymentseva,<sup>a</sup> Adeliya G. Gilyazeva,<sup>a</sup> Sergey V. Kazakov,<sup>b</sup> Elena I. Shagimardanova,<sup>a</sup> Sofia Yu Khaitlina,<sup>c</sup> Margarita R. Sharipova<sup>a</sup>

Institute of Fundamental Medicine and Biology, Kazan (Volga region) Federal University, Kazan, Russia<sup>a</sup>; Computer Technologies Laboratory, Information Technologies, Mechanics & Optics University, Saint Petersburg, Russia<sup>b</sup>; Institute of Cytology Russian Academy of Science, Saint Petersburg, Russia<sup>c</sup>

We report the first draft genome assembly of *Serratia grimesii* strain A2, previously identified as *Escherichia coli* strain A2, which produces protease ECP32 with a high specificity toward actin. *S. grimesii* strain A2 has multidrug resistance associated with a number of efflux pump genes.

Received 18 August 2014 Accepted 20 August 2014 Published 18 September 2014

Citation Mardanova AM, Toymentseva AA, Gilyazeva AG, Kazakov SV, Shagimardanova El, Khaitlina SY, Sharipova MR. 2014. Draft genome sequence of Serratia grimesii strain A2. Genome Announc. 2(5):e00937-14. doi:10.1128/genomeA.00937-14.

Copyright © 2014 Mardanova et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 3.0 Unported license.

Address correspondence to Ayslu M. Mardanova, mardanovaayslu@mail.ru, or Elena I. Shagimardanova, rjuka@mail.ru.

Gram-negative bacterium from the family Enterobacteriaceae that produces a new metalloproteinase, ECP32, with a high specificity toward actin was previously isolated (1–3). The bacterium was identified as atypical strain Escherichia coli A2 (1, 2). With the use of the Vitek 2 system (bioMérieux, France) and partial sequencing of the 16S rRNA gene, the strain A2 was reidentified as Serratia grimesii A2 (4). S. grimesii was classified as a part of the Serratia liguefaciens complex, along with Serratia proteamaculans (5). It was shown that ECP32 metalloprotease is homologous to actin-specific grimelysin from S. grimesii DSMZ30063 (4). Protease ECP32/grimelysin was successfully used in studies of actin structure/function relationships and mechanisms of actin polymerization (6, 7). Moreover, it was shown that introduction of the ECP32 protease gene to nonpathogenic E. coli results in acquisition of the ability to invade eukaryotic cells (8).

S. grimesii A2 genomic DNA, isolated using the GeneJET genomic DNA purification kit (Fermentas, Lithuania), was whole-genome shotgun sequenced using the 454 GS Junior (Roche) platform. The sequencing resulted in 5,137,381 highquality reads with an average read length of 500 bp and total 12× coverage. Testing different de novo assemblers (GS Junior, Roche; Geneious R7; SPAdes 3.0; ABySS 1.3.2; and MIRA 3-9-17), we applied the SPAdes platform (http://bioinf.spbau.ru/en/spades) and obtained 149 contigs (99 contigs larger than 500 bp;  $N_{50}$ , 108,561 bp). The chromosome has an overall G+C content of 52.85%. Nontranslated genes were predicted using tRNAscan-SE (9) and RNAmmer (10), which identified 69 tRNAs and 11 rRNAs. Open reading frame (ORF) prediction and annotation were performed through the Rapid Annotation using Subsystems Technology (RAST) pipeline (11). The functional comparison of genome sequences available on the RAST server revealed that the closest neighbors of S. grimesii A2 are S. proteamaculans 568 (score 525) followed by Serratia plymutica A30 (score 493), Serratia odorifera 4Rx13 (score 489), and Serratia marcescens Db11 (score 462). A total of 5,006 protein-coding sequences (CDSs) were identified, among which 2,883 CDSs (58% of the genome) were assigned to 1 of the 579 RAST subsystems. The annotated genome has 126 genes of the virulence, disease, and defense subsystem, among which 93

genes were predicted as antibiotic resistance genes. Thus, the *S. grimesii* A2 strain possesses multidrug resistance, and it is probably associated with a number of efflux pump genes.

Nucleotide sequence accession numbers. The *S. grimesii* A2 whole-genome shotgun project has been deposited in DDBJ/ENA/GenBank under the accession no. JGVP00000000. The version described in this paper is the first version.

## **ACKNOWLEDGMENTS**

The work was performed according to the Russian Government Program of Competitive Growth of Kazan Federal University. Some of the study was conducted with the support of a subsidy allocated to the Kazan Federal University for state assignment in the sphere of scientific activities (project 14-83). Part of this work was supported by the Russian Foundation for Basic Research and Republic of Tatarstan Government (grant 13-04-97130).

We thank all Genome Center members of the Interdisciplinary Center for Proteomic studies of the Institute of Fundamental Medicine and Biology (Kazan [Volga region] Federal University, Kazan) for 454 sequencing. We are grateful to Oleg A. Gusev for correction of the manuscript.

## **REFERENCES**

- Khaitlina SY, Smirnova TD, Usmanova AM. 1988. Limited proteolysis of actin by a specific bacterial protease. FEBS Lett. 228:172–174.
- Usmanova AM, Khaitlina SY. 1989. Protease from a strain of bacteria E. coli A2, specifically cleaving actin. Biokhimiia 54:1308–1314.
- 3. Matveyev VV, Usmanova AM, Morozova AV, Collins JH, Khaitlina SY. 1996. Purification and characterization of the proteinase ECP 32 from *Escherichia coli* A2 strain. Biochim. Biophys. Acta 1296:55–62. http://dx.doi.org/10.1016/0167-4838(96)00053-2.
- Bozhokina E, Khaitlina S, Adam T. 2008. Grimelysin, a novel metalloprotease from *Serratia grimesii*, is similar to ECP32. Biochem. Biophys. Res. Commun. 367:888–892. http://dx.doi.org/10.1016/j.bbrc.2008.01.003.
- 5. Grimont PAD, Grimont F, Irino K. 1983. Biochemical characterization of Serratia liquefaciens sensu strict, Serratia proteamaculans, and Serratia grimesii sp. nov. Curr. Microbiol. 7:69–74.
- Khaitlina SY, Hinssen H. 1997. Conformational changes in actin induced by its interaction with gelsolin. Biophys. J. 73:929–937. http://dx.doi.org/ 10.1016/S0006-3495(97)78125-6.
- Khaitlina SY, Strzelecka-Gołaszewska H. 2002. Role of DNase-1-binding loop in dynamic properties of actin filament. Biophys. J. 82:321–334. http://dx.doi.org/10.1016/S0006-3495(02)75397-6.

- Bozhokina ES, Tsaplina OA, Efremova TN, Kever LV, Demidyuk IV, Kostrov SV, Adam T, Komissarchik YY, Khaitlina SY. 2011. Bacterial invasion of eukaryotic cells can be mediated by actin-hydrolysing metalloproteases grimelysin and protealysin. Cell Biol. Int. 35:111–118. http:// dx.doi.org/10.1042/CBI20100314.
- Lowe TM, Eddy SR. 1997. tRNAscan-SE: a program for improved detection of transfer RNA genes in genomic sequence. Nucleic Acids Res. 25: 955–964. http://dx.doi.org/10.1093/nar/25.5.0955.
- 10. Lagesen K, Hallin P, Rødland EA, Staerfeldt HH, Rognes T, Ussery
- DW. 2007. RNAmmer: consistent and rapid annotation of ribosomal RNA genes. Nucleic Acids Res. 35:3100–3108. http://dx.doi.org/10.1093/nar/gkm160.
- Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: rapid annotations using subsystems technology. BMC Genomics 9:75. http://dx.doi.org/10.1186/1471-2164-9-75.