




Complete Genome Sequence of *Streptomyces* Siphophage Sitrop

Victor Portillo,^a Haydee Diaz,^a James Clark,^b Isla Hernandez,^b Mei Liu,^b  Ben Burrowes^b

^aDepartment of Biochemistry and Biophysics, Texas A&M University, College Station, Texas, USA

^bCenter for Phage Technology, Texas A&M University, College Station, Texas, USA

ABSTRACT *Streptomyces* sp. strain Mg1 is a Gram-positive soil bacterium capable of causing cell lysis and degradation of *Bacillus subtilis* colonies. Here, we report the 48,481-bp genome of *Streptomyces* sp. Mg1 siphophage Sitrop. With 77 predicted protein-coding genes and one tRNA, Sitrop shares 77% nucleotide sequence identity with the *Streptomyces* phage Verse.

Streptomyces sp. strain Mg1 is a Gram-positive, saprotrophic bacterium predominantly found in soil (1, 2). This filamentous organism is known to produce the antibiotic chalcomycin A, which plays a role in competition with and inhibition of *Bacillus subtilis* (2). Streptomycetes produce several useful secondary metabolites, including approximately 80% of today's antibiotics (3). Studying the genomes of phages of industrially important bacterial species, such as *Streptomyces* sp. Mg1 siphophage Sitrop, may be useful for improving bioproduction technologies.

Bacteriophage Sitrop was isolated in February 2019 from a soil sample taken from Lincoln, Nebraska. Sitrop was plaque purified as described elsewhere (4) using *Streptomyces* sp. Mg1 as its host and cultured at 30°C on nutrient broth or agar supplemented with 10 mM MgCl₂, 8 mM Ca(NO₃)₂, and 0.5% glucose. Sitrop was found to be chloroform sensitive. DNA was purified using a modified Wizard DNA clean-up kit (Promega) protocol (5) and prepared as Illumina TruSeq libraries. Sequencing was performed on an Illumina iSeq 100 instrument with paired-end 300-bp reads using a TruSeq Nano DNA kit. Quality control of the 254,194 resulting sequence reads was done using FastQC (<http://www.bioinformatics.babraham.ac.uk/projects/fastqc>) and trimmed manually with FastX 0.0.14 (http://hannonlab.cshl.edu/fastx_toolkit/download.html). The genome was assembled into a single contig at 109.8-fold coverage using SPAdes v3.5.0 (6) and closed by PCR and Sanger sequencing of the resulting product using primers GGACGTTGAACCTGTTGAGGA (forward) and GTCCTCCAGGTGTGAAGAAG (reverse). Structural annotations of protein-coding genes were initially predicted by GLIMMER v3 (7) and MetaGeneAnnotator v1.0 (8), while tRNAs were found using ARAGORN v2.36 (9). Conserved domains, sequence similarity, and transmembrane domains were found using InterProScan v5.33 (10), BLAST v2.9.0 (11), and TMHMM v2.0 (12), respectively, to predict gene function. BLAST similarity searches used a 0.001 maximum expectation value cutoff against the NCBI nonredundant, Swiss-Prot, and TrEMBL databases (13) (accessed 8 April 2020). The DNA sequence similarity of the entire genome was calculated with progressiveMauve v2.4 (14). Annotation tools used in Galaxy and Web Apollo are hosted at <https://cpt.tamu.edu/galaxy-pub> (15) by the Center for Phage Technology. Phage virion morphology was visualized as 2% (wt/vol) uranyl acetate negatively stained samples by transmission electron microscopy at the Texas A&M Microscopy and Imaging Center and determined to be a siphovirus (data not shown). All tools were run with default parameters unless otherwise specified.

Sitrop is a siphophage with a genome of 48,481 bp and a G+C content of 65.6%,

Citation Portillo V, Diaz H, Clark J, Hernandez I, Liu M, Burrowes B. 2021. Complete genome sequence of *Streptomyces* siphophage Sitrop. Microbiol Resour Announc 10:e01337-20. <https://doi.org/10.1128/MRA.01337-20>.

Editor Simon Roux, DOE Joint Genome Institute

Copyright © 2021 Portillo et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Ben Burrowes, benburrowes@tamu.edu.

Received 23 November 2020

Accepted 3 December 2020

Published 7 January 2021

compared to 72.17% of its host. The genome was composed of 1 tRNA gene and 77 predicted protein-coding genes, 41 being assigned putative functions, with a total coding density of 89.7%. Sitrop is most closely related to *Streptomyces* phage Verse (GenBank accession number [KT186229.1](https://doi.org/10.1128/genomeA.00535-13)), having 76.93% nucleotide identity and 67 shared proteins (16). Sitrop also shared close similarity to other *Streptomyces* phages within the *Camvirus* genus. Most of the protein-coding genes predicted to be tail proteins appear to be novel, sharing significant amino acid sequence similarity only with *Streptomyces* phages Alsaber ([MG298964.1](https://doi.org/10.1093/nar/gky092)) and Saftant ([MN204498.1](https://doi.org/10.1093/nar/gky092)), two novel phages with close similarity to Sitrop overall. There were four genes predicted to be involved in lysis, including an endolysin endopeptidase, a holin, and a separated two-component spanin complex gene (17).

Data availability. The genome of Sitrop is available in GenBank under accession number [MT701598.1](https://doi.org/10.1128/genomeA.00535-13). The associated BioProject, SRA, and BioSample accession numbers are [PRJNA222858](https://doi.org/10.1128/genomeA.00535-13), [SRR11558341](https://doi.org/10.1128/genomeA.00535-13), and [SAMN14609631](https://doi.org/10.1128/genomeA.00535-13), respectively.

ACKNOWLEDGMENTS

We thank Paul Straight, Texas A&M University, for providing *Streptomyces* sp. Mg1.

This work was supported with funding from the National Science Foundation (awards EF-0949351 and DBI-1565146) and additional support from the Center for Phage Technology (CPT). This announcement was prepared in partial fulfillment of the requirements for BICH464 Phage Genomics, an undergraduate course at Texas A&M University.

REFERENCES

- Hoefler BC, Konganti K, Straight PD. 2013. De novo assembly of the *Streptomyces* sp. strain Mg1 genome using PacBio single-molecule sequencing. *Genome Announc* 1:e00535-13. <https://doi.org/10.1128/genomeA.00535-13>.
- Barger SR, Hoefler BC, Cubillos-Ruiz A, Russell WK, Russell DH, Straight PD. 2012. Imaging secondary metabolism of *Streptomyces* sp. Mg1 during cellular lysis and colony degradation of competing *Bacillus subtilis*. *Antonie Van Leeuwenhoek* 102:435–445. <https://doi.org/10.1007/s10482-012-9769-0>.
- de Lima Procópio RE, da Silva IR, Martins MK, de Azevedo JL, de Araújo JM. 2012. Antibiotics produced by *Streptomyces*. *Braz J Infect Dis* 16:466–471. <https://doi.org/10.1016/j.bjid.2012.08.014>.
- van Charante F, Holtappels D, Blasdel B, Burrowes B. 2019. Isolation of bacteriophages. In Harper D, Abedon S, Burrowes B, McConville M (ed), *Bacteriophages*. Springer, Cham, Switzerland.
- Summer EJ. 2009. Preparation of a phage DNA fragment library for whole genome shotgun sequencing, p 27–46. In Clokie MRJ, Kropinski AM (ed), *Bacteriophages: methods and protocols, volume 2: molecular and applied aspects*. Humana Press, Totowa, NJ.
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J Comput Biol* 19:455–477. <https://doi.org/10.1089/cmb.2012.0021>.
- Delcher AL, Harmon D, Kasif S, White O, Salzberg SL. 1999. Improved microbial gene identification with GLIMMER. *Nucleic Acids Res* 27:4636–4641. <https://doi.org/10.1093/nar/27.23.4636>.
- Noguchi H, Taniguchi T, Itoh T. 2008. MetaGeneAnnotator: detecting species-specific patterns of ribosomal binding site for precise gene prediction in anonymous prokaryotic and phage genomes. *DNA Res* 15:387–396. <https://doi.org/10.1093/dnares/dsn027>.
- Laslett D, Canback B. 2004. ARAGORN, a program to detect tRNA genes and tmRNA genes in nucleotide sequences. *Nucleic Acids Res* 32:11–16. <https://doi.org/10.1093/nar/gkh152>.
- Jones P, Binns D, Chang H-Y, Fraser M, Li W, McAnulla C, McWilliam H, Maslen J, Mitchell A, Nuka G, Pesseat S, Quinn AF, Sangrador-Vegas A, Scheremetjew M, Yong S-Y, Lopez R, Hunter S. 2014. InterProScan 5: genome-scale protein function classification. *Bioinformatics* 30:1236–1240. <https://doi.org/10.1093/bioinformatics/btu031>.
- Camacho C, Coulouris G, Avagyan V, Ma N, Papadopoulos J, Bealer K, Madden TL. 2009. BLAST+: architecture and applications. *BMC Bioinformatics* 10:421–421. <https://doi.org/10.1186/1471-2105-10-421>.
- Krogh A, Larsson B, von Heijne G, Sonnhammer ELL. 2001. Predicting transmembrane protein topology with a hidden Markov model: application to complete genomes. *J Mol Biol* 305:567–580. <https://doi.org/10.1006/jmbi.2000.4315>.
- The UniProt Consortium. 2018. UniProt: the universal protein knowledgebase. *Nucleic Acids Res* 46:2699–2699. <https://doi.org/10.1093/nar/gky092>.
- Darling AE, Mau B, Perna NT. 2010. progressiveMauve: multiple genome alignment with gene gain, loss and rearrangement. *PLoS One* 5:e11147. <https://doi.org/10.1371/journal.pone.0011147>.
- Afgan E, Baker D, Batut B, van den Beek M, Bouvier D, Cech M, Chilton J, Clements D, Coraor N, Grüning BA, Guerler A, Hillman-Jackson J, Hiltmann S, Jalili V, Rasche H, Soranzo N, Goecks J, Taylor J, Nekrutenko A, Blankenberg D. 2018. The Galaxy platform for accessible, reproducible and collaborative biomedical analyses: 2018 update. *Nucleic Acids Res* 46:W537–W544. <https://doi.org/10.1093/nar/gky379>.
- Layton SR, Hemenway RM, Munyoki CM, Barnes EB, Barnett SE, Bond AM, Narvaez JM, Sirisakd CD, Smith BR, Swain J, Syed O, Bowman CA, Russell DA, Bhuiyan S, Donegan-Quick R, Benjamin RC, Hughes LE. 2016. Genome sequences of *Streptomyces* phages Amela and Verse. *Genome Announc* 4:e01589-15. <https://doi.org/10.1128/genomeA.01589-15>.
- Kongari R, Rajaure M, Cahill J, Rasche E, Mijalis E, Berry J, Young R. 2018. Phage spanins: diversity, topological dynamics and gene convergence. *BMC Bioinformatics* 19:326–326. <https://doi.org/10.1186/s12859-018-2342-8>.