

## PROKARYOTES



# Genome Sequence of *Weissella cibaria* DmW\_103, Isolated from Wild Drosophila

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**ABSTRACT** Lactic acid bacteria are commonly associated with *Drosophila* spp. Here, we report on the isolation of a strain of *Weissella cibaria* and the sequencing, assembly, and annotation of its genome. A total of 35 contigs were generated, with 2,349 coding sequences found.

Weissella cibaria is a lactic acid-producing bacterium that has potential uses in many industries, ranging from food production to medicine. For example, *W. cibaria* produces bacteriocins that have been studied for potential use as meat and dairy preservatives (1). Also, *W. cibaria* isolates have been screened for both general probiotic properties (adhesion and bile salt resistance) (2) and for conferring protection against a specific skin disease, atopic dermatitis (3). Beyond these properties, lactic acid bacteria are often associated with *Drosophila* spp. and influence their life history traits (4, 5). Since most genome sequences for lactic acid bacteria isolated from *Drosophila* spp. are restricted to a single genus (*Lactobacillus*), we sought to expand this taxonomic range by sequencing the genome of *W. cibaria* isolated from wild *Drosophila*.

DNA from W. cibaria was extracted and assembled using the following methods. Wild Drosophila samples were collected from compost in Ithaca, New York, USA (42.427447°N, 76.464339°W), homogenized, and diluted onto modified MRS medium (6). A single colony was isolated, and its identity was verified by Sanger sequencing of the 16S rRNA gene. To prepare the DNA for whole-genome shotgun sequencing with a 1,200-bp insert size, DNA was extracted using the Qiagen DNeasy Blood & Tissue Kit and fragmented by NEB fragmentase, and the adapters were ligated using components of the NEBNext Ultra II Ligation Module Kit according to manufacturer's instructions. The library was sequenced using paired-end 250-bp sequencing chemistry on an Illumina HiSeq 2500; 11,001,772 reads passed quality filtering, and the sequence assembly proceeded using Velvet version 1.2.10, as in our previous work (7, 8). The reads were randomly divided into 13 bins, each representing  $200 \times$  genome coverage, and for each bin a separate genome sequence was assembled by selecting the k-mer length between 191 and 211 that maximized the  $N_{50}$  (range of 150,723 to 160,231 bp, mean 157,625 bp). A consensus W. cibaria DmW\_103 genome was assembled using the representative contig file produced in each bin's separate assembly. A final assembly of 2,458,382 nucleotides in 35 contigs had an  $N_{50}$  of 160,221 and a max contig length of 434,968. ANIm analysis against Weissella spp. in the JSpeciesWS database in April 2017 confirmed the genome was from a W. cibaria isolate (>95% identity) (9). The data were submitted to GenBank and annotated by the NCBI Prokaryotic Genome Annotation Pipeline, yielding 2,349 genes.

Preliminary investigation using annotations produced in RAST (10–12) identified metabolic pathways unique to *W. cibaria*. Related to the other two *Leuconostocaceae* spp. with publically available genomes in RAST, only *W. cibaria* had annotated genes with functions in glycerate, glycogen, or sialic acid metabolism, or in fructose utiliza-

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tion. It would be interesting to test in a future work if any of these differentially present pathways influence *D. melanogaster* metabolism.

Accession number(s). The whole-genome shotgun data have been deposited in GenBank under the accession number NDXJ00000000. The version described in this paper is the first version, NDXJ01000000.

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## REFERENCES

- Li SW, Chen YS, Lee YS, Yang CH, Srionnual S, Wu HC, Chang CH. 2017. Comparative genomic analysis of bacteriocin-producing *Weissella cibaria* 110. Appl Microbiol Biotechnol 101:1227–1237. https://doi.org/10.1007/ s00253-016-8073-8.
- Lee KW, Park JY, Jeong HR, Heo HJ, Han NS, Kim JH. 2012. Probiotic properties of *Weissella* strains isolated from human faeces. Anaerobe 18:96–102. https://doi.org/10.1016/j.anaerobe.2011.12.015.
- Lim SK, Kwon MS, Lee J, Oh YJ, Jang JY, Lee JH, Park HW, Nam YD, Seo MJ, Roh SW, Choi HJ. 2017. *Weissella cibaria* WIKIM28 ameliorates atopic dermatitis-like skin lesions by inducing tolerogenic dendritic cells and regulatory T cells in BALB/c mice. Sci Rep 7:40040. https://doi.org/10 .1038/srep40040.
- Storelli G, Defaye A, Erkosar B, Hols P, Royet J, Leulier F. 2011. Lactobacillus plantarum promotes Drosophila systemic growth by modulating hormonal signals through TOR-dependent nutrient sensing. Cell Metab 14:403–414. https://doi.org/10.1016/j.cmet.2011.07.012.
- Téfit MA, Leulier F. 2017. Lactobacillus plantarum favors the early emergence of fit and fertile adult Drosophila upon chronic undernutrition. J Exp Biol 220:900–907. https://doi.org/10.1242/jeb.151522.
- Newell PD, Douglas AE. 2014. Interspecies interactions determine the impact of the gut microbiota on nutrient allocation in *Drosophila melanogaster*. Appl Environ Microbiol 80:788–796. https://doi.org/10.1128/ AEM.02742-13.
- Newell PD, Chaston JM, Wang Y, Winans NJ, Sannino DR, Wong AC, Dobson AJ, Kagle J, Douglas AE. 2014. *In vivo* function and comparative genomic analyses of the *Drosophila* gut microbiota identify candidate

symbiosis factors. Front Microbiol 5:576. https://doi.org/10.3389/fmicb.2014.00576.

- Zerbino DR, Birney E. 2008. Velvet: algorithms for de novo short read assembly using de Bruijn graphs. Genome Res 18:821–829. https://doi .org/10.1101/gr.074492.107.
- Richter M, Rosselló-Móra R, Oliver Glöckner F, Peplies J. 2016. JSpeciesWS: a web server for prokaryotic species circumscription based on pairwise genome comparison. Bioinformatics 32:929–931. https://doi .org/10.1093/bioinformatics/btv681.
- Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: rapid annotations using subsystems technology. BMC Genomics 9:75. https://doi.org/10.1186/1471-2164-9-75.
- Overbeek R, Olson R, Pusch GD, Olsen GJ, Davis JJ, Disz T, Edwards RA, Gerdes S, Parrello B, Shukla M, Vonstein V, Wattam AR, Xia FF, Stevens R. 2014. The SEED and the rapid annotation of microbial genomes using subsystems technology (RAST). Nucleic Acids Res 42:D206–D214. https://doi.org/10.1093/nar/gkt1226.
- Brettin T, Davis JJ, Disz T, Edwards RA, Gerdes S, Olsen GJ, Olson R, Overbeek R, Parrello B, Pusch GD, Shukla M, Thomason JA, Stevens R, Vonstein V, Wattam AR, Xia FF. 2015. RASTtk: a modular and extensible implementation of the RAST algorithm for building custom annotation pipelines and annotating batches of genomes. Sci Rep 5:8365. https:// doi.org/10.1038/srep08365.