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Review

# Insights into Gut Dysbiosis: Inflammatory Diseases, Obesity, and Restoration Approaches

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Abstract: The gut microbiota is one of the most critical factors in human health. It involves numerous physiological processes impacting host health, mainly via immune system modulation. A balanced microbiome contributes to the gut's barrier function, preventing the invasion of pathogens and maintaining the integrity of the gut lining. Dysbiosis, or an imbalance in the gut microbiome's composition and function, disrupts essential processes and contributes to various diseases. This narrative review summarizes key findings related to the gut microbiota in modern multifactorial inflammatory conditions such as ulcerative colitis or Crohn's disease. It addresses the challenges posed by antibiotic-driven dysbiosis, particularly in the context of *C. difficile* infections, and the development of novel therapies like fecal microbiota transplantation and biotherapeutic drugs to combat these infections. An emphasis is given to restoration of the healthy gut microbiome through dietary interventions, probiotics, prebiotics, and novel approaches for managing gut-related diseases.

Keywords: gut microbiota; obesity; ulcerative colitis; Crohn's disease; Clostridioides difficile



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## 1. The Importance of the Gut Microbiota for Human Health

The human gut microbiota is a complex community of microorganisms that co-evolved with us and reside in the gastrointestinal tract. This co-evolution with human cells was done in the context of many changes, including lifestyle, such as medications, urbanization, and dietary habits. The gut microbiome includes bacteria, archaea, relatively less-known fungi, and viruses. Bacteria are well surveyed and are the focus of our review. The development of high-throughput DNA and RNA sequencing technologies, along with advanced computational methodologies, has allowed scientists to catalog microorganisms comprehensively in an unprecedented manner. Various body habitats host distinct microbial communities and microbiomes that vary in microbial composition and function, including metabolic modules and pathways [1]. We now know that each body site has its own distinct composition of microbial communities depending on the physiological characteristics of the body niche [2]. Poor diet, antibiotic use, infections, and stress can lead to dysbiosis, characterized by a loss of beneficial microbes, microbial diversity, and the appearance of harmful bacteria.

The biogeography of the gut compartments (small and large intestines) affects the composition of the microbiota. Following the upper digestive tract, like the oral cavity and esophagus, the duodenum and jejunum have lower diversity than the ileum or the proximal colon [3]. The small intestine is predominantly populated by Gram-positive Firmicutes, including genera such as *Streptococcus*, *Veillonella*, and *Clostridium* [3]. In contrast, the colon hosts different dominant bacterial groups, especially strict anaerobes. The distal small intestine has higher abundances of bacterial phyla such as Bacteroidetes and Proteobacteria. The large intestine, composed of the cecum, ascending, transverse, descending, and sigmoid colon, rectum, and anus, harbors rich and highly diverse microbial communities, primarily

consisting of two bacterial phyla in healthy individuals: Bacteroidetes and Firmicutes. At the genus level, *Prevotella*, *Clostridium*, *Lactobacillus*, *Ruminococcus*, or *Odoribacter* [4], as well as *Roseburia*, *Faecalibacterium prausnitzii*, *Bacteroides vulgatus*, or *Fusicatenibacter saccharivorans*, are reduced in colon cancer and important anti-inflammatory taxa of the distal gut [5]. Various intestinal bacteria transform simple sugars into organic acids like lactate, acetate, propionate, and butyrate (short-chain fatty acids (SCFAs), which impact the proliferation and virulence of pathogens. For instance, acetate produced by *Bifidobacterium* spp. can suppress the virulence of Shiga-like toxins from verotoxigenic *E. coli* [6,7]. Dysbiosis has been implicated in several gastrointestinal disorders, including inflammatory bowel disease (IBD) [8], irritable bowel syndrome (IBS) [9], and other problems including *Clostridioides difficile* infections. For instance, patients with IBD often show reduced levels of beneficial bacteria like *Faecalibacterium prausnitzii* and increased levels of pro-inflammatory microbes. Beyond the gut, dysbiosis is linked to systemic conditions such as obesity [10] and even neurological disorders like depression and anxiety [11], and often, dietary changes and probiotic consumption help improve mood and overall health.

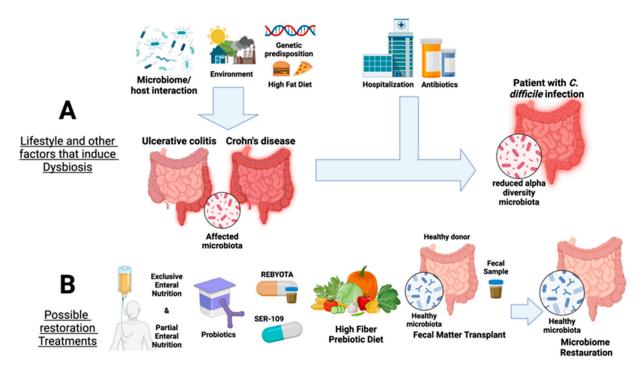
Restoring a healthy gut microbiome to counteract dysbiosis involves dietary interventions, probiotics, prebiotics, and fecal microbiota transplantation (FMT). Dietary changes, such as increased fiber intake, can promote the growth of beneficial bacteria. Below are some specific cases where microbiota modulation has been proven to improve phenotypic and symptomatic outcomes within gut-microbiome-related problems.

## 2. Inflammatory Bowel Diseases: Ulcerative Colitis and Crohn's Disease

### 2.1. Crohn's Disease Etiology and Pathogenesis

Crohn's disease is a chronic gastrointestinal inflammatory condition that can affect a patient's digestive system, anywhere from the mouth to the anus. By generating an exaggerated immune response throughout the digestive tract in inconsistent patches of tissue inflammation, it demonstrates symptoms such as abdominal pain, diarrhea, bleeding, nausea and vomiting, loss of appetite, weight loss, and fatigue [12]. These symptoms all depend on the location of the disease and the host's respective immune response, meaning that the condition can vary greatly between patients, and having an equal chance of affecting males and females aged between 15 and 35. Prolonged inflammatory activity along the digestive tract can lead to disease aggravation, further resulting in strictures (narrowing of the gut wall due to scar tissue), abscesses, and fistulas [13]. In 2019, both inflammatory bowel diseases (IBDs), Crohn's disease and ulcerative colitis, were reported to have a prevalence of approximately 24.51, 20.03, and 16.94 out of 100,000 people in North America, Australasia, and Western Europe, respectively [14]. The mechanisms by which this disease originates in a person are still unknown. Still, research indicates genetic, immunological, and environmental factors contribute to the onset of these intestinal perturbations [15].

Modern lifestyles have been well linked with the development of inflammatory diseases (Figure 1A). The Western lifestyle brings about practices such as cigarette smoking, antibiotic usage, and reduced exposure to microbes throughout childhood by urbanization and hygiene practices, which are known to increase the risks of developing Crohn's disease [16]. Recently an important study has shown how rural people in China and Israel shift their microbiota and metabolic profiles towards a Crohn's disease profile in urban settings [17]. The study compared the microbiome and metabolome of healthy and CD patients in developing rural and modern urban Chinese communities with a Westernized Israeli population. In rural settings, higher exposure to bacteria is shown with the presence of cattle and farms, extended family cohabitation, less hygiene, and a less processed diet that incorporates more vegetables and fruits. Meanwhile, the opposite is observed in urban settings, showing that the reduction in microbial diversity correlates with increased disease incidence. It demonstrated the significant impact of a Western diet involving high fats and processed foods on the gut microbial taxa, showing an increase in communities such as Fusobacteriaceae-like taxa linked with the Crohn's disease profile [17].



**Figure 1.** Overview of factors that affect the gut microbiota and current available therapies for IBD, namely ulcerative colitis and Crohn's disease, as well as in *Clostridioides difficile* infection. Panel (**A**) summarizes the factors that induce dysbiosis in the microbial gut communities, such as microbiome/host interaction, environmental factors, high-fat diet, and genetic predisposition leading to a reduction in gut alpha diversity. Panel (**B**) shows available microbiota restoration therapies for UC and CD and treatments against *C. difficile* colonization (probiotic supplementation, Rebyota/SER-109 Partial Enteral Nutrition, high-fiber prebiotic diet, and fecal microbiota transplants). Created with Biorender.

## 2.2. Clinical Significance of Crohn's Disease and Diagnostic Procedures

There is no straightforward screening method for Crohn's disease. The current analysis detects hematological biomarkers, such as C-reactive protein (CRP) and clinical history. CRP is produced by the liver and used as a marker of inflammation. Testing for CRP is used to try to distinguish between types of IBD conditions. Another way to survey disease progression is by studying microbiota metabolites, which are molecule end-products of metabolic processes, such as peptides and lipids [18]. Among some of the important metabolic markers are loss of SCFAs, and an increase in bile acid, fecal amino acid, and lipopolysaccharide (LPS) levels in blood, the functions of which can regulate microbial communities and maintain homeostasis of gut bacteria. These metabolites are actively present in healthy intestinal flora compared to the dysregulated quantities found in Crohn's patients [19]. Metabolites, such as bile acids, can be used to understand the presence of commensal gut bacteria and how disorders in these types of intestinal acids can negatively impact the composition of microbes present in this system. Lipopolysaccharides in high quantities are known to induce chronic inflammation. With damage to gut tissues and permeability, these molecules then enter the bloodstream, cause systemic inflammation, and can further progress IBD [20]. LPS triggers key immune system components, such as TLR4, which stimulates the NFkB pathway known for inducing inflammatory responses and activates other molecules like IFN- $\gamma$  and TNF- $\alpha$ , which further enhance inflammation pathways. SCFAs, including acetate, propionate, and butyrate, are produced by gut bacteria, like Bacteroidetes, that select for and ferment dietary fiber and prebiotics as a source of energy for the host [21]. These SCFAs reinforce intestinal barrier functions, maintain proper immune responses with regulatory T cells, and promote gut microbial homeostasis and mucosal tissue regeneration [22]. These hematological and metabolic

molecules that modulate diverse immune responses and gut functions are key biomarkers in CD diagnostics.

Dysbiosis in the microbiota can lead to many negative changes, as the gut plays a role in the developing immune system. In addition, the usage of antibiotics has been repeatedly shown to have a detrimental impact on the intestinal microbiota by allowing the displacement of a healthy flora (Table 1). An example of these circumstances can be seen in escalating health conditions such as cancer, rheumatoid arthritis, cardiovascular diseases, and obesity. This is a result of gut microbiota disturbances that can diminish the colonization resistance of beneficial commensal bacteria and provide an opportunity to pathogenic microbes residing in the intestines to harm the digestive system [23]. With the rising prevalence of disease, disrupting symptoms of health, and complicated origin pathways, it is highly important to provide viable treatments for these patients. A cure for Crohn's disease is currently unknown. Meanwhile, present treatments offer ways to handle and cope with the symptoms. Anti-inflammatory medication such as corticosteroids can mainly reduce swelling and inflammation in the digestive tract. Recently, biologic agents have been more readily implemented as lower-risk medication. These medication types only suppress certain gut immune system components, such as interleukins, integrins, and TNF, avoiding inhibiting other useful immune pathways [12]. However, even with a concurrent and stable treatment, surgery procedures may be required in more advanced cases due to the formation of bowel obstructions or strictures [12].

## 2.3. Treatment Options for Crohn's Disease

One recommended method for treating these conditions is the use of probiotics, as they have been shown to promote a healthier intestinal flora, inducing remission, and maintaining the remission period in Crohn's disease patients. The predominant presence of bacteria such as Bacteroides, Faecalibacterium, Bifidobacterium, and Lactobacillus species have been proposed as beneficial in maintaining a colonized gut, augmenting gut microbial diversity in a reduced environment, and improving other organ functions affected by dysbiosis [24,25]. Studies have found that implementing the use of probiotics decreases inflammation markers throughout the usage period, presents less-recurrent flare-ups, and maintains more extended remission periods, keeping symptoms at bay [24]. However, further studies need to be conducted to better identify a proper probiotic and prebiotic treatment plan, since not all studies present reproducible and concurrent data. Below, we summarize several published approaches shown to improve gut symptoms. To reinforce the usage of probiotics, supplementing with a proper precise diet can help combine individual benefits. Exclusive enteral nutrition (EEN) is a fully liquid-based diet, mainly aimed at children, to bypass any issues with nutrient absorption and benefit weight gain, facilitating gut mucosal healing [26]. Many reports in the last 30 years indicate high remission rates above 70% in various studies implementing EEN in children, concordant with clinical medication effects, demonstrating significant similarity in anti-inflammatory treatment while being less impactful on the body [26]. Solid foods are slowly incorporated at the end of the treatment period to avoid the return of symptoms, although this is not always successful. Meanwhile, a partial enteral nutrition (PEN) method has recently been used to incorporate a half-liquid/half-solid diet with higher tolerance rates than a liquid-only diet. A final significant therapy under active clinical studies and pending FDA approval in Crohn's disease patients is the fecal microbiota transplant (FMT); this is mainly seen in a pediatric setting to treat mild to moderate disease early on [27,28]. Details of this procedure and other prebiotic and probiotic restoration methods are provided in Section 5.

## 2.4. An Overview of Ulcerative Colitis and the Microbiota

Ulcerative colitis (UC) is an autoimmune inflammatory disease that targets the large intestine and progresses gradually. It can be diagnosed at any age and increases the risk of developing colon cancer by 4.5% after 20 years of diagnosis [29]. Still, it is more common in people aged 15 to 30 years [30]. Factors including microbiome/host interactions, the

environment, and genetic predisposition can affect the pathogenesis of this disease [31]. The clinical manifestations include weight loss, bloody stool, diarrhea, shortened colon, and inflammatory infiltration [32]. The World Health Organization has cataloged UC as one of the most complex diseases to treat nowadays [32].

Previous research has demonstrated a decrease in microbial diversity and an alteration of homeostasis associated with UC [33,34]. Studies found a significant reduction in the Phylum Firmicutes, and an increase mostly in Proteobacteria, but also in some members of Bacteroidetes [35,36]. Within Firmicutes, an important loss of *Roseburia*, *Faecalibacterium* [33,37,38], and *Lactobacillus* [39,40] is notorious, as these protective taxa are key for maintaining a healthy gut (Table 1). Studies related to bacterial and fungi interactions could uncover more details about the role of the microbiome in a UC profile. A study conducted by Sovran et al. in 2018 investigated the relationship between fungi and certain intestinal bacteria that play a role in the development of UC. *S. boulardii* and *C. albicans*, which can have either beneficial or detrimental effects, respectively, depending on the bacterial diversity within the gut microbiome [41].

Different species of Lactobacillus are associated with a positive reduction in the severe symptoms provoked by this syndrome, like colon shortening and rectal bleeding [39]. Species like Lactobacillus johnsonii can produce anti-inflammatory metabolites resulting in the induction of macrophage M2-mediated responses. This leads to an enhancement of the production of IL-10, an anti-inflammatory cytokine [40]. Lactobacillus acidophilus helps support the intestinal barrier, reduces the magnitude of proinflammatory cytokines, and enhances the production of acetate [42]. Likewise, Lactobacillus casei triggers an antiinflammatory reaction by suppressing the pro-inflammatory response and helping restore the intestinal barrier [32]. Additionally, Bifidobacterium, a genus from the phylum Actinobacteria, is downregulated. Bifidobacterium lactis (BLa80) helps to reduce excessive weight loss, colon shortening, and inflammation [38,43]. Research has demonstrated that B. Lactis species can restore homeostasis [44]. Faecalibacterium prausnitzii can similarly help reduce symptoms produced by UC, as it increases the length of the colon and reduces inflammatory cytokines in a pre-clinical model [45]. Additionally, the Faecalibacterium genus produces large amounts of butyrate, the primary energy source for colonocytes—the epithelial cells on the surface of the colon—aiding in the restoration of these cells [46].

Regarding phyla that are upregulated, it is crucial to highlight the increase in the Bacteroidetes because approximately 50% of the proteins secreted by microorganisms related to UC are secreted or derived from the genus of Bacteroides. Bacteroides can generate proteases which are correlated with proteolysis, aggravating the severity of ulcerative colitis. Remarkably, Bacteroides vulgatus and Bacteroides dorei produce proteases that break down serine and/or cysteine [47], and these enzymes are correlated with the initiation of this illness. A study has proposed the inhibition of the indicated serine proteases produced by Bacteroides vulgarus as a treatment for UC because approximately 40% of the patients with ulcerative colitis are prone to have these bacteria expressing serine proteases in large amounts, and this can be one of the reasons for the disruption of the colon [47]. Moreover, Enterobacteriaceae within phylum Proteobacteria have been observed in more significant amounts during inflammatory bowel diseases, especially the genera Escherichia, Klebsiella, and Citrobacter (Table 1) [43,48]. Escherichia coli is a commensal microorganism that, depending on the conditions and immune system of the host, can switch to being a harmful pathogen capable of triggering inflammation in the host [48]. It can also produce serine proteases enzymes that have been previously described as harmful in the case of Bacteroides [49]. In addition, studies in mice have shown that Citrobacter rodentium decreases P-glycoprotein transporters in the colon; these are efflux transporters that serve to export out of the cell toxic compound. A decrease in this molecule can induce an accumulation of toxins in the intestinal epithelial cells that can later be a factor inducing the alteration of the epithelial integrity and the gut microbiota [50]. Furthermore, other factors like inflammatory cytokines negatively affect the expression of P-glycoprotein transporters [50]. Indeed, it is important to mention that in both mice and humans, gut

dysbiosis is characterized by an imbalance in the microbiota (share in ~90% in both groups), and changes in flora are closely associated with the development and progression of inflammatory diseases.

Depending on the species and conditions, some metabolites are secreted or generated by microorganisms. Amino acid metabolites like Glutamine and Arginine have been demonstrated to be at higher levels in inflammatory bowel diseases [38]. A study conducted in pre-clinical models (rats) has revealed that the administration of Evodiamine—a quinolone alkaloid extracted from a Chinese tree—can help to reduce the levels of Glutamine in the serum and enhance the beneficial microbiota of the gut that are downregulated in UC [42]. The precise mechanism has not been elucidated, but the article indicated that a treatment using this quinolone can increase levels of Lactobacillus, specifically Lactobacillus acidophilus, which previously has been described as probiotic gut bacteria [42]. Furthermore, Evodiamine can downregulate IL-1 $\beta$  and IL-6 cytokines (pro-inflammatory), and upregulate IL-10 (anti-inflammatory), in experimental models with induced UC [42,51]. Lastly, metabolic products like pyruvic acid, fumaric acid, malonic acid, and oxoglutaric acid have been detected at higher levels in models with UC [38]. It is suggested that the increased energy production can cause the increase in pyruvic acid via glycolysis in hypoxic conditions induced by these diseases. This, in turn, leads to the accumulation of pyruvates that cannot be used in the tricarboxylic acid cycle [52], and in hypoxic conditions, an excess of pyruvate can induce an overflow of lactate, which causes an acidic environment and dysbiosis [53]. Understanding these metabolic changes can be crucial in determining potential targets for therapeutic intervention to treat UC.

Due to the large number of individuals affected by UC nowadays, there is a constant search for an effective treatment that enables these individuals to lead normal lives. An approved treatment is Vedolizumab, a monoclonal antibody that targets  $\alpha 4\beta 7$  integrin expressed on the gut lymphocyte surface [54]. This action inhibits the migration of these cells to the gut, thereby reducing inflammation [54]. Scientists are searching for a treatment beyond that of modulating the host's immune response but instead focusing on the microbiome, and its own ability to provoke anti-inflammatory reactions. Kaleido Biosciences is developing KB295 Metabolic Therapy based on the microbiota for mild-to-moderate ulcerative colitis, having successfully completed phase 1 clinical trials [52]. This treatment targets the microbiota directly, focusing on the bacteria's capacity to use different types of glycans as a substrate for their development [52]. KB295 is a synthetic glycan that selectively supports beneficial bacteria. In the in vitro experiments, this artificial glycan increased short-chain fatty acids selected from Bacteroidetes and Firmicutes, and downregulated Enterobacteriaceae [52]. Overall, KB295 can induce an upregulation of the beneficial bacteria and a downregulation of the harmful bacteria, making it a promising new approach for UC treatment.

**Table 1.** Summary of microbiota associated with health and gut dysbiosis. Note that upward arrows indicate increase and downward arrows indicate a decrease in taxa.

Gut Health and Disease/Dysbiotic Phenotypes	Significant Taxa	Country	References
Normal gut microbiome	↑Firmicutes	Korea Germany/Lithuania/India	[3,55,56]

Table 1. Cont.

Gut Health and Disease/Dysbiotic Phenotypes	Significant Taxa	Country	References
Ulcerative colitis	↑Proteobacteria	Netherlands Italy China Germany/Lithuania/India	[33,35,38,43,56–58]
	↓Actinobacteria ↓Bifidobacterium longum	Netherlands	[58]
	↓Cyanobacteria ↓Fusobactera ↑Verrucomicrobia	Italy	[59]
Chron's disease	↑Fusobacteria ↑Fusobacteriaaceae	Southern China Israel	[17]
	↓Firmicutes ↓Eubacterium rectale ↓Faecalibacterium prausnitzii ↓Roseburia intestinalis ↓Roseburia inulinivorans ↓Blautia faecis ↑Bacteroidetes ↑Bacteroides fragilis	Netherlands Korea Germany/Lithuania/India	[55,56,58]
Clostridioides difficile infection	↑Clostridioides difficile  ↓Bacteroidetes ↓Bacteroides ↓Firmicutes ↓Lactobacilllus ↓Enterococcus ↓Bacillus ↓Faecalibacterium ↓Ruminococcus	France	[60]
	↓Actinobacteria ↓Bifidobacterium longum subsp. longum ↓Bifidobacterium bifidum	Italy Brazil	[61,62]
Obesity	↑Firmicutes  ↑Eubacterium  ↑Roseburia  ↓Faecalibaterium  ↓Clostridiaceae  ↑Bacteroides	Japan Korea Mexico	[63–66]

## 3. Clostridioides Difficile—A Special Case of Antibiotic-Driven Dysbiosis

Clostridioides difficile is a Gram-positive anaerobic bacterium that forms spores associated with gastrointestinal infections. It is one of the most pressing threats to human antimicrobial resistance and gut inflammation [43]. The gut microbiome is significantly impacted by *C. difficile*, a member of the Firmicutes phylum. *C. difficile* spores are tolerant to extreme environments and utilize this to colonize disrupted or altered microbiomes [67].

This bacterium has two lethal toxins [Enterotoxin (Toxin A, tcdA) and Cytotoxin (Toxin B, tcdB)] that together are the causative agents of *C. difficile* infections in the gut [68–70]. Clostridioides difficile infections develop due to the inflammation and cellular events these toxins produce [71,72]. Up until 2017, CDI-related hospitalizations were continuously increasing in healthcare- and community-associated infections in the United States [73]. A study of the burden of C. difficile infections showed that healthcare-associated infections were higher than community-associated infections and antibiotic exposure, the most common risk factor for acquiring C. difficile infections [74]. CDI can cause a variety of symptoms, from intense diarrhea to chronic infections known as inflammatory bowel diseases (IBDs), underscoring the gravity of this condition [75]. Colonization resistance is crucial for the gut microbiota in the fight against pathogens and opportunistic bacteria like C. difficile [76]. A balanced gut with protective bacteria consists of taxa such as *Bacteroides*, *Lactobacillus*, Enterococcus, Bacillus, Faecalibacterium, and Ruminococcus [60]. Dysbiosis has several community profiles but can be mostly identified as a decrease in the probiotic bacteria. The reduction in alpha diversity and an increase in C. difficile infections have been related to the development of IBD illnesses such as ulcerative colitis (UC) and Crohn's disease. Lifestyle changes, mostly diet, can alter the gut microbiota as previously mentioned [71,76]. Dysbiosis in the gut through the colonization of *C. difficile* is facilitated by toxins A and B by proliferation and disruption, causing diarrhea and diarrhea-like symptoms [73]. A recent study in 2024 confirmed that lower alpha diversity is a risk factor for CDI. This study found a significant association between individuals with a history of CDI and their stature (CDI patients were shorter) compared to those without CDI history, and this association was mediated by alpha diversity [77].

## 4. Inflammation Is a Matter of Microbial Dysbiosis Resulting in Obesity

Obesity is a matter of microbes and their associated inflammatory pathways. Research conducted at Washington University in St. Louis demonstrated that obesity can be transferred with fecal matter [78]. This incredible and simple experiment showed how stool samples from an obese twin, placed into the gut of a gnotobiotic mouse, transferred the obese phenotype to the animal. The same occurred with fecal matter from the lean twin [78]. Obesity might even be considered to be an infectious disease. It was originally expected by 2022 for obesity to affect nearly 890 million adults and 160 million children [78], and a recent update declared that between 2020 and 2035, the prevalence of severe obesity in richer communities is predicted to double from 10 to 20%, posing a serious danger to healthcare systems [79]. Obesity may differ with ethnicity and correlate with socioeconomic status as well as changes in diet, such as high-fat diets [80]. Obesity is thus associated with an inflamed gut, fluctuation in the gut microbiota, and developing diseases such as diabetes, cardiovascular diseases, cancer, and digestive disorders [81,82]. Obese patients have been found to have a reduced abundance of Bifidobacterium longum subsp. longum, which contributes to glucose and lipid metabolism, and Bifidobacterium bifidum, which contributes to decreasing weight and insulin resistance [61]. For type 1 diabetic patients, an increase in Bifidobacterium adolescentis has been found [61] and has been suggested to be implemented in diets for nutritional modifications for these diseases. Pro-inflammatory responses in obese individuals may affect gut permeability, and the effect of high-fat diets associated with the intake of processed foods leads to increased levels of pro-inflammatory macrophages [81]. Adding Bacteroides uniformis CECT 7771 has been shown to help regulate mice's responses. Mice given a high-fat diet supplemented with fructose and administered B. uniformis were found to reduce body weight gain and restore immune cells by reducing B cells and macrophages and increasing Tregs; the pro-inflammatory cytokines decreased to increase anti-inflammatory cytokines [83] (Figure 2A,C). Lastly, the expression of Tolllike receptor 5 (TLR5) was restored to regular expression levels. TLR5 is important for metabolism since its deficiency can lead to hyperlipidemia, insulin resistance, weight gain, and changes in gut microbiota [83,84]. An essential component of inflammation in obesity is a T cell-mediated immune response (Figure 2). Previous research in pre-clinical models

showed how a deficiency in the innate adaptor molecule Myd88 leads to impaired development of T follicular helper cells and reduced IgA production in the gut, subsequently impacting the microbiota and leading to a metabolic syndrome [63]. A study observed that T-Myd88<sup>-/-</sup> mice would increase weight with aging and were more prone to develop obesity. Compared to a high-fat diet, it would accelerate the development of a metabolic syndrome. To ensure that microbiota is an influential factor in T-Myd88-/- mice in the high-fat diet, they were treated with antibiotics, and it was realized that the mice did not increase in weight [85]. Another observed factor was that T-Myd88<sup>-/-</sup> mice in the high-fat diet had lower microbial diversity in the ileum and fecal contents, specifically Clostridiaceae. Later, in mice enriched in Clostridiaceae and Lachnospiracea, it was found that when Desulfrovibrio colonization occurs, there was a reduction in Clostridium that led to weight gain. Lastly, it was demonstrated that mice Tcrb-/- (essential in T cell function) given T-Myd88-/- CD4+ T cells, gained a lot of weight, and a low percent of bacteria were coated by IgA compared with the Tcrb<sup>-/-</sup> mice given WT CD4+ T cells [63,64]. This emphasizes that the development of T follicular helper cells is essential for IgA production in the gut. IgA production is another essential component for the gut microbiota, which decreases in a high-fat diet. It has been demonstrated with a mice model that plasma cells producing IgA and B cell IgA were reduced during a high-fat diet, thus decreasing the amount of immunoglobulin. IgA deficiency in mice leads to increased T cells, which means inflammatory responses. Lastly, the IgA-deficient mice were observed to have major intestinal permeability and microbes closer to the epithelial cells with high endotoxin levels in the serum, establishing the importance of IgA for the mucous layer [86]. IgA's significance in the gut is in its prevention of the flagellated bacteria from getting to the epithelial cells. A study demonstrated that mice administered a high-fat diet and flagellin immunized showed decreased intestinal inflammation [87], indicating flagellin—a potent immunomodulatory agent—was as a possible way to reduce inflammation (Figure 2). Table 2 compares the dysbiosis in different phenotypes in both mice and humans.

**Table 2.** References of animal and human studies supporting the role of the microbiome in gut dysbiosis.

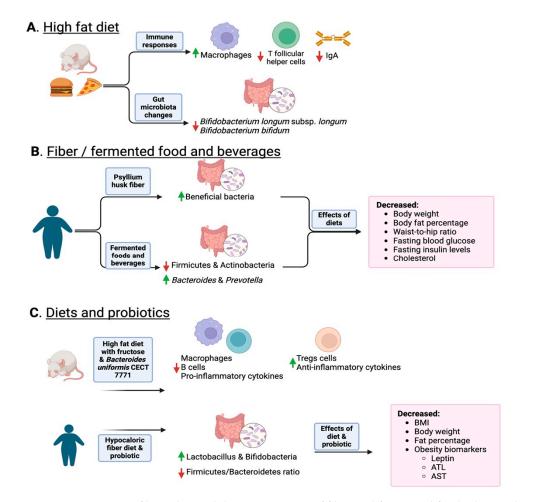
Gut Health and Disease/Dysbiotic Phenotypes	Animal Studies	References	Human Studies	References
Normal gut microbiome	In mammals, opportunistic bacteria that can thrive in dysbiotic habitats displace the microbes that contribute to a healthy ecosystem, influencing the environmental metagenome; as such, a clear difference in microbial populations is seen between captive and wild animals. In the mouse research model, Firmicutes, Bacteroidetes, Proteobacteria, Deferribacteres, Actinobacteria, Tenericutes, and Verrucomicrobia are shown to be commensal inhabitants of the gut microbiota, with the first three being the most abundant taxa.	[88–91]	An abundance of microbes such as Bacteroides, Lactobacillus, Bacillus, Enterococcus, Faecalibacterium, Bifidobacterium and Ruminococcus spp. are characteristic of a healthy and diverse gut microbiota.	[24,25,60]

 Table 2. Cont.

Gut Health and Disease/Dysbiotic Phenotypes	Animal Studies	References	<b>Human Studies</b>	References
Ulcerative colitis	In mice with UC, dysbiosis in their microbiota is closely related to disease pathogenesis and symptomatology. Increased populations of Proteobacteria, Actinobacteria, and <i>Clostridium</i> spp. were observed in mice with mucosal inflammation correlating to a UC profile.  Enterobacteriaceae taxa, <i>R. gnavus</i> , and especially <i>E. coli</i> are shown to drive colonization of pathobionts and inflammation in colitis mice models.	[92–94]	The interactions of the host and their microbiome, a decrease in Firmicutes, an increase in Proteobacteria, reduction in microbial diversity, and alterations in homeostasis are associated with UC pathogenesis. The connection of fungi with certain intestinal bacteria could lead to developing UC. S. boulardii and C. albicans are seen to confer positive or negative effects, respectively, depending on the bacterial diversity present in the gut microbiota.	[31,33,34,41,95]
Crohn's disease	The role of the microbiota can be seen in how the transference of microbes from a dysbiotic animal model can influence a healthy system. The transferrence of an altered microbiota from actively diseased mice induced CD symptomatology in healthy mice.  Research by Schaubeck et al. points to Bacteroidaceae, Erysipelotrichaceae, Peptostreptococcaceae and Verrucomicrobiaceae taxa in mice that simulate a CD profile.  Specifically, B. acidifaciens and B. sartorii were found in greater abundance. Bacteroides spp. are more prevalent when transferring human microbiota into mice.	[94,96,97]	Reduction in bacterial exposure and lower microbial diversity correlates with increased disease incidence.  Increase in Fusobacteriaceae-like taxa is linked with the Crohn's disease profile. CD is also characterized by a decrease in fungal and bacterial interactions. An abundance of fungi, like <i>C. tropicalis</i> , can give rise to more opportunistic pathogenic bacteria.	[17,95]
Clostridioides difficile	Antibiotics can diminish microbial communities for prolonged periods and eliminate species that could provide colonization resistance against <i>C. difficile</i> . A combination of antibiotics and CDI can further alter the intestinal microbiota and increase mucosal inflammation in mice, demonstrating increased abundance in Proteobacteria and Enterococci and a loss of microbial commensal diversity, such as Enterobacteriaceae. Even after the removal of antibiotics, their dysbiotic microbial communities remain susceptible to recurrence of CDI.	[98–100]	Antibiotics are known to generate dysbiosis in the human microbiome; in the same manner, they are heavily linked to the pathogenesis of CDI. Beneficial bacteria such as Bacteroides, Lactobacillus, and Ruminococcus, among related taxa, provide resistance for the gut microbiota against developing CDI in a perturbed intestinal system.	[60,76,101]

Table 2. Cont.

Gut Health and Disease/Dysbiotic Phenotypes	Animal Studies	References	Human Studies	References
Obesity	There exist links between the immune system and the gut microbiome. NOD2, as well as TLR5-deficient, mice are prone to increased adiposity. Dysbiotic microbiota transference into germ-free mice resulted in inflammation and gain in adipose tissue.	[78,102,103]	Diet has been proven to influence the microbes in the gut and modulate the levels of inflammation and obesity in a human host, especially in a Westernized high-fat diet selecting for less beneficial bacteria. The unbalanced ratio between Firmicutes and Bacteroidetes in dysbiosis correlates with increased adiposity, weight gain, and BMI.	[10,102,104]



**Figure 2.** Overview of how diet and the consumption of fiber and fermented foods change the gut microbiota. Panel (**A**) shows how a high-fat diet induces a reduction in IgA and permeability as well as a reduction in probiotic Bifidobacteria. Panel (**B**) shows how fiber and fermented foods such as kimchi increase protective bacteria and lean phenotypes. Panel (**C**) shows how combining diet and probiotics helps decrease inflammation and obese phenotype markers. In the figure, upward green arrows indicate an increase in specific taxa, while red downward arrows indicate a decrease. Created with Biorender.

## 5. Restoration Procedures for Gut Dysbiosis

The emergence of dysbiosis and gastrointestinal diseases, including the inflammatory patterns discussed above or the antibiotic-driven *Clostridioides difficile* infections, has spurred the development of therapies and strategies to maintain healthy gut flora and combat these infections. Treatment with antibiotics like Vancomycin and Fidaxomicin [105] has been the most successful at fighting inflammatory bowel diseases and *C. difficile* infections, but these therapies have not been entirely successful at treating *C. difficile* spores [105]. It is important to note that antibiotic therapies for CDI are strongly associated with increased antibiotic resistance and a less diverse bacterial community in the gut [106], highlighting the potential risks and the need to consider their use with utmost care and caution.

## 5.1. Fecal Microbiota Transplantation (FMT)

Fecal microbiota transplantation (FMT) is a promising approach to restore the gut microbiome and reduce *C. difficile* colonization [75]. The first study back in 2010 reported treating a patient with chronic diarrhea with the fecal matter from a healthy donor (husband) for a successful restoration [107]. This first report shows how natural fecal microbial communities positively impact the balance of the gut microbiome [107]. This method involves the transfer of stool from a person with a healthy diverse microbiota to a patient who presents intestinal dysbiosis or IBD symptoms. It has been found in some cases to treat and reverse *Clostridioides difficile* infections that are resistant to antibiotics [108,109]. The donor's beneficial microbes potentially colonize the damaged mucosa of the patient to regulate intestinal flora and start to reverse dysbiosis (Figure 1B). A study conducted at Tongji Pediatric Hospital in child Crohn's disease patients demonstrated that the combination of FMT with partial enteral nutrition (PEN) resulted in higher clinical remission rates, lower CRP inflammation markers, mucosal healing, and disappearance of ulcers in comparison to patients who only underwent PEN therapy [109].

Other studies have shown that FMT can lower antimicrobial resistance and inhibit strains of C. difficile. With the continuous improvement of FMT as a treatment for C. difficile infections, donor selection has also been an emerging topic. The concept of the "superdonor" is being used to describe those whose donations have been highly successful for patients [110]. Even though FMT for microbiota restoration is simpler for CDI patients, there is still ongoing research on the factors that influence the super-donor stool samples, such as microbial and genetic interactions [110]. This innovative treatment, approved by the Food and Drug Administration (FDA) in 2022, can significantly improve the management of CDI, offering a ray of hope in the fight against this infection. The first biotherapeutic drug, REBYOTA (RBL), consists of live microorganisms from the Bacteroidetes and Firmicutes phyla collected from healthy human stool samples [111,112]. In a randomized, placebocontrolled, pivotal phase 3 trial, the drug's efficacy and safety analysis demonstrated that RBL improves microbiome diversity and prevents C. difficile infections at increasing rates [80]. Another recent and approved therapy is the spore-delivering SER-109 drug, which consists of bacterial spores of healthy individuals and is used to combat recurrent C. difficile infection (rCDI) [113]. It was demonstrated that SER-109 administration in highrisk patients of rCDI increased Firmicutes bacteria and decreased C. difficile spores [113]. Another study that supports the efficacy of live fecal microbiota oral therapy determined that administration of this drug significantly reduces rCDI when compared with control groups with recurrent infections [114].

## 5.2. FMT Safety Concerns and Alternative Therapies

FMT is typically regarded as safe, particularly when thorough donor screening and testing are performed. Most immediate risks are related to the colonoscopy procedure if involved [115]. The long-term risks are not as well understood, but ongoing follow-up of treated patients is being conducted to monitor this, with few complications reported [115]. Among the most common risks are the following: (1) transmission of pathogens, including resistant bacteria, infectious agents, viruses, and parasites, previously not identified

from the donor to the recipient; (2) abdominal discomfort irritation and adverse cramping, which range from mild gastrointestinal symptoms to more severe symptoms; and (3) severe adverse effects leading to inflammatory diseases and even death (less than 1% of patients) [116]. The long-term effects of FMT are not yet fully understood. There is concern about the possibility of transferring not just beneficial but also harmful microorganisms, as well as resistant bacteria that could have unknown long-term consequences, including alterations in the recipient's microbiome that might increase the risk of chronic conditions such as autoimmune diseases or obesity [115,117].

Regulatory processes can still be considered a little blurry. FMTs in the UK are medicinal products; in the rest of Europe, they are human tissue products, while in the US, they are considered biological products [117]. The lack of standardized methods and regulations creates restrictions and confusion when implementing stool banks, and discourages medical doctors from seeking these innovative therapies [117].

There are also challenges for FMT development, especially the burden of donor screening, the costs associated with pathogen identification in patients, and the difficulty in finding suitable donors. Race and ethnicity account for a statistically significant proportion of the variation in human gut microbiome composition. Even after accounting for all demographic, dietary, and other relevant factors significantly linked to the gut microbiome, the impact of ethnicity on the gut microbiota remained evident [118]. This important variation advocates for and justifies local collections as essential health keepers to deliver the most appropriate stool/FMT from the standpoint of genetics and physiology. Alternatives to FMT that are becoming popular include (1) creating bacterial community consortia designed to mimic healthy communities to overcome transmission of pathogens [119]; (2) delivery of small drugs such as reactive sulfur species (R-SSH) that can selectively modulate the gut microbiome [120]; and (3) phage therapy aimed at targeting bacteria to be eliminated or at neutralizing the expression of a virulence factor [121], as well as probiotics and prebiotic fibers, which we discuss below.

The term probiotic, introduced by Elie Metchnikoff, came about from fermented milk that presented a high abundance of Lactobacilli and was proposed to be a good source of the firmicutes that are beneficial to the GI tract [122]. Although a recent study also showed that probiotic supplementation may delay microbiota restoration compared to fecal matter transplants [123], many studies still show the improvement of gut microbiomes through probiotic consumption. Probiotics have been demonstrated as a restoring therapy against recurrent Clostridioides difficile infections and antibiotic resistance [76], and have also been useful for inflammatory diseases, as discussed above, as well as in obesity. Living microorganisms of the gut flora, such as Lactobacillus acidophilus, Lactobacillus casei, Saccharomyces boulardii, and Bifidobacterium species [124], are suggested for the administration of probiotic therapy against *C. difficile* [125]. In a study of the suppression of *C. difficile* with the use of Bifidobacteria, it was shown that the synthesis of oligosaccharides to grow beneficial bacteria such as B. infantis inhibited C. difficile in mice. Probiotic strains, such as Lactocaseibacillus rhamnosus ATCC 53103 and Limosilactobacillus reuteri have been well studied, providing good evidence against abdominal pain and IBS [126], as recently mentioned in the World Gastroenterology Organization's Probiotics and Prebiotics Guidelines [127].

Prebiotics, which are often cheaper alternatives to probiotics, are being suggested as great fiber sources, or sources of good natural microbial communities that can select for fermentative microbes. Gut microbiota composition is altered due to nutrition changes, and it has been proven that healthy diets that select beneficial bacteria like *Bifidobacteria* and *Lactobacillus* promote a healthier gut environment [128] (Table 3).

Prebiotics are mostly non-digestible foods that promote the growth and activity of beneficial microorganisms, as well as healthy dietary fibers that select for fermentative bacteria. Prebiotics have the capacity to suppress the growth of harmful bacteria and promote the growth of beneficial bacteria, thereby helping to restore the balance in the gut microbiome [129] (Figure 1B). One such prebiotic is stachyose, which is extracted from plants. Studies have shown that this prebiotic can decrease the presence of inflammatory

cytokines and reduce the shortening of the colon in mice [130]. Other prebiotics, such as fructooligosaccharide (FOS) and galactooligosaccharide (GOS), have the capacity to induce a positive selection in genera like *Faecalibacterium*, *Lactobacillus*, and *Bifidobacterium* [131]. These genera have been previously described as beneficial for the host and downregulated in UC. Moreover, FOS prebiotics can reduce oxidative stress, possess antioxidant activity, and contribute to overall gut health. On the other hand, GOS can modulate immune function, increasing anti-inflammatory cytokines [131]. Overall, these prebiotics induce a positive regulation of the beneficial bacteria and possess other functions that maintain intestinal integrity and homeostasis. Treatment strategies based on probiotics and prebiotics previously described as beneficial or inhibiting metabolic pathways for the over-expressed metabolites could offer promising options for this illness [131,132].

A fiber-rich diet may include the consumption of psyllium, a water-soluble fiber that improves intestinal flora. Abdominal discomfort with intense epigastric pain can be improved with psyllium husk fiber and lifestyle modifications [132]. The implemented lifestyle modifications include diet (consuming more fruits and vegetables and reducing sugar intake) and physical activity (30 min walks). The study compared gender-wise groups, showing that females improved their symptoms [132] (Figure 2B). Another study supported the use of psyllium husk fiber to improve the microbial composition of constipated patients. It was observed that the use of fiber increased beneficial microbiota, including three butyrate-producers: *Lachnospira*, *Roseburia*, and *Faecalibacterium* [133].

Fermented foods are special cases of prebiotic supplementation. A study in Korea demonstrated the effects of kimchi on obese women. Women moved from having fewer Firmicutes and Actinobacteria and higher levels of Bacteroides, to higher levels of Actinobacteria after kimchi consumption [65] (Table 3). Indeed, after eight weeks of administering kimchi, clinical parameters, such as body weight, body fat percentage, and cholesterol decreased, with the exceptions of triglyceride and pulse rate [65]. In another study, the group was gender-matched and results confirmed that the intervention of fresh kimchi or fermented kimchi for 4 weeks had the ability to decrease weight, BMI, and body fat [134]. The administration of fermented kimchi reduced waist-to-hip ratio, blood glucose, and insulin levels [134]. A hypocaloric fiber regimen combined with physical exercise and probiotic administration was used in a group of obese women [135]. Fiber diet and probiotic administration increased the abundance of protective Lactobacillus and Bifidobacteria and reduced the ratio of Firmicutes/Bacteroidetes (Figure 2C). Similar research on obese people with type 2 diabetes, after supplementation with specific strains of Lacticaseibacillus paracasei and Bifidobacterium breve as probiotics, and galactooligosaccharides as prebiotics, showed there was a rise in the abundance of protective Bifidiobacteriaceae [136]. In obese children and adolescents, a synbiotic composed of Lactobacillus coagulans SC-208 and Lactobacillus indicus HU36 coupled with a prebiotic short-chain fructooligosaccharide resulted in a decrease in the waist-height ratio [137]. The use of probiotics and prebiotics in obese people suggests that this is an important factor in modulating the gut microbiota to restore the beneficial bacteria and enhance the reduction in weight, BMI, or body fat.

Yogurt is another known probiotic implemented in everyday diets that may modulate the gut microbiota [138]. A specific case of a commercial yogurt—known as 166 yogurt by Sanyuan Foods Co.—was designed to contain fermented milk with a culture of *Lactobacillus bulgaricus* and *Streptococcus thermophilus* with an additional supplementation with *L. acidophilus*, *L. plantarum*, *L. paracasei*, and three *Bifidobacterium strains: breve*, *lactis*, and *longum* (Table 3). To this probiotic mix, they also added fructooligosaccharides, inulin polydextrose, galactooligosaccharide, isomaltooligosacharides, and xylooligosaccharides. This yogurt symbiotic was demonstrated to improve constipation in mice and humans [138]. This improvement in alleviating the constipation symptoms may be attributed to the metabolites that are produced by the bacteria, through stimulation of the prebiotic fibers [138]. In another study, the effect of fresh yogurt and heat-treated yogurt was assessed on immune function in healthy individuals. Fresh yogurt was demonstrated to have a better effect on the immune system since it stimulated the synthesis of IFN-gamma, phagocytic cells,

proinflammatory cytokines, and lymphocytes such as Th1 that contribute to IFN-gamma synthesis. The heat-treated yogurt demonstrated an increase in NK cells, Th2 lymphocytes in response to IL-5, and B cells [139].

Fermented beverages are also known to be important items for gut-diversity-focused dietary plans. Kefir is a milk beverage fermented by yeasts and bacteria. This beverage supplies an ample community of more than 100 lactobacilli species, allowing a resistance response to gut colonization by pathogenic bacteria [140]. Among the microbes present, species of Lactobacillus, Bifidobacterium, Acetobacter, Streptococcus, Candida, and Saccharomyces are mainly abundant [140,141] (Table 3). Most notably Lactobacillus kefiranofaciens and Lactobacillus kefiri, which are only found in this fermented milk, demonstrate a strong capability to inhabit the gut and a capacity to modulate the intestinal microbiome [141]. Even if the quality and microbial composition of Kefir differs by production and respective brands, research suggests it still possesses the capabilities to alleviate weight gain and support metabolic processes while under Western high-fat diets [142]. A recent clinical carried out at Mayo Clinic Rochester with 53 critically ill patients in an ICU provided Kefir supplementation to treat gut dysbiosis despite antibiotic treatment [143]. Results demonstrated a decrease in microbial diversity due to antibiotic treatment; however, the Gut Microbiome Wellness Index (GWMI), indicating the overall health of the gut microbiota, was shown to increase exponentially after 72 h of Kefir implementation, reducing pathogenic bacteria and easing dysbiosis in a short period [143]. The findings also suggest Kefir's anti-cancerous, anti-oxidant, and anti-bacterial functions, which are further enhanced by fermentation modulation [144]. As such, studies demonstrate a significant role of the benefits of Kefir products for a variety of health profiles in restoring gut microbial diversity and granting whole body benefits.

Another beverage that is currently being added to diets is Kombucha. This fermented tea consists of sugar, tea leaves, and previous fermentation [145–147], which leads to a symbiotic culture of yeast and bacteria that act as prebiotics and probiotics [145–148]. A study conducted in 2020 demonstrated that Kombucha, as a beneficial and antioxidant drink, needed to be further analyzed due to the findings of the benefits depending on the fermentation type and tea being used [147].

Kombucha can be prepared with any type of tea. Chemical profiles of these teas show that green tea has the highest antioxidant activity compared to white, red, and black teas. In its fermentation period, the tea also grows healthy bacteria and yeast, such as Acetobacter spp. and Gluconobacter spp. [145,147,149], Zygosaccharomyces spp. [145,149], and Schizosaccharomyces pombe [147,149] (Table 3). To demonstrate Kombucha's benefits for healthy diets, a pre-post intervention study revealed that obese individuals who consumed fermented tea showed a decrease in insulin in their bodies [145]. Dietary habits, such as increasing vegetable consumption, complemented with Kombucha, have also been studied in studies aimed at reducing levels of cholesterol. In New Zealand, a study divided rabbits into groups with high-cholesterol diets and others with high-cholesterol diets with Kombucha treatment, and intestinal bacteria were analyzed through stool collection [150]. This study concluded that the Kombucha treatment reduced the high cholesterol levels [150]. Kombucha treatment has also been used for non-alcoholic fatty liver disease in mice. In 2018, Jung et al. demonstrated that mice fed with Kombucha tea had decreased levels of Clostridium and reduced fat accumulation compared to groups that did not receive the treatment [151]. Kombucha tea-fed groups also showed increased beneficial bacteria like E. coli, Bacteroidetes, and Lactobacillus [151]. Due to its antioxidant and symbiotic benefits, Kombucha has been a popular drink implemented into multiple diets for its anti-inflammatory effects and reduction in obesity risks and secondary diseases such as diabetes [145]. Spent coffee grounds are also an emerging prebiotic [152]. In vitro studies have shown that mannanoligosaccharides from coffee grounds stimulate the growth in beneficial bacteria such as Bifidobacterium, Ruminococcus, and Blautia, and short-chain fatty acids [153], thus showing the prebiotic activity in coffee grounds and the potential effects on glucose metabolism [153]. Lastly, two far less popular fermented beverages are

mauby (made of woody plants) and Tepache (fermented from pineapple rinds), which are traditional drinks in the Caribbean. A pilot study revealed that these drinks are rich in beneficial bacteria, particularly from a diverse array of multi-strain species of Lactobacillus and other probiotics. The relative abundance of probiotic changes depending on the source material used for mauby fermentation (different species of wood) [154]. A study of fermented beverages in Mexico demonstrated that Tepache contains lactic bacteria such as E. faecium and L. lactis. Although more studies are needed, this characterization presents the potential of this drink as an antimicrobial and a prebiotic [155]. Among fruits, kiwi stands out as a major prebiotic fiber source, influencing gut microbiota profiles towards an enrichment of Lactobacilli and Bifidobacteria (as compared to the same individual before consumption) [156]. Other fruits, such as black raspberries, have also been demonstrated to have prebiotic compounds [157]. In 2019, Gu et al. fed mice black raspberries and analyzed the mice for colon microbiome composition [158]. Mice showed greater Bacteroidetes levels and lower levels of Firmicutes, such as Clostridium species, in the gut's lumen after the raspberry diet. This study also confirmed the improvement in metabolite levels in the gut after the fiber-rich fruit diet [158]. Overall, many fruit and fermented foods are sources of important fermentative taxa that can improve gut biodiversity and antiinflammatory pathways.

**Table 3.** Summary of probiotics/prebiotics and protective taxa associated with improved gut health outcomes.

Probiotic/Prebiotic	Functions	Outcomes	References
Psyllium	Increased levels of <i>Lachnospira</i> , <i>Roseburia</i> , and <i>Faecalibacterium</i>	Improved abdominal discomfort, epigastric pain, and constipation symptoms	[132,133]
Kimchi	Increased levels of Actinobacteria	Improved cholesterol levels, insulin levels, body weight, body fat percentage, and BMI	[65,134]
Lacticaseibacillus paracasei YIT 9029 and Bifidobacterium breve YIT 12272	Increased levels of Bifidobacteriaceae and <i>Lactobacillus</i>	Improved glucose metabolism	[136]
Yogurt	Increased levels of <i>Bacteroides</i> , Streptococcus, Blautia, and Saccharomyces; NK cells, B cells, IL-5, and Th2	Improved constipation symptoms and immune responses	[138,139]
Kefir	Increased levels of Lactobacillus	Improved constipation symptoms, cholesterol levels, and obesity risks	[141-143]
Kombucha	Increased levels of beneficial <i>E. coli</i> , Bacteroidetes and <i>Lactobacillus</i>	Improved glucose levels, obesity risks, and cholesterol levels.	[145,150,151]
Kiwi	Increased levels of <i>Lactobacilli</i> , and <i>Bifidobacteria</i>	Improved the growth of intestinal lactid acid bacteria and perturbation of <i>Clostridium</i>	[156]
Black raspberries	Increased levels of Bacteroidetes	Improved colon microbial $\alpha$ -diversity	[157,158]
Coffee spent grounds	Increased levels of <i>Bifidobacterium</i> , <i>Jamie Quinton</i> , and <i>Ruminococcus</i>	Improved levels of microbial α-diversity in fecal samples	[152,153]
Bifidobacterium animalis and Lactobacillus paracasei	Increased levels of Bacteroidetes	Improved gut microbiota and risk for metabolic disorder	[159,160]

## 6. Conclusions/Summary

The development of sequencing technologies, improvement in bioinformatic pipelines, and the spread of knowledge on computational methodologies has allowed microbial ecologists and other health experts to survey the global human microbiome. Factors such as poor diet, antibiotic use, infections, and stress can lead to dysbiosis, characterized by a loss of beneficial microbes, an overgrowth of harmful microorganisms, or a loss of microbial diversity. Dysbiosis, an imbalance in the gut microbiota, disrupts essential processes and contributes to diseases such as ulcerative colitis, Crohn's disease, and *Clostridioides difficile* infections. Challenges posed by antibiotic-driven dysbiosis, particularly concerning *C. difficile* infections, call for novel therapeutic approaches like fecal microbiota transplantation (FMT) and biotherapeutic drugs. Restoration of a healthy gut microbiome through dietary interventions, probiotics, and prebiotics presents itself as a promising strategy to manage gut-related diseases. The connection between gut dysbiosis and obesity suggests that

microbial imbalances contribute to inflammatory pathways and associated conditions such as cardiovascular diseases and diabetes.

New integrative approaches are proposed to combine dietary modifications and emerging therapeutic techniques like FMT and synthetic glycans targeting beneficial bacteria. In conclusion, this review underlines the importance of maintaining gut microbiota balance to prevent and manage various inflammatory diseases, proposing integrative approaches that include dietary modifications and emerging therapeutic techniques like FMT and synthetic glycans targeting beneficial bacteria.

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#### References

- 1. Godoy-Vitorino, F. Human Microbial Ecology and the Rising New Medicine. Ann. Transl. Med. 2019, 7, 342. [CrossRef]
- 2. Dominguez-Bello, M.G.; Godoy-Vitorino, F.; Knight, R.; Blaser, M.J. Role of the Microbiome in Human Development. *Gut* **2019**, 68, 1108–1114. [CrossRef] [PubMed]
- 3. Kastl, A.J., Jr.; Terry, N.A.; Wu, G.D.; Albenberg, L.G. The Structure and Function of the Human Small Intestinal Microbiota: Current Understanding and Future Directions. *Cell Mol. Gastroenterol. Hepatol.* **2020**, *9*, 33–45. [CrossRef]
- Adamberg, S.; Adamberg, K. Prevotella Enterotype Associates with Diets Supporting Acidic Faecal pH and Production of Propionic Acid by Microbiota. Heliyon 2024, 10, e31134. [CrossRef] [PubMed]
- 5. He, T.; Cheng, X.; Xing, C. The Gut Microbial Diversity of Colon Cancer Patients and the Clinical Significance. *Bioengineered* **2021**, 12, 7046–7060. [CrossRef]
- 6. Mann, E.R.; Lam, Y.K.; Uhlig, H.H. Short-Chain Fatty Acids: Linking Diet, the Microbiome and Immunity. *Nat. Rev. Immunol.* **2024**, 24, 577–595. [CrossRef]
- 7. Nishida, A.; Imaeda, H.; Inatomi, O.; Bamba, S.; Sugimoto, M.; Andoh, A. The Efficacy of Fecal Microbiota Transplantation for Patients with Chronic Pouchitis: A Case Series. *Clin. Case Rep.* **2019**, 7, 782–788. [CrossRef] [PubMed]
- 8. Serban, D.E. Microbiota in Inflammatory Bowel Disease Pathogenesis and Therapy: Is It All About Diet? *Nutr. Clin. Prac.* **2015**, 30, 760–779. [CrossRef]
- 9. Kassinen, A.; Krogius-Kurikka, L.; Makivuokko, H.; Rinttila, T.; Paulin, L.; Corander, J.; Malinen, E.; Apajalahti, J.; Palva, A. The Fecal Microbiota of Irritable Bowel Syndrome Patients Differs Significantly from That of Healthy Subjects. *Gastroenterology* **2007**, 133, 24–33. [CrossRef]
- 10. Indiani, C.M.D.S.P.; Rizzardi, K.F.; Castelo, P.M.; Ferraz, L.F.C.; Darrieux, M.; Parisotto, T.M. Childhood Obesity and Firmicutes/Bacteroidetes Ratio in the Gut Microbiota: A Systematic Review. *Child Obes.* 2018, 14, 501–509. [CrossRef]
- 11. Guo, Y.; Xie, J.-P.; Deng, K.; Li, X.; Yuan, Y.; Xuan, Q.; Xie, J.; He, X.-M.; Wang, Q.; Li, J.-J.; et al. Prophylactic Effects of Bifidobacterium Adolescentis on Anxiety and Depression-Like Phenotypes After Chronic Stress: A Role of the Gut Microbiota-Inflammation Axis. *Front. Behav. Neurosci.* **2019**, *13*, 126. [CrossRef] [PubMed]
- 12. Seyedian, S.S.; Nokhostin, F.; Malamir, M.D. A Review of the Diagnosis, Prevention, and Treatment Methods of Inflammatory Bowel Disease. *J. Med. Life* **2019**, *12*, 113–122. [CrossRef] [PubMed]
- 13. Cushing, K.; Higgins, P.D.R. Management of Crohn Disease: A Review. JAMA 2021, 325, 69–80. [CrossRef]
- 14. Zhou, J.-L.; Bao, J.-C.; Liao, X.-Y.; Chen, Y.-J.; Wang, L.-W.; Fan, Y.-Y.; Xu, Q.-Y.; Hao, L.-X.; Li, K.-J.; Liang, M.-X.; et al. Trends and Projections of Inflammatory Bowel Disease at the Global, Regional and National Levels, 1990–2050: A Bayesian Age-Period-Cohort Modeling Study. *BMC Public Health* 2023, 23, 2507. [CrossRef]
- 15. Kaczmarek-Ryś, M.; Hryhorowicz, S.T.; Lis, E.; Banasiewicz, T.; Paszkowski, J.; Borejsza-Wysocki, M.; Walkowiak, J.; Cichy, W.; Krokowicz, P.; Czkwianianc, E.; et al. Crohn's Disease Susceptibility and Onset Are Strongly Related to Three NOD2 Gene Haplotypes. *J. Clin. Med.* **2021**, *10*, 3777. [CrossRef]
- 16. Kaplan, G.G.; Ng, S.C. Understanding and Preventing the Global Increase of Inflammatory Bowel Disease. *Gastroenterology* **2017**, 152, 313–321.e2. [CrossRef] [PubMed]
- 17. Braun, T.; Feng, R.; Amir, A.; Levhar, N.; Shacham, H.; Mao, R.; Hadar, R.; Toren, I.; Algavi, Y.; Abu-Saad, K.; et al. Diet-Omics in the Study of Urban and Rural Crohn Disease Evolution (SOURCE) Cohort. *Nat. Commun.* **2024**, *15*, 3764. [CrossRef]

18. Metwaly, A.; Dunkel, A.; Waldschmitt, N.; Raj, A.C.D.; Lagkouvardos, I.; Corraliza, A.M.; Mayorgas, A.; Martinez-Medina, M.; Reiter, S.; Schloter, M.; et al. Integrated Microbiota and Metabolite Profiles Link Crohn's Disease to Sulfur Metabolism. *Nat. Commun.* 2020, 11, 4322. [CrossRef]

- 19. Zheng, L.; Wen, X.-L.; Duan, S.-L. Role of Metabolites Derived from Gut Microbiota in Inflammatory Bowel Disease. *World J. Clin. Cases* **2022**, *10*, 2660–2677. [CrossRef]
- 20. Candelli, M.; Franza, L.; Pignataro, G.; Ojetti, V.; Covino, M.; Piccioni, A.; Gasbarrini, A.; Franceschi, F. Interaction between Lipopolysaccharide and Gut Microbiota in Inflammatory Bowel Diseases. *Int. J. Mol. Sci.* **2021**, 22, 6242. [CrossRef]
- 21. Markowiak-Kopeć, P.; Śliżewska, K. The Effect of Probiotics on the Production of Short-Chain Fatty Acids by Human Intestinal Microbiome. *Nutrients* **2020**, *12*, 1107. [CrossRef]
- 22. Shin, Y.; Han, S.; Kwon, J.; Ju, S.; Choi, T.G.; Kang, I.; Kim, S.S. Roles of Short-Chain Fatty Acids in Inflammatory Bowel Disease. *Nutrients* **2023**, *15*, 4466. [CrossRef]
- 23. Vandana, U.K.; Barlaskar, N.H.; Gulzar, A.B.M.; Laskar, I.H.; Kumar, D.; Paul, P.; Pandey, P.; Mazumder, P.B. Linking Gut Microbiota with the Human Diseases. *Bioinformation* **2020**, *16*, 196–208. [CrossRef]
- 24. Vakadaris, G.; Stefanis, C.; Giorgi, E.; Brouvalis, M.; Voidarou, C.; Kourkoutas, Y.; Tsigalou, C.; Bezirtzoglou, E. The Role of Probiotics in Inducing and Maintaining Remission in Crohn's Disease and Ulcerative Colitis: A Systematic Review of the Literature. *Biomedicines* 2023, 11, 494. [CrossRef] [PubMed]
- 25. Virk, M.S.; Virk, M.A.; He, Y.; Tufail, T.; Gul, M.; Qayum, A.; Rehman, A.; Rashid, A.; Ekumah, J.-N.; Han, X.; et al. The Anti-Inflammatory and Curative Exponent of Probiotics: A Comprehensive and Authentic Ingredient for the Sustained Functioning of Major Human Organs. *Nutrients* **2024**, *16*, 546. [CrossRef] [PubMed]
- 26. Day, A.S.; Lopez, R.N. Exclusive Enteral Nutrition in Children with Crohn's Disease. *World J. Gastroenterol.* **2015**, *21*, 6809–6816. [CrossRef] [PubMed]
- Allegretti, J.R. Update on Fecal Microbiota Transplantation for the Treatment of Inflammatory Bowel Disease. Gastroenterol. Hepatol. 2021, 17, 31–34.
- 28. Kahn, S.A. A Phase I/II, Double Blinded, Placebo Controlled, Single-Center Study of Fecal Microbiota Transplant (FMT) for the Treatment of Active Pediatric Ulcerative Colitis and Pediatric Active Crohn's Colitis. 2023. Available online: https://clinicaltrials.gov/ (accessed on 24 May 2024).
- 29. Voelker, R. What Is Ulcerative Colitis? *JAMA* **2024**, 331, 716. [CrossRef]
- 30. Definition & Facts of Ulcerative Colitis—NIDDK; National Institute of Diabetes and Digestive and Kidney Diseases: Bethesda, MD, USA, 2020.
- 31. Goldinova, A.; Tan, C.X.W.; Bouma, G.; Duijvestein, M.; Brand, H.S.; de Boer, N.K. Oral Health and Salivary Function in Ulcerative Colitis Patients. *United Eur. Gastroenterol. J.* **2020**, *8*, 1067–1075. [CrossRef]
- 32. Zhu, L.; Qiao, L.; Dou, X.; Song, X.; Chang, J.; Zeng, X.; Xu, C. Lactobacillus Casei ATCC 393 Combined with Vasoactive Intestinal Peptide Alleviates Dextran Sodium Sulfate-Induced Ulcerative Colitis in C57BL/6 Mice via NF-κB and Nrf2 Signaling Pathways. *Biomed. Pharmacother.* **2023**, *165*, 115033. [CrossRef]
- 33. Yang, Z.-H.; Liu, F.; Zhu, X.-R.; Suo, F.-Y.; Jia, Z.; Yao, S.-K. Altered Profiles of Fecal Bile Acids Correlate with Gut Microbiota and Inflammatory Responses in Patients with Ulcerative Colitis. *World J. Gastroenterol.* **2021**, 27, 3609–3629. [CrossRef] [PubMed]
- 34. Inciuraite, R.; Gedgaudas, R.; Lukosevicius, R.; Tilinde, D.; Ramonaite, R.; Link, A.; Kasetiene, N.; Malakauskas, M.; Kiudelis, G.; Jonaitis, L.V.; et al. Constituents of Stable Commensal Microbiota Imply Diverse Colonic Epithelial Cell Reactivity in Patients with Ulcerative Colitis. *Gut Pathog.* 2024, 16, 16. [CrossRef]
- 35. Imhann, F.; Vila, A.V.; Bonder, M.J.; Fu, J.; Gevers, D.; Visschedijk, M.C.; Spekhorst, L.M.; Alberts, R.; Franke, L.; van Dullemen, H.M.; et al. The Interplay of Host Genetics and the Gut Microbiota Underlying the Onset and Clinical Presentation of Inflammatory Bowel Disease. *Gut* 2019, 67, 108–119. [CrossRef] [PubMed]
- 36. Zhang, W.; Zou, G.; Li, B.; Du, X.; Sun, Z.; Sun, Y.; Jiang, X. Fecal Microbiota Transplantation (FMT) Alleviates Experimental Colitis in Mice by Gut Microbiota Regulation. *J. Microbiol. Biotechnol.* **2020**, *30*, 1132–1141. [CrossRef] [PubMed]
- 37. Sun, S.; Xu, X.; Liang, L.; Wang, X.; Bai, X.; Zhu, L.; He, Q.; Liang, H.; Xin, X.; Wang, L.; et al. Lactic Acid-Producing Probiotic Saccharomyces Cerevisiae Attenuates Ulcerative Colitis via Suppressing Macrophage Pyroptosis and Modulating Gut Microbiota. *Front. Immunol.* 2021, 12, 777665. [CrossRef] [PubMed]
- 38. Ning, L.; Zhou, Y.-L.; Sun, H.; Zhang, Y.; Shen, C.; Wang, Z.; Xuan, B.; Zhao, Y.; Ma, Y.; Yan, Y.; et al. Microbiome and Metabolome Features in Inflammatory Bowel Disease via Multi-Omics Integration Analyses across Cohorts. *Nat. Commun.* 2023, 14, 7135. [CrossRef]
- 39. Cheng, H.; Liu, J.; Zhang, D.; Wang, J.; Tan, Y.; Feng, W.; Peng, C. Ginsenoside Rg1 Alleviates Acute Ulcerative Colitis by Modulating Gut Microbiota and Microbial Tryptophan Metabolism. *Front. Immunol.* **2022**, *13*, 817600. [CrossRef]
- 40. Jia, D.-J.-C.; Wang, Q.-W.; Hu, Y.-Y.; He, J.-M.; Ge, Q.-W.; Qi, Y.-D.; Chen, L.-Y.; Zhang, Y.; Fan, L.-N.; Lin, Y.-F.; et al. Lactobacillus Johnsonii Alleviates Colitis by TLR1/2-STAT3 Mediated CD206+ macrophagesIL-10 Activation. *Gut Microbes* 2022, 14, 2145843. [CrossRef]
- 41. Sovran, B.; Planchais, J.; Jegou, S.; Straube, M.; Lamas, B.; Natividad, J.M.; Agus, A.; Dupraz, L.; Glodt, J.; Da Costa, G.; et al. Enterobacteriaceae Are Essential for the Modulation of Colitis Severity by Fungi. *Microbiome* **2018**, *6*, 152. [CrossRef]

42. Wang, M.-X.; Lin, L.; Chen, Y.-D.; Zhong, Y.-P.; Lin, Y.-X.; Li, P.; Tian, X.; Han, B.; Xie, Z.-Y.; Liao, Q.-F. Evodiamine Has Therapeutic Efficacy in Ulcerative Colitis by Increasing Lactobacillus Acidophilus Levels and Acetate Production. *Pharmacol. Res.* **2020**, 159, 104978. [CrossRef]

- 43. Wan, J.; Zhang, Y.; He, W.; Tian, Z.; Lin, J.; Liu, Z.; Li, Y.; Chen, M.; Han, S.; Liang, J.; et al. Gut Microbiota and Metabolite Changes in Patients With Ulcerative Colitis and *Clostridioides difficile* Infection. *Front. Microbiol.* **2022**, *13*, 802823. [CrossRef]
- 44. Dong, Y.; Liao, W.; Tang, J.; Fei, T.; Gai, Z.; Han, M. Bifidobacterium BLa80 Mitigates Colitis by Altering Gut Microbiota and Alleviating Inflammation. *AMB Express* **2022**, *12*, 67. [CrossRef]
- 45. Kawade, Y.; Sakai, M.; Okamori, M.; Morita, M.; Mizushima, K.; Ueda, T.; Takagi, T.; Naito, Y.; Itoh, Y.; Shimada, T. Administration of Live, but Not Inactivated, Faecalibacterium Prausnitzii Has a Preventive Effect on Dextran Sodium Sulfate-induced Colitis in Mice. *Mol. Med. Rep.* 2019, 20, 25–32. [CrossRef] [PubMed]
- 46. Martín, R.; Rios-Covian, D.; Huillet, E.; Auger, S.; Khazaal, S.; Bermúdez-Humarán, L.G.; Sokol, H.; Chatel, J.-M.; Langella, P. Faecalibacterium: A Bacterial Genus with Promising Human Health Applications. *FEMS Microbiol. Rev.* **2023**, 47, fuad039. [CrossRef] [PubMed]
- 47. Mills, R.H.; Dulai, P.S.; Vázquez-Baeza, Y.; Sauceda, C.; Daniel, N.; Gerner, R.R.; Batachari, L.E.; Ochoa, M.M.; Zhu, Q.; Weldon, K.; et al. Multi-Omics Analyses of the Ulcerative Colitis Gut Microbiome Link Bacteroides Vulgatus Proteases with Disease Severity. *Nat. Microbiol.* 2022, 7, 262–276. [CrossRef]
- 48. Khorsand, B.; Asadzadeh Aghdaei, H.; Nazemalhosseini-Mojarad, E.; Nadalian, B.; Nadalian, B.; Houri, H. Overrepresentation of Enterobacteriaceae and Escherichia Coli Is the Major Gut Microbiome Signature in Crohn's Disease and Ulcerative Colitis; a Comprehensive Metagenomic Analysis of IBDMDB Datasets. Front. Cell Infect. Microbiol. 2022, 12, 1015890. [CrossRef] [PubMed]
- 49. Wei, J.; Chen, C.; Feng, J.; Zhou, S.; Feng, X.; Yang, Z.; Lu, H.; Tao, H.; Li, L.; Xv, H.; et al. Muc2 Mucin O-Glycosylation Interacts with Enteropathogenic Escherichia Coli to Influence the Development of Ulcerative Colitis Based on the NF-kB Signaling Pathway. *J. Transl. Med.* 2023, 21, 793. [CrossRef] [PubMed]
- 50. Patel, M.; Kumar, A.; Jayawardena, D.; Priyamvada, S.; Anbazhagan, A.N.; Alrefai, W.A.; Gill, R.K.; Dudeja, P.K.; Saksena, S. Citrobacter Rodentium Infection Inhibits Colonic P-Glycoprotein Expression. *Gene Rep.* **2020**, *18*, 100549. [CrossRef]
- 51. Zhang, Y.; Zhao, Y.; Wu, W.; Meng, W.; Zhou, Y.; Qiu, Y.; Li, C. Protection against Ulcerative Colitis and Colorectal Cancer by Evodiamine via Anti-Inflammatory Effects. *Mol. Med. Rep.* **2022**, 25, 188. [CrossRef]
- 52. Meisner, J.; Bransford, T.; Miller, K.; Lee, J.; Jose, A.; Giuggio, M.; McComb, M.; Humphries, E.; Rosini, M.; Wingertzahn, M.; et al. The Synthetic Glycan KB295 Optimizes Microbiome Composition and Function in Ulcerative Colitis—Results from a Proof of Principle Human Study. *Gastroenterology* 2022, 162, S70–S71. [CrossRef]
- 53. Rabinowitz, J.D.; Enerbäck, S. Lactate: The Ugly Duckling of Energy Metabolism. Nat. Metab. 2020, 2, 566–571. [CrossRef]
- 54. Luzentales-Simpson, M.; Pang, Y.C.F.; Zhang, A.; Sousa, J.A.; Sly, L.M. Vedolizumab: Potential Mechanisms of Action for Reducing Pathological Inflammation in Inflammatory Bowel Diseases. *Front. Cell Dev. Biol.* **2021**, *9*, 612830. [CrossRef] [PubMed]
- 55. Eun, C.S.; Kwak, M.-J.; Han, D.S.; Lee, A.R.; Park, D.I.; Yang, S.-K.; Kim, Y.S.; Kim, J.F. Does the Intestinal Microbial Community of Korean Crohn's Disease Patients Differ from That of Western Patients? *BMC Gastroenterol.* **2016**, *16*, 28. [CrossRef]
- 56. Rehman, A.; Rausch, P.; Wang, J.; Skieceviciene, J.; Kiudelis, G.; Bhagalia, K.; Amarapurkar, D.; Kupcinskas, L.; Schreiber, S.; Rosenstiel, P.; et al. Geographical Patterns of the Standing and Active Human Gut Microbiome in Health and IBD. *Gut* 2016, 65, 238–248. [CrossRef] [PubMed]
- 57. Lopetuso, L.R.; Petito, V.; Graziani, C.; Schiavoni, E.; Paroni Sterbini, F.; Poscia, A.; Gaetani, E.; Franceschi, F.; Cammarota, G.; Sanguinetti, M.; et al. Gut Microbiota in Health, Diverticular Disease, Irritable Bowel Syndrome, and Inflammatory Bowel Diseases: Time for Microbial Marker of Gastrointestinal Disorders. *Dig. Dis.* 2017, *36*, 56–65. [CrossRef]
- 58. Vich Vila, A.; Imhann, F.; Collij, V.; Jankipersadsing, S.A.; Gurry, T.; Mujagic, Z.; Kurilshikov, A.; Bonder, M.J.; Jiang, X.; Tigchelaar, E.F.; et al. Gut Microbiota Composition and Functional Changes in Inflammatory Bowel Disease and Irritable Bowel Syndrome. *Sci. Transl. Med.* **2018**, *10*, eaap8914. [CrossRef]
- 59. Santoru, M.L.; Piras, C.; Murgia, A.; Palmas, V.; Camboni, T.; Liggi, S.; Ibba, I.; Lai, M.A.; Orrù, S.; Blois, S.; et al. Cross Sectional Evaluation of the Gut-Microbiome Metabolome Axis in an Italian Cohort of IBD Patients. *Sci. Rep.* **2017**, *7*, 9523. [CrossRef]
- 60. Martinez, E.; Crevecoeur, S.; Thirion, C.; Grandjean, J.; Fall, P.A.; Hayette, M.-P.; Michel, M.; Taminiau, B.; Louis, E.; Daube, G. Gut Microbiota Associated with *Clostridioides difficile* Carriage in Three Clinical Groups (Inflammatory Bowel Disease, *C. difficile* Infection and Healthcare Workers) in Hospital Field. *Microorganisms* 2023, 11, 2527. [CrossRef] [PubMed]
- 61. Nobili, A.; Pane, M.; Skvortsova, M.; Salem, M.B.; Morgenthaler, S.; Jamieson, E.; Stefano, M.D.; Bathrellou, E.; Mamalaki, E.; Ramos-Garcia, V.; et al. Innovative Biomarkers for Obesity and Type 1 Diabetes Based on Bifidobacterium and Metabolomic Profiling. *Microorganisms* **2024**, *12*, 931. [CrossRef]
- 62. Teixeira, T.F.S.; Grześkowiak, Ł.M.; Salminen, S.; Laitinen, K.; Bressan, J.; Peluzio, M.D.C.G. Faecal Levels of Bifidobacterium and Clostridium Coccoides but Not Plasma Lipopolysaccharide Are Inversely Related to Insulin and HOMA Index in Women. *Clin. Nutr.* 2013, 32, 1017–1022. [CrossRef]
- 63. Kubinak, J.L.; Petersen, C.; Stepens, W.Z.; Soto, R.; Bake, E.; O'Connell, R.M.; Round, J.L. MyD88 Signaling in T Cells Directs IgA-Mediated Control of the Microbiota to Promote Health. *Cell Host Microbe* **2015**, *17*, 153–163. [CrossRef] [PubMed]
- 64. Kameyama, K.; Itoh, K. Intestinal Colonization by a Lachnospiraceae Bacterium Contributes to the Development of Diabetes in Obese Mice. *Microbes Env.* **2014**, *29*, 427–430. [CrossRef] [PubMed]

65. Han, K.; Bose, S.; Wang, J.; Kim, B.-S.; Kim, M.J.; Kim, E.-J.; Kim, H. Contrasting Effects of Fresh and Fermented Kimchi Consumption on Gut Microbiota Composition and Gene Expression Related to Metabolic Syndrome in Obese Korean Women. *Mol. Nutr. Food Res.* **2015**, *59*, 1004–1008. [CrossRef]

- 66. Andoh, A.; Nishida, A.; Takahashi, K.; Inatomi, O.; Imaeda, H.; Bamba, S.; Kito, K.; Sugimoto, M.; Kobayashi, T. Comparison of the Gut Microbial Community between Obese and Lean Peoples Using 16S Gene Sequencing in a Japanese Population. *J. Clin. Biochem. Nutr.* 2016, 59, 65–70. [CrossRef]
- 67. Paredes-Sabja, D.; Shen, A.; Sorg, J.A. Clostridium Difficile Spore Biology: Sporulation, Germination, and Spore Structural Proteins. *Trends Microbiol.* **2014**, *22*, 406–416. [CrossRef]
- 68. Lyerly, D.M.; Krivan, H.C.; Wilkins, T.D. Clostridium Difficile: Its Disease and Toxins. *Clin. Microbiol. Rev.* **1988**, *1*, 1–18. [CrossRef]
- 69. Gerding, D.N.; Johnson, S.; Rupnik, M.; Aktories, K. Clostridium Difficile Binary Toxin CDT: Mechanism, Epidemiology, and Potential Clinical Importance. *Gut Microbes* **2014**, *5*, 15–27. [CrossRef]
- 70. Young, M.K.; Leslie, J.L.; Madden, G.R.; Lyerly, D.M.; Carman, R.J.; Lyerly, M.W.; Stewart, D.B.; Abhyankar, M.M.; Petri, W.A. Binary Toxin Expression by *Clostridioides difficile* Is Associated With Worse Disease. *Open Forum Infect. Dis.* **2022**, *9*, ofac001. [CrossRef] [PubMed]
- 71. Wan, S.; You, P.; Shi, Q.; Hu, H.; Zhang, L.; Chen, L.; Wu, Z.; Lin, S.; Song, X.; Luo, Y.; et al. Gut Microbiome Changes in Mouse, Mongolian Gerbil, and Hamster Models Following *Clostridioides difficile* Challenge. *Front. Microbiol.* **2024**, *15*, 1368194. [CrossRef]
- 72. Rao, K.; Malani, P.N. Diagnosis and Treatment of Clostridioides (Clostridium) Difficile Infection in Adults in 2020. *JAMA* 2020, 323, 1403. [CrossRef]
- 73. Gawey, B.J.; Khanna, S. Clostridioides difficile Infection: Landscape and Microbiome Therapeutics. *Gastroenterol. Hepatol.* **2023**, 19, 319–328.
- 74. Guh, A.Y.; Mu, Y.; Winston, L.G.; Johnston, H.; Olson, D.; Farley, M.M.; Wilson, L.E.; Holzbauer, S.M.; Phipps, E.C.; Dumyati, G.K.; et al. Trends in U.S. Burden of *Clostridioides difficile* Infection and Outcomes. N. Engl. J. Med. 2020, 382, 1320–1330. [CrossRef]
- Yunita, B.; Fauzi, A. Current Diagnostic and Treatment Approach of Clostridioides difficile Infection. Acta Medica Indones. 2023, 55, 231.
- 76. Pike, C.M.; Theriot, C.M. Mechanisms of Colonization Resistance Against *Clostridioides difficile*. *J. Infect. Dis.* **2020**, 223, S194–S200. [CrossRef] [PubMed]
- 77. Sarmiento, K.R.; Carr, A.; Diener, C.; Locey, K.J.; Gibbons, S.M. Island Biogeography Theory Provides a Plausible Explanation for Why Larger Vertebrates and Taller Humans Have More Diverse Gut Microbiomes. *ISME J.* **2024**, *18*, wrae114. [CrossRef] [PubMed]
- 78. Ridaura, V.K.; Faith, J.J.; Rey, F.E.; Cheng, J.; Duncan, A.E.; Kau, A.L.; Griffin, N.W.; Lombard, V.; Henrissat, B.; Bain, J.R.; et al. Gut Microbiota from Twins Discordant for Obesity Modulate Metabolism in Mice. *Science* **2013**, *341*, 1241214. [CrossRef]
- 79. Koliaki, C.; Dalamaga, M.; Liatis, S. Update on the Obesity Epidemic: After the Sudden Rise, Is the Upward Trajectory Beginning to Flatten? *Curr. Obes. Rep.* **2023**, *12*, 514–527. [CrossRef]
- 80. Stanislawski, M.A.; Dabelea, D.; Lange, L.A.; Wagner, B.D.; Lozupone, C.A. Gut Microbiota Phenotypes of Obesity. NPJ Biofilms Microbiomes 2019, 5, 18. [CrossRef] [PubMed]
- 81. Rohm, T.V.; Fuchs, R.; Müller, R.L.; Keller, L.; Baumann, Z.; Bosch, A.J.T.; Schneider, R.; Labes, D.; Langer, I.; Pilz, J.B.; et al. Obesity in Humans Is Characterized by Gut Inflammation as Shown by Pro-Inflammatory Intestinal Macrophage Accumulation. *Front. Immunol.* **2021**, *12*, 668654. [CrossRef]
- 82. Djalalinia, S.; Qorbani, M.; Peykari, N.; Kelishadi, R. Health Impacts of Obesity. Pak. J. Med. Sci. 2015, 31, 239–242.
- 83. Fabersani, E.; Portune, K.; Campillo, I.; López-Almela, I.; la Paz, S.M.; Romaní-Pérez, M.; Benítez-Páez, A.; Sanz, Y. Bacteroides Uniformis CECT 7771 Alleviates Inflammation within the Gut-adipose Tissue Axis Involving TLR5 Signaling in Obese Mice. *Nat. Portf.* 2021, 11, 11788. [CrossRef]
- 84. Vijay-Kumar, M.; Aitken, J.D.; Carvalho, F.A.; Cullender, T.C.; Mwangi, S.; Srinivasan, S.; Sitaraman, S.V.; Knight, R.; Ley, R.E.; Gewirtz, A.T. Metabolic Syndrome and Altered Gut Microbiota in Mice Lacking Toll-Like Receptor 5. *Science* **2010**, *328*, 228–231. [CrossRef]
- 85. Petersen, C.; Bell, R.; Klag, K.A.; Lee, S.-H.; Soto, R.; Ghazaryan, A.; Buhrke, K.; Ekiz, H.A.; Ost, K.S.; Boudina, S.; et al. T Cell–Mediated Regulation of the Microbiota Protects against Obesity. *Science* **2019**, *365*, eaat9351. [CrossRef] [PubMed]
- 86. Luck, H.; Khan, S.; Kim, J.H.; Copeland, J.K.; Revelo, X.S.; Tsai, S.; Chakraborty, M.; Cheng, K.; Chan, Y.T.; Nøhr, M.K.; et al. Gut-Associated IgA+ Immune Cells Regulate Obesity-Related Insulin Resistance. *Nat. Commun.* **2019**, *10*, 3650. [CrossRef]
- 87. Tran, H.Q.; Ley, R.E.; Gewirtz, A.T.; Chassaing, B. Flagellin-Elicited Adaptive Immunity Suppresses Flagellated Microbiota and Vaccinates against Chronic Inflammatory Diseases. *Nat. Commun.* **2019**, *10*, 5650. [CrossRef] [PubMed]
- 88. Mafra, D.; Borges, N.A.; Baptista, B.G.; Martins, L.F.; Borland, G.; Shiels, P.G.; Stenvinkel, P. What Can the Gut Microbiota of Animals Teach Us about the Relationship between Nutrition and Burden of Lifestyle Diseases? *Nutrients* 2024, 16, 1789. [CrossRef]
- 89. Gogarten, J.F.; Davies, T.J.; Benjamino, J.; Gogarten, J.P.; Graf, J.; Mielke, A.; Mundry, R.; Nelson, M.C.; Wittig, R.M.; Leendertz, F.H.; et al. Factors Influencing Bacterial Microbiome Composition in a Wild Non-Human Primate Community in Taï National Park, Côte d'Ivoire. *ISME J.* 2018, 12, 2559–2574. [CrossRef]
- 90. McKenzie, V.J.; Song, S.J.; Delsuc, F.; Prest, T.L.; Oliverio, A.M.; Korpita, T.M.; Alexiev, A.; Amato, K.R.; Metcalf, J.L.; Kowalewski, M.; et al. The Effects of Captivity on the Mammalian Gut Microbiome. *Integr. Comp. Biol.* 2017, 57, 690–704. [CrossRef] [PubMed]

91. Clavel, T.; Lagkouvardos, I.; Blaut, M.; Stecher, B. The Mouse Gut Microbiome Revisited: From Complex Diversity to Model Ecosystems—ClinicalKey. Available online: https://www.clinicalkey.com/#!/content/playContent/1-s2.0-S143842211630017 0?returnurl=null&referrer=null (accessed on 29 August 2024).

- 92. Pei, L.; Ke, Y.; Zhao, H.; Wang, L.; Jia, C.; Liu, W.; Fu, Q.; Shi, M.; Cui, J.; Li, S. Role of Colonic Microbiota in the Pathogenesis of Ulcerative Colitis. *BMC Gastroenterol.* **2019**, *19*, 10. [CrossRef]
- 93. Okayasu, I.; Hatakeyama, S.; Yamada, M.; Ohkusa, T.; Inagaki, Y.; Nakaya, R. A Novel Method in the Induction of Reliable Experimental Acute and Chronic Ulcerative Colitis in Mice. *Gastroenterology* **1990**, *98*, 694–702. [CrossRef]
- 94. Gray, S.M.; Moss, A.D.; Herzog, J.W.; Kashiwagi, S.; Liu, B.; Young, J.B.; Sun, S.; Bhatt, A.P.; Fodor, A.A.; Balfour Sartor, R. Mouse Adaptation of Human Inflammatory Bowel Diseases Microbiota Enhances Colonization Efficiency and Alters Microbiome Aggressiveness Depending on the Recipient Colonic Inflammatory Environment. *Microbiome* 2024, 12, 147. [CrossRef]
- 95. Rashed, R.; Valcheva, R.; Dieleman, L.A. Manipulation of Gut Microbiota as a Key Target for Crohn's Disease. *Front. Med.* **2022**, 9, 887044. [CrossRef] [PubMed]
- 96. Núñez-Sánchez, M.A.; Melgar, S.; O'Donoghue, K.; Martínez-Sánchez, M.A.; Fernández-Ruiz, V.E.; Ferrer-Gómez, M.; Ruiz-Alcaraz, A.J.; Ramos-Molina, B. Crohn's Disease, Host–Microbiota Interactions, and Immunonutrition: Dietary Strategies Targeting Gut Microbiome as Novel Therapeutic Approaches. *Int. J. Mol. Sci.* 2022, 23, 8361. [CrossRef] [PubMed]
- 97. Schaubeck, M.; Clavel, T.; Calasan, J.; Lagkouvardos, I.; Haange, S.B.; Jehmlich, N.; Basic, M.; Dupont, A.; Hornef, M.; von Bergen, M.; et al. Dysbiotic Gut Microbiota Causes Transmissible Crohn's Disease-like Ileitis Independent of Failure in Antimicrobial Defence. *Gut* 2016, 65, 225–237. [CrossRef] [PubMed]
- 98. Buffie, C.G.; Jarchum, I.; Equinda, M.; Lipuma, L.; Gobourne, A.; Viale, A.; Ubeda, C.; Xavier, J.; Pamer, E.G. Profound Alterations of Intestinal Microbiota Following a Single Dose of Clindamycin Results in Sustained Susceptibility to Clostridium Difficile-Induced Colitis. *Infect. Immun.* **2012**, *80*, 62–73. [CrossRef]
- 99. Antonopoulos, D.A.; Huse, S.M.; Morrison, H.G.; Schmidt, T.M.; Sogin, M.L.; Young, V.B. Reproducible Community Dynamics of the Gastrointestinal Microbiota Following Antibiotic Perturbation. *Infect. Immun.* **2009**, 77, 2367–2375. [CrossRef]
- 100. Piccioni, A.; Rosa, F.; Manca, F.; Pignataro, G.; Zanza, C.; Savioli, G.; Covino, M.; Ojetti, V.; Gasbarrini, A.; Franceschi, F.; et al. Gut Microbiota and Clostridium Difficile: What We Know and the New Frontiers. *Int. J. Mol. Sci.* **2022**, *23*, 13323. [CrossRef]
- 101. Sehgal, K.; Khanna, S. Gut Microbiome and *Clostridioides difficile* Infection: A Closer Look at the Microscopic Interface. *Ther. Adv. Gastroenterol.* **2021**, *14*, 1756284821994736. [CrossRef]
- 102. van der Vossen, E.W.J.; de Goffau, M.C.; Levin, E.; Nieuwdorp, M. Recent Insights into the Role of Microbiome in the Pathogenesis of Obesity. *Ther. Adv. Gastroenterol.* **2022**, *15*, 17562848221115320. [CrossRef]
- 103. Chauhan, S.; Jena, K.K.; Mehto, S.; Chauhan, N.R.; Sahu, R.; Dhar, K.; Yadav, R.; Krishna, S.; Jaiswal, P.; Chauhan, S. Innate Immunity and Inflammophagy: Balancing the Defence and Immune Homeostasis. *FEBS J.* **2022**, *289*, 4112–4131. [CrossRef]
- 104. Murphy, E.A.; Velazquez, K.T.; Herbert, K.M. Influence of High-Fat Diet on Gut Microbiota: A Driving Force for Chronic Disease Risk. *Curr. Opin. Clin. Nutr. Metab. Care* **2015**, *18*, 515–520. [CrossRef]
- 105. Feuerstadt, P.; Allegretti, J.R.; Dubberke, E.R.; Guo, A.; Harvey, A.; Yang, M.; Garcia-Horton, V.; Fillbrunn, M.; Tillotson, G.; Bancke, L.L.; et al. Efficacy and Health-Related Quality of Life Impact of Fecal Microbiota, Live-Jslm: A Post Hoc Analysis of PUNCH CD3 Patients at First Recurrence of *Clostridioides difficile* Infection. *Infect. Dis. Ther.* **2024**, *13*, 221–236. [CrossRef]
- 106. Binyamin, D.; Nitzan, O.; Azrad, M.; Hamo, Z.; Koren, O.; Peretz, A. The Microbial Diversity Following Antibiotic Treatment of *Clostridioides difficile* Infection. *BMC Gastroenterol.* **2021**, 21, 166. [CrossRef] [PubMed]
- 107. Khoruts, A.; Dicksved, J.; Jansson, J.K.; Sadowsky, M.J. Changes in the Composition of the Human Fecal Microbiome After Bacteriotherapy for Recurrent Clostridium Difficile-Associated Diarrhea. *J. Clin. Gastroenterol.* **2010**, *44*, 354. [CrossRef] [PubMed]
- 108. Roshan, N.; Clancy, A.K.; Borody, T.J. Faecal Microbiota Transplantation Is Effective for the Initial Treatment of Clostridium Difficile Infection: A Retrospective Clinical Review. *Infect. Dis. Ther.* **2020**, *9*, 935–942. [CrossRef]
- 109. Zou, B.; Liu, S.; Liu, S.; Liu, S.; Liu, S.; Dong, C.; Ruan, M.; Huang, Z.; Shu, S. Repeated and Multiple Fecal Microbiota Transplantations plus Partial Enteral Nutrition as the First-Line Treatment in Active Pediatric Crohn's Disease. *Front. Cell. Infect. Microbiol.* **2023**, 13, 1083236. [CrossRef] [PubMed]
- 110. Wilson, B.C.; Vatanen, T.; Cutfield, W.S.; O'Sullivan, J.M. The Super-Donor Phenomenon in Fecal Microbiota Transplantation. *Front. Cell. Infect. Microbiol.* **2019**, *9*, 2. [CrossRef]
- 111. Khanna, S.; Assi, M.; Lee, C.; Yoho, D.; Louie, T.; Knapple, W.; Aguilar, H.; Garcia-Diaz, J.; Wang, G.P.; Berry, S.M.; et al. Efficacy and Safety of RBX2660 in PUNCH CD3, a Phase III, Randomized, Double-Blind, Placebo-Controlled Trial with a Bayesian Primary Analysis for the Prevention of Recurrent Clostridioides difficile Infection. Drugs 2022, 82, 1527–1538. [CrossRef]
- 112. Lee, C.; Louie, T.; Bancke, L.; Guthmueller, B.; Harvey, A.; Feuerstadt, P.; Khanna, S.; Orenstein, R.; Dubberke, E.R. Safety of Fecal Microbiota, Live-Jslm (REBYOTA<sup>TM</sup>) in Individuals with Recurrent *Clostridioides difficile* Infection: Data from Five Prospective Clinical Trials. *Ther. Adv. Gastroenterol.* **2023**, *16*, 17562848231174277. [CrossRef]
- 113. Feuerstadt, P.; Louie, T.J.; Lashner, B.; Wang, E.E.L.; Diao, L.; Bryant, J.A.; Sims, M.; Kraft, C.S.; Cohen, S.H.; Berenson, C.S.; et al. SER-109, an Oral Microbiome Therapy for Recurrent *Clostridioides difficile* Infection. N. Engl. J. Med. 2022, 386, 220–229. [CrossRef]
- 114. Berenson, C.S.; Lashner, B.; Korman, L.Y.; Hohmann, E.; Deshpande, A.; Louie, T.J.; Sims, M.; Pardi, D.; Kraft, C.S.; Wang, E.E.L.; et al. Prevalence of Comorbid Factors in Patients With Recurrent *Clostridioides difficile* Infection in ECOSPOR III, a Randomized Trial of an Oral Microbiota–Based Therapeutic. *Clin. Infect. Dis.* 2023, 77, 1504–1510. [CrossRef] [PubMed]

115. Chuang, C.; Lee, K.C.; Wang, Y.P.; Lee, P.C.; Chang, T.E.; Huang, Y.H.; Lin, Y.T.; Hou, M.C. High Carriage Rate of Extended-Spectrum Beta-Lactamase Enterobacterales and Diarrheagenic Escherichia Coli in Healthy Donor Screening for Fecal Microbiota Transplantation. *Eur. J. Clin. Microbiol. Infect. Dis.* 2023, 42, 1103–1113. [CrossRef] [PubMed]

- 116. Rapoport, E.A.; Baig, M.; Puli, S.R. Adverse Events in Fecal Microbiota Transplantation: A Systematic Review and Meta-Analysis. *Ann. Gastroenterol.* **2022**, *35*, 150–163. [CrossRef] [PubMed]
- 117. Merrick, B.; Allen, L.; Masirah, M.Z.N.; Forbes, B.; Shawcross, D.L.; Goldenberg, S.D. Regulation, Risk and Safety of Faecal Microbiota Transplant. *Infect. Prev. Prac.* **2020**, *2*, 100069. [CrossRef]
- 118. Dwiyanto, J.; Hussain, M.H.; Reidpath, D.; Ong, K.S.; Qasim, A.; Lee, S.W.H.; Lee, S.M.; Foo, S.C.; Chong, C.W.; Rahman, S. Ethnicity Influences the Gut Microbiota of Individuals Sharing a Geographical Location: A Cross-Sectional Study from a Middle-Income Country. *Sci. Rep.* **2021**, *11*, 2618. [CrossRef]
- 119. van Leeuwen, P.T.; Brul, S.; Zhang, J.; Wortel, M.T. Synthetic Microbial Communities (SynComs) of the Human Gut: Design, Assembly, and Applications. *FEMS Microbiol. Rev.* **2023**, 47, fuad012. [CrossRef]
- 120. Dillon, K.M.; Morrison, H.A.; Powell, C.R.; Carrazzone, R.J.; Ringel-Scaia, V.M.; Winckler, E.W.; Council-Troche, R.M.; Allen, I.C.; Matson, J.B. Targeted Delivery of Persulfides to the Gut: Effects on the Microbiome. *Angew. Chem. Int. Ed. Engl.* **2021**, *60*, 6061–6067. [CrossRef] [PubMed]
- 121. Hsu, B.B.; Way, J.C.; Silver, P.A. Stable Neutralization of a Virulence Factor in Bacteria Using Temperate Phage in the Mammalian Gut. *mSystems* **2020**, *5*, 10–1128. [CrossRef]
- 122. Metchnikoff, E. *The Prolongation of Life: Optimistic Studies*; English translation edited by P. Chalmers Mitchell; Heinemann: London, UK, 1907.
- 123. Suez, J.; Zmora, N.; Zilberman-Schapira, G.; Mor, U.; Dori-Bachash, M.; Bashiardes, S.; Zur, M.; Regev-Lehavi, D.; Ben-Zeev Brik, R.; Federici, S.; et al. Post-Antibiotic Gut Mucosal Microbiome Reconstitution Is Impaired by Probiotics and Improved by Autologous FMT. *Cell* 2018, 174, 1406–1423.e16. [CrossRef]
- 124. Lau, C.S.M.; Chamberlain, R.S. Probiotics Are Effective at Preventing Clostridium Difficile-Associated Diarrhea: A Systematic Review and Meta-Analysis. *Int. J. Gen. Med.* **2016**, *9*, 27–37. [CrossRef]
- 125. Chen, K.; Zhu, Y.; Zhang, Y.; Hamza, T.; Yu, H.; Fleur, A.S.; Galen, J.; Yang, Z.; Feng, H. A Probiotic Yeast-Based Immunotherapy against *Clostridioides difficile* Infection. *Sci. Transl. Med.* **2020**, *12*, eaax4905. [CrossRef] [PubMed]
- 126. Hojsak, I. Probiotics in Functional Gastrointestinal Disorders. Adv. Exp. Med. Biol. 2019, 1125, 121–137. [CrossRef] [PubMed]
- 127. Guarner, F.; Sanders, M.E.; Szajewska, H.; Cohen, H.; Eliakim, R.; Herrera-deGuise, C.; Karakan, T.; Merenstein, D.; Piscoya, A.; Ramakrishna, B.; et al. World Gastroenterology Organisation Global Guidelines: Probiotics and Prebiotics. *J. Clin. Gastroenterol.* **2024**, *58*, 533–553. [CrossRef] [PubMed]
- 128. Rinninella, E.; Raoul, P.; Cintoni, M.; Franceschi, F.; Miggiano, G.A.D.; Gasbarrini, A.; Mele, M.C. What Is the Healthy Gut Microbiota Composition? A Changing Ecosystem across Age, Environment, Diet, and Diseases. *Microorganisms* 2019, 7, 14. [CrossRef] [PubMed]
- 129. Guo, J.; Li, L.; Cai, Y.; Kang, Y. The Development of Probiotics and Prebiotics Therapy to Ulcerative Colitis: A Therapy That Has Gained Considerable Momentum. *Cell Commun. Signal* **2024**, 22, 268. [CrossRef]
- 130. Wang, C.; Bai, J.; Wang, B.; Yu, L.; Tian, F.; Zhao, J.; Zhang, H.; Suo, H.; Chen, W.; Zhai, Q. Stachyose Modulates Gut Microbiota and Alleviates DSS-Induced Ulcerative Colitis in Mice. *Food Sci. Hum. Wellness* **2023**, *12*, 2211–2220. [CrossRef]
- 131. Guarino, M.P.L.; Altomare, A.; Emerenziani, S.; Di Rosa, C.; Ribolsi, M.; Balestrieri, P.; Iovino, P.; Rocchi, G.; Cicala, M. Mechanisms of Action of Prebiotics and Their Effects on Gastro-Intestinal Disorders in Adults. *Nutrients* **2020**, *12*, 1037. [CrossRef]
- 132. Bacha, A.A.; Suhail, M.; Awwad, F.A.; Ismail, E.A.A.; Ahmad, H. Role of Dietary Fiber and Lifestyle Modification in Gut Health and Sleep Quality. *Front. Nutr.* **2024**, *11*, 1324793. [CrossRef]
- 133. Jalanka, J.; Major, G.; Murray, K.; Singh, G.; Nowak, A.; Kurtz, C.; Silos-Santiago, I.; Johnston, J.M.; de Vos, W.M.; Spiller, R. The Effect of Psyllium Husk on Intestinal Microbiota in Constipated Patients and Healthy Controls. *Int. J. Mol. Sci.* **2019**, *20*, 433. [CrossRef]
- 134. Kim, E.K.; An, S.-Y.; Lee, M.-S.; Kim, T.H.; Lee, H.-K.; Hwang, W.S.; Choe, S.J.; Kim, T.-Y.; Han, S.J.; Kim, H.J.; et al. Fermented Kimchi Reduces Body Weight and Improves Metabolic Parameters in Overweight and Obese Patients. *Nutr. Res.* 2011, 31, 436–443. [CrossRef]
- 135. Hassan, N.E.; El-Masry, S.A.; Shebini, S.M.E.; Ahmed, N.H.; Mehanna, N.S.; Wahed, M.M.A.; Amine, D.; Hashish, A.; Selim, M.; Afify, M.A.S.; et al. Effect of Weight Loss Program Using Prebiotics and Probiotics on Body Composition, Physique, and Metabolic Products: Longitudinal Intervention Study. *Sci. Rep.* **2024**, *14*, 10960. [CrossRef]
- 136. Kanazawa, A.; Aida, M.; Yoshida, Y.; Kaga, H.; Katahira, T.; Suzuki, L.; Tamaki, S.; Sato, J.; Goto, H.; Azuma, K.; et al. Effects of Synbiotic Supplementation on Chronic Inflammation and the Gut Microbiota in Obese Patients with Type 2 Diabetes Mellitus: A Randomized Controlled Study. *Nutrients* 2021, 13, 558. [CrossRef] [PubMed]
- 137. Atazadegan, M.A.; Heidari-Beni, M.; Entezari, M.H.; Sharifianjazi, F.; Kelishadi, R. Effects of Synbiotic Supplementation on Anthropometric Indices and Body Composition in Overweight or Obese Children and Adolescents: A Randomized, Double-Blind, Placebo-Controlled Clinical Trial. *World J. Pediatr. WJP* **2023**, *19*, 356–365. [CrossRef]
- 138. Li, Y.; Yu, Y.; Wu, X.; Liu, B.; Ma, H.; Zhao, X.; Cao, S.; Ding, S.; Li, T.; Wang, X.; et al. Specially Designed Yogurt Supplemented with Combination of Pro- and Prebiotics Relieved Constipation in Mice and Humans. *Nutrition* **2022**, *103–104*, 111802. [CrossRef] [PubMed]

139. Rivero-Pino, F.; Casquete, M.; Castro, M.J.; Redondo Del Rio, P.; Gutierrez, E.; Mayo-Iscar, A.; Nocito, M.; Corell, A. Prospective, Randomized, Double-Blind Parallel Group Nutritional Study to Evaluate the Effects of Routine Intake of Fresh vs. Pasteurized Yogurt on the Immune System in Healthy Adults. *Nutrients* **2024**, *16*, 1969. [CrossRef] [PubMed]

- 140. Lazda, I.; Krūmiņa, A.; Zeltiņa, I.; Krūmiņa, N.; Ķibilds, J.; Siksna, I.; Vīksna, L.; Derovs, A. Microbial Community of Kefir and Its Impact on the Gastrointestinal Microbiome in Health and Disease. *Proc. Latv. Acad. Sci. Sect. B. Nat. Exact Appl. Sci.* 2020, 74, 58–64. [CrossRef]
- 141. Bourrie, B.C.T.; Ju, T.; Fouhse, J.M.; Forgie, A.J.; Sergi, C.; Cotter, P.D.; Willing, B.P. Kefir Microbial Composition Is a Deciding Factor in the Physiological Impact of Kefir in a Mouse Model of Obesity. *Br. J. Nutr.* **2021**, *125*, 129–138. [CrossRef]
- 142. Bourrie, B.C.T.; Cotter, P.D.; Willing, B.P. Traditional Kefir Reduces Weight Gain and Improves Plasma and Liver Lipid Profiles More Successfully than a Commercial Equivalent in a Mouse Model of Obesity. *J. Funct. Foods* **2018**, *46*, 29–37. [CrossRef]
- 143. Gupta, V.K.; Rajendraprasad, S.; Ozkan, M.; Ramachandran, D.; Ahmad, S.; Bakken, J.S.; Laudanski, K.; Gajic, O.; Bauer, B.; Zec, S.; et al. Safety, Feasibility, and Impact on the Gut Microbiome of Kefir Administration in Critically Ill Adults. *BMC Med.* 2024, 22, 80. [CrossRef]
- 144. Hatmal, M.M.; Nuirat, A.; Zihlif, M.A.; Taha, M.O. Exploring the Influence of Culture Conditions on Kefir's Anticancer Properties. *J. Dairy Sci.* **2018**, *101*, 3771–3777. [CrossRef]
- 145. Fraiz, G.M.; Costa, M.A.C.; Cardoso, R.R.; Hébert, J.R.; Zhao, L.; Corich, V.; Giacomini, A.; Milagro, F.I.; Barros, F.A.R. Fermentation | Free Full-Text | Black Tea Kombucha Consumption: Effect on Cardiometabolic Parameters and Diet Quality of Individuals with and without Obesity. Fermentation 2024, 10, 384. [CrossRef]
- 146. Greenwalt, C.J.; Steinkraus, K.H.; Ledford, R.A. Kombucha, the Fermented Tea: Microbiology, Composition, and Claimed Health Effects. *J. Food Prot.* **2000**, *63*, 976–981. [CrossRef]
- 147. Jakubczyk, K.; Kałduńska, J.; Kochman, J.; Janda, K. Chemical Profile and Antioxidant Activity of the Kombucha Beverage Derived from White, Green, Black and Red Tea. *Antioxidants* **2020**, *9*, 447. [CrossRef] [PubMed]
- 148. Kim, J.; Bhattarai, U.; Adhikari, K. The Healthy Eater's Idea and Related Behavior of a Healthy Diet—A Case Study with Kombucha Drinkers. *Beverages* **2022**, *8*, 25. [CrossRef]
- 149. Antolak, H.; Piechota, D.; Kucharska, A. Kombucha Tea—A Double Power of Bioactive Compounds from Tea and Symbiotic Culture of Bacteria and Yeasts (SCOBY). *Antioxidants* **2021**, *10*, 1541. [CrossRef]
- 150. Alaei, Z.; Doudi, M.; Setorki, M. The Protective Role of Kombucha Extract on the Normal Intestinal Microflora, High-Cholesterol Diet Caused Hypercholesterolemia, and Histological Structures Changes in New Zealand White Rabbits. *Avicenna J. Phytomed.* **2020**, *10*, 604–614. [PubMed]
- 151. Jung, Y.; Kim, I.; Mannaa, M.; Kim, J.; Wang, S.; Park, I.; Kim, J.; Seo, Y.-S. Effect of Kombucha on Gut-Microbiota in Mouse Having Non-Alcoholic Fatty Liver Disease. *Food Sci. Biotechnol.* **2018**, *28*, 261–267. [CrossRef]
- 152. Bamigbade, G.B.; Subhash, A.J.; Kamal-Eldin, A.; Nyström, L.; Ayyash, M. An Updated Review on Prebiotics: Insights on Potentials of Food Seeds Waste as Source of Potential Prebiotics. *Molecules* **2022**, *27*, 5947. [CrossRef]
- 153. Pérez-Burillo, S.; Pastoriza, S.; Fernández-Arteaga, A.; Luzón, G.; Jiménez-Hernández, N.; D'Auria, G.; Francino, M.P.; Rufián-Henares, J.Á. Spent Coffee Grounds Extract, Rich in Mannooligosaccharides, Promotes a Healthier Gut Microbial Community in a Dose-Dependent Manner. *J. Agric. Food Chem.* **2019**, *67*, 2500–2509. [CrossRef]
- 154. Godoy-Vitorino, F.; Vilanova-Cuevas, B. *Probiotic Microbiota of Fermented Tropical Wood Drinks: Mauby and Tepache*; ResearchGate: Santiago, Chile, 2019.
- 155. de la Fuente-Salcido, N.; Castañeda-Ramirez, J.; García-Almendárez, B.; Bideshi, D.; Salcedo-Hernández, R.; Barbosa-Corona, J. Isolation and Characterization of Bacteriocinogenic Lactic Bacteria from M-Tuba and Tepache, Two Traditional Fermented Beverages in México. Available online: https://onlinelibrary.wiley.com/doi/epdf/10.1002/fsn3.236 (accessed on 27 August 2024).
- 156. Lee, Y.K.; Low, K.Y.; Siah, K.; Drummond, L.M.; Gwee, K.A. Kiwifruit (Actinidia Deliciosa) Changes Intestinal Microbial Profile. *Microb. Ecol. Health Dis.* **2012**, 23, 18572. [CrossRef]
- 157. You, S.; Ma, Y.; Yan, B.; Pei, W.; Wu, Q.; Ding, C.; Huang, C. The Promotion Mechanism of Prebiotics for Probiotics: A Review. *Front. Nutr.* **2022**, *9*, 1000517. [CrossRef] [PubMed]
- 158. Gu, J.; Thomas-Ahner, J.M.; Riedl, K.M.; Bailey, M.T.; Vodovotz, Y.; Schwartz, S.J.; Clinton, S.K. Dietary Black Raspberries Impact the Colonic Microbiome and Phytochemical Metabolites in Mice. *Mol. Nutr. Food Res.* **2019**, *63*, 1800636. [CrossRef] [PubMed]
- 159. Chandrasekaran, P.; Weiskirchen, S.; Weiskirchen, R. Effects of Probiotics on Gut Microbiota: An Overview. *Int. J. Mol. Sci.* **2024**, 25, 6022. [CrossRef]
- 160. Ke, X.; Walker, A.; Haange, S.-B.; Lagkouvardos, I.; Liu, Y.; Schmitt-Kopplin, P.; von Bergen, M.; Jehmlich, N.; He, X.; Clavel, T.; et al. Synbiotic-Driven Improvement of Metabolic Disturbances Is Associated with Changes in the Gut Microbiome in Diet-Induced Obese Mice. *Mol. Metab.* **2019**, 22, 96–109. [CrossRef] [PubMed]

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