

ORIGINAL RESEARCH

A non-synonymous polymorphism in NBS I is associated with progression from chronic hepatitis B virus infection to hepatocellular carcinoma in a Chinese population

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Jie Li Department of General Surgery, Qianfoshan Hospital, Shandong University, 16766 Jingshi Road, Jinan 250014, People's Republic of China Email lijie0503@126.com **Purpose:** Nijmegen breakage syndrome 1 (NBS1) has a vital role in DNA double-strand break (DSB) repair, functioning as a sensor to identify and repair DNA damage and maintaining genomic stability by participating in the intra-S-phase checkpoint. Polymorphisms of NBS1 have been investigated in multiple cancers with variable results. To our best knowledge, no previous study has focused on the association between *NBS1* single-nucleotide polymorphisms (SNPs) and hepatitis B virus (HBV)-related hepatocellular carcinoma (HCC).

Patients and methods: Five *NBS1* SNPs were selected based on their potential functional impact. A hospital-based cohort, comprising 481 patients with HBV-related HCC, 508 patients with chronic hepatitis B virus infection (CHB), and 581 healthy controls, was recruited for genotyping analysis.

Results: After quality control, four SNPs were successfully genotyped (rs10464867, rs1063053, rs1805794, and rs709816), none of which were significantly associated with HCC or CHB compared with those of healthy controls. Similarly, the combined HBV-infected group (including the HCC and CHB groups) exhibited no significant associations with these SNPs compared with healthy controls. In contrast, comparison of the frequency of rs1805794 between patients with CHB and those with HCC identified a significant association (P=2.99E-03, odds ratio =1.31, 95% confidence interval =1.10-1.56).

Conclusion: These findings suggest that, as a non-synonymous SNP, the rs1805794 C/G polymorphism may play a role in the progression from CHB to HCC.

Keywords: NBS1, polymorphism, hepatitis B virus, Nijmegen breakage syndrome 1

Introduction

Primary liver cancer (PLC) is one of the most common causes of cancer death worldwide.¹ Half of PLC cases are estimated to occur in China,² and hepatocellular carcinoma (HCC) accounts for ~90% of disease. The majority of HCC cases are diagnosed with advanced liver cancer, for which the 5-year survival rate is <10%.³ In China, ~80% of HCC patients have a history of hepatitis B virus (HBV) infection.⁴ The high HCC rates in parts of Asia and sub-Saharan Africa largely reflect the elevated prevalence of chronic hepatitis B virus infection (CHB), with >8% of the populations in these regions chronically infected with the virus.⁵ The step from viral hepatitis and cirrhosis to HCC is considered as one of the most important processes in hepatocarcinogenesis; however, only a proportion of chronic HBV carriers develop HCC, indicating that host genetic factors may also be involved in the process;⁶

this hypothesis has been proven by several genome-wide association studies (GWASs).7-9 DNA damage occurs when harmful factors, such as HBV infection, aflatoxin, excessive drinking, and toxic metabolites, act on liver cell DNA, and the most detrimental form of DNA damage is DNA doublestrand breaks (DSBs). The important mechanisms of DSB repair in human beings are homologous recombination (HR) and non-homologous end joining (NHEJ), and the protein Nijmegen breakage syndrome 1 (NBS1) plays a vital role in identifying and repairing DSBs and maintaining genome stability in both pathways. 10 As a DNA repair gene, NBS1 was first identified in the rare autosomal recessive inherited disease, Nijmegen breakage syndrome (NBS), which is characterized by elevated sensitivity to ionizing radiation, chromosome instability, abnormal cell cycle checkpoint, and a high frequency of malignancies.11

The results of a recent study showed that expression of NBS1 was significantly increased in liver tumors compared with corresponding adjacent nontumor tissue, and higher levels were detected in poorly differentiated HCCs,¹² indicating that increased expression of NBS1 may have a significant role in liver cancer progression. The c-Myc protooncogene encodes a ubiquitous transcription factor involved in the control of cell growth and plays an essential role in carcinogenesis. NBS1 is a direct transcriptional target of c-Myc, linking the function of c-Myc to DNA DSB repair, and increased NBS1 can induce cell growth promotion.¹³ Increased NBS1 expression can promote the phosphorylation of Akt and increase the expression of the downstream mTOR protein. Activation of the Akt-mTOR-p70S6K pathway has a significant role in HCC progression through promotion of neoangiogenesis.14 Studies in head and neck cancer have generated similar results, indicating that increased expression of NBS1 is a significant prognostic marker of advanced head and neck squamous cell carcinoma and that its overexpression is an oncogenic event that contributes to transformation through activation of the PI3-kinase/Akt pathway.¹⁵

Functional genetic variation of *NBS1* may influence the susceptibility to tumor development as a result of DNA repair deficiency. Genetic mutation of *NBS1* can promote the occurrence of liver cancer in mice, ¹⁶ and multiple single-nucleotide polymorphisms (SNPs) of *NBS1* have been reported as associated with risk for several cancers, including those of the breast, ¹⁷ nasopharynx, ¹⁸ liver, ¹⁹ colorectum, ²⁰ lung, ²¹ and prostate, ²² in addition to leukemia. ²³ However, to our best knowledge, no previous study has focused on the association between *NBS1* SNPs and HBV-related HCC. We hypothesized that five SNPs in *NBS1* may be positively associated

with susceptibility among patients with HBV-related HCC, given their strong association with cancer susceptibility; therefore, we investigated associations between *NBS1* SNPs and risk of HBV-related HCC.

Patients and methods

Study subjects

Our study population was recruited in a population-based case—control study in Jinan, Shandong Province, and all subjects were ethnic Han Chinese. Patients with HBV-related HCC (HCC patients; n=481), sex- and age (±5 years)-matched patients with CHB (n=508), and sex- and age-matched healthy controls (n=581) were recruited. HBV-related patients with HCC and CHB were recruited between January 2014 and December 2016 at the Affiliated Hospital of Shandong Academy of Medical Sciences (Jinan, Shandong Province, China). Control subjects were randomly selected from among individuals attending for regular physical examination at the same hospital during the same time period when patients were collected.

All HCC patients were diagnosed by pathological examination and computerized tomography and those with any another malignant tumor were excluded from the study. Patients with CHB and HBV-related HCC were defined according to the following serological parameters: HBsAg positive for >6 months, anti-HBs negative, and anti-HBc positive. All individuals were negative for antibodies to hepatitis C virus, hepatitis D virus, and human immunodeficiency virus. Healthy controls were individuals without any of the virus infections mentioned earlier. Individuals who smoked for ≥6 months during their lifetime were considered as smokers and those who drank at least once per week for >1 year were classified as drinkers. This study was approved by the ethics committee of the Affiliated Hospital of Shandong Academy of Medical Sciences. Written informed consent was obtained from each subject at recruitment.

SNP selection

SNP markers were selected from the previous GWASs and candidate—gene association studies of *NBS1* gene.^{19,21,24,25} SNPs were selected based on the following criteria: 1) previously reported to be associated with any kind of cancer; 2) rare variants (MAF <0.01) in the Chinese population were excluded; 3) all variants located in the coding region (synonymous [SYN] or non-synonymous [NSM]); 4) selection of potential regulatory elements, according to the Encyclopedia of DNA Elements (ENCODE) database (www.encodeproject.org); and 5) inclusion of only one

SNP in high linkage disequilibrium (LD; $r^2 > 0.8$). Finally, five SNPs were selected for the subsequent validation analysis: rs10464867, rs1063053, rs1061302, rs1805794, and rs709816.

Genotyping

Genomic DNA samples were extracted from 500 μ L ethylenediaminetetraacetic acid (EDTA)-anticoagulated peripheral blood samples using a DNA extraction kit (Lab-Aid 820, Zeesan Corp., Xiamen, Fujian Province, China), according to the manufacturer's instructions. DNA sample quantification was performed using a NanoDrop 2000 Spectrophotometer (Thermo Fisher Scientific, Waltham, MA, USA) and diluted to a final concentration of 20 ng/ μ L.

Genotyping analysis of all samples was conducted using TaqMan custom genotyping assays on a 7500 Real-Time PCR System (Thermo Fisher Scientific), according to the manufacturer's instructions. All five SNPs were genotyped using TaqMan SNP Genotyping Assay kits on the GeneAmp 7500 Sequence Detection System using specific TaqMan probes. SNPs with a call rate <90% or generating results that deviated from the Hardy–Weinberg equilibrium (P<0.01) in control subjects were excluded from further analysis. Of the five SNPs included in the validation study, only one (rs1061302) was excluded from genotyping analysis because of the generation of inadequate cluster plots.

Statistical analysis

Two-sided chi-square tests were used to assess differences in the distribution of age, sex, smoking, and drinking status between patients and controls; values of P < 0.05 were considered as statistically significant. Data analyses and summarizations were conducted using SPSS 19.0 for Windows (IBM Corporation, Armonk, NY, USA). The Cochran–Armitage trend test was used to analyze the genotype–phenotype association in each group using Plink v1.07 software (Center for Human Genetic Research, Massachusetts General Hospital and the Broad Institute of Harvard and MIT, Cambridge, MA, USA). Bonferroni correction for multiple SNP testing was performed with P < 0.0125 considered as a significant association.

Results

The demographic and clinical characteristics of recruited patients are given in Table 1. The mean age of patients with HCC was 56 years, while that of patients with CHB and healthy controls were 51 and 53 years, respectively. In all, 79% of patients with HCC were males, while 77.4%

Table I Demographic and clinical characteristics of patients with HCC and CHB and healthy controls

Characteristics	cteristics HCC cases (n=481)		CHB cases I) (n=508)		Healthy controls (n=581)		HCC vs	HCC vs controls	
							CHB		
	n	%	n	%	n	%	P-value	P-value	
Age (years)							0.416	0.385	
≤56	267	55.5	295	58. I	307	52.8			
>56	214	44.5	213	41.9	274	47.2			
Sex							0.533	0.684	
Male	380	79.0	393	77.4	453	78.0			
Female	101	21.0	115	22.6	128	22.0			
Smoking status							0.302	< 0.001	
Smoker	306	63.6	307	60.4	282	48.5			
Nonsmoker	175	36.4	201	39.6	299	51.5			
Drinking status							0.728	< 0.001	
Drinker	294	61.1	305	60.0	232	39.9			
Nondrinker	187	38.9	203	40.0	349	60.1			

Abbreviations: HCC, hepatocellular carcinoma; CHB, chronic hepatitis B virus infection.

and 78% of patients with CHB and healthy controls were males, respectively. There were no significant differences in age and sex among groups (P>0.05). Of HCC patients with HBV, 63.6% and 61.1% had histories of smoking and drinking, respectively, which were significantly higher than that of healthy controls but not significantly different to that of patients with CHB.

Based on the previous publications and the criteria described in the "Patients and methods" section, we selected five candidate SNPs (rs10464867, rs1063053, rs1061302, rs1805794, and rs709816) for validation analysis. After quality control, data from 481 HCC, 508 CHB, and 581 controls samples were available for validation analysis, and four SNPs (rs10464867, rs1063053, rs1805794, and rs709816) were successfully genotyped.

Genotyping results are presented in Table 2. First, the association of the SNPs was evaluated in patients with HCC or CHB compared with healthy controls. None of the four SNPs showed significant associations in these two comparisons. As the HCC and CHB group patients were all infected with HBV, we then compared the HBV-infected group (consisting of the HCC and CHB groups combined) with healthy controls. Similarly, none of the four SNPs showed significant associations. Finally, we compared patients with HCC having HBV infection with those from the CHB group who had no HCC involvement. A significant association was identified with the *NBS1* SNP rs1805794, (*P*=2.99E–03) with an odds ratio of 1.31, suggesting a role for this SNP in the progression to HCC from CHB. The rs1805794 SNP is

Table 2 Association results of four candidate SNPs in HCC and CHB patients and healthy controls

SNP	Minor allele/	Group	Frequency_	Frequency_	P-value	OR	L95	U95
information major allele			Cases	Controls				
rs1805794	C/G	HCC vs control	51.48%	47.16%	4.44E-02	1.19	1.00	1.41
chr8: 90990479		CHB vs control	44.80%	47.16%	1.90E-01	0.89	0.75	1.06
NSM	HCC + CHB vs control	47.11%	47.16%	9.77E-01	1.00	0.86	1.15	
		HCC vs CHB	51.48%	44.80%	2.99E-03*	1.31	1.10	1.56
rs10464867	T/C	HCC vs control	14.96%	13.17%	2.29E-01	1.16	0.91	1.48
chr8: 90945598		CHB vs control	12.06%	13.17%	4.20E-01	0.90	0.69	1.17
U3	HCC + CHB vs control	13.48%	13.17%	8.03E-01	1.03	0.83	1.27	
		HCC vs CHB	14.96%	12.06%	5.94E-02	1.28	0.99	1.66
rs1063053	A/G	HCC vs control	43.58%	46.47%	1.76E-01	0.89	0.75	1.05
chr8: 90947537		CHB vs control	47.71%	46.47%	4.56E-01	1.07	0.90	1.27
U3	HCC + CHB vs control	46.53%	46.47%	9.74E-01	1.00	0.87	1.16	
		HCC vs CHB	43.58%	47.71%	6.52E-02	0.85	0.71	1.01
rs709816	A/G	HCC vs control	32.58%	30.12%	2.17E-01	1.12	0.94	1.34
chr8: 90967711		CHB vs control	30.35%	30.12%	9.48E-01	0.99	0.82	1.21
SYN		HCC + CHB vs control	30.53%	30.12%	8.12E-01	1.02	0.87	1.19
		HCC vs CHB	32.58%	30.35%	2.87E-01	1.11	0.92	1.34

Note: *P<0.0125 was considered as significant (Bonferroni correction for four tested SNPs).

Abbreviations: SNP, single-nucleotide polymorphism; HCC, hepatocellular carcinoma; CHB, chronic hepatitis B virus infection; OR, odds ratio; NSM, non-synonymous; U3, 3'-untranslated region; SYN, synonymous; L95, lower-tail 95%; U95, upper-tail 95%.

located in the fifth exon of the *NBS1* gene and encodes an NSM substitution of glutamate (Glu) to glutamine (Gln).

Discussion

In this study, we investigated the association between NBS1 polymorphisms and HBV-related HCC using a case-control study to identify genetic factors that confer individual susceptibility to HBV-related HCC. The results showed that the functional polymorphism, rs1805794 C/G in NBS1, was associated with an increased risk for developing HBV-related HCC among CHB patients. A Chinese study by Huang et al¹⁹ previously investigated the association between genetic polymorphisms in NBS1 and HCC risk, analyzing genotypes in 865 patients with HCC and 900 cancer-free controls, and found that the rs1805794 C/G polymorphism in NBS1 was associated with an increased risk of developing HCC in an allele dose-response manner. There was no independent CHB group in that study; however, interestingly, 83.1% of patients with HCC had HBV infection and 13% of controls had HBV infection; hence, the conclusions of that report are consistent with those of our study. In China, HBV infection is the most important risk factor for HCC, and HCC usually occurs in patients with HBV-associated cirrhosis.4 In the US and several other Western developed countries, alcoholrelated cirrhosis is thought to account for the majority of liver cancers.1 Harmful factors, such as HBV infection and alcohol abuse, cause DNA and cell damage in the liver. If such damage is not repaired in a timely manner, the accumulation

of abnormalities may lead to liver cancer. In patients with CHB, integration of HBV into the host genome has been found to lead to novel fusion transcripts and/or local genomic instability, resulting in secondary deletions, rearrangements, duplications, or inversions of the host and/or viral genomic sequences.²⁶

The NBS1 gene, mapping to human chromosome 8q21, is ~50 kb in length, including 16 exons, and encodes the 754 amino acid (AA) protein NBS1 (p95).27 NBS1 is an important component of the MRE-RAD50-NBS1 (MRN) complex that has a critical role in DNA DSB repair. MRN functions as a sensor for DNA damage in NHEJ and participates in DNA repair and the intra-S-phase cell cycle checkpoint in HR.²⁸ It binds directly to phosphorylated histone, H2AX, located around DSBs, to maintain genomic stability and prevent cells from undergoing telomeric fusion.²⁹ The NBS1 protein has three major functional regions: the N-terminus (1-196 AAs), the central region (278-343 AAs), and the C-terminus (665–693 AAs).²⁸ The N-terminal region contains a forkhead-associated (FHA) domain (24-108 AAs) and two breast cancer carboxy-terminus (BRCT) domains (BRCT1: 114-183 AAs and BRCT2: 221-291 AAs). The FHA and BRCT domains are widely conserved in eukaryotic nuclear proteins related to cell cycle checkpoints or DNA repair, and the central region functions in signal transduction in response to damage.²⁷ The C-terminal region contains a binding site for the MRN,28 which is crucial for detection and repair of DSBs via the NHEJ and HR pathways.

Animal experiments have provided experimental evidence that NBS1 heterozygosity predisposes cells to malignancy, most likely because of chromosomal instability and defects in DNA repair. In the study, heterozygous knockout (NBS1±) mice developed a wide array of solid tumors, including those of the liver, mammary gland, prostate, and lung, in addition to lymphomas.¹⁶ Other studies have demonstrated that the overexpression of NBS1 is associated with the occurrence and progression of some cancers, including head and neck squamous cell carcinomas, 15 prostate cancer,³⁰ liver cancer,³¹ and esophageal malignancies.³² The study showed that overexpressed NBS1 interacts directly, through its C-terminal region, with the N-terminal domain (1–108 AAs) of the p110α catalytic subunit of PI3-kinase to stimulate PI3-kinase activity, 33 and the activated PI3K/ Akt pathway can promote angiogenesis, which is involved in tumor development.³⁴

As an NSM SNP, the NBS1 rs1805794 C/G polymorphism leads to a substitution of Glu with Gln. This SNP site is in the BRCT domain, through which NBS1 can interact with BRCA1 (one of two familial breast cancers' mutated genes) to form the BRCA1-associated genome surveillance complex (BASC),35 which is responsible for the recognition and repair of aberrant DNA. Since either one or two missense changes in the dominant genetic model may affect the function of NBS1 and interfere with protein-protein interaction,36 the rs1805794 C/G polymorphism could potentially affect the function of the NBS1 protein and the protein-protein interaction between NBS1 and BRCA1. The NBS1 rs1805794 C/G polymorphism has frequently been investigated in different cancer types and geographical areas, and its variant genotype is associated with increased morbidity of several cancers, including nasopharyngeal carcinoma, 18 hepatic cancer, 19 acute lymphoblastic leukemia,³⁷ lung cancer,³⁸ and prostate cancer.²² The findings of our present study suggest that the rs1805794 C/G polymorphism also increases the risk of HCC in patients who have been infected with HBV, rather than increasing the risk of HBV infection in individuals, and that the rs1805794 C/G polymorphism may affect the DNA damage repair function of NBS1 in hepatitis-infected liver cells, increasing the risk of HCC.

Along with *NBS1*, *RAD52* is another key gene involved in the HR pathway, and the association between its genetic polymorphisms and HBV-related HCC has also been investigated.³⁹ In that study, genotypes were determined in two independent case—control cohorts from two regions of China. The results indicated that the RAD52 rs7963551 A/C

polymorphism contributes to susceptibility to developing HBV-related HCC. Furthermore, genotype–phenotype correlation analyses were carried out using 44 liver tissue samples, and rs7963551 CC or AC genotypes were identified as associated with a statistically significant increase in *RAD52* mRNA expression, consistent with a functional correlation between allelic regulation of *RAD52* expression and the rs7963551 SNP in tumor cells.

Moreover, along with NBS1, another direct transcriptional target gene of c-Myc, EPB41, is also a HCC susceptibility gene; the same researchers identified the EPB41 SNP rs157224 G/T as associated with susceptibility to HBVrelated HCC in Chinese populations. Binding site prediction and in vivo and vitro experiments to determine the function of this SNP revealed that rs157224 G/T affected binding of the transcription factor, c-Myc, to a site involved in the regulation of the EPB41 gene. 40 The current study has some limitations. As it is a hospital-based study, there may be inherent selection bias. Moreover, the statistical power of our study is limited by the sample size, particularly for statistical analyses of gene-environment interactions; however, the fact that the genotype frequencies among controls fitted the Hardy-Weinberg equilibrium suggests that subject selection was sufficiently random. Finally, based on relevant publications, 39,40 further experiments are required to test the influence of the rs1805794 C/G polymorphism on NBS1 regulation.

Conclusion

Our study suggests that, as an NSM substitution, the rs1805794 C/G polymorphism in *NBS1* may be a genetic susceptibility factor for the risk of HBV-related HCC developing from CHB in a Chinese population. These findings require verification using larger, preferably population-based, case—control studies and well-designed mechanistic experiments.

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Disclosure

The authors report no conflicts of interest in this work.

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