

Genome Sequence of *Serratia plymuthica* Strain S13, an Endophyte with Germination- and Plant-Growth-Promoting Activity from the Flower of Styrian Oil Pumpkin

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The bacterium *Serratia plymuthica* strain S13 was demonstrated to colonize various plant-associated microhabitats and to suppress damping-off diseases. The completed genome sequence has a size of 5.5 Mb, containing 4,957 putative proteinencoding regions, and will be used to identify genetic determinants enabling the bacterium to escort a plant's entire life cycle.

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embers of the enterobacterial genus Serratia have been reported to be associated with many plants, where they perform beneficial functions by suppressing phytopathogens and promoting growth (1). The strain Serratia plymuthica S13 was isolated from the anthosphere of Styrian oil pumpkin (Cucurbita pepo L. subsp. pepo var. styriaca) but was also shown to colonize the rhizosphere, endosphere, and seeds of the same plant species (2). S13 was selected as a strong antagonist toward *Didymella* bryoniae, the causal agent of black rot in pumpkins. When applied as a seed treatment, S. plymuthica enhanced the germination rate and controlled damping-off diseases under field conditions with efficiency comparable to that of chemical fungicides containing copper, metalaxyl-M, or fludioxonil (3). The genome was sequenced to reveal the genetic predisposition that facilitates versatile microniche colonization and seedprotecting capacity.

The *S. plymuthica* S13 genome sequence was obtained using the Roche/454 GS-FLX Titanium sequencing platform. A draft assembly based on 339,698 reads of a standard shotgun library and 427,170 reads of an 8-kbp paired-end library (LGC Genomics, Berlin, Germany) with a total of 191.6 Mb (35-fold coverage) was generated with Newbler assembler (software release 2.6) (Roche Diagnostics GmbH, Mannheim, Germany). This assembly consisted of 34 contigs, 27 of which could be joined into a single circular scaffold. Gaps resulting from repetitive sequences were resolved by *in silico* gap filling; remaining gaps were closed by PCR followed by Sanger sequencing, yielding a final genome of 5,467,306 bp.

Genes were identified with the software tool Prodigal 1/1/20 (4). Functional annotation of the predicted genes was performed by use of the GenDB system 2.2 (5) and JCoast 1.7 (6), which provided annotations with respect to clusters of orthologous groups (COG) (7), Pfam (8), and gene ontology (GO) (9). The final genome includes 5.4 Mb, with a GC content of 56.19%. The number of putative genes totals 4,957, of which 4,866 are protein

coding. There are seven instances of the 5S-23S-to-16S rRNA cluster and 84 tRNAs.

We identified putative genetic elements enabling beneficial interaction with the plant as well as antagonism toward fungal pathogens. Bacterium-plant interaction was assumed to be conferred by genes involved in plant growth hormone synthesis/signaling (auxin, salicylic acid, and butandiol) and root colonization (type I secretion system, adhesin, and hemagglutinin). Other genes encoding fungal cell-wall-degrading enzymes (chitinases and β -1,3 glucanases) and nonribosomal peptides (syringomycin and siderophores) are probably involved in the suppression of fungi.

Nucleotide sequence accession number. The *Serratia plym-uthica* S13 genome sequence and annotation data have been deposited in GenBank under the accession number CP006566.

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REFERENCES

- 1. Berg G. 2000. Diversity of antifungal and plant-associated *Serratia plym-uthica* strains. J. Appl. Microbiol. 88:952–960.
- Fürnkranz M, Lukesch B, Müller H, Huss H, Grube M, Berg G. 2012. Microbial diversity inside pumpkins: microhabitat-specific communities display a high antagonistic potential against phytopathogens. Microb. Ecol. 63:418–428.
- Fürnkranz M, Adam E, Müller H, Grube M, Huss H, Winkler J, Berg G. 2012. Promotion of growth, health and stress tolerance of Styrian oil pumpkins by bacterial endophytes. Eur. J. Plant Pathol. 134:– 509–519.
- Hyatt D, Chen GL, Locascio PF, Land ML, Larimer FW, Hauser LJ. 2010. Prodigal: prokaryotic gene recognition and translation initiation site identification. BMC Bioinformatics 11:119. doi:10.1186/1471-2105-11-119.
- 5. Meyer F, Goesmann A, McHardy AC, Bartels D, Bekel T, Clausen J, Kalinowski J, Linke B, Rupp O, Giegerich R, Pühler A. 2003. GenDB—an

open source genome annotation system for prokaryote genomes. Nucleic Acids Res. **31**:2187–2195.

- Richter M, Rosselló-Móra R. 2009. Shifting the genomic gold standard for the prokaryotic species definition. Proc. Natl. Acad. Sci. U. S. A. 106: 19126–19131.
- Tatusov RL, Fedorova ND, Jackson JD, Jacobs AR, Kiryutin B, Koonin EV, Krylov DM, Mazumder R, Mekhedov SL, Nikolskaya AN, Rao BS, Smirnov S, Sverdlov AV, Vasudevan S, Wolf YI, Yin JJ, Natale DA. 2003.

The COG database: an updated version includes eukaryotes. BMC Bioinformatics 4:41. doi:10.1186/1471-2105-4-41.

- Finn RD, Mistry J, Tate J, Coggill PC, Heger A, Pollington JE, Gavin OL, Gunasekaran P, Ceric G, Forslund K, Holm L. 2010. The Pfam protein families database. Nucleic Acids Res. 38:D211–D222. doi:10.1093/nar/gkp 985.
- 9. Gene Ontology Consortium. 2001. Creating the gene ontology resource: design and implementation. Genome Res. 11:1425–1433.