



# **Programming of Embryonic Development**

Carl R. Dahlen<sup>1</sup>, Pawel P. Borowicz<sup>1</sup>, Alison K. Ward<sup>1</sup>, Joel S. Caton<sup>1</sup>, Marta Czernik<sup>2</sup>, Luca Palazzese<sup>3</sup>, Pasqualino Loi<sup>2</sup> and Lawrence P. Reynolds<sup>1,\*</sup>

- <sup>1</sup> Center for Nutrition and Pregnancy, Department of Animal Sciences, North Dakota State University, Fargo, ND 58108, USA; carl.dahlen@ndsu.edu (C.R.D.); pawel.borowicz@ndsu.edu (P.P.B.); alison.ward@ndsu.edu (A.K.W.); joel.caton@ndsu.edu (J.S.C.)
- <sup>2</sup> Faculty of Veterinary Medicine, University of Teramo, 64100 Teramo, Italy; mczernik@unite.it (M.C.); ploi@unite.it (P.L.)
- <sup>3</sup> Institute of Genetics and Animal Biotechnology of the Polish Academy of Sciences, Warsaw, Jastrzębiec, 05-552 Magdalenka, Poland; Ipalazzese@unite.it
- Correspondence: larry.reynolds@ndsu.edu

**Abstract:** Assisted reproductive techniques (ART) and parental nutritional status have profound effects on embryonic/fetal and placental development, which are probably mediated via "programming" of gene expression, as reflected by changes in their epigenetic landscape. Such epigenetic changes may underlie programming of growth, development, and function of fetal organs later in pregnancy and the offspring postnatally, and potentially lead to long-term changes in organ structure and function in the offspring as adults. This latter concept has been termed developmental origins of health and disease (DOHaD), or simply developmental programming, which has emerged as a major health issue in animals and humans because it is associated with an increased risk of non-communicable diseases in the offspring, including metabolic, behavioral, and reproductive dysfunction. In this review, we will briefly introduce the concept of developmental programming and its relationship to epigenetics. We will then discuss evidence that ART and periconceptual maternal and paternal nutrition may lead to epigenetic alterations very early in pregnancy, and how each pregnancy experiences developmental programming based on signals received by and from the dam. Lastly, we will discuss current research on strategies designed to overcome or minimize the negative consequences or, conversely, to maximize the positive aspects of developmental programming.

**Keywords:** maternal nutrition; assisted reproductive techniques; developmental programming; epigenetics; early pregnancy; reproductive function

# 1. Introduction—Developmental Programming

An important concept in developmental biology and medicine is that of "developmental programming," often termed developmental origins of health and disease (DOHaD), which is the concept that "insults" during embryonic, fetal, and postnatal life can have long-lasting effects on the offspring. These effects result in a much greater risk of acquiring various so-called "non-communicable diseases," or NCDs, as infants and adults. Such NCDs include abnormal growth and body composition, as well as behavioral, cardiovascular, endocrine, gastro-intestinal, immune system, metabolic, and reproductive dysfunction, the latter of which is a major socioeconomic issue because of its role in infertility [1–6] The initial observations that led to the concept of developmental programming were based on epidemiological studies in humans. These studies suggested not only that an adverse intrauterine environment may lead to a greater incidence of NCDs in the offspring as adults, but also that poor maternal nutrition was " ... an obvious suspect [7,8]." The initial observations also noted that developmental programming was associated with "Discordance between placental and birth weights ... "

Both assisted reproductive techniques (ART; e.g., embryo transfer [ET], in vitro fertilization [IVF], in vitro production of embryos [IVP], and cloning), and maternal nutritional



Citation: Dahlen, C.R.; Borowicz, P.P.; Ward, A.K.; Caton, J.S.; Czernik, M.; Palazzese, L.; Loi, P.; Reynolds, L.P. Programming of Embryonic Development. *Int. J. Mol. Sci.* 2021, 22, 11668. https://doi.org/ 10.3390/ijms222111668

Academic Editor: Pilar Coy

Received: 1 September 2021 Accepted: 24 September 2021 Published: 28 October 2021

**Publisher's Note:** MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



**Copyright:** © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). status (e.g., over- or underfeeding or feeding specific nutrients) have profound effects on embryonic development during the periconceptual period (i.e., around the time of fertilization/conception) and continuing into early pregnancy, which is the period of embryonic/fetal organogenesis and placentation [1,5,6,9–11]. As discussed in several recent reviews [12–14], paternal nutritional status, via effects on sperm gene expression and epigenetic landscape, extracellular vesicles, and seminal plasma content, has been shown to contribute to embryo quality and affect embryonic/fetal and placental development in cattle [15,16], rodents [17–20], and sheep [21,22] models.

Importantly for the topic of this review, functional defects due to maternal underor overnutrition or ART that have been documented during the periconceptual period include reduced embryonic development to blastocyst stage after IVF and embryo culture [23], reduced embryo–fetal growth [23,24], and altered nutrient supply to the developing fetus [25–29]. Assisted reproductive techniques also altered placental angiogenesis [9,30–32], global DNA methylation [24], and expression of steroid receptor mRNA and proteins [11,33–35]. Additionally, assisted reproductive techniques and moderate maternal nutrient restriction altered expression of gap junctions and chemokines and chemokine receptors in placenta [36,37]; altered expression of endogenous retroviruses, interferon-tau, and pregnancy-specific protein-B, as well as nutrient transporters, in placenta [25,27,38,39]; and altered gene expression during early development of fetal organs [40].

As recently reviewed [1,6,41–43], the effects of ART and nutritional status of the parents are probably mediated via "programming" of embryonic/fetal and placental gene expression, as reflected by changes in their epigenetic landscape [17,44]. Such epigenetic changes may underlie developmental programming of growth, development, and function of organs of the fetus later in pregnancy and the offspring postnatally. As discussed in the opening paragraph, they also potentially lead to long-term changes in organ structure and function in the offspring during infancy and continuing into adulthood, leading to behavioral, cardiovascular, immune system, metabolic, and reproductive dysfunction.

Some of the programming effects of ART and periconceptual parental nutrition likely depend on the epigenetic profile the oocyte or the sperm. The oocyte [10] and sperm [14,15,17,22,45–51] both contribute mRNAs, small non-coding RNAs, and proteins to the embryo, and these have profound influences on embryonic development before activation of the embryonic genome [10,52–54]. In addition, both the maternal (oocyte) and paternal (sperm) genomes can "transfer" their DNA methylation status to the embryo, and this is influenced by ART and nutritional status [10,15].

In this review, we will discuss the evidence that ART and periconceptual maternal and paternal nutritional status may lead to epigenetic alterations very early in pregnancy, ultimately determining events that lead to developmental programing, both during fetal life and postnatally. We also will discuss the effects of ART and maternal nutrition on reproductive function in the male offspring that may lead to transgenerational programming of development. Lastly, we will discuss current research strategies designed to overcome or minimize the negative consequences or, conversely, to maximize the positive aspects of developmental programming.

## 2. Evidence for Programming by Assisted Reproductive Techniques

Comparison of placental development in natural pregnancies and pregnancies achieved by various ART, such as after transfer of embryos created through cloning or IVF, has demonstrated numerous significant effects of ART on placental and fetal growth and development, as well as offspring outcomes in several species [9,11,34]. In various animals, including mice, cattle, and sheep, impaired placental steroid metabolism, abnormal offspring syndrome, increased duration of gestation, and altered placental vascular development, have been reported with the use of ART [9,11,23,24,32–35]. In addition, in sheep, use of ART significantly decreased fetal size at the third week of pregnancy and this was accompanied by decreased placental vascularity and cell proliferation, and altered expression of genes involved in epigenetic processes (e.g., 5-methyl cytosine and DNA methyl transferases), as well as factors involved in regulation of placental growth (e.g., steroid hormone receptors) and angiogenesis (e.g., several angiogenic and growth factors [24,32,33,35,55]). For early pregnancy in cows, both greater and lesser crown-rump length have been reported for fetuses created in vitro and then transferred compared to fetuses created in vivo [56,57].

In sheep, we, and others, have recently demonstrated that utero–placental vascularization, expression of several angiogenic factors, receptors for progesterone and estrogen, and DNA methyl transferases, as well as global DNA methylation and markers of growth all were altered in the developing placenta during early pregnancy after transfer of embryos obtained through IVF or in vitro activation (IVA; parthenotes, which have only a maternal genome [9,24,32–35,58]). One of the very interesting observations from this work is that even when they are derived from naturally mated ewes and transferred to recipients, the embryos show many of the same defects as IVP embryos [9,24,32,35], implying that improvements in embryo collection, culturing, and transfer methods are sorely needed.

These observations also suggest that changes in DNA methylation or histone modifications observed in early embryos of several species (cattle, mice, sheep) after ART [59–63] continue during the critical period of placentation in early pregnancy. In addition, based on observations later in pregnancy, we have suggested that these placental developmental defects probably contribute to poor placental vascularization and function later in pregnancy, ultimately leading to altered fetal growth and development and the poor pregnancy outcomes associated with ART [9,11,34].

Such epigenetic defects leading to altered placental development and gene transcription have been observed after IVF, but especially in clones, not only in sheep but also in cattle and mice [64]. These defects include altered transcription of imprinted genes, which are thought to be critical for normal placental development [65,66].

## 3. Evidence for Programming by Maternal and Paternal Nutrition

In adult women, extremes of pre-pregnancy body mass index (BMI), or, more commonly, BMI at the first prenatal exam, and both inadequate and excessive gestational weight gains are widely associated with a variety of negative obstetrical outcomes [42,67–73]. While few of these clinical studies have been able to cleanly study the separate and joint effects of initial BMI and gestational intake on pregnancy outcome, the broad consensus is that a very high pre-pregnancy BMI and/or high gestational weight gains are associated with an increased risk of preeclampsia, gestational diabetes, preterm and caesarian delivery, stillbirth, fetal macrosomia, and, in some cases, fetal growth restriction. In contrast, a low pre-pregnancy BMI and/or low gestational weight gains are commonly associated with an increased risk of preterm delivery and fetal growth restriction.

Despite these observations in adults, there is a paucity of similar data evaluating the effects of BMI and nutritional extremes on pregnancy outcome in pregnant adolescent girls. Nevertheless, there is some consensus that dietary intakes of pregnant adolescents are often lower than recommended in terms of energy and a range of micronutrients [71,74]. In addition, there also is evidence that pregnant adolescents, particularly those less than 15 years of age, are at risk of excessive gestational weight gain [75], similar in magnitude to the weight change measured in the overnourished and rapidly growing adolescent dams in one of our nutritional models of the pregnant adolescent ewe lamb [71]. Moreover, fetal growth restriction has been reported in mothers who are still growing or in mothers who have not achieved their adult height at the time of conception [76–78].

Maternal nutritional status during the periconceptual period and early pregnancy has profound effects on embryonic/fetal and placental development. For example, maternal under- or overnutrition during the 6 weeks preceding oocyte collection dramatically affect the rates of IVF and subsequent development of embryos to blastocyst stage in vitro [23,79]. In addition, underfed and overfed sheep had altered serum insulin, insulin-like growth factor-1, leptin, progesterone, and (or) estradiol concentrations, indicating they were affected metabolically as well [23,80]. Other studies reported decreased rates of fertilization

with IVF and (or) decreased rates of early embryonic development in ewes or cows fed a low- (nutrient-restricted) or high- (ad libitum intake) energy diet during the periconceptual period [81–85]. Conversely, others have shown that an altered plane of nutrition during the periconceptual period had either positive or no effects on blastocyst formation and/or embryo health in sheep [83,86]. Interestingly, premating nutrition altered mRNA expression in oocytes and follicular cells, which may account for reductions in reproductive performance of ewes fed restricted diets [87]. Maternal undernutrition and folate supplementation during the periconceptual period also altered the methylome of sperm in the offspring [22]. In addition, maternal undernutrition reduced the proportion of embryos reaching the blastocyst stage after IVF, but folate supplementation reversed these effects (Table 1).

**Table 1.** Outcomes of in vitro fertilization (IVF) using sperm from lambs born to Control, Undernourished or Folic acid-Supplemented ewes.

Nutritional Treatment	Oocytes (n)	2-Cell Stage Embryos (%)	Blastocysts (%)
Control	178	56/178 (31.5)	26/178 (14.6)
Undernourished	186	44/186 (23.6) <sup>b</sup>	10/186 (5.4) <sup>a,b</sup>
Folic acid-supplemented	118	46/118 (39)	16/118 (13.5)

<sup>a</sup> p < 0.005 between Undernourished and Control; <sup>b</sup> p < 0.05 between Undernourished and Folic acidsupplemented; Fisher's exact test. Table from [22].

In cows, the effects of diet before and during the periconceptual period on oocyte quality measured by rates of fertilization with IVF and blastocyst formation, and cell number per blastocyst, depend on body condition (i.e., adiposity) and diet composition [88,89]. High levels of food intake improved postfertilization embryonic development for animals in low body condition, but reduced postfertilization embryonic development for animals in good body condition [87]. In addition, hyperinsulinemic cows with good body condition that were fed a high-energy diet produced fewer oocytes and had lower blastocyst yield after IVF [89]. Moreover, dairy cows that were fed a high-protein and high-energy diet exhibited altered endocrine and metabolic function and were frequently in negative energy balance, which is associated with low fertility, perhaps due to low-quality oocytes and embryos [90–92]. It also has been suggested that genetic factors also may contribute to diminished oocyte and embryo quality [91,92].

Maternal undernutrition during the periconceptual period in sheep results in altered fetal growth trajectory, altered fetal hypothalamic–pituitary–adrenal axis development, accelerated maturation of fetal adrenal glands, altered fetal pancreatic development, altered insulin signaling, and altered amino acid metabolism, all associated with an increased rate of premature birth and high rates of postnatal mortality [93–96]. How maternal underfeeding during the periconceptual period affects offspring growth and development is unknown. Overall, the available data indicate that the maternal diet during the periconceptual period alters the epigenetic status of the oocyte or early embryo [97], leading to short-term (e.g., effects on oocyte quality and thus the rates of fertilization and early embryonic development) and/or long-term (e.g., effects on offspring development and timing of birth, and thus developmental programming) effects. Recently, Sinclair and Singh [97] suggested that specific maternal dietary components such as vitamin B12 and methionine fed to ewes during the periconceptual period affect DNA methylation, insulin resistance, and blood pressure in the offspring.

Data from human and rodent models clearly show that developmental programming effects arise from paternal contributions. Authors reviewing 45 publications reporting data from rodent models presented overwhelming evidence that developmental programming extends to the F2 generation and beyond, with several of the papers reviewed presenting evidence of a paternal contribution to developmental programming [98]. Paternal diet has been shown to affect growth, metabolism, and reproductive traits in the offspring of rodents and sheep, potentially across multiple generations [12,14,17–21]. Messages carried in a father's semen after exposure to alcohol, drugs, or an obese state result in altered

development of their children [99,100]. The impacts of sire obesity can be long lasting, with effects on post-pubertal semen development in male offspring and the negative effects further exacerbated when offspring also received a high-fat diet [101].

At least some of the programming effects of paternal nutrition may involve epigenetic alterations in the father's sperm [15,16,21,102]. Small non-coding RNAs are implicated in paternal programming by targeting maternal RNAs, or by entering the nucleus to directly alter transcriptional programming of the embryo [103]. Injecting purified sperm RNAs from male mice exposed to high-fat diets into normal zygotes resulted in offspring with impaired glucose tolerance and elevated blood glucose and insulin levels [50]. Injection of sperm microRNAs into wildtype oocytes resulted in metabolic and behavioral changes in the offspring [104]. When microRNA from stressed males was injected into fertilized oocytes of control females, emotional response and microRNA in the hippocampus (the emotional control center of the brain) were altered in the offspring [105].

In rodent models, fat and protein content of paternal diets have altered gene expression and function of key metabolic tissues in the offspring (liver and muscle; [12,20]) and feeding elevated concentrations of methyl donors (Choline, Betaine, Vit B6, Folate, and Vit B12) to boars resulted in offspring with differential methylation patterns in liver and muscle and altered body composition (reduced backfat and greater percentage of shoulder) [106]. Targeted feeding of rumen-protected methionine to rams resulted in changes in sperm methylation and was associated with altered weight at puberty and scrotal circumference of the offspring [21]. Diet of bulls during the early postpubertal period (1 year of age) affected sperm characteristics and concentrations of minerals and metabolites in seminal plasma [107,108], and a model of divergent nutrition in mature bulls providing either global nutrient restriction or excess resulted in differential expression of 769 transcripts in their sperm, including genes related to de novo methylation and histone to protamine transition [109].

## 4. Strategies to Improve Embryo Development and Pregnancy Outcomes

#### 4.1. Reproductive Fluids

Coy and colleagues have shown that oviductal fluid, as well as specific proteins secreted by the oviduct, improve the rates of fertilization and embryonic development in vitro and in vivo, resulting in greater pregnancy rates and improved pregnancy outcomes with ART [110–113]. This technology holds great potential to overcome poor fertility and also to maximize the impact of elite genetic stock, and thus to dramatically improve the efficiency of livestock production. Whether maternal diet affects oviductal gene or protein expression, and whether such effects influence the ability of oviductal secretions to improve fertilization, embryonic development, or pregnancy outcomes has received little attention. However, we recently showed that restricted dietary intake does not affect protein expression of a major protein secreted by the oviduct in beef cows, namely oviductal glycoprotein 1 (OVGP1; [114]).

### 4.2. Supplementation

Targeted dietary supplementation of nutritionally compromised parents also is an important strategy being pursued by several groups of investigators. For example, Bazer et al. [115] have suggested that dietary supplementation of specific dietary amino acids that are present in uterine secretions (histotroph) could improve embryonic development. Such amino acids include serine and methionine, both of which are involved in one-carbon metabolism, as well as arginine, lysine, and histidine, which are targets of methylation when incorporated into histones. Arginine also improves feto-placental development in rodents [116–118], sheep [119–123], swine [124–127], and humans [128–130], potentially due to its involvement in generation of nitric oxide and polyamines [115,131].

The placenta transports amino acids from the maternal circulation and histotroph across the trophoblast/chorion via amino acid transporters ([115]. The cationic amino acid transporters (SLC7A1, A2, and A3) are expressed in uterine luminal and glandular

epithelia and also in trophoblasts, and expression of two of these is stimulated by treatment of ewes with progesterone (SLC7A1 and A2) or, in the case of SLC7A2, by administration of interferon-tau (a conceptus product [132]). Expression of SLC1A5, a neutral amino acid transporter, in uterine luminal and glandular epithelia also is stimulated by both progesterone and interferon-tau treatment [133]. Progesterone treatment also increases the concentrations of fructose in uterine luminal fluid (histotroph; [134]).

Recently, our research group has shown not only that hexose and amino acid transporters are expressed in bovine utero-placental tissues, but also that expression of several of these transporters is influenced by maternal dietary restriction during early pregnancy (days 16 to 50 after mating [25,27,135–137]). In addition, maternal dietary intake affects the concentrations of hexoses (glucose and fructose) and several amino acids in allantoic and amniotic fluids, as well as homocysteine in maternal blood (Figure 1 [1]), all of which imply a profound effect of maternal dietary intake on maternal and embryo/fetal metabolism, including 1-carbon metabolism.

Another promising strategy is dietary supplementation with vitamins and minerals. In a recent series of studies, we have shown that providing a dietary vitamin–mineral supplements to beef heifers before mating and continuing for the first trimester of pregnancy affected maternal hormonal and mineral status, fetal organ weight, concentrations of amino acids in fetal fluids, abundance of fat soluble vitamins in fetal liver, placental nutrient transporter expression, and placental gene expression [28,29,135–141].



**Figure 1.** Comparison of (**A**) glucose concentrations in allantoic and amniotic fluid, (**B**) glutamine concentrations in allantoic and amniotic fluid, (**C**) methionine concentrations in allantoic fluid, and (**D**) homocysteine concentrations in maternal serum of heifers receiving control or restricted dietary treatment from the day of mating (d 0) until d 50 of gestation. Treatments provided for 0.5 kg of gain/hd daily vs. -0.08 kg of gain/hd daily between d 0 and d 50 of gestation for control vs. restricted heifers, respectively. Figure from Caton et al. [1]. a,b and y,z for (**A**,**B**) bars significantly different, with *p*-value as indicated; a,b for (**C**,**D**), bars differ significantly (*p* < 0.10).

Targeted nutritional supplementation of one-carbon micronutrients to either the parents or cultured embryos is another strategy to "rescue" compromised embryos. This approach is based on in vivo and in vitro studies and the fact that the early embryo undergoes dramatic changes in methylation of the majority of its genes [17,142–149]. For example, in zebrafish, a parental diet deficient in one-carbon micronutrients altered oocyte production and one-carbon metabolism in the parents, as well as embryonic gene expression of pathways involved in the one-carbon cycle and lipid transport [150]. In dairy cows, maternal methionine supplementation caused significant changes in the transcriptome of the preimplantation embryos [151]. Recently, we have shown that supplementation of one-carbon metabolites during culture of bovine embryonic tracheal fibroblasts improves their mitochondrial respiration and growth rates and alters their DNA methylation status [26].

However, dietary supplementation of 1-carbon metabolites may also have detrimental effects [17,19]. For example, male mice fed folate-deficient diets had altered DNA methylation in their sperm; in addition, their offspring exhibited differential gene expression in their placentas as well as increased incidence of birth defects, including craniofacial and musculoskeletal malformations [17].

### 5. Conclusions and Future Directions

It seems clear that both ART and parental nutritional status, and probably a host of other "stressors" as well (e.g., parental age; environmental conditions, including temperature/humidity, elevation, and contaminants; relational or economic stress, etc.) can affect developmental competence of gametes and embryos [3–6,9,11,42,43,152]. It also seems clear that the periconceptual period is particularly important in terms of programming of embryonic and feto-placental development [1,85,93,95,153]. Thus, research on factors that affect gametes, including those during IVP of embryos, such as reproductive fluids or culture media supplements, should receive much more emphasis in the future. In addition, research to improve embryo culture, storage, and transfer methods should also be an area of emphasis.

Recent models by our group have gone beyond the extremes of dietary restriction and excess, and focused on a range of experimental conditions that are normally found in production scenarios. Differences in offspring outcomes, nutrients found in fetal tissues and fluids, and gene expression in placenta and fetal organs were observed in these models of moderate dietary alterations in maternal nutrient supply [28,29,137–141], which indicates to us that developmental programming may be present in a much larger proportion of pregnancies than given credence to in the literature.

We, therefore, propose that there is a continuum of developmental programming and that each pregnancy is uniquely programmed; with alterations in epigenetic signals generated in response to dietary environmental factors, stress, and other signals experienced by the dam. Related to this, an area that should receive much more attention in the future is management and therapeutic strategies to take advantage of the positive effects of developmental programming. This idea is based on the suggestion that developmental programming ultimately is adaptive and should improve the fitness of the individual [154,155]. In keeping with this focus, our studies of maternal nutritional status during the first 50 days of pregnancy in cattle have shown that the majority of genes in the fetal liver, muscles, and the brain were upregulated in fetuses from nutrient-restricted dams. We, therefore, suggested that upregulation of genes may represent an adaptive, rather than a negative, response to maternal nutrient restriction [40]. Consistent with this observation, cattle and other ruminants (deer, goats, sheep, etc.) that graze rangelands or other "extensive," pasture-based systems are often nutrient restricted during early pregnancy [156–159]. Thus, they are well adapted to yearly cycles of low forage quality and availability and, therefore, to severe, short-term fluctuations in body weight and condition.

This idea of adaptive versus maladaptive responses to maternal malnutrition also has relevance to humans, as low caloric intake and malnutrition, usually associated with poverty, are experienced by many pregnant women worldwide, with severe consequences for pregnancy outcomes. Such consequences include low and very low birth weights, high rates of postnatal morbidity and mortality, and long-term consequences for health and productivity of the offspring, including stunting and wasting [67,160–164]. Thus, a much better understanding is needed of whether changes in expression of fetal and placental

genes are adaptive, and therefore beneficial, or maladaptive, and therefore detrimental, to the offspring in both the short and long term.

Lastly, research on potential solutions to the negative consequences associated with programming of embryonic development has shown great promise. For example, the recent studies in which media were supplemented with oviductal or uterine luminal fluids have shown improvements in survival and development of embryos produced in vitro. Similarly, providing targeted dietary supplements to nutritionally compromised parents has shown improved developmental outcomes of offspring, including their reproductive function, thereby impacting subsequent generations. More work should be done, however, on supplementation of embryo culture media with various nutrients and hormones, and supplementing nutrients that provide substrates for key epigenetic processes. Due to their potential long-term benefits across generations and across species, we believe these efforts should continue and even be expanded.

**Funding:** The research from NDSU that provided the basis for much of this review was funded by grants from the National Institutes of Health (grants HD22559, HL64141, HD45784, HD61532, and HD76073) and the National Institute of Food and Agriculture (grant 2007-01215).

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Conflicts of Interest: The authors declare no conflict of interest.

## References

- Caton, J.S.; Crouse, M.S.; McLean, K.J.; Dahlen, C.R.; Ward, A.K.; A Cushman, R.; Grazul-Bilska, A.T.; Neville, B.W.; Borowicz, P.P.; Reynolds, L.P. Maternal periconceptual nutrition, early pregnancy, and developmental outcomes in beef cattle. *J. Anim. Sci.* 2020, *98*. [CrossRef]
- Reynolds, L.P.; Caton, J.S. Role of the pre- and post-natal environment in developmental programming of health and productivity. *Mol. Cell. Endocrinol.* 2012, 354, 54–59. [CrossRef]
- Reynolds, L.P.; Vonnahme, K.A. Triennial Reproduction Symposium: Developmental programming of fertility1. J. Anim. Sci. 2016, 94, 2699–2704. [CrossRef]
- 4. Reynolds, L.P.; Vonnahme, K.A. Livestock as models for developmental programming. Anim. Front. 2017, 7, 12–17. [CrossRef]
- Reynolds, L.P.; Borowicz, P.P.; Caton, J.S.; Vonnahme, K.A.; Luther, J.S.; Hammer, C.J.; Maddock Carlin, K.R.; Grazul-Bilska, A.T.; Redmer, D.A. Developmental programming: The concept, large animal models, and the key role of utero-placental vascular development. Invited review. J. Anim. Sci. 2010, 88, E61–E72. [CrossRef]
- 6. Reynolds, L.P.; Borowicz, P.P.; Caton, J.S.; Crouse, M.; Dahlen, C.R.; Ward, A.K. Developmental Programming of Fetal Growth and Development. *Invited Review. Veter. Clin. North Am. Food Anim. Pr.* **2019**, *35*, 229–247. [CrossRef]
- 7. Barker, D.J.P. Fetal and Infant Origins of Adult Disease; BMJ Publishing Group: London, UK, 1992.
- 8. Barker, D.J.P. Developmental origins of well being. *Philos. Trans. R. Soc. Lond.* 2004, 359, 1359–1366. [CrossRef]
- Bairagi, S.; Quinn, K.E.; Crane, A.R.; Ashley, R.L.; Borowicz, P.P.; Caton, J.S.; Redden, R.R.; Grazul-Bilska, A.T.; Reynolds, L.P. Maternal environment and placental vascularization in small ruminants. *Theriogenology* 2016, *86*, 288–305. [CrossRef]
- Fleming, T.P.; Watkins, A.J.; Velazquez, M.A.; Mathers, J.C.; Prentice, A.M.; Stephenson, J.; Barker, M.; Saffery, R.; Yajnik, C.S.; Eckert, J.J.; et al. Origins of lifetime health around the time of conception: Causes and consequences. *Lancet* 2018, 391, 1842–1852. [CrossRef]
- 11. Reynolds, L.P.; Borowicz, P.P.; Palmieri, C.; Grazul-Bilska, A.T. Placental Vascular Defects in Compromised Pregnancies: Effects of Assisted Reproductive Technologies and Other Maternal Stressors. *Adv. Exp. Med. Biol.* **2014**, *814*, 193–204. [CrossRef]
- 12. Li, J.; Tsuprykov, O.; Yang, X.; Hocher, B. Paternal programming of offspring cardiometabolic diseases in later life. *J. Hypertens.* **2016**, *34*, 2111–2126. [CrossRef]
- 13. Morgan, C.P.; Chan, N.; Bale, T.L. Driving the Next Generation: Paternal Lifetime Experiences Transmitted via Extracellular Vesicles and Their Small RNA Cargo. *Biol. Psychiatry* **2019**, *85*, 164–171. [CrossRef]
- 14. Morgan, H.L.; Eid, N.; Khoshkerdar, A.; Watkins, A.J. Defining the male contribution to embryo quality and offspring health in assisted reproduction in farm animals. *Anim. Reprod.* **2020**, *17*, e20200018. [CrossRef] [PubMed]
- 15. Gross, N.; Strillacci, M.G.; Peñagaricano, F.; Khatib, H. Characterization and functional roles of paternal RNAs in 2–4 cell bovine embryos. *Sci. Rep.* **2019**, *9*, 1–9. [CrossRef] [PubMed]
- Kropp, J.; Carrillo, J.A.; Namous, H.; Daniels, A.; Salih, S.M.; Song, J.; Khatib, H. Male fertility status is associated with DNA methylation signatures in sperm and transcriptomic profiles of bovine preimplantation embryos. *BMC Genom.* 2017, 18, 1–15. [CrossRef]

- 17. Lambrot, R.; Xu, C.; Saintphar, S.; Chountalos, G.; Cohen, T.V.; Paquet, M.; Suderman, M.; Hallett, M.; Kimmins, S. Low paternal dietary folate alters the mouse sperm epigenome and is associated with negative pregnancy outcomes. *Nat. Commun.* **2013**, *4*, 2889. [CrossRef]
- 18. Ng, S.F.; Lin, R.C.; Laybutt, D.R.; Barres, R.; Owens, J.A.; Morris, M.J. Chronic high-fat diet in fathers programs β-cell dysfunction in female rat offspring. *Nature* **2010**, *467*, 963–966. [CrossRef]
- Ryan, D.P.; Henzel, K.S.; Pearson, B.L.; Siwek, M.E.; Papazoglou, A.; Guo, L.; Paesler, K.; Yu, M.; Müller, R.; Xie, K.; et al. A paternal methyl donor-rich diet altered cognitive and neural functions in offspring mice. *Mol. Psychiatry* 2018, 23, 1345–1355. [CrossRef]
- Watkins, A.J.; Sirovica, S.; Stokes, B.; Isaacs, M.; Addison, O.; Martin, R.A. Paternal low protein diet programs preimplantation embryo gene expression, fetal growth and skeletal development in mice. *Biochim. Biophys. Acta (BBA) Mol. Basis Dis.* 2017, 1863, 1371–1381. [CrossRef] [PubMed]
- 21. Gross, N.; Taylor, T.; Crenshaw, T.; Khatib, H. The intergenerational impacts of paternal diet on DNA methylation and offspring phenotypes in sheep. *Front. Genet.* 2020, *11*, 597943. [CrossRef]
- Toschi, P.; Capra, E.; Anzalone, D.A.; Lazzari, B.; Turri, F.; Pizzi, F.; Scapolo, P.A.; Stella, A.; Williams, J.L.; Ajmone Marsan, P.; et al. Maternal peri-conceptional undernourishment perturbs offspring sperm methylome. *Reproduction* 2020, 159, 513–523. [CrossRef] [PubMed]
- Grazul-Bilska, A.T.; Borowczyk, E.; Bilski, J.J.; Reynolds, L.P.; Redmer, D.A.; Caton, J.S.; Vonnahme, K.A. Overfeeding and underfeeding have detrimental effects on oocyte quality measured by in vitro fertilization and early embryonic development in sheep. *Domest. Anim. Endocrinol.* 2012, 43, 289–298. [CrossRef]
- Grazul-Bilska, A.T.; Johnson, M.L.; Borowicz, P.P.; Baranko, L.; Redmer, D.A.; Reynolds, L.P. Placental development during early pregnancy in sheep: Effects of embryo origin on fetal and placental growth and global methylation. *Theriogenology* 2013, 79, 94–102. [CrossRef] [PubMed]
- Crouse, M.S.; McLean, K.J.; Greseth, N.P.; Crosswhite, M.R.; Pereira, N.N.; Ward, A.K.; Reynolds, L.P.; Dahlen, C.R.; Neville, B.W.; Borowicz, P.P.; et al. Maternal nutrition and stage of early pregnancy in beef heifers: Impacts on expression of glucose, fructose, and cationic amino acid transporters in utero-placental tissues. J. Anim. Sci. 2017, 95, 5563–5572. [CrossRef]
- Crouse, M.S.; Caton, J.S.; Menezes, A.C.B.; Reynolds, L.P.; Dahlen, C.R.; Borowicz, P.P.; Ward, A.K. One-carbon metabolite supplementation improves mitochondrial respiration, growth rates, and alters DNA methylation of bovine embryonic fibroblast cells cultured in divergent energy supply. *Transl. Anim. Sci.* 2019, *3*, 1705–1709. [CrossRef] [PubMed]
- Greseth, N.P.; Crouse, M.S.; McLean, K.J.; Crosswhite, M.R.; Pereira, N.N.; Dahlen, C.R.; Borowicz, P.P.; Reynolds, L.P.; Ward, A.K.; Neville, B.W.; et al. The effects of maternal nutrition on the messenger ribonucleic acid expression of neutral and acidic amino acid transporters in bovine uteroplacental tissues from day sixteen to fifty of gestation1. *J. Anim. Sci.* 2017, 95, 4668–4676. [CrossRef] [PubMed]
- Menezes, A.C.B.; McCarthy, K.L.; Kassetas, C.J.; Baumgaertner, F.; Kirsch, J.D.; Dorsam, S.; Neville, T.L.; Ward, A.K.; Borowicz, P.P.; Reynolds, L.P.; et al. Vitamin and mineral supplementation and rate of gain during the first trimester of gestation affect concentrations of amino acids in maternal serum and allantoic fluid of beef heifers. J. Anim. Sci. 2021, 99, skab024. [CrossRef]
- Menezes, A.C.B.; McCarthy, K.L.; Kassetas, C.J.; Baumgaertner, F.; Kirsch, J.D.; Dorsam, S.; Neville, T.L.; Ward, A.K.; Borowicz, P.P.; Reynolds, L.P.; et al. Vitamin and mineral supplementation and rate of gain in beef heifers: Effects on fetal trace mineral reserves at day 83 of gestation. J. Anim. Sci. 2021, in press. [CrossRef]
- Grazul-Bilska, A.T.; Borowicz, P.P.; Johnson, M.L.; A Minten, M.; Bilski, J.J.; Wroblewski, R.; Redmer, D.A.; Reynolds, L.P. Placental development during early pregnancy in sheep: Vascular growth and expression of angiogenic factors in maternal placenta. *Reproduction* 2010, 140, 165–174. [CrossRef] [PubMed]
- Grazul-Bilska, A.T.; Johnson, M.L.; Borowicz, P.P.; Minten, M.; Bilski, J.J.; Wroblewski, R.; Velimirovich, M.; Coupe, L.R.; Redmer, D.A.; Reynolds, L.P. Placental development during early pregnancy in sheep: Cell proliferation, global methylation, and angiogenesis in the fetal placenta. *Reproduction* 2011, 141, 529–540. [CrossRef]
- 32. Grazul-Bilska, A.T.; Johnson, M.L.; Borowicz, P.P.; Bilski, J.J.; Cymbaluk, T.; Norberg, S.; Redmer, D.A.; Reynolds, L.P. Placental development during early pregnancy in sheep: Effects of embryo origin on vascularization. *Reproduction* **2014**, 147, 639–648. [CrossRef]
- Grazul-Bilska, A.T.; Bairagi, S.; Kraisoon, A.; Dorsam, S.T.; Reyaz, A.; Navanukraw, C.; Borowicz, P.P.; Reynolds, L.P. Placental development during early pregnancy in sheep: Nuclear estrogen and progesterone receptor mRNA expression in the uteroplacental compartments. *Domest. Anim. Endocrinol.* 2019, *66*, 27–34. [CrossRef] [PubMed]
- 34. Reynolds, L.P.; Vonnahme, K.A.; Lemley, C.O.; Redmer, D.A.; Grazul-Bilska, A.T.; Borowicz, P.P.; Caton, J.S. Maternal Stress and Placental Vascular Function and Remodeling. *Curr. Vasc. Pharmacol.* **2013**, *11*, 564–593. [CrossRef] [PubMed]
- Reynolds, L.P.; Haring, J.S.; Johnson, M.L.; Ashley, R.L.; Redmer, D.A.; Borowicz, P.P.; Grazul-Bilska, A. Placental development during early pregnancy in sheep: Estrogen and progesterone receptor messenger RNA expression in pregnancies derived from in vivo–produced and in vitro–produced embryos. *Domest. Anim. Endocrinol.* 2015, 53, 60–69. [CrossRef]
- Johnson, M.L.; Redmer, D.A.; Reynolds, L.P.; Grazul-Bilska, A.T. Gap junctional connexin messenger RNA expression in the ovine uterus and placenta: Effects of estradiol-17β-treatment, early pregnancy stages, and embryo origin. *Domest. Anim. Endocrinol.* 2017, 58, 104–112. [CrossRef]

- 37. Quinn, K.E.; Reynolds, L.P.; Grazul-Bilska, A.T.; Borowicz, P.P.; Ashley, R.L. Placental development during early pregnancy: Effects of embryo origin on expression of chemokine ligand twelve (CXCL12). *Placenta* **2016**, *43*, 77–80. [CrossRef]
- McLean, K.J.; Crouse, M.; Crosswhite, M.R.; Pereira, N.N.; Dahlen, C.R.; Borowicz, P.P.; Reynolds, L.P.; Ward, A.K.; Neville, B.W.; Caton, J.S. Impacts of maternal nutrition on uterine and placental vascularity and mRNA expression of angiogenic factors during the establishment of pregnancy in beef heifers. *Transl. Anim. Sci.* 2017, *1*, 160–167. [CrossRef]
- McLean, K.J.; Crouse, M.S.; Crosswhite, M.R.; Pereira, N.N.; Dahlen, C.R.; Borowicz, P.P.; Reynolds, L.P.; Ward, A.K.; Neville, B.W.; Caton, J.S. The effects of nutrient restriction on mRNA expression of endogenous retroviruses, interferon-tau, and pregnancyspecific protein-B during the establishment of pregnancy in beef heifers. J. Anim. Sci. 2018, 96, 950–963. [CrossRef]
- 40. Crouse, M.S.; Caton, J.S.; A Cushman, R.A.; McLean, K.J.; Dahlen, C.R.; Borowicz, P.P.; Reynolds, L.P.; Ward, A.K. Moderate nutrient restriction of beef heifers alters expression of genes associated with tissue metabolism, accretion, and function in fetal liver, muscle, and cerebrum by day 50 of gestation. *Transl. Anim. Sci.* **2019**, *3*, 855–866. [CrossRef]
- 41. Goyal, D.; Limesand, S.W.; Goyal, R. Epigenetic responses and the developmental origins of health and disease. *J. Endocrinol.* **2019**, 242, T105–T119. [CrossRef] [PubMed]
- 42. Rajasingam, D.; Seed, P.T.; Briley, A.L.; Shennan, A.H.; Poston, L. A prospective study of pregnancy outcome and biomarkers of oxidative stress in nulliparous obese women. *Am. J. Obstet. Gynecol.* 2009, 200, 395.e1–. [CrossRef] [PubMed]
- Reynolds, L.P.; Ward, A.K.; Caton, J.S. Epigenetics and developmental programming in ruminants—Long-term impacts on growth and development. In *Biology of Domestic Animals*; Scanes, C.G., Hill, R., Eds.; CRC Press/Taylor & Francis Group: Milton Park, UK, 2017; pp. 85–121. ISBN 9781498747851.
- 44. Anckaert, E.; Fair, T. DNA methylation reprogramming during oogenesis and interference by reproductive technologies: Studies in mouse and bovine models. *Reprod. Fertil. Dev.* **2015**, *27*, 739–754. [CrossRef]
- 45. Avendaño, C.; Franchi, A.; Jones, E.; Oehninger, S. Pregnancy-specific β-1-glycoprotein 1 and human leukocyte antigen-E mRNA in human sperm: Differential expression in fertile and infertile men and evidence of a possible functional role during early development. *Hum. Reprod.* 2009, 24, 270–277. [CrossRef]
- 46. Ostermeier, G.C.; Miller, D.; Huntriss, J.D.; Diamond, M.; Krawetz, S.A. Delivering spermatozoan RNA to the oocyte. *Nature* **2004**, *429*, 154. [CrossRef]
- 47. Krawetz, S.A. Paternal contribution: New insights and future challenges. Nat. Rev. Genet. 2005, 6, 633-642. [CrossRef]
- 48. MacLeod, G.; Varmuza, S. The application of proteomic approaches to the study of mammalian spermatogenesis and sperm function. *FEBS J.* **2013**, *280*, 5635–5651. [CrossRef] [PubMed]
- Fang, P.; Zeng, P.; Wang, Z.; Liu, M.; Xu, W.; Dai, J.; Zhao, X.; Zhang, N.; Liang, D.; Chen, X.; et al. Estimated Diversity of Messenger RNAs in Each Murine Spermatozoa and Their Potential Function During Early Zygotic Development. *Biol. Reprod.* 2014, 90, 1–11. [CrossRef]
- 50. Chen, Q.; Yan, M.; Cao, Z.; Li, X.; Zhang, Y.; Shi, J.; Feng, G.-H.; Peng, H.; Zhang, X.; Qian, J.; et al. Sperm tsRNAs contribute to intergenerational inheritance of an acquired metabolic disorder. *Science* **2016**, *351*, 397–400. [CrossRef]
- 51. Immler, S. The sperm factor: Paternal impact beyond genes. *Heredity* 2018, 121, 239–247. [CrossRef]
- 52. Kempisty, B.; Antosik, P.; Bukowska, D.; Jackowska, M.; Lianeri, M.; Jaśkowski, J.M.; Jagodziński, P.P. Analysis of selected transcript levels in porcine spermatozoa, oocytes, zygotes and two-cell stage embryos. *Reprod. Fertil. Dev.* **2008**, *20*, 513–518. [CrossRef]
- 53. Kawano, M.; Kawaji, H.; Grandjean, V.; Kiani, J.; Rassoulzadegan, M. Novel small noncoding RNAs in mouse spermatozoa, zygotes and early embryos. *PLoS ONE* **2012**, *7*, e44542. [CrossRef]
- 54. Saunders, C.M.; Larman, M.G.; Parrington, J.; Cox, L.J.; Royse, J.; Blayney, L.M.; Swann, K.; Lai, F.A. PLC zeta: A sperm-specific trigger of Ca (2+) oscillations in eggs and embryo development. *Development* 2002, *129*, 3533–3544. [CrossRef] [PubMed]
- Bairagi, S.; Grazul-Bilska, A.T.; Borowicz, P.P.; Reyaz, A.; Valkov, V.; Reynolds, L.P. Placental development during early pregnancy in sheep: Progesterone and estrogen receptor protein expression. *Theriogenology* 2018, *114*, 273–284. [CrossRef]
- 56. Bertolini, M.; Mason, J.B.; Beam, S.W.; Carneiro, G.F.; Sween, M.L.; Kominek, D.J.; Moyer, A.L.; Famula, T.R.; Sainz, R.D.; Anderson, G.B. Morphology and morphometry of in vivo- and in vitro-produced bovine concepti from early pregnancy to term and association with high birth weights. *Theriogenology* 2002, *58*, 973–994. [CrossRef]
- 57. Farin, P.W.; Piedrahita, J.A.; Farin, C.E. Errors in development of fetuses and placentas from in vitro-produced bovine embryos. *Theriogenology* **2006**, *65*, 178–191. [CrossRef]
- Ptak, G.E.; D'Agostino, A.; Toschi, P.; Fidanza, A.; Zacchini, F.; Czernik, M.; Monaco, F.; Loi, P. Post-implantation mortality of in vitro produced embryos is associated with DNA methyltransferase 1 dysfunction in sheep placenta. *Hum. Reprod.* 2012, 28, 298–305. [CrossRef]
- 59. Beaujean, N.; Taylor, J.; Gardner, J.; Wilmut, I.; Meehan, R.; Young, L. Effect of limited DNA methylation reprogramming in the normal sheep embryo on somatic cell nuclear transfer. *Biol. Reprod.* **2004**, *71*, 185–193. [CrossRef]
- 60. Bourc'His, D.; Le Bourhis, D.; Patin, D.; Niveleau, A.; Comizzoli, P.; Renard, J.-P.; Viegas-Péquignot, E. Delayed and incomplete reprogramming of chromosome methylation patterns in bovine cloned embryos. *Curr. Biol.* **2001**, *11*, 1542–1546. [CrossRef]
- 61. De Waal, E.; Yamazaki, Y.; Ingale, P.; Bartolomei, M.S.; Yanagimachi, R.; McCarrey, J.R. Gonadotropin stimulation contributes to an increased incidence of epimutations in ICSI-derived mice. *Hum. Mol. Genet.* **2012**, *21*, 4460–4472. [CrossRef]

- 62. De Waal, E.; Yamazaki, Y.; Ingale, P.; Bartolomei, M.; Yanagimachi, R.; McCarrey, J.R. Primary epimutations introduced during intracytoplasmic sperm injection (ICSI) are corrected by germline-specific epigenetic reprogramming. *Proc. Natl. Acad. Sci. USA* **2012**, *109*, 4163–4168. [CrossRef]
- 63. Suteevun, T.; Parnpai, R.; Smith, S.L.; Chang, C.-C.; Muenthaisong, S.; Tian, X.C. Epigenetic characteristics of cloned and in vitro-fertilized swamp buffalo (*Bubalus bubalis*) embryos. *J. Anim. Sci.* **2006**, *84*, 2065–2071. [CrossRef] [PubMed]
- 64. Palmieri, C.; Loi, P.; Ptak, G.; Della Salda, L. Review paper: A review of the pathology of abnormal placentae of somatic cell nuclear transfer clone pregnancies in cattle, sheep, and mice. *Veter. Pathol.* **2008**, *45*, 865–880. [CrossRef]
- 65. Bressan, F.F.; De Bem, T.H.C.; Perecin, F.; Lopes, F.L.; Ambrosio, C.E.; Meirelles, F.V.; Miglino, M. Unearthing the roles of imprinted genes in the placenta. *Placenta* **2009**, *30*, 823–834. [CrossRef]
- 66. Mesquita, F.S.; Machado, S.A.; Drnevich, J.; Borowicz, P.; Wang, Z.; Nowak, R.A. Influence of cloning by chromatin transfer on placental gene expression at Day 45 of pregnancy in cattle. *Anim. Reprod. Sci.* **2013**, *136*, 231–244. [CrossRef]
- 67. Abenhaim, H.A.; Kinch, R.A.; Morin, L.; Benjamin, A.; Usher, R. Effect of prepregnancy body mass index categories on obstetrical and neonatal outcomes. *Arch. Gynecol. Obstet.* **2007**, 275, 39–43. [CrossRef]
- 68. Bhattacharya, S.; Campbell, D.M.; Liston, W.A.; Bhattacharya, S. Effect of Body Mass Index on pregnancy outcomes in nulliparous women delivering singleton babies. *BMC Public Health* **2007**, *7*, 168. [CrossRef]
- 69. Crane, J.M.G.; White, J.; Murphy, P.; Burrage, L.; Hutchens, D. The effect of gestational weight gain by body mass index on maternal and neonatal outcomes. *J. Obstet. Gynaecol. Can.* **2009**, *31*, 28–35. [CrossRef]
- 70. Ehrenberg, H.M.; Dierker, L.; Milluzzi, C.; Mercer, B.M. Low maternal weight, failure to thrive in pregnancy, and adverse pregnancy outcomes. *Am. J. Obs. Gynecol.* **2003**, *189*, 1726–1730. [CrossRef]
- Luther, J.S.; Redmer, D.A.; Reynolds, L.P.; Wallace, J.M. Nutritional paradigms of ovine fetal growth restriction: Implications for human pregnancy. *Hum. Fertil.* 2005, *8*, 179–187. [CrossRef]
- Nohr, E.A.; Vaeth, M.; Baker, J.L.; Sorensen, T.I.A.; Olsen, J.; Rasmussen, K.M. Combined associations of prepregnancy body mass index and gestation weight gain with the outcome of pregnancy. *Am. J. Clin. Nut.* 2008, *87*, 1750–1759. [CrossRef] [PubMed]
- Rode, L.; Hegaard, H.K.; Kjtaergaard, H.; Moller, L.F.; Tabor, A.; Ottesen, B. Association between maternal weight gain and birth weight. Obs. Gynecol. 2007, 109, 1309–1315. [CrossRef]
- 74. Moran, V.H. A systemic review of dietary assessments of pregnant adolescents in industrialised countries. *Br. J. Nutr.* 2007, *97*, 411–425. [CrossRef]
- 75. Howie, L.J.D.; Parker, J.D.; Schoendorf, M.D. Excessive maternal weight patterns in adolescents. J. Am. Diet. Assoc. 2003, 103, 1653–1657. [CrossRef]
- Frisancho, A.R.; Matos, J.; Leonard, W.R.; Yaroch, L.A. Developmental and nutritional determinants of pregnancy outcome among teenagers. *Am. J. Phys. Anthropol.* 1985, 66, 247–261. [CrossRef] [PubMed]
- Kirchengast, S.; Hartmann, B. Impact of maternal age and maternal somatic characteristics on newborn size. *Am. J. Hum. Biol.* 2003, 15, 220–228. [CrossRef] [PubMed]
- 78. Scholl, T.O.; Hediger, M.L.; Schall, J.I. Maternal growth and foetal growth: Pregnancy course and outcome in the Camden Study. *Ann. N.Y. Acad. Sci.* **1997**, *81*, 292–301. [CrossRef]
- Borowczyk, E.; Caton, J.S.; Redmer, D.A.; Bilski, J.J.; Weigl, R.M.; Vonnahme, K.A.; Borowicz, P.P.; Kirsch, J.D.; Kraft, K.C.; Reynolds, L.P.; et al. Effects of plane of nutrition on in vitro fertilization and early embryonic development in sheep1. *J. Anim. Sci.* 2006, 84, 1593–1599. [CrossRef]
- Kaminski, S.L.; Redmer, D.A.; Bass, C.S.; Keisler, D.H.; Carlson, L.S.; Vonnahme, K.A.; Dorsam, S.T.; Grazul-Bilska, A.T. The effects of diet and arginine treatment on serum metabolites and selected hormones during the estrous cycle in sheep. *Theriogenology* 2014, *83*, 808–816. [CrossRef]
- 81. Abdelatty, A.M.; Iwaniuk, M.E.; Potts, S.B.; Gad, A. Influence of maternal nutrition and heat stress on bovine oocyte and embryo development. *Int. J. Vet. Sci. Med.* 2018, *6*, S1–S5. [CrossRef]
- 82. D'Occhio, M.J.; Baruselli, P.S.; Campanile, G. Influence of nutrition, body condition, and metabolic status on reproduction in female beef cattle: A review. *Theriogenology* **2018**, *125*, 277–284. [CrossRef]
- 83. Lozano, J.M.; Lonergan, P.; Boland, M.P.; O'Callaghan, D. Influence of nutrition on the effectiveness of superovulation programmes in ewes: Effect on oocyte quality and post-fertilization development. *Reproduction* 2003, 125, 543–553. [CrossRef] [PubMed]
- 84. Papadopoulos, S.; Lonergan, P.; Gath, V.; Quinn, K.M.; Evans, A.C.; O'Callaghan, D.; Boland, M.P. Effect of diet quantity and urea supplementation on oocyte and embryo quality in sheep. *Theriogenology* **2001**, *55*, 1059–1069. [CrossRef]
- Van Eetvelde, M.; Heras, S.; Leroy, J.L.M.R.; Van Soom, A.; Opsomer, G. The importance of the periconception period: Immediate effects in cattle breeding and in assisted reproduction such as artificial insemination and embryo transfer. *Adv. Exp. Med. Biol.* 2017, 1014, 41–68. [CrossRef]
- Kakar, M.A.; Maddocks, S.; Lorimer, M.F.; Kleemann, D.O.; Rudiger, S.R.; Hartwich, K.M.; Walker, S. The effect of peri-conception nutrition on embryo quality in the superovulated ewe. *Theriogenology* 2005, 64, 1090–1103. [CrossRef]
- Pisani, L.F.; Antonini, S.; Pocar, P.; Ferrari, S.; Brevini, T.A.; Rhind, S.M.; Gandolfi, F. Effects of pre-mating nutrition on mRNA levels of developmentally relevant genes in sheep oocytes and granulosa cells. *Reproduction* 2008, 136, 303–312. [CrossRef] [PubMed]
- Adamiak, S.J.; Mackie, K.; Watt, R.G.; Webb, R.; Sinclair, K.D. Impact of Nutrition on Oocyte Quality: Cumulative Effects of Body Composition and Diet Leading to Hyperinsulinemia in Cattle. *Biol. Reprod.* 2005, 73, 918–926. [CrossRef] [PubMed]

- 89. Adamiak, S.J.; Powell, K.; A Rooke, J.A.; Webb, R.; Sinclair, K.D. Body composition, dietary carbohydrates and fatty acids determine post-fertilisation development of bovine oocytes in vitro. *Reproduction* **2006**, *131*, 247–258. [CrossRef] [PubMed]
- 90. Butler, W. Energy balance relationships with follicular development, ovulation and fertility in postpartum dairy cows. *Livest. Prod. Sci.* **2003**, *83*, 211–218. [CrossRef]
- 91. Leroy, J.L.; Van Soom, A.; Opsomer, G.; Bols, P.E. The consequences of metabolic changes in high-yielding dairy cows on oocyte and embryo quality. *Animal* **2008**, *2*, 1120–1127. [CrossRef]
- Valckx, S.D.M.; De Pauw, I.; De Neubourg, D.; Inion, I.; Berth, M.; Fransen, E.; Bols, P.E.J.; Leroy, J.L.M.R. BMI-related metabolic composition of the follicular fluid of women undergoing assisted reproductive treatment and the consequences for oocyte and embryo quality. *Hum. Reprod.* 2012, 27, 3531–3539. [CrossRef]
- 93. Bloomfield, F.H.; Oliver, M.H.; Hawkins, P.; Campbell, M.; Phillips, D.J.; Gluckman, P.D.; Challis, J.R.G.; Harding, J.E. A periconceptional nutritional origin for noninfectious preterm birth. *Science* **2003**, *300*, 606. [CrossRef] [PubMed]
- Jaquiery, A.L.; Oliver, M.H.; Rumball, C.W.; Bloomfield, F.H.; Harding, J.E. Undernutrition before mating in ewes impairs the development of insulin resistance during pregnancy. *Obs. Gynecol.* 2009, 114, 869–876. [CrossRef]
- Kumarasamy, V.; Mitchell, M.D.; Bloomfield, F.H.; Oliver, M.H.; Campbell, M.E.; Challis, J.R.G.; Harding, J.E. Effects of periconceptional undernutrition on the initiation of parturition in sheep. *Am. J. Physiol.* 2005, 288, R67–R72. [CrossRef]
- Oliver, M.H.; Jaquiery, A.L.; Bloomfield, F.H.; Harding, J.E. The effects of maternal nutrition around the time of conception on the health of the offspring. Soc. Reprod. Fertil. Suppl. 2007, 64, 397–410. [CrossRef] [PubMed]
- 97. Sinclair, K.D.; Singh, R. Modelling the developmental origins of health and disease in the early embryo. *Theriogenology* **2007**, *67*, 43–53. [CrossRef] [PubMed]
- 98. Aiken, C.E.; Ozanne, S.E. Transgenerational developmental programming. Hum. Reprod. Update 2013, 20, 63–75. [CrossRef]
- 99. Baber, M.; Koren, G. Investigating the fetal and postnatal effects of paternal alcohol exposure in mouse offspring: A review. *J. Popul. Clin. Pharm.* **2015**, *22*, e57–e58.
- Fullston, T.; McPherson, N.O.; Zander-Fox, D.; Lane, M. The most common vices of men can damage fertility and the health of the next generation. *J. Endocrinol.* 2017, 234, F1–F6. [CrossRef] [PubMed]
- 101. Fullston, T.; McPherson, N.O.; Owens, J.A.; Kang, W.X.; Sandeman, L.Y.; Lane, M. Paternal obesity induces metabolic and sperm disturbances in male offspring that are exacerbated by their exposure to an "obesogenic" diet. *Physiol. Rep.* 2015, *3*, e12336. [CrossRef]
- 102. Slyvka, Y.; Zhang, Y.; Nowak, F.V. Epigenetic effects of paternal diet on offspring: Emphasis on obesity. *Endocrine* **2015**, *48*, 36–46. [CrossRef]
- 103. Hur, S.S.J.; E Cropley, J.E.; Suter, C.M. Paternal epigenetic programming: Evolving metabolic disease risk. *J. Mol. Endocrinol.* 2017, 58, R159–R168. [CrossRef]
- 104. Gapp, K.; Jawaid, A.; Sarkies, P.; Bohacek, J.; Pelczar, P.; Prados, J.; Farinelli, L.; Miska, E.; Mansuy, I.M. Implication of sperm RNAs in transgenerational inheritance of the effects of early trauma in mice. *Nat. Neurosci.* 2014, 17, 667–671. [CrossRef] [PubMed]
- 105. Constantinof, A.; Moisiadis, V.G.; Matthews, S.G. Programming of stress pathways: A transgenerational perspective. J. Steroid Biochem. Mol. Biol. 2016, 160, 175–180. [CrossRef]
- Braunschweig, M.; Jagannathan, V.; Gutzwiller, A.; Bee, G. Investigations on Transgenerational Epigenetic Response Down the Male Line in F2 Pigs. *PLoS ONE* 2012, 7, e30583. [CrossRef] [PubMed]
- 107. Kassetas, C.J.; Caton, J.S.; Kirsch, J.D.; Dorsam, S.T.; McCarthy, K.L.; Crouse, M.S.; Sedivec, K.K.; Neville, B.W.; Dahlen, C.R. Effects of feeding 60% dried corn distillers grains plus solubles or the equivalent sulfur as CaSO4 on performance and reproductive traits of yearling Angus bulls. *Theriogenology* 2021, 162, 6–14. [CrossRef]
- 108. Kassetas, C.J.; Caton, J.S.; Kirsch, J.D.; Dorsam, S.T.; McCarthy, K.L.; Crouse, M.S.; Sedivec, K.K.; Neville, B.W.; Dahlen, C.R. Effects of feeding bulls dried corn distiller's grains plus solubles or CaSO4 on mineral and metabolite concentrations in serum and seminal plasma. *Anim. Reprod. Sci.* 2021, 226, 106703. [CrossRef] [PubMed]
- 109. Diniz, W.J.S.; Kassetas, C.J.; Dorsam, S.T.; Underdahl, S.R.; Crouse, M.S.; McCarthy, K.L.; Baumgaertner, F.; Kirsch, J.D.; Sedivec, K.K.; Caton, J.S.; et al. Effects of managing mature beef bulls on divergent planes of nutrition on sperm transcriptome. In Proceedings of the 2021 WSASAS Meeting, Fort Collins, CO, USA, 18–20 October 2021.
- Coy, P.; Lloyd, R.; Romar, R.; Satake, N.; Matas, C.; Gadea, J.; Holt, W.V. Effects of porcine pre-ovulatory oviductal fluid on boar sperm function. *Theriogenology* 2010, 74, 632–642. [CrossRef] [PubMed]
- 111. Coy, P.; Yanagimachi, R. The Common and Species-Specific Roles of Oviductal Proteins in Mammalian Fertilization and Embryo Development. *BioScience* 2015, 65, 973–984. [CrossRef]
- Lopera-Vasquez, R.; Hamdi, M.; Maillo, V.; Lloreda, V.; Coy, P.; Gutierrez-Adan, A.; Bermejo-Alvarez, P.; Rizos, D. Effect of bovine oviductal fluid on development and quality of bovine embryos produced in vitro. *Reprod. Fertil. Dev.* 2015, 29, 621. [CrossRef] [PubMed]
- 113. Canha Gouveia, A.; Prieto-Sánchez, M.T.; Sanchez-Ferrer, M.L.; Molla, M.; Martínez Soto, J.C.; París-Oller, E.; Soriano-Úbeda, C.; Landeras Gutiérrez, J.; Coy Fuster, P. Physicochemical and functional characterization of female reproductive fluids: A report of the first two infants born following addition of their mother's fluids to the embryo culture media. *Front. Physiol. Dev. Physiol.* 2021, in press. [CrossRef]

- 114. Reynolds, L.P.; McCarthy, K.L.; Crouse, M.S.; Kassetas, C.J.; Baumgaertner, F.; Dorsam, S.T.; Kirsch, J.D.; Neville, T.L.; Coy, P.; Caton, J.; et al. PSII-31 Effects of restricted dietary intake on protein expression of oviductal glycoprotein 1 (OVGP1) in the oviductal ampulla of beef cows. J. Anim. Sci. 2020, 98, 376. [CrossRef]
- 115. Bazer, F.; Johnson, G.; Wu, G. Amino Acids and conceptus development during the peri-implantation period of pregnancy. *Adv. Exp. Med. Biol.* **2015**, *843*, 23–52. [PubMed]
- 116. Zeng, X.; Wang, F.; Fan, X.; Yang, W.; Zhou, B.; Li, P.; Yin, Y.; Wu, G.; Wang, J. Dietary Arginine Supplementation during Early Pregnancy Enhances Embryonic Survival in Rats. *J. Nutr.* **2008**, *138*, 1421–1425. [CrossRef] [PubMed]
- 117. Zeng, X.; Huang, Z.; Mao, X.; Wang, J.; Wu, G.; Qiao, S. *N*-carbamylglutamate enhances pregnancy outcome in rats through activation of the PI3K/PKB/mTOR signaling pathway. *PLoS ONE* **2012**, 7, e41192. [CrossRef]
- 118. Zeng, X.; Mao, X.; Huang, Z.; Wang, F.; Wu, G.; Qiao, S. Arginine enhances embryo implantation in rats through PI3K/PKB/mTOR/NO signaling pathway during early pregnancy. *Reproduction* **2013**, *145*, 1–7. [CrossRef]
- 119. Thureen, P.J.; A Baron, K.A.; Fennessey, P.V.; Hay, W.W., Jr. Ovine Placental and Fetal Arginine Metabolism at Normal and Increased Maternal Plasma Arginine Concentrations. *Pediatr. Res.* **2002**, *51*, 464–471. [CrossRef] [PubMed]
- Kim, J.; Burghardt, R.C.; Wu, G.; Johnson, G.A.; Spencer, T.E.; Bazer, F.W. Select Nutrients in the ovine uterine lumen: VII. Effects of arginine, leucine, glutamine and glucose on trophectodem cell signaling, proliferation and migration. *Biol. Reprod.* 2011, *84*, 70–78. [CrossRef]
- 121. Kim, J.; Burghardt, R.C.; Wu, G.; Johnson, G.A.; Spencer, T.E.; Bazer, F.W. Select Nutrients in the ovine uterine lumen: VIII. Arginine stimulates proliferation of ovine trophectoderm cells through mTOR RPS6 K RPS6 signaling cascade and synthesis of nitric oxide and polyamines. *Biol. Reprod.* 2011, 84, 62–69. [CrossRef]
- 122. Kim, J.; Burghardt, R.C.; Wu, G.; Johnson, G.A.; Spencer, T.E.; Bazer, F.W. Select Nutrients in the ovine uterine lumen: IX. Differential effects of arginine, leucine, glutamine and glucose on interferon tau, orinithine decarboxylase and nitric oxide synthase in the ovine conceptus. *Biol. Reprod.* 2011, *84*, 1139–1147. [CrossRef]
- Peine, J.L.; Jia, G.; Van Emon, M.; Neville, T.L.; Kirsch, J.D.; Hammer, C.J.; O'Rourke, S.T.; Reynolds, L.P.; Caton, J.S. Effects of maternal nutrition and rumen-protected arginine supplementation on ewe performance and postnatal lamb growth and internal organ mass1. J. Anim. Sci. 2018, 96, 3471–3481. [CrossRef]
- 124. Bérard, J.; Bee, G. Effects of dietary l-arginine supplementation to gilts during early gestation on foetal survival, growth and myofiber formation. *Animal* **2010**, *4*, 1680–1687. [CrossRef]
- 125. Gao, K.G.; Jiang, Z.Y.; Lin, Y.C.; Zheng, C.; Zhou, G.; Chen, F.; Yang, L.; Wu, G. Dietary l-arginine supplementation enhances placental growth and reproductive performance in sows. *Amino Acids* **2012**, *42*, 2207–2214. [CrossRef] [PubMed]
- Kim, J.Y.; Song, G.H.; Wu, G.; Gao, H.J.; Johnson, G.A.; Bazer, F.W. Arginine, Leucine, and Glutamine Stimulate Proliferation of Porcine Trophectoderm Cells Through the MTOR-RPS6K-RPS6-EIF4EBP1 Signal Transduction Pathway1. *Biol. Reprod.* 2013, 88, 113–120. [CrossRef]
- 127. Mateo, R.D.; Wu, G.; Bazer, F.W.; Park, J.C.; Shinzato, I.; Kim, S.W. Dietary I-Arginine Supplementation Enhances the Reproductive Performance of Gilts. J. Nutr. 2007, 137, 652–656. [CrossRef] [PubMed]
- 128. Gui, S.; Jia, J.; Niu, X.; Bai, Y.; Zou, H.; Deng, J.; Zhou, R. Arginine supplementation for improving maternal and neonatal outcomes in hypertensive disorder of pregnancy: A systematic review. J. Renin-Angiotensin-Aldosterone Syst. 2014, 15, 88–96. [CrossRef] [PubMed]
- 129. Shen, S.-F.; Hua, C.-H. Effect of L-arginine on the expression of Bcl-2 and Bax in the placenta of fetal growth restriction. *J. Matern. Fetal Neonatal. Med.* 2011, 24, 822–826. [CrossRef] [PubMed]
- 130. Xiao, X.M.; Li, L.P. L-arginine treatment for asymmetric fetal growth restriction. Int. J. Gynecol. Obstet. 2005, 88, 15–18. [CrossRef]
- 131. Wu, G.; Bazer, F.W.; Satterfield, M.C.; Li, X.L.; Wang, X.Q.; Johnson, G.A.; Burghardt, R.C.; Dai, Z.L.; Wang, J.J.; Wu, Z.L. Impacts of arginine nutrition on embryonic and fetal development in mammals. *Amino Acids* **2013**, *45*, 241–256. [CrossRef]
- 132. Gao, H.; Wu, G.; Spencer, T.E.; Johnson, G.A.; Bazer, F.W. Select Nutrients in the Ovine Uterine Lumen. III. Cationic Amino Acid Transporters in the Ovine Uterus and Peri-Implantation Conceptuses1. *Biol. Reprod.* 2009, *80*, 602–609. [CrossRef]
- Gao, H.; Wu, G.; Spencer, T.E.; Johnson, G.A.; Bazer, F.W. Select nutrients in the ovine uterine lumen: IV. Expression of neutral and acidic amino acid transporters in ovine uteri and periimplantation conceptuses. *Biol. Reprod.* 2009, *80*, 1196–1208. [CrossRef] [PubMed]
- Simintiras, C.A.; Sanchez, J.M.; McDonald, M.; Martins, T.; Binelli, M.; Lonergan, P. Biochemical characterization of progesteroneinduced alterations in bovine uterine fluid amino acid and carbohydrate composition during the conceptus elongation window. *Biol. Reprod.* 2019, 100, 672–685. [CrossRef]
- 135. Crouse, M.S.; McLean, K.J.; Greseth, N.P.; Ward, A.K.; Reynolds, L.P.; Dahlen, C.R.; Neville, B.W.; Borowicz, P.P.; Caton, J.S. The effects of maternal nutrient restriction and day of early pregnancy on the location and abundance of neutral amino acid transporters in beef heifer utero-placental tissues. *J. Anim. Sci.* **2020**, *98*, skaa197. [CrossRef]
- 136. Crouse, M.S.; McLean, K.J.; Dwamena, J.; Neville, T.L.; Menezes, A.C.B.; Ward, A.K.; Reynolds, L.P.; Dahlen, C.R.; Neville, B.W.; Borowicz, P.P.; et al. The effects of maternal nutrition during the first 50 d of gestation on the location and abundance of hexose and cationic amino acid transporters in beef heifer uteroplacental tissues. *J. Anim. Sci.* **2021**, *99*, skaa386. [CrossRef] [PubMed]
- 137. Crouse, M.S.; McCarthy, K.L.; Menezes, A.C.B.; Kassetas, C.J.; Baumgaertner, F.; Kirsch, J.D.; Dorsam, S.; Neville, T.L.; Ward, A.K.; Borowicz, P.P.; et al. Vitamin and mineral supplementation and rate of gain during the first trimester of gestation affect the abundance of fat-soluble vitamins in fetal liver at d 83 of gestation. *J. Anim. Sci.* 2021, in press.

- 138. McCarthy, K.L.; Nestande, J.; Kassetas, C.J.; Baumgaertner, F.; Kirsch, J.D.; Dorsam, S.T.; Neville, T.L.; Ward, A.K.; Borowicz, P.P.; Reynolds, L.P.; et al. 201 Effects of feeding vitamin and mineral and (or) energy supplements to beef heifers during the first 83 days of gestation on progesterone concentrations, corpus luteum size, and fetal body measurements. *J. Anim. Sci.* 2020, *98*, 161–162. [CrossRef]
- 139. Menezes, A.C.B.; McCarthy, K.L.; Kassetas, C.J.; Baumgaertner, F.; Kirsch, J.D.; Dorsam, S.T.; Neville, T.L.; Ward, A.K.; Borowicz, P.P.; Reynolds, L.P.; et al. PSVIII-37 Late-Breaking Abstract: Effects of feeding a vitamin and mineral supplement and (or) an energy supplement on the abundance of SLC7A5 transporter in beef heifer placentomes at d 83 of gestation. *J. Anim. Sci.* 2020, *98*, 346–347. [CrossRef]
- 140. Diniz, W.J.S.; Reynolds, L.P.; Borowicz, P.P.; Ward, A.K.; Sedivec, K.K.; McCarthy, K.L.; Kassetas, C.L.; Baumgaertner, F.; Kirsch, J.D.; Dorsam, S.T.; et al. Maternal vitamin and mineral supplementation and rate of maternal weight gain affects placental expression of energy metabolism and transport-related genes. *Genes* 2021, *12*, 385. [CrossRef]
- 141. Diniz, W.J.S.; Reynolds, L.P.; Borowicz, P.P.; Ward, A.K.; Sedivec, K.K.; McCarthy, K.L.; Kassetas, C.L.; Baumgaertner, F.; Kirsch, J.D.; Dorsam, S.T.; et al. Maternal vitamin and mineral supplementation affect fetal hepatic expression of genes underlying mineral homeostasis and lipid metabolism in early pregnancy. *J. Anim. Sci* **2021**, in press.
- 142. Clare, C.E.; Pestinger, V.; Kwong, W.Y.; Tutt, D.A.R.; Xu, J.; Byrne, H.M.; Barrett, D.A.; Emes, R.D.; Sinclair, K.D. Interspecific Variation in One-Carbon Metabolism within the Ovarian Follicle, Oocyte, and Preimplantation Embryo: Consequences for Epigenetic Programming of DNA Methylation. *Int. J. Mol. Sci.* 2021, 22, 1838. [CrossRef] [PubMed]
- 143. Dean, W.; Santos, F.; Reik, W. Epigenetic reprogramming in early mammalian development and following somatic nuclear transfer. *Semin. Cell Dev. Biol.* 2003, 14, 93–100. [CrossRef]
- 144. Dobbs, K.B.; Rodriguez, M.; Sudano, M.J.; Ortega, M.S.; Hansen, P.J. Dynamics of DNA Methylation during Early Development of the Preimplantation Bovine Embryo. *PLoS ONE* **2013**, *8*, e66230. [CrossRef]
- 145. Ikeda, S.; Koyama, H.; Sugimote, M.; Kume, S. Review: Roles of one-carbon metabolism in preimplantation period—Effects on short-term development and long-term programming. J. Reprod. Dev. 2012, 58, 38–43. [CrossRef]
- 146. Ispada, J.; De Lima, C.B.; Sirard, M.-A.; Fontes, P.K.; Nogueira, M.F.G.; Annes, K.; Milazzotto, M.P. Genome-wide screening of DNA methylation in bovine blastocysts with different kinetics of development. *Epigenet. Chromatin* **2018**, *11*, 1–13. [CrossRef]
- 147. Salilew-Wondim, D.; Saeed-Zidane, M.; Hoelker, M.; Gebremedhn, S.; Poirier, M.; Pandey, H.O.; Tholen, E.; Neuhoff, C.; Held, E.; Besenfelder, U.; et al. Genome-wide DNA methylation patterns of bovine blastocysts derived from in vivo embryos subjected to in vitro culture before, during or after embryonic genome activation. *BMC Genom.* **2019**, *19*, 1–19. [CrossRef] [PubMed]
- 148. Santos, F.; Hendrich, B.; Reik, W.; Dean, W. Dynamic reprogramming of DNA methylation in the early mouse embryo. *Dev. Biol.* **2002**, 241, 172–182. [CrossRef]
- 149. Xu, J.; Sinclair, K.D. Review: One-carbon metabolism and epigenetic regulation of embryo development. *Reprod. Fertil. Dev.* **2015**, 27, 667–676. [CrossRef] [PubMed]
- 150. Skjaerven, K.H.; Jakt, L.M.; Dahl, J.A.; Espe, M.; Aanes, H.; Hamre, K.; Fernandez, J.M.O. 2016. Parental vitamin deficiency affects the embryonic gene expression of immune-, lipid transport- and apolipoprotein genes. *Sci. Rep.* 2016, *6*, 34535. [CrossRef]
- 151. Peñagaricano, F.; Souza, A.H.; Carvalho, P.D.; Driver, A.M.; Gambra, R.; Kropp, J.; Hackbart, K.S.; Luchini, D.; Shaver, R.D.; Wiltbank, M.C.; et al. Effect of Maternal Methionine Supplementation on the Transcriptome of Bovine Preimplantation Embryos. *PLoS ONE* 2013, 8, e72302. [CrossRef] [PubMed]
- 152. Reynolds, L.P.; McLean, K.J.; McCarthy, K.L.; Diniz, W.J.S.; Menezes, A.C.B.; Forcherio, J.C.; Scott, R.R.; Borowicz, P.P.; Ward, A.K.; Dahlen, C.R.; et al. Nutritional regulation of embryonic survival, growth and development. *Invited Review*. In *Recent Advances in Animal Nutrition and Metabolism*; Advances in Experimental Medicine and Biology 1354; Wu, G., Ed.; Springer Nature: Cham, Switzerland, 2022; ISBN 978-3-030-85686-1. [CrossRef]
- 153. Zhu, L.; Marjani, S.L.; Jiang, Z. The Epigenetics of Gametes and Early Embryos and Potential Long-Range Consequences in Livestock Species—Filling in the Picture with Epigenomic Analyses. *Front. Genet.* **2021**, 12. [CrossRef]
- 154. Mueller, C.A.; Eme, J.; Burggren, W.W.; Roghair, R.D.; Rundle, S.D. Challenges and opportunities in developmental integrative physiology. *Comp. Biochem. Physiol. Part A Mol. Integr. Physiol.* **2015**, *184*, 113–124. [CrossRef]
- 155. Nettle, D.; Frankenhuis, W.E.; Rickard, I.J. The evolution of predictive adaptive responses in human life history. *Proc. R. Soc. B Biol. Sci.* 2013, *280*, 20131343. [CrossRef] [PubMed]
- 156. Cline, H.J.; Neville, B.W.; Lardy, G.P.; Caton, J.S. Influence of advancing season on dietary composition, intake, site of digestion, and microbial efficiency in beef steers grazing a native range in western North Dakota1. *J. Anim. Sci.* **2009**, *87*, 375–383. [CrossRef] [PubMed]
- 157. Cline, H.J.; Neville, B.W.; Lardy, G.P.; Caton, J.S. Influence of advancing season on dietary composition, intake, site of digestion, and microbial efficiency in beef steers grazing season-long or twice-over rotation native range pastures in western North Dakota1. *J. Anim. Sci.* **2010**, *88*, 2812–2824. [CrossRef]
- 158. Johnson, J.A.; Caton, J.S.; Poland, W.; Kirby, D.R.; Dhuyvetter, D.V. Influence of season on dietary composition, intake, and digestion by beef steers grazing mixed-grass prairie in the northern Great Plains. J. Anim. Sci. 1998, 76, 1682–1690. [CrossRef] [PubMed]
- Krysl, L.J.; Galyean, M.L.; Wallace, J.D.; McCollum, F.T.; Judkins, M.B.; Branine, M.E.; Caton, J.S. Cattle Nutrition on Blue Grama Rangeland in New Mexico; Bulletin 727; New Mexico State University Agricultural Experiment Station: Las Cruces, NM, USA, 1987.

- 160. Bhutta, Z.A.; Salam, R.A. Global Nutrition Epidemiology and Trends. Ann. Nutr. Metab. 2012, 61, 19–27. [CrossRef]
- 161. Latham, M.C. *Human Nutrition in the Developing World*; Food and Nutrition Series No. 29; Food and Agriculture Organization of the United Nations: Rome, Italy, 1997.
- UNICEF. The State of the World's Children 2019. Children, Food and Nutrition: Growing Well in a Changing World; United Nations Children's Fund: New York, NY, USA. Available online: https://www.unicef.org/media/60806/file/SOWC-2019.pdf (accessed on 22 October 2021).
- 163. UNSCN. The UN Decade of Action on Nutrition 2016–2025; United Nations System Standing Committee on Nutrition: 2021. Available online: https://www.unscn.org/en/topics/un-decade-of-action-on-nutrition (accessed on 22 October 2021).
- 164. WHO. *Good Maternal Nutrition: The Best Start in Life;* WHO Regional Office for Europe: Copenhagen, Denmark, 2016. Available online: https://www.euro.who.int/\_\_data/assets/pdf\_file/0008/313667/Good-maternal-nutrition-The-best-start-in-life.pdf (accessed on 22 October 2021).