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Accepted: 2019.07.21 Published: 2019.08.30		Procalcitonin Levels Are Intrauterine Growth			
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Corresponding Author: Source of support:	Pervin Karlı, e-mail: parpi2300@hotmail.com Departmental sources				
Background: Material/Methods: Results:	blood samples of patients with idiopathic intraute priate for gestational age (AGA) infants. The present prospective study included 43 patients infants at similar gestational ages (control group). <i>N</i> from the control group and IUGR group at time of ternal blood. Procalcitonin and CRP levels were and The median value of CRP levels in maternal blood AGA group (p=0.001). The mdian CRP level in cord and 10.1 mg/dl (range, 4.07–16.5) in the control gro	was 47.5 mg/dl in the IUGR group and 15.255 mg/dl in the blood was 36.4 mg/dl (range, 17.3–47.2) in the IUGR group up, and the difference was statistically significant (p=0.001).			
Conclusions:	and the difference was not statistically significant (was 0.06 μ g/l in the IUGR group and 0.04 μ g/l in t nificant (p=0.741).	0.05 μg/l in the IUGR group and 0.04 μg/l in the AGA group, p=0.435). The median procalcitonin value in fetal cord blood the AGA group, and the difference was not statistically sig-			
MeSH Keywords:	procalcitonin, which is another inflammatory indica	ator, between the groups.			
Full-text PDF:	https://www.medscimonit.com/abstract/index/idArt/917397				
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Maternal Serum and Fetal Cord Blood C-Reactive



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Background

Intrauterine growth restriction is a serious problem that causes perinatal mortality and morbidity [1–3]. Intrauterine growth restriction is also known as fetal growth restriction (FGR). The causes for IUGR include hypertensive diseases during pregnancy, genetic disorders, infections, maternal malnutrition, toxins, and drug use [4]. However, there is no known cause in 70% of these patients and this condition is defined as idiopathic IUGR [5]. Idiopathic IUGR usually presents with placental failure [6].

Procalcitonin is a calcitonin-progenitor polypeptide acting in calcium homeostasis. It is produced in parafollicular C cells of the thyroid gland, as well as in neuroendocrine cells of the lungs and intestines [7]. It is also used as an indicator for inflammation in the body [8]. Increasing inflammatory cytokines in the maternal blood negatively affect fetal development [9]. Therefore, we hypothesized that levels of procalcitonin, an inflammatory marker, are increased in patients with IUGR.

Procalcitonin has been studied in many inflammatory processes. There are studies indicating elevated procalcitonin levels in preeclampsia [10–12]. However, to the best of our knowledge, the present study is the first to evaluate procalcitonin levels in IUGR. At present, there is no diagnostic marker for IUGR. The inflammatory process is considered to be active in IUGR. Therefore, the aim of the present study was to assess procalcitonin levels in maternal blood and cord blood of patients with IUGR and to compare the results with a control group.

Therefore, this prospective, single-center study compared procalcitonin and CRP levels in maternal serum and fetal cord blood samples from pregnancies associated with idiopathic IUGR vs. appropriate for gestational age (AGA) births.

Material and Methods

This prospective study was carried out at the Obstetrics Clinic of Ondokuz Mayis University on 27 patients with idiopathic IUGR as the patient group and 26 appropriate for gestational age (AGA) infants as the control group. The study was approved by our Local Ethics Committee approval (no. B.30.2.ODM.0.20.08/213-315), and written informed consent was obtained from all women who participated in the study. All authors and the study protocol complied with the World Medical Association Declaration of Helsinki regarding ethical conduct of research involving human subjects. IUGR was diagnosed in infants with predicted fetal body weight below the 10th percentile according to the gestational week through ultrasonographic measurements. Gestational age was calculated from the first day of the last menstruation and was confirmed by the first fetal ultrasound measurements.

The causes of termination of pregnancy before 38 weeks by caesarean section were events that required termination of pregnancy prematurely due to maternal conditions such as maternal lumbar discopathy or maternal cardiac diseases. Patients at gestational week 36 weeks and over were included in the study because steroids that are administrated for lung maturation can change the inflammation levels in earlier weeks.

Exclusion criteria were: gestational and pre-gestational diabetes, hypertension, multiple pregnancy, fetal abnormalities including trisomies, thyroid diseases, drug use during pregnancy (smoking, alcohol, or illegal drugs) premature membrane rupture, autoimmune disease, systemic lupus erythematosus, fever at least 37.5°C, elevated white blood cells before delivery, suspicion of infectious diseases, and receiving antibiotic therapy.

After diagnosis of IUGR through obstetric ultrasound scan, were recorded patient age, gravidity, parity, gestational week, and body mass index (BMI). Doppler ultrasound scan of the umbilical artery was performed for all patients, and diastolic flow was classified as present or absent. Amniotic fluid index was calculated on 4 quadrants for amniotic fluid measurement. If the amniotic fluid index was below 50 in 4 quadrants, the case was evaluated as oligohydramnios.

Collection and storage of serum samples

Maternal blood was collected from the mother and cord blood was collected from the newborn after birth, then the samples were centrifuged. Serum samples obtained were placed into Eppendorf tubes and stored at -80° C. Serum samples were stored at -40° C, -20° C, and at $2-8^{\circ}$ C, then the samples were kept at room temperature and analysis was performed.

Procalcitonin and CRP measurement

Procalcitonin measurement was performed by electrochemiluminescence (ECLIA) method through original Roche diagnostic kits on a Roche Cobas E601 hormone device. This method uses a procalcitonin-specific monoclonal antibody, a biotinlabeled procalcitonin-specific monoclonal antibody, and a ruthenium complex.

CRP measurement was performed by immunoturbidimetric method using original Roche diagnostic kits on a Roche Cobas C501 device. We used human-originated CRP agglutinate with latex particles coated with monoclonal anti-CRP antibodies. The aggregates were analyzed turbidimetrically.

Table 1. Demographic criteria.

	Patient group (IUGR) Median (IQR)	Control group (AGA) Median (IQR)	P value
Age (year)	31 (27–34)	28.5 (24.7–32)	0.095
BMI (kg/m²)	28.4±3.1	28.3±2.7	0.05
Pregnancy period (days)	258 (255–262)	266 (265–271)	0.4
Birth weight (grams)	2100 (2000–2440)	3230 (3000–3545)	0.001**
Gender	15 (55.6%)/12 (44.4%) boy/girl	14 (53.8%)/12 (46.2%) boy/girl	

Table 2. Test results.

	Patient (IUGR) Median (IQR)		Control (AGA) Median (IQR)		P value
Apgar score 1 st min.	8	(7.2–9.1)	9	(8.25–9.25)	0.033*
Diastolic flow loss	4/27		0/26		0.298
Number of the patients with Oligohydramnios	4/27		0/26		0.02
CRP mg/dL	47.5	(25–60)	15.25	(8–22.45)	0.001**
Serum Procalcitonin (µg/L)	0.05	(0.02–0.07)	0.04	(0.02–0.06)	0.435
Cord Procalcitonin (µg/L)	0.06	(0.02–0.07)	0.04	(0.03–0.08)	0.741
CRP/Procalcitonin (mg/dL/µg/L)	853.3	(527.8–2200)	312.7	(163.3–870)	0.004*
Cord CRP mg/dL	36.4	(17.3–47.2)	10.1	(4.07–16.5)	0.001**
Cord CRP/procalcitonin (mg/dL/µg/L)	735.0	(274.6–1126.3)	178.5	(104.1–419.8)	0.001**

Alpha significance level p<0.05; IQR – interquartile range.

Statistical analysis

Statistical analysis of the data was performed using SPSS 15 for Windows. Definitive statistical data were presented as median, interquartile range (IQR), and percent (%). Skewness and kurtosis values of normal distribution were detected by Kolmogorov-Smirnov test (with Lilliefors significance correction), Shapiro-Wilk tests, and histogram distribution graphs. Comparison of independent groups that did not meet normal distribution criteria for numeric variables was performed through Mann Whitney U test, whereas Spearman correlation analysis, which is used for non-parametric tests, was performed for correlation analysis. The alpha significance level was accepted as p<0.05.

Results

There were no significant differences detected between the groups in terms of age, BMI, sex of the infant, and pregnancy period (p=0.095, p>0.05, p>0.05). The median birth weight was

2100 g in the IUGR group and 3230 g in the AGA group, and the difference was statistically significant (p=0.001) (Table 1).

The median Apgar score was 8 in the IUGR group and 9 in the AGA group. There was no significant difference between the groups in first-minute Apgar scores (p=0.033). Diastolic flow loss is detected in 4 patients in the IUGR group, and this was not statistically significant between groups (0.298). There were 4 patients with oligohydroaminosis in the patient group only, and this was statistically significant (p=0.02). The median value of CRP level in maternal blood was 47.5 mg/dl in the IUGR group and 15.255 mg/dl in the AGA group (p=0.001). CRP level in the cord blood was 36.4 mg/dl (17.3-47.2) in the IUGR group and 10.1 mg/dl (4.07-16.5) in the control group, and the difference was statistically significant (p=0.001). The median maternal serum procalcitonin level was 0.05 µg/l in the IUGR group and 0.04 µg/l in the AGA group, and the difference was not statistically significant (p=0.435). The median procalcitonin value in fetal cord blood was 0.06 μ g/l in the IUGR group and 0.04 μ g/l in the AGA group, and the difference was not statistically significant (p=0.741). The CRP/procalcitonin ratio was 853.3

	Serum Procalcitonin (µg/L)	Pregnancy period (days)	Newborn weight (gram)	Apgar score	CRP (mg/dL)	Cord Procalcitonin (µg/L)	CRP/ Procalcitonin (mg/dL/µg/L)
Serum Procalcitonin (µg/L)	_	r: –0.078 p=0.699	r: -0.436 p=0.023*	r: 0.237 p=0.234	r: –0.279 p=0.159	r: -0.014 p=0.946	r: -0.814 p=0.001*
Pregnancy period (days)	r: -0.078 p=0.699	-	r: 0.738 p=0.001*	r: 0.195 p=0.331	r: 0.105 p=0.603	r: 0.063 p=0.755	r: 0.125 p=0.535
Newborn weight (gram)	r: -0.436 p=0.023*	r: 0.738 p=0.001*	-	r: 0.391 p=0.044*	r: 0.324 p=0.100	r: -0.011 p=0.957	r: 0.452 p=0.018*
Apgar score	r: 0.237 p=0.234	r: 0.195 p=0.331	r: 0.391 p=0.044*	-	r: 0.402 p=0.037*	r: -0.169 p=0.401	r: 0.054 p=0.790
CRP (mg/dL)	r: -0.279 p=0.159	r: 0.195 p=0.331	r: 0.324 p=0.100	r: 0.402 p=0.037*	-	r: 0.063 p=0.756	r: 0.717 p=0.001*
Cord Procalcitonin (µg/L)	r: -0.014 p=0.946	r: 0.063 p=0.755	r: -0.011 p=0.957	r: -0.169 p=0.401	r: 0.063 p=0.756	_	r: -0.008 p=0.970
CRP/Procalcitonin (mg/dL/µg/L)	r: -0.814 p=0.001*	r: 0.125 p=0.535	r: 0.452 p=0.018*	r: 0.054 p=0.790	r: 0.717 p=0.001*	r: -0.008 p=0.970	-
Oligohydramnios	r: -0.050 p=0.806	r: 0.067 p=0.739	r: 0.104 p=0.607	r: 0.073 p=0.718	r: 0.057 p=0.777	r: -0.281 p=0.156	r: 0.121 p=0.548
Diastolic flow loss	r: -0.124 p=0.537	r: 0.218 p=0.274	r: 0.264 p=0.184	r: 0.004 p=0.986	r: 0.309 p=0.116	r: 0.409 p=0.034*	r: 0.289 p=0.143
Cord CRP mg/dL	r: -0.247 p=0.213	r: 0.116 p=0.565	r: 0.333 p=0.090	r: 0.350 p=0.073	r: 0.851 p=0.01*	r: 0.05 p=0.803	
Cord CRP/Procalcitonin (mg/dL/µg/L)	r: -0.092 p=0.650	r: 0.139 p=0.489	r: 0.240 p=0.228	r: 0.342 p=0.080	r: 0.462 p=0.015*	r: 0.526 p=0.005*	

Table 3. IUGR group, results of Spearman correlation analysis.

* Weakly significant at 0.05 level; * strongly significant at 0.01 level.

mg/dl in the IUGR group and 312.7 mg/dl in the control group, and the difference was statistically significant (p=0.004). The CRP/procalcitonin ratio in cord blood was 735 mg/dl (range, 274.6–1126.3) in the IUGR group and 178.5 mg/dl (range, 104.1–419.8) in the control group, and the difference was statistically significant (p=0.001) (Table 2).

Correlation results in the IUGR group are presented in Table 3. A negative correlation was detected between serum procalcitonin and birth weight (r: -0.436, p=0.023). A positive correlation was detected between Apgar score and CRP level (r: 0.402, p=0.037). A positive correlation was found between diastolic flow loss and procalcitonin level of cord blood by Doppler scan of the umbilical artery (r: 0.409 p=0.034).

Discussion

Intrauterine growth restriction is defined as fetal weight below 10% according to the population at the same gestational age. IUGR is an untreatable pathology in intrauterine life except for delivery at a suitable time after diagnosis. However, this condition has many effects on fetal life and later. These effects include bronchopulmonary dysplasia, cerebral palsy, cardiac, metabolic, and renal diseases [13–16].

We found no detectable causes of IUGR in 70% of the patients. Previous studies indicated that increasing inflammation can cause IUGR during pregnancy. Levels of IL-6, IL1 beta, and TNF alpha were reported to be higher in amniotic fluid and cord blood of small for gestational age (SGA) infants [17,18]. Placental mRNA levels of IL-8, IL-6, interferon-g, and TNF alpha were reported to be higher in infants with IUGR [19,20]. Inflammatory cytokine levels in the serum were found to be higher in mothers of infants with IUGR compared with normal pregnant women; inflammatory cytokines were higher in the patients with IUGR and placental failure [21]. Ernst et al. showed that maternal CRP levels in early pregnancy are associated with lower birth weight and increased risk of neonatal complications [22].

Fat tissue-secreted hsCRP and the increase in fat tissue is an aseptic inflammatory process [23,24]. Boutsikou et al. found that serum CRP levels were similar in IUGR and AGA fetuses [25].

Although IUGR patients has less fat tissue, CRP levels were reported to be similar to those in the control group. Such results were considered as higher CRP levels in the patients with IUGR according to the fat tissue quantity. The above study results are different from those of our study. Boutsikou et al. included patients with preeclampsia, hypertension, or other IUGR reasons, but we included only idiopathic IUGR patients. Thus, inflammation may be more important in idiopathic IUGR than in preeclampsia or hypertensive disorders.

We detected significantly higher CRP levels in blood samples of pregnant women whose infants were diagnosed with IUGR. However, despite higher procalcitonin levels in maternal serum and cord blood in the IUGR group, the difference was not statistically significant. CRP levels gradually increase during pregnancy and peak at birth; CRP cannot pass the placental barrier and affect the fetus [26–28]. In contrast to CRP, no difference was shown in procalcitonin levels between pregnant and nonpregnant women [29]. It was demonstrated that procalcitonin was not associated with gestational week or birth [30]. Ruiz Gonzalez et al. showed that serum CRP and procalcitonin levels were not different between IUGR and AGA neonates [31]. We therefore assessed procalcitonin levels in patients with IUGR and combined this analysis with procalcitonin levels. In fact, procalcitonin, as an inflammatory marker, was higher in pregnant women with preeclampsia than in normal pregnant women [32,33]. However, there is no relevant study about IUGR in the literature.

References:

- 1. Brodsky D, Christou H: Current concepts in intrauterine growth restriction. J Intensive Care Med, 2004; 19: 307–19
- 2. Sankaran S, Kyle PM: Aetiology and pathogenesis of IUGR. Best Pract Res Clin Obstet Gynaecol, 2009; 23: 765–77
- ACOG Practice Bulletin No. 204: Fetal Growth Restriction. Obstet Gynecol, 2019; 133(2): e97–e109
- 4. Kalanithi LE, Illuzzi JL, Nossov VB et al: Intrauterine growth restriction and placental location. J Ultrasound Med, 2007; 26: 1481–89
- Suhag A, Berghella V. Intrauterine growth restriction (IUGR): Etiology and diagnosis. Curr Obstet Gynecol Rep, 2013; 2: 102–11
- Salafia CM, Minior VK, Pezzullo JC et al: Intrauterine growth restriction in infants of less than thirty-two weeks' gestation: Associated placental pathologic features. Am J Obstet Gynecol, 1995; 173: 1049–57
- 7. Schneider HG, Lam QT: Procalcitonin for the clinical laboratory: A review. Pathology, 2007; 39: 383–90
- Meisner M: Update on procalcitonin measurements. Ann Lab Med, 2014; 34: 263–73
- 9. Roman A, Desai N, Rochelson B et al: Maternal magnesium supplementation reduces intrauterine growth restriction and suppresses inflammation in a rat model. Am J Obstet Gynecol, 2013; 208: 383.e1–7
- Kucukgoz Gulec U, Ozgunen FT, Guzel AB et al: An analysis of C-reactive protein, procalcitonin, and D-dimer in pre-eclamptic patients. Am J Reprod Immunol, 2012; 68: 331–37
- Montagnana M, Lippi G, Albiero A et al: Procalcitonin values in preeclamptic women are related to severity of disease. Clin Chem Lab Med, 2008; 46: 1050–51

Procalcitonin levels are significantly higher in people with bacterial infections [34]; it increases and decreases more rapidly than does the erythrocyte sedimentation rate and CRP. The halflife is 25–30 h [35]. Although CRP is significantly higher in IUGR patients due to such increase and decrease, procalcitonin may not be significant.

The present study has some limitations. It was performed in a single center, with a small study population, and compared only 2 biomarkers that are usually associated with inflammation. There were no clinical details provided for the women or infants that could have affected the levels of these markers, such as acute or chronic infection or inflammatory diseases, prolonged labor, method of delivery, and medications used.

Conclusions

To the best of our knowledge, this is the first study assessing procalcitonin levels in patients with IUGR. However, we believe that similar studies in larger patient populations are needed. Our results indicated that increased levels of CRP contribute to the inflammatory process in idiopathic IUGR. Similar results were not found for procalcitonin, which is a less affected marker in pregnancy-associated cases. Therefore, prospective studies of IUGR in larger patient populations are needed.

Conflict of interest

None.

- 12. Can M, Sancar E, Harma M et al: Inflammatory markers in preeclamptic patients. Clin Chem Lab Med, 2011; 49: 1469–72
- 13. Wu YW, Escobar GJ, Grether JK et al: Chorioamnionitis and cerebral palsy in term and near-term infants. JAMA, 2003; 290: 2677–84
- 14. Jarvis S, Glinianaia SV, Torrioli MG et al: Cerebral palsy and intrauterine growth in single births: European collaborative study. Lancet, 2003; 362: 1106–11
- Zeitlin J, El Ayoubi M, Jarreau PH et al: Impact of fetal growth restriction on mortality and morbidity in a very preterm birth cohort. J Pediatr, 2010; 157: 733–9e1
- Amarilyo G, Oren A, Mimouni FB et al: Increased cord serum inflammatory markers in small for gestational age neonates. J Perinatol, 2011; 31: 30–32
- Heyborne KD, Witkin SS, McGregor JA: Tumor necrosis factor alpha in midtrimester amniotic fluid is associated with impaired intra uterine fetal growth. Am J Obstet Gynecol, 1992; 167: 920–25
- Street ME, Seghini P, Fieni S et al: Changes in interleukin-6 and IGF system and their relationships in placenta and cord blood in newborns with fetal growth restriction compared with controls. Eur J Endocrinol, 2006; 155: 567–74
- 19. Simmons RA: Developmental origins of adult disease. Pediatr Clin North Am, 2009; 56: 449–66
- Bartha JL, Romero-Carmona R, CominoDelgado R: Inflammatory cytokines in intrauterine growth retardation. Acta Obstet Gynecol Scand, 2003; 82: 1099–102

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- Al-Azemi M, Raghupathy R, Azizieh F: Pro-inflammatory and anti-inflammatory cytokine profiles in fetal growth restriction. Clin Exp Obstet Gynecol, 2017; 44(1): 98–103
- 22. Ernst GD, de Jonge LL, Hofman A et al: C-reactive protein levels in early pregnancy, fetal growth patterns, and the risk for neonatal complications: The Generation R Study. Am J Obstet Gynecol, 2011; 205(2): 132.e1–12
- Ouchi N, Kihara S, Funahashi T et al: Reciprocal association of C-reactive protein with adiponectin in blood stream and adipose tissue. Circulation, 2003; 107: 671–74
- 24. Das UN: Is obesity an inflammatory condition? Nutrition, 2001: 17: 953-66
- 25. Boutsikou T, Mastorakos G, Kyriakakou M et al: Levels of inflammatory markers in intrauterine growth restriction. Mediators Inflamm, 2010; 2010: 790605
- Belo L, Santos-Silva A, Rocha S et al: Fluctuations in C-reactive protein concentration and neutrophil activation during normal human pregnancy. Eur J Obstet Gynecol Reprod Biol, 2005; 123: 46–51
- 27. Watts DH, Krohn MA, Wener MH, Eschenbach DA: C-reactive protein in normal pregnancy. Obstet Gynecol. 1991; 77: 176–80

- Nielsen FR, Bek KM, Rasmussen PE et al: C-reactive protein during normal pregnancy. Eur J Obstet Gynecol Reprod Biol, 1990; 35: 23–27
- 29. Paccolat C, Harbarth S, Courvoisier D et al: Procalcitonin levels during pregnancy, delivery and postpartum. J Perinat Med, 2011; 39: 679–83
- Assumma M, Signore F, Pacifico L et al: Serum procalcitonin concentrations in term delivering mothers and their healthy offspring: A longitudinal study. Clin Chem, 2000; 46: 1583–87
- Ruiz-González MD, Cañete MD, Gómez-Chaparro JL et al: Morbility, clinical data and proteomic analysis of IUGR and AGA newborns at different gestational ages. Data Brief, 2016; 9: 438–47
- Montagnana M, Lippi G, Albiero A et al: Procalcitonin values in preeclamptic women are related to severity of disease. Clin Chem Lab Med, 2008; 46: 1050–51
- Gulec UK, Ozgunen FT, Guzel AB et al: An analysis of C-reactive protein, procalcitonin, and D-dimer in pre-eclamptic patients. Am J Reprod Immunol, 2012; 68: 331–37
- 34. Meisner M: Update on procalcitonin measurements. Ann Lab Med, 2014; 34: 263-73
- 35. Schneider HG, Lam QT: Procalcitonin for the clinical laboratory: A review. Pathology, 2007; 39: 383–90