



Article

Common Genetic Aberrations Associated with Metabolic Interferences in Human Type-2 Diabetes and Acute Myeloid Leukemia: A Bioinformatics Approach

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Abstract: Type-2 diabetes mellitus (T2D) is a chronic metabolic disorder, associated with an increased risk of developing solid tumors and hematological malignancies, including acute myeloid leukemia (AML). However, the genetic background underlying this predisposition remains elusive. We herein aimed at the exploration of the genetic variants, related transcriptomic changes and disturbances in metabolic pathways shared by T2D and AML, utilizing bioinformatics tools and repositories, as well as publicly available clinical datasets. Our approach revealed that rs11709077 and rs1801282, on *PPARG*, rs11108094 on *USP44*, rs6685701 on *RPS6KA1* and rs7929543 on *AC118942.1* comprise common SNPs susceptible to the two diseases and, together with 64 other co-inherited proxy SNPs, may affect the expression patterns of metabolic genes, such as *USP44*, *METAP2*, *PPARG*, *TIMP4* and *RPS6KA1*, in adipose tissue, skeletal muscle, liver, pancreas and whole blood. Most importantly, a set of 86 AML/T2D common susceptibility genes was found to be significantly associated with metabolic cellular processes, including purine, pyrimidine, and choline metabolism, as well as insulin, AMPK, mTOR and PI₃K signaling. Moreover, it was revealed that the whole blood of AML patients exhibits deregulated expression of certain T2D-related genes. Our findings support the existence of common metabolic perturbations in AML and T2D that may account for the increased risk for AML in T2D patients. Future studies may focus on the elucidation of these pathogenetic mechanisms in AML/T2D patients, as well as on the assessment of certain susceptibility variants and genes as potential biomarkers for AML development in the setting of T2D. Detection of shared therapeutic molecular targets may enforce the need for repurposing metabolic drugs in the therapeutic management of AML.

Keywords: acute myeloid leukemia (AML); type-2 diabetes mellitus (T2D); metabolic pathways; single-nucleotide polymorphisms (SNPs)



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1. Introduction

Type-2 diabetes mellitus (T2D) is a chronic metabolic disorder, nowadays considered a global epidemic, with ever-increasing prevalence and high cardiovascular mortality rates [1]. Metabolic disturbances in T2D are associated with chronic hyperglycemia due to deficient insulin secretion by pancreatic β -cells and decreased insulin sensitivity in the skeletal muscle, liver, and adipose tissue [2]. During the last two decades, 85 genome-wide association studies (GWAS) have revealed 1894 single-nucleotide polymorphisms (SNPs) in 1294 genes involved in the aforementioned processes [3]. Interestingly, it was recently shown that certain T2D susceptibility genes exhibit deregulated mRNA expression in the peripheral blood of patients and predisposed individuals, possibly mirroring the aberrant regulation in disease-target organs [4].

T2D also has been associated with the development of various types of human neoplasia, including both solid tumors and hematological malignancies [5]. A recent study on 804,100 new cancer patients bearing different tumor types reported that 5.7% of their development was attributable to diabetes and high body mass index (BMI) [6]. Moreover, observational and Mendelian randomization studies support a strong epidemiological link between T2D and cancer [7]. Common pathophysiological background includes: (a) risk factors such as aging, obesity and physical inactivity; (b) biological processes including hyperinsulinemia, hyperglycemia, oxidative stress and chronic low-grade inflammation and (c) molecular pathways such as the insulin/insulin-like growth factor (IGF) and interleukin (IL)-6/signal transducer and activator of transcription 3 (STAT3) axes [5]. Importantly, the first-line anti-diabetic drug metformin is known to lower the risk of cancer development in T2D patients and improve the response to anti-cancer therapies in diabetic or non-diabetic individuals bearing certain tumor types [8]. At the cellular level, the drug exerts its anti-cancer function by interfering with mitochondrial respiration and activating the AMP-activated protein kinase (AMPK) pathway [8]. At the systemic level, metformin suppresses insulin/IGF-1 and nuclear factor- κ B (NF- κ B) signaling pathways, downregulates the release of proinflammatory cytokines and augments CD8⁺ T cell anti-tumor responses [8].

Among hematological malignancies, acute and chronic leukemias have been associated with a previous history of T2D. A recent meta-analysis of 18 studies involving 10,516 leukemia cases within a total of more than 4 million individuals with diabetes showed that the risk for the disease is increased in patients with T2D but not in patients with type 1 diabetes [9]. Especially for acute myeloid leukemia (AML), a life-threatening hematological malignancy with critical survival rates [10], it has been described that the standard incidence ratio in a cohort of 641 T2D individuals is 1.36 (95% CI: 1.26–1.47), significantly higher than in the general population [11]. Furthermore, various studies have detected BMI as an independent adverse prognostic factor for AML [12–14], which aggravates the relative risk for the disease in T2D [9,15]. Additionally, metformin has been associated with improved outcomes also in patients with leukemias [16]. On the other hand, *in vitro* studies have described that AML cells exhibit a hyper-metabolic phenotype that involves upregulations in basal and maximal respiration [17] and perturbations in glycolysis and oxidative phosphorylation processes [18,19]. These clinical and *in vitro* data suggest that repurposing metformin could possibly modify leukemic cells' metabolism, indicating a promising option for the management of AML [16].

Despite the identified epidemiological association of AML with T2D, the genetic and molecular links between the two disorders remain unclear. The possible existence of common metabolic interferences that may underlie the development and perpetuation of the disease has not yet been investigated. Neither is it known whether these are attributed to aberrations in the genomic, transcriptional, or post-transcriptional level. To this end, we herein investigated a network of common genetic alterations (single-nucleotide polymorphisms, SNPs) and co-inherited variants, related mRNA deviations and pathway deregulations in the two conditions, utilizing appropriate bioinformatic tools and publicly available clinical datasets. Priority was given to the identification of gene sets and pathways associated with possible metabolic disturbances, perchance known to be related to T2D, that may control the development of AML. To the best of our knowledge, our results provide the first information regarding common genetic predisposition and connected mechanisms that may lead to the development of AML in the setting of T2D.

2. Results

2.1. Common Susceptibility SNPs in AML and T2D

Data on all SNPs associated with AML or T2D development were downloaded from the NHGRI-EBI Catalog (Supplementary Table S1). The numbers of SNPs listed and further processed were 5321 for AML and 1894 for T2D, as depicted in Figure 1A. Of these, five SNPs (rs11108094, rs1801282, rs7929543, rs11709077, rs6685701) were found

to be linked with the development of both AML and T2D. All of them exerted a p -value for the association with either disease of $<5 \times 10^{-8}$, which was set as a threshold of significance. These five SNPs were included in the subsequent analyses of this study as significantly associated with both AML and T2D. Corresponding information on these SNPs is summarized in Table 1. In addition, information regarding their frequency in the general population is reported in Supplementary Figure S1.

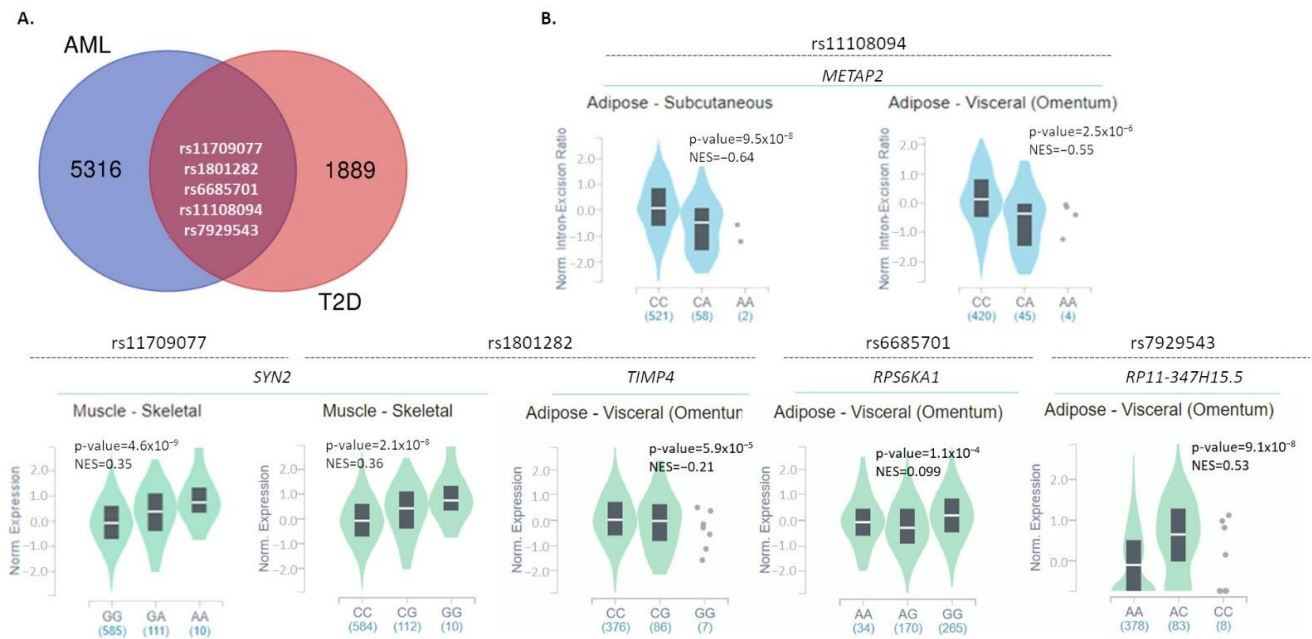


Figure 1. Common SNPs between AML and T2D and their impact on gene expression in disease-associated tissues. **(A)** Venn diagrams reporting the number of common and specific SNPs significantly associated with AML or T2D, based on data downloaded from the NHGRI-EBI GWAS Catalog. **(B)** Violin plots depicting the impact of the five common SNPs on the expression levels of associated or other genes, in disease-associated tissues (subcutaneous or visceral adipose tissue, skeletal muscle, liver, pancreas, whole blood) (GTex portal, May 2021). NES: normalized effect size.

Two of these SNPs (rs11709077, rs1801282) lie in the *PPARG* (peroxisome proliferator-activated receptor gamma) gene, exerting the following p -values: for rs11709077 5×10^{-11} for AML and 2×10^{-36} for T2D, and for rs1801282 5×10^{-11} for AML and 2×10^{-19} for T2D. Another common SNP, the rs6685701, is found in the gene encoding for the ribosomal protein S6 kinase A1 (*RPS6KA1*) and exhibits a significant association with AML ($p = 6 \times 10^{-18}$) and T2D ($p = 1 \times 10^{-8}$). *USP44* (Ubiquitin Specific Peptidase 44) also bears an SNP (rs11108094) significantly related to both AML and T2D development ($p = 2 \times 10^{-10}$ and 6×10^{-10} , respectively). Last, rs7929543, located in *AC118942.1* (NADPH oxidase 4 pseudogene), is also significantly associated with both AML ($p = 7 \times 10^{-9}$) and T2D ($p = 2 \times 10^{-9}$). It is important to note that all SNPs are in non-coding regions except SNP rs1801282 which is a missense variant in *PPARG*, also known as Pro12Ala. The more common C allele encodes for the Pro amino acid at the SNP position [20].

Table 1. Information about the five common SNPs associated with both AML and T2D, as obtained upon search in the NHGRI-EBI Catalog of genome-wide association studies (GWAS) (May 2021) [3]. Variant ID, chromosomal location, cytogenetic region, mapped genes, risk alleles, *p*-values detected in each study, study accession numbers and the corresponding traits are reported.

SNP	Chromosomal Location	Cytogenetic Region	Mapped Gene	Risk Allele	<i>p</i> -Value	Study Accession Number	Trait
rs11709077	3:12295008	3p25.2	<i>PPARG</i>	G	2×10^{-36} 1×10^{-8}	GCST009379 GCST005047	T2D
				A	5×10^{-11}	GCST008413	AML
rs1801282	3:12351626	3p25.2	<i>PPARG</i>	C	3×10^{-19} 1×10^{-17}	GCST007516 GCST007515	T2D
					1×10^{-12} 5×10^{-12}	GCST005047 GCST007517	
				G	2×10^{-14} 2×10^{-19}	GCST004894 GCST004894	AML
					5×10^{-11}	GCST008413	
rs6685701	1:26542148	1p36.11	<i>RPS6KA1</i>	G	6×10^{-18} 1×10^{-8}	GCST008413 GCST010555	T2D
				A	1×10^{-10}	GCST008413	AML
rs11108094	12:95534337	12q22	<i>USP44</i>	C	1×10^{-10} 1×10^{-10}	GCST010557 GCST010555	T2D
					2×10^{-10}	GCST008413	AML
rs7929543	11:49329474	11p11.12	<i>AC118942.1</i>	C	2×10^{-9}	GCST006867	T2D
				A	7×10^{-9} 6×10^{-6}	GCST008413 GCST008413	AML

To investigate whether these genetic variants affect the expression levels of associated or other genes in disease-related tissues (adipose, skeletal muscle, liver, pancreas, whole blood), we searched for eQTLs through the GTex and Blood eQTL Browser databases [21,22]. All results obtained are reported in Table 2. Moreover, graphical data from the GTex portal are shown in Figure 1B; corresponding data from Blood eQTL Browser were not available. Rs11709077 (allele: G/A; minor allele: A) and rs1801282 (G/C; minor: G), on the *PPARG* gene, were found to affect the mRNA expression levels of *SYN2* (synapsin II) in the skeletal muscle (Figure 1B and Table 2) and whole blood (Table 2). In the skeletal muscle, the presence of the minor alleles correlates with increased *SYN2* expression (normalized effect size (NES): 0.35 and 0.36 for rs11709077 and rs1801282, respectively) (Figure 1B and Table 2), whereas in the whole blood, they are correlated with decreased levels (z-score: -3.61 , for both) (Table 2). In addition, rs1801282 was found to negatively impact the expression of the *GATA3* transcription factor in whole blood (z-score = -4.54) (Table 2) and of *TIMP4* (TIMP metalloproteinase inhibitor 4) (NES = -0.21) in visceral adipose tissue (Figure 1B and Table 2). The rs11108094 variant (C/A; minor allele: A) on *USP44* was associated with decreased expression of *METAP2* (methionine aminopeptidase 2) in subcutaneous and visceral adipose tissue (NES: -0.64 and -0.55 , respectively) (Figure 1B and Table 2). Finally, in visceral adipose tissue, rs6685701 (A/G; minor allele: G) in *RPS6KA1* negatively affects its own expression levels (NES: -0.099), while rs7929543 (A/C; minor allele: C) on *AC118942.1* is positively associated with the expression levels of *RP11-347H15.5* (clone-based (Vega) gene) (NES: 0.53) (Figure 1B and Table 2).

Table 2. eQTL associated with the five common disease susceptibility SNPs described in AML and/or T2D target tissues, as well as with their 64 proxies, as deposited in the GTEx project and Blood eQTL Browser. The SNP ID, SNP alleles, associated and affected genes and tissue(s), as well as corresponding *p*-values and the effect sizes, are reported.

SNP	Associated Gene	SNP Alleles	Affected Gene	Tissue	<i>p</i> -Value	Effect Size	Database
Five (5) common AML/T2D susceptibility SNPs							
rs11108094	USP44	C/A	METAP2	Subcutaneous adipose	9.50×10^{-8}	NES = −0.64	GTEx project
				Visceral adipose	2.50×10^{-6}	NES = −0.55	GTEx project
rs11709077	PPARG	G/A	SYN2	Whole blood	3.09×10^{-4}	Z-score = −3.61	Blood eQTL Browser
				Skeletal muscle	5.90×10^{-5}	NES = −0.21	GTEx project
rs1801282	PPARG	G/C	GATA3	Whole blood	5.70×10^{-6}	Z-score = −4.54	Blood eQTL browser
				SYN2	Whole blood	3.09×10^{-4}	Z-score = −3.61
			TIMP4	Skeletal muscle	2.10×10^{-8}	NES = 0.36	GTEx project
rs6685701	RPS6KA1	A/G	RPS6KA1	Visceral adipose	1.10×10^{-4}	NES = −0.099	GTEx project
rs7929543	AC118942.1	A/C	RP11-347H15.5	Visceral adipose	9.10×10^{-8}	NES = 0.53	GTEx project
Sixty-four (64) proxies of the five common AML/T2D susceptibility SNPs							
rs10839264	FOLH1, AC118942.1	C/T	RP11-347H15.5	Visceral adipose	7.90×10^{-8}	NES = 0.51	GTEx project
rs10859889	USP44, METAP2	A/T	METAP2	Subcutaneous adipose	5.20×10^{-8}	NES = −0.65	GTEx project
				Visceral adipose	2.30×10^{-6}	NES = −0.54	
rs11040352	FOLH1, AC118942.1	A/C	RP11-347H15.5	Visceral adipose	5.10×10^{-13}	NES = 0.69	GTEx project
rs11040365	FOLH1, AC118942.1	C/A	RP11-347H15.5	Visceral adipose	1.40×10^{-11}	NES = 0.65	GTEx project
rs11108070	USP44	T/A	METAP2	Subcutaneous adipose	5.20×10^{-8}	NES = −0.65	GTEx project
				Visceral adipose	2.30×10^{-6}	NES = −0.54	
rs11108072	USP44, METAP2	T/C	METAP2	Subcutaneous adipose	5.20×10^{-8}	NES = −0.65	GTEx project
				Visceral adipose	2.30×10^{-6}	NES = −0.54	
rs11108076	USP44, METAP2	G/A	METAP2	Subcutaneous adipose	5.20×10^{-8}	NES = −0.65	GTEx project
				Visceral adipose	2.30×10^{-6}	NES = −0.54	
rs11108079	USP44, METAP2	G/A	METAP2	Subcutaneous adipose	5.20×10^{-8}	NES = −0.65	GTEx project
				Visceral adipose	2.30×10^{-8}	NES = −0.54	
rs11108086	USP44	T/C	METAP2	Subcutaneous adipose	5.20×10^{-8}	NES = −0.65	GTEx project
				Visceral adipose	1.60×10^{-6}	NES = −0.56	
rs11108087	USP44	A/G	METAP2	Subcutaneous adipose	9.50×10^{-8}	NES = −0.64	GTEx project
				Visceral adipose	1.70×10^{-6}	NES = −0.56	
rs11519597	USP44, METAP2	T/C	METAP2	Subcutaneous adipose	5.20×10^{-8}	NES = −0.65	GTEx project
				Visceral adipose	2.30×10^{-6}	NES = −0.54	
rs11522874	USP44, METAP2	G/A	METAP2	Subcutaneous adipose	5.20×10^{-8}	NES = −0.65	GTEx project
				Visceral adipose	2.30×10^{-6}	NES = −0.54	
rs11580180	RPS6KA1	A/G	RPS6KA1	Visceral adipose	1.40×10^{-4}	NES = 0.098	GTEx project
rs11603576	FOLH1, AC118942.1	G/A	RP11-347H15.5	Visceral adipose	9.10×10^{-8}	NES = 0.53	GTEx project
rs11607791	FOLH1, AC118942.1	T/C	RP11-347H15.5	Visceral adipose	7.90×10^{-8}	NES = 0.51	GTEx project
rs11709077	PPARG	G/A	SYN2	Whole blood	3.09×10^{-4}	Z-score = −3.61	Blood eQTL Browser
				Skeletal muscle	4.60×10^{-9}	NES = 0.35	GTEx project
rs11712037	PPARG, TIMP4	C/G	TIMP4	Visceral adipose	7.30×10^{-5}	NES = −0.21	GTEx project
				Skeletal muscle	2.20×10^{-9}	NES = 0.35	
rs12146719	USP44, METAP2	C/A	METAP2	Subcutaneous adipose	5.20×10^{-8}	NES = −0.65	GTEx project
				Visceral adipose	2.30×10^{-6}	NES = −0.54	

Table 2. Cont.

SNP	Associated Gene	SNP Alleles	Affected Gene	Tissue	p-Value	Effect Size	Database
Sixty-four (64) proxies of the five common AML/T2D susceptibility SNPs							
rs12369757	USP44	G/A	METAP2	Subcutaneous adipose	5.20×10^{-8}	NES = -0.65	GTEx project
				Visceral adipose	2.30×10^{-6}	NES = -0.54	
rs13064760	PPARG	T/C	SYN2	Whole blood	2.55×10^{-4}	Z-score = -3.66	Blood eQTL Browser
				Skeletal muscle	4.10×10^{-9}	NES = 0.35	GTEx project
				Visceral adipose	7.50×10^{-5}	NES = -0.21	GTEx project
rs13083375	PPARG	G/T	SYN2	Skeletal muscle	4.10×10^{-9}	NES = 0.35	GTEx project
				Visceral adipose	7.50×10^{-5}	NES = -0.21	
rs143400372	USP44	G/GA	METAP2	Subcutaneous adipose	9.50×10^{-8}	NES = -0.64	GTEx project
				Visceral adipose	2.50×10^{-6}	NES = -0.55	
rs150732434	PPARG, TIMP4	TG/T	TIMP4	Visceral adipose	7.50×10^{-5}	NES = -0.21	GTEx project
				Skeletal muscle	4.10×10^{-9}	NES = 0.35	
rs17036160	PPARG, TIMP4	C/T	TIMP4	Visceral adipose	8.50×10^{-5}	NES = -0.21	GTEx project
				Skeletal muscle	6.50×10^{-9}	NES = 0.34	
rs1801282	PPARG	G/C	SYN2	Whole blood	3.09×10^{-4}	Z-score = -3.61	Blood eQTL Browser
				Skeletal muscle	2.10×10^{-8}	NES = 0.36	GTEx project
				Visceral adipose	5.90×10^{-5}	NES = -0.21	
rs1843628	FOLH1, AC118942.1	A/G	RP11-347H15.5	Visceral adipose	3.40×10^{-9}	NES = -0.55	GTEx project
rs1880436	FOLH1, AC118942.1	A/G	RP11-347H15.5	Visceral adipose	2.70×10^{-9}	NES = 0.55	GTEx project
rs2012444	PPARG	C/T	SYN2	Skeletal muscle	4.10×10^{-9}	NES = 0.35	GTEx project
				Visceral adipose	7.50×10^{-5}	NES = -0.21	
rs2278978	RPS6KA1	G/A	RPS6KA1	Whole blood	1.96×10^{-4}	Z-score = -3.72	Blood eQTL Browser
				Whole blood	2.41×10^{-3}	Z-score = -3.03	Blood eQTL Browser
rs2305293	USP44, METAP2	C/T	METAP2	Subcutaneous adipose	5.20×10^{-8}	NES = -0.65	GTEx project
				Visceral adipose	2.30×10^{-6}	NES = -0.54	
rs35000407	PPARG, TIMP4	T/G	TIMP4	Visceral adipose	7.50×10^{-5}	NES = -0.21	GTEx project
				Skeletal muscle	4.60×10^{-9}	NES = 0.35	
rs35788455	PPARG	CTTG/C	SYN2	Skeletal muscle	1.80×10^{-9}	NES = 0.36	GTEx project
				Visceral adipose	8.20×10^{-5}	NES = -0.21	
rs4443935	RPS6KA1	G/A	RPS6KA1	Whole blood	2.45×10^{-4}	Z-score = -3.67	Blood eQTL Browser
rs4684847	USP44, METAP2	C/T	TIMP4	Visceral adipose	8.20×10^{-5}	NES = -0.21	GTEx project
				Skeletal muscle	1.80×10^{-9}	NES = 0.36	
rs4762563	USP44, METAP2	G/C	METAP2	Subcutaneous adipose	5.20×10^{-8}	NES = -0.65	GTEx project
				Visceral adipose	2.30×10^{-6}	NES = -0.54	
rs61939476	USP44, METAP2	A/C	METAP2	Subcutaneous adipose	5.20×10^{-8}	NES = -0.65	GTEx project
				Visceral adipose	2.30×10^{-6}	NES = -0.54	
rs61939479	USP44, METAP2	C/T	METAP2	Subcutaneous adipose	5.20×10^{-8}	NES = -0.65	GTEx project
				Visceral adipose	1.60×10^{-6}	NES = -0.54	
rs61939481	USP44	T/C	METAP2	Subcutaneous adipose	9.50×10^{-8}	NES = -0.64	GTEx project
				Visceral adipose	6.40×10^{-6}	NES = -0.52	
rs71304101	PPARG, TIMP4	G/A	TIMP4	Visceral adipose	5.80×10^{-5}	NES = -0.21	GTEx project
				Skeletal muscle	9.30×10^{-10}	NES = 0.36	

Table 2. Cont.

SNP	Associated Gene	SNP Alleles	Affected Gene	Tissue	p-Value	Effect Size	Database
Sixty-four (64) proxies of the five common AML/T2D susceptibility SNPs							
rs737465	<i>RPS6KA1</i>	C/T	<i>DHDDS</i>	Whole blood	1.88×10^{-3}	Z-score = -3.11	Blood eQTL Browser
			<i>RPS6KA1</i>	Whole blood	2.04×10^{-4}	Z-score = -3.71	Blood eQTL Browser
				Visceral adipose	1.40×10^{-4}	NES = 0.098	GTEx project
rs75781920	<i>FOLH1</i> , <i>AC118942.1</i>	T/G	<i>RP11-347H15.5</i>	Visceral adipose	2.70×10^{-9}	NES = 0.55	GTEx project
rs76218798	<i>FOLH1</i> , <i>AC118942.1</i>	T/C	<i>RP11-347H15.5</i>	Visceral adipose	7.90×10^{-8}	NES = 0.51	GTEx project
rs76427006	<i>FOLH1</i> , <i>AC118942.1</i>	T/A	<i>RP11-347H15.5</i>	Visceral adipose	2.70×10^{-9}	NES = 0.55	GTEx project
rs79067108	<i>USP44</i>	GCT/G	<i>METAP2</i>	Subcutaneous adipose	5.20×10^{-8}	NES = -0.65	GTEx project
				Visceral adipose	2.30×10^{-6}	NES = -0.54	

2.2. Proxy SNPs of the Five Common AML/T2D Susceptibility SNPs

Apart from the SNPs directly identified to be associated with a disease, other co-inherited SNPs may also lead to its development [23]. Based on this, we searched for the proxy SNPs of the five common AML/T2D susceptibility SNPs, utilizing the LDLink tool [24]. The selection criterion for a proxy SNP was to possess a squared correlation measure (R^2) of LD greater than 0.8. Data are shown in Figure 2 and Table 3. Sixty-six (66) unique proxy SNPs that lie in the *USP44*, *METAP2*, *PPARG*, *TIMP4*, *FOLH1* (folate hydrolase 1), *AC118942.1* and *RPS6KA1* genes were identified; some of them were detected as proxies for more than one of the five common SNPs. Through this analysis, it was also revealed that two of the common AML/T2D susceptibility genes (rs1801282, rs11709077) on the *PPARG* gene were mutual proxy SNPs (Table 3; bold/italics highlighted). Moreover, Venn diagram analysis revealed that one of the 64 SNPs (rs11519597) is an AML-specific disease susceptibility SNP, while two of them (rs71304101, rs17036160) are T2D-specific disease susceptibility SNPs (data not shown).

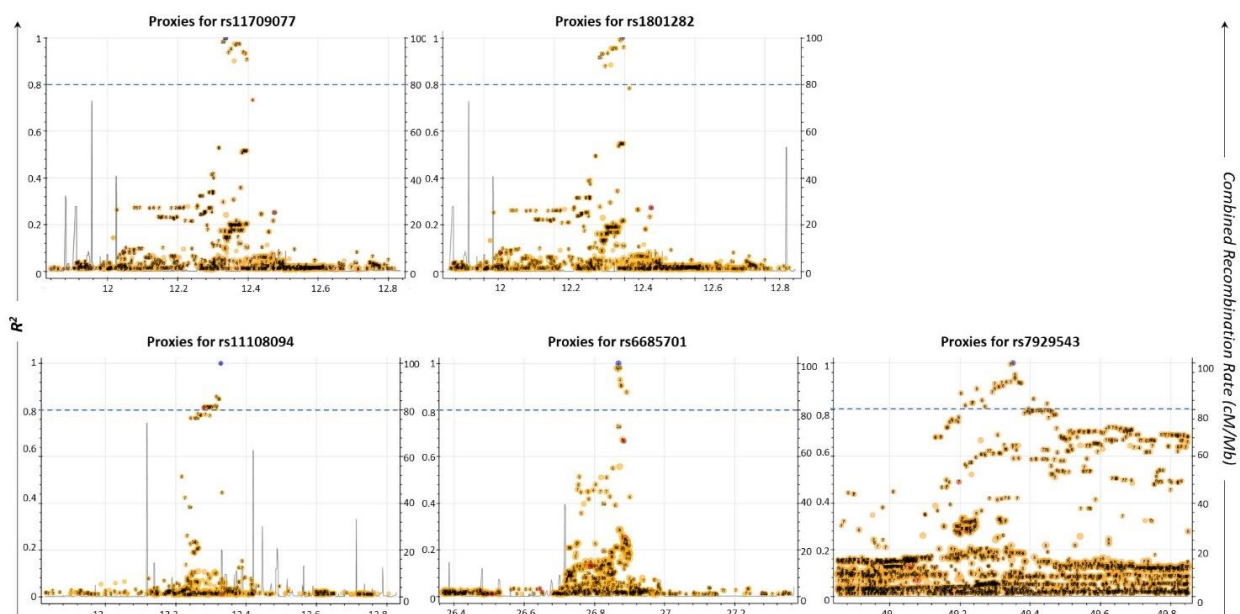


Figure 2. Regional LD plots of five commonly associated SNPs generated using the LDLink web tool (May 2021). Each dot represents the pairwise LD level between two individual SNPs. X-axis depicts the chromosomal coordinates. Left y-axis represents the pairwise R^2 value with the query variant; R^2 threshold greater than or equal to 0.8 was considered as a cut-off for selected proxies (blue dashed line). Right y-axis indicates the combined recombination rate (cM/Mb) from HapMap. Recombination rate is the rate at which the association between the two loci is changed. It combines the genetic (cM) and physical positions (Mb) of the marker by an interactive plot.

Table 3. Summary of the proxy SNPs ($R^2 \geq 0.8$) for each common AML/T2D susceptibility SNP, along with their chromosomal location, correlated alleles and associated genes, as collected from LDLink tool [24] (May 2021).

	Proxy SNPs	Chr	Position	Alleles	R^2	Correlated Alleles	Associated Genes
rs11709077	rs17036160	3	12329783	(C/T)	0.9844	G = C,A = T	PPARG
	rs2012444	3	12375956	(C/T)	0.9751	G = C,A = T	
	rs13064760	3	12369401	(C/T)	0.9751	G = C,A = T	
	rs150732434	3	12360884	(G/-)	0.9751	G = G,A = -	
	rs13083375	3	12365308	(G/T)	0.972	G = G,A = T	
	rs35000407	3	12351521	(T/G)	0.9539	G = T,A = G	
	rs4684847	3	12386337	(C/T)	0.9391	G = C,A = T	
	rs11712037	3	12344730	(C/G)	0.9379	G = C,A = G	
	rs35788455	3	12388908	(TTG/-)	0.9362	G = TTG,A = -	
	rs1801282	3	12393125	(C/G)	0.9334	G = C,A = G	
	rs71304101	3	12396913	(G/A)	0.9083	G = G,A = A	
rs35408322	3	12360357	(-/T)	0.9021	G = -,A = T		
rs1801282	rs4684847	3	12386337	(C/T)	0.9939	C = C,G = T	PPARG, TIMP4
	rs35788455	3	12388908	(TTG/-)	0.9908	C = TTG,G = -	
	rs71304101	3	12396913	(G/A)	0.9613	C = G,G = A	
	rs150732434	3	12360884	(G/-)	0.9573	C = G,G = -	
	rs13064760	3	12369401	(C/T)	0.9573	C = C,G = T	
	rs2012444	3	12375956	(C/T)	0.9573	C = C,G = T	
	rs13083375	3	12365308	(G/T)	0.9543	C = G,G = T	
	rs35000407	3	12351521	(T/G)	0.9365	C = T,G = G	
	rs11709077	3	12336507	(G/A)	0.9334	C = G,G = A	
	rs17036160	3	12329783	(C/T)	0.9183	C = C,G = T	
	rs35408322	3	12360357	(-/T)	0.8855	C = -,G = T	
rs11712037	3	12344730	(C/G)	0.8806	C = C,G = G		
rs6685701	rs4970486	1	26871669	(C/T)	0.9826	A = C,G = T	RPS6KA1
	rs737465	1	26862939	(T/C)	0.9814	A = T,G = C	
	rs11580180	1	26867453	(A/G)	0.9746	A = A,G = G	
	rs2278978	1	26873245	(A/G)	0.9311	A = A,G = G	
	rs4443935	1	26875433	(A/G)	0.9072	A = A,G = G	
	rs10902750	1	26876245	(G/T)	0.9052	A = G,G = T	
	rs389548	1	26891697	(C/A)	0.8777	A = C,G = A	
rs11108094	rs11108087	12	95915763	(A/G)	0.8578	C = A,A = G	USP44, METAP2
	rs61939481	12	95921998	(T/C)	0.8477	C = T,A = C	
	rs143400372	12	95923620	(-/A)	0.8477	C = -,A = A	
	rs11108086	12	95914758	(T/C)	0.8187	C = T,A = C	
	rs79067108	12	95881761	(CT/-)	0.8141	C = CT,A = -	
	rs11108070	12	95881787	(T/A)	0.8141	C = T,A = A	
	rs12369757	12	95888603	(G/A)	0.8141	C = G,A = A	
	rs11108072	12	95890218	(T/C)	0.8141	C = T,A = C	
	rs10859889	12	95890413	(A/T)	0.8141	C = A,A = T	
	rs11522874	12	95893609	(G/A)	0.8141	C = G,A = A	
	rs61939476	12	95894581	(A/C)	0.8141	C = A,A = C	
	rs11108076	12	95897348	(G/A)	0.8141	C = G,A = A	
	rs11108079	12	95899173	(G/A)	0.8141	C = G,A = A	
	rs12146719	12	95901434	(C/A)	0.8141	C = C,A = A	
	rs61939479	12	95905364	(C/T)	0.8141	C = C,A = T	
	rs2305293	12	95879734	(C/T)	0.8095	C = C,A = T	
	rs11519597	12	95894247	(T/C)	0.8095	C = T,A = C	
	rs61939477	12	95896692	(A/G)	0.8095	C = A,A = G	
	rs4762563	12	95915341	(G/C)	0.805	C = G,A = C	

Table 3. Cont.

Proxy SNPs	Chr	Position	Alleles	R ²	Correlated Alleles	Associated Genes	
rs11603576	11	49344126	(G/A)	0.9947	A = G,C = A		
rs10839264	11	49356806	(C/T)	0.9511	A = C,C = T		
rs76218798	11	49356186	(T/C)	0.9366	A = T,C = C		
rs11607791	11	49358347	(T/C)	0.9339	A = T,C = C		
rs1880436	11	49344775	(A/G)	0.92	A = A,C = G		
rs148517532	11	49332611	(A/G)	0.9188	A = A,C = G		
rs144550850	11	49366641	(T/C)	0.9175	A = T,C = C		
rs1843629	11	49319195	(G/A)	0.9161	A = G,C = A		
rs75781920	11	49371482	(T/G)	0.9152	A = T,C = G		
rs76427006	11	49375021	(T/A)	0.9149	A = T,C = A		
rs7932396	11	49299282	(A/G)	0.9112	A = A,C = G		
rs1843628	11	49323039	(A/G)	0.9033	A = A,C = G		
rs7939300	11	49311134	(C/A)	0.8985	A = C,C = A		
rs7929543	rs7939316	11	49311208	(A/G)	0.8985	A = A,C = G	FOLH1, AC118942.1
rs11040313	11	49299786	(A/G)	0.8915	A = A,C = G		
rs11040291	11	49248150	(C/T)	0.8898	A = C,C = T		
rs61350355	11	49292311	(G/A)	0.8757	A = G,C = A		
rs16906190	11	49203487	(A/G)	0.8709	A = A,C = G		
rs11040354	11	49409798	(G/A)	0.847	A = G,C = A		
rs10839244	11	49263085	(A/G)	0.8406	A = A,C = G		
rs74380550	11	49236977	(C/T)	0.8301	A = C,C = T		
rs59386222	11	49235409	(G/A)	0.8288	A = G,C = A		
rs4091958	11	49234514	(T/C)	0.8286	A = T,C = C		
rs11040365	11	49448078	(C/A)	0.826	A = C,C = A		
rs10839237	11	49215635	(C/T)	0.8187	A = C,C = T		
rs76002284	11	49271829	(A/G)	0.8145	A = A,C = G		
rs11040352	11	49395272	(A/C)	0.8039	A = A,C = C		

Furthermore, to pinpoint possible deregulation at the mRNA levels, attributed to the 64 proxy SNPs, we performed analysis using the GTex and Blood eQTL databases for the identification of eQTLs in disease-affected tissues (Table 2).

2.3. Common Susceptibility Genes in AML and T2D

Beyond the identification of specific genetic variants associated with both AML and T2D, we proceeded to the detection of common susceptibility genes between the two disorders. Analysis using combined data from the GWAS Catalog and the GTex portal showed that 86 genes bear SNPs that have been significantly associated with the development of both diseases, as per GWAS performed (Figure 3A). These include the five genes with common SNPs and another 81 disease-specific genes. Notably, most of the genes contain a significantly higher number of SNPs associated with AML compared to T2D (Table 4).

To investigate whether these genes comprise eGenes, which have at least one eQTL located near the gene of origin (*cis*-eQTL) acting upon them, affected by AML or T2D-specific SNPs in-disease target tissues, we searched through the GTex and eQTL Browsers. Analysis using Venn diagrams identified AML- or T2D-specific SNPs/eQTLs in certain susceptibility genes in adipose, muscle tissue, liver, pancreas and/or whole blood (Figure 3B). In adipose tissue, 6517 eQTLs on common AML/T2D susceptibility genes were detected, of which 79 were AML- and 8 T2D-specific. In skeletal muscle, 4220 were identified—28 AML- and 5 T2D-specific. In liver, 602 were detected—seven AML- and none T2D-specific. In pancreas, 3507 were found—36 AML- and 5 T2D-specific. Finally, in whole blood, 7187 were identified—55 AML- and 10 T2D-specific. A complementary analysis of the same data revealed the distribution of the AML- or T2D- SNPs/eQTLs in disease-target tissues and identified common and tissue-specific ones (Figure 3C and Table 5). All identified eQTLs affecting the 86 common disease susceptibility genes are included in Supplementary Table S2.

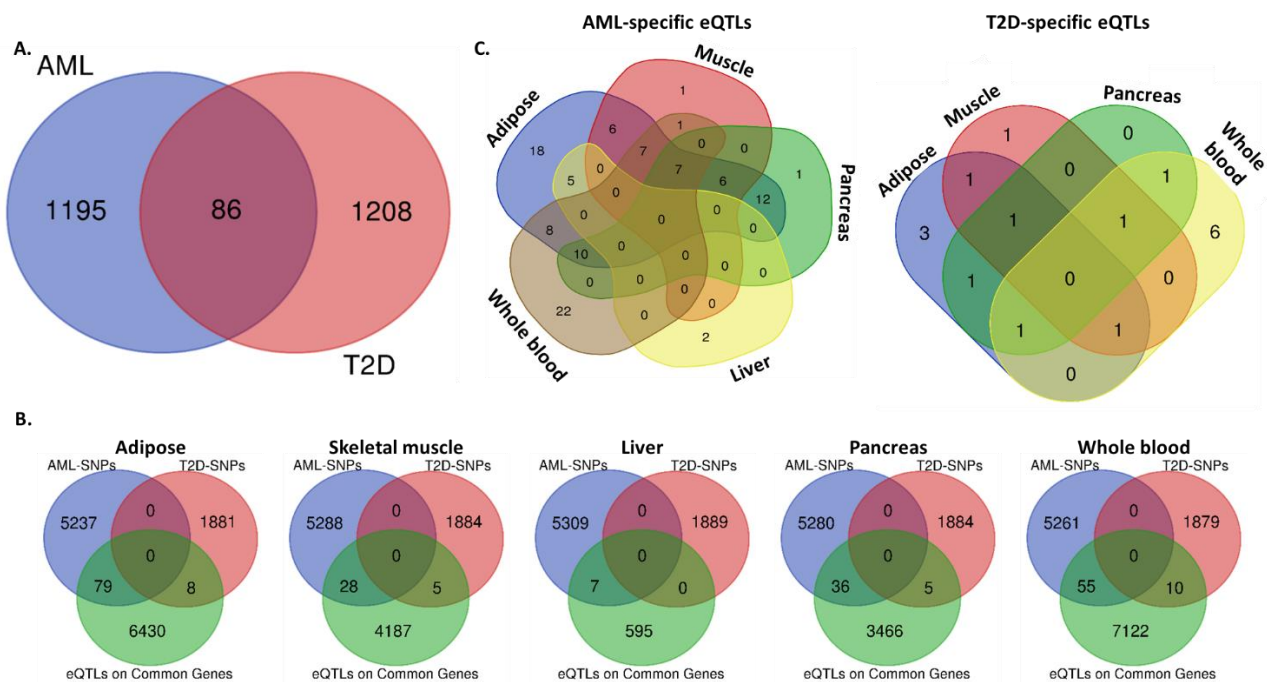


Figure 3. Common and disease-specific SNPs and eQTLs per target tissue. Venn diagrams reporting: (A) the number of common and disease-specific susceptibility genes between AML and T2D, (B) the numbers of AML- or T2D-specific SNPs that act as eQTLs upon the expression of common AML/T2D susceptibility genes, in adipose, skeletal muscle, liver, pancreas and whole blood, (C) the number of tissue-specific and common AML- or T2D- SNPs. Analysis was performed combining data from the NHGRI-EBI Catalog of GWAS and GTex portal.

Table 4. Common genes with common or different disease susceptibility SNPs for AML and T2D, as analyzed using data downloaded from the NHGRI-EBI Catalog of human GWAS [3] (May 2021).

	Gene Symbol	Full Gene Name	AML SNPs	T2D SNPs
1	AC003681.1	-	rs3788418, rs12627929, rs8139217, rs7285751, rs737903, rs36600, rs5752972, rs11090584, rs36608, rs5763609, rs39713, rs2051764, rs9614125, rs9625870, rs737904, rs737911, rs41170, rs5763681, rs36605, rs41158, rs4823058, rs41164, rs3788421, rs713718, rs5763559, rs737909, rs41159, rs3788425, rs5763688, rs7284538, rs5997546	rs41278853
2	AC006041.1	-	rs13225661, rs10242655, rs12113983, rs17348974, rs7811500, rs12532826, rs17169090, rs10950583	rs38221
3	AC010967.1	-	rs10204358, rs903230, rs745685, rs17044784, rs9677678, rs985549, rs903229, rs17044786, rs903231, rs17044787	rs9309245
4	AC016903.2	-	rs1545378	rs4482463
5	AC022414.1	-	rs10942819, rs10061629, rs6453303, rs11750661, rs17671389, rs9293712, rs9784696, rs6453304	rs7732130, rs4457053, rs6878122
6	AC022784.1	-	rs17656706, rs330003, rs6984551, rs11777846, rs75527, rs17149618, rs330035, rs330033, rs17656431, rs735449	rs17662402
7	AC034195.1	-	rs11717189, rs6768756	rs9842137
8	AC069157.2	-	rs10204358, rs903230, rs745685, rs17044784, rs9677678, rs985549, rs903229, rs17044786, rs903231, rs17044787	rs9309245
9	AC073176.2	-	rs950718	rs827237
10	AC087311.2	-	rs12227331, rs11052394	rs10844518, rs10844519
11	AC093675.1	-	rs4567941	rs34589210

Table 4. Cont.

	Gene Symbol	Full Gene Name	AML SNPs	T2D SNPs
12	AC093898.1	-	rs1503886, rs1039539, rs7673064, rs7681205, rs11934728, rs2320289, rs1847400, rs11941617	rs2169033
13	AC097634.4	-	rs9844845, rs17653411, rs9840264	rs844215, rs853866
14	AC098588.2	-	rs11100859, rs2719340, rs6817612	rs200995462
15	AC098588.3	-	rs11100859, rs2719340, rs6817612	rs200995462, rs75686861
16	AC098650.1	-	rs6549877, rs1350867, rs2371341, rs6549876, rs4258916, rs1381392, rs1563981, rs6549878	rs9869477
17	AC114971.1	-	rs10067455	rs73167517
18	AC118942.1	-	rs10501324, rs7929543, rs7115281, rs3960835, rs1164681, rs1164673, rs1164666, rs10769572, rs12806588, rs2204366, rs7930322, rs2205020, rs11040338, rs11040339, rs10839257, rs7118379, rs598101, rs10839272, rs7925896, rs7924782, rs7114817, rs588295	rs7929543
19	AFF3	AF4/FMR2 Family Member 3	rs6707538, rs7423759, rs17023314, rs4449188, rs7577040, rs17436893	rs34506349
20	AL135878.1	-	rs10138733, rs4981687, rs8016028, rs8022374, rs1951540, rs17114593, rs3950100, rs8022457, rs8016946, rs17560052, rs8020665	rs8005994
21	AL135923.2	-	rs10815796, rs10815795, rs10815793	rs10758950
22	AL136114.1	-	rs2065140, rs1885645, rs3131325, rs1923640, rs2065141, rs10494504, rs1885644	rs532504, rs539515
23	AL136962.1	-	rs7552571	rs9316706
24	AL359922.1	-	rs10965197, rs2027938, rs10757261, rs9657608	rs1063192
25	AL391117.1	-	rs10811816, rs10811815, rs1350996	rs11793831, rs7029718
26	ASAH1	N-Acylsphingosine Amidohydrolase (Acid Ceramidase)	rs17692377, rs382752, rs11782529	rs34642578
27	AUTS2	Activator of Transcription and Developmental Regulator	rs7459368, rs7791651, rs2057913, rs1557970, rs4718971, rs3922333, rs1008584, rs11772435, rs17578487, rs2057914, rs2057911, rs10486866	rs2103132, rs6947395, rs6975279, rs12698877, rs10618080, rs610930
28	CACNA2D3	Calcium Voltage-Gated Channel Auxiliary Subunit Alpha2delta3	rs11711040, rs6805548	rs76263492
29	CHMP4B	Charged Multivesicular Body Protein 4B	rs2050209, rs6088343, rs2092475, rs17091328	rs7274168
30	CPNE4	Copine 4	rs3851353, rs1010900, rs17341291, rs1850941, rs16838814, rs3900591, rs9853646, rs16838856, rs10512856, rs12636272, rs6792708, rs11708369, rs1505811, rs4522813, rs3914303, rs2369466, rs3922808, rs10934990, rs9876304, rs7626343	rs9857204, rs1225052
31	CRTC1	CREB-regulated transcription coactivator 1	rs2023878, rs17757406, rs6510997, rs12462498, rs6510999, rs2240887, rs7256986	rs10404726
32	CSMD1	CUB and Sushi Multiple Domains 1	rs592700, rs11779410, rs13277378, rs4876060, rs596332, rs673465	rs117173251
33	DGKB	Diacylglycerol Kinase Beta	rs10244653, rs10486042, rs17167995	rs17168486, rs10281892, rs11980500
34	EIF2S2P7	Eukaryotic Translation Initiation Factor 2 Subunit Beta	rs2193632, rs6714162, rs2870503, rs768329	rs1116357
35	EML6	EMAP-Like 6	rs10496035, rs4625954, rs13394146	rs5010712
36	ERBB4	Erb-B2 Receptor Tyrosine Kinase 4	rs10207288, rs10174084, rs13019783, rs4673628, rs4423543, rs6759039	rs3828242, rs13005841

Table 4. Cont.

Gene Symbol	Full Gene Name	AML SNPs	T2D SNPs	
37	FAM86B3P	Family with sequence similarity 86, member A pseudogene	rs13274039, rs2980417, rs2945230, rs2980422, rs10095669, rs2980420	rs7841082
38	FSD2	Fibronectin type III and SPRY domain containing 2	rs4779064	rs36111056
39	GP2	Glycoprotein 2	rs8046269, rs12930599, rs11642182, rs9937721, rs4383154	rs117267808
40	GRID1	Glutamate Ionotropic Receptor Delta Type Subunit 1	rs1991426, rs4933387, rs7084960, rs1896526, rs17096224, rs11201974, rs1896527, rs1896525, rs7918205	rs11201999, rs11201992
41	GRK5	G Protein-Coupled Receptor Kinase 5	rs12357403, rs17606601, rs4752269, rs10787945, rs7903013, rs12264832, rs17098576, rs12358835, rs12244897, rs10886439, rs4752276, rs17098586, rs10510056	rs10886471
42	HPSE2	Heparanase 2	rs12219674, rs527822, rs592142, rs10748739, rs657442, rs537851, rs521390, rs10883130, rs650527, rs526877, rs7907389, rs551674, rs10509724, rs523205, rs10883134, rs558398, rs526698, rs2018085, rs17538604, rs621644, rs552644, rs489611, rs552436, rs625777, rs11189692, rs563937, rs660426, rs17459507, rs898892, rs541519	rs524903
43	KCNB2	Potassium Voltage-Gated Channel Subfamily B Member 2	rs2251899	rs349359
44	KCNQ1	Potassium Voltage-Gated Channel Subfamily Q Member 1	rs10832134, rs12576156, rs11523905	rs2283159, rs163184, rs2237896, rs2283228, rs2237897, rs2237892, rs2237895, rs231362, rs2283220, rs231361, rs231349, rs163182, rs233450, rs77402029, rs2106463, rs463924, rs231356, rs233449, rs8181588, rs234853
45	LCORL	Ligand-Dependent Nuclear Receptor Corepressor-Like	rs1503886, rs1039539, rs7673064, rs7681205, rs11934728, rs2320289, rs1847400, rs11941617	rs2169033, rs2011603
46	LDLRAD4	Low-Density Lipoprotein Receptor Class A Domain Containing 4	rs7241766, rs6505821, rs7230189, rs8091352, rs7230276	rs11662800
47	LHFPL3	LHFPL Tetraspan Subfamily Member 3	rs2106504, rs17136882, rs13234807, rs6958831, rs7794181, rs979522, rs7787976, rs7787988	rs73184014
48	LINC00424	Long Intergenic Non-Protein Coding RNA 424	rs9316684, rs7320437, rs9316683, rs17074792	rs9316706
49	LINC01234	Long Intergenic Non-Protein Coding RNA 1234	rs4766686, rs10850140	rs7307263
50	LINC02641	Long Intergenic Non-Protein Coding RNA 2641	rs845083, rs2282015, rs1219960, rs845084, rs11597044, rs7091877, rs6599698	rs705145
51	LINGO2	Leucine-Rich Repeat and Ig Domain Containing 2	rs1452338, rs10511822, rs1349638, rs10124164, rs16912518	rs1412234
52	MERTK	MER Proto-Oncogene, Tyrosine Kinase	rs11684476	rs34589210
53	MLIP	Muscular LMNA-Interacting Protein	rs9357785, rs1325831, rs16884633, rs12191362, rs9464019, rs1359563, rs1325833, rs9637973, rs7750294, rs9370259	rs9370243

Table 4. Cont.

Gene Symbol	Full Gene Name	AML SNPs	T2D SNPs
54	MTMR3 <i>Myotubularin-Related Protein 3</i>	rs3788418, rs12627929, rs8139217, rs7285751, rs737903, rs36600, rs5752972, rs11090584, rs36608, rs5763609, rs39713, rs2051764, rs9614125, rs9625870, rs737904, rs737911, rs411170, rs5763681, rs36605, rs41158, rs4823058, rs41164, rs3788421, rs713718, rs5763559, rs737909, rs41159, rs3788425, rs5763688, rs7284538, rs5997546	rs41278853
55	NELL1 <i>Neural EGFL-Like 1</i>	rs4412753, rs11025959, rs1377744, rs4923393, rs4576820, rs7119634, rs7948285, rs10500896, rs10833472, rs1945321	rs16907058
56	NFATC2 <i>Nuclear Factor of Activated T Cells 2</i>	rs17791950, rs4396773, rs4811167, rs6021170, rs1123479, rs959996	rs6021276
57	NLGN1 <i>Neuroigin 1</i>	rs9809489, rs6782940, rs16829698, rs1502461, rs6776485, rs16829573	rs686998, rs247975
58	OARD1 <i>O-Acyl-ADP-Ribose Deacylase 1</i>	rs6912013, rs9296355, rs7760860	rs7841082
59	PAM <i>Peptidylglycine Alpha-Amidating Monooxygenase</i>	rs888801, rs467186, rs258132, rs462957, rs458256, rs2657459, rs401114, rs438126, rs451819, rs442443, rs382964, rs382946, rs647343	rs78408340
60	PARD3B <i>Par-3 Family Cell Polarity Regulator Beta</i>	rs4673320, rs1990667, rs10179357, rs849207, rs16837235, rs907462, rs2160455, rs849250, rs12620034, rs10490293, rs10490292, rs4673324, rs4595957, rs4673329, rs2668152	rs4482463
61	PCSK6 <i>Proprotein convertase subtilisin/kexin type 6</i>	rs9806369, rs12905649, rs11858490, rs12719737, rs2047219, rs2047220, rs4965873, rs903552, rs11852310, rs11858491	rs6598475
62	PKHD1 <i>Polycystic kidney and hepatic disease 1</i>	rs1326570, rs41412044, rs9370050, rs728996, rs11754532, rs6458777, rs2104522, rs2894788, rs2397061, rs9474070, rs4715233, rs2104521, rs6922497, rs6940892-	rs1819564
63	POLR1D <i>RNA Polymerase I And III Subunit D</i>	rs12584838, rs9551373, rs531950, rs10492484, rs7337722, rs667374, rs12876263, rs12870355, rs17821569, rs9507915, rs634035, rs542610, rs6491221, rs12050009	rs9319382
64	PPARG <i>Peroxisome Proliferator Activated Receptor Gamma</i>	rs10517032, rs10517031, rs2324237, rs16874420, rs10020457, rs10517030, rs2324241	rs17036160
65	PPP2R2C <i>Protein Phosphatase 2 Regulatory Subunit B gamma</i>	rs11946417, rs4505896, rs4689469, rs6446507, rs10937739, rs11938118, rs4689011, rs4689462, rs4076293, rs7654321, rs4234751, rs4689465	rs35678078
66	PRAG1 <i>PEAK1 Related, Kinase-Activating Pseudokinase 1</i>	rs13274039, rs2980417, rs2945230, rs2980422, rs10095669, rs2980420	rs7841082
67	PTPRD <i>Protein Tyrosine Phosphatase Receptor Type D</i>	rs10815796, rs10815795, rs10815793	rs10758950, rs17584499
68	RBMS3 <i>RNA Binding Motif Single-Stranded Interacting Protein 3</i>	rs6549877, rs1350867, rs2371341, rs6549876, rs4258916, rs1381392, rs1563981, rs6549878	rs9869477
69	RELN <i>Reelin</i>	rs6961175, rs10235204, rs2106283, rs2106282, rs6465955, rs6955789, rs6465954	rs39328
70	RPL12P33 <i>Ribosomal protein L12 pseudogene 33</i>	rs10774577, rs6489785, rs7300612, rs7969196, rs11065341, rs2701179, rs868795	rs118074491
71	RPS6KA1 <i>Ribosomal Protein S6 Kinase A1</i>	rs3127011, rs12094989, rs12723046, rs6685701, rs1982525, rs11576300, rs4659444, rs6670311	rs6685701
72	RPTOR <i>Regulatory Associated Protein of MTOR Complex 1</i>	rs8065459, rs9915426, rs2333990, rs2589133, rs2138125, rs734338	rs11150745
73	RREB1 <i>Ras Responsive Element Binding Protein 1</i>	rs10458204, rs4960285, rs12196079, rs17142726, rs12197730, rs552188, rs7759330, rs3908470, rs6597246	rs9505085, rs9505097, rs9379084
74	SEPTIN9 <i>Septin 9</i>	rs8079522, rs1075457, rs3744069, rs9916143, rs312907, rs11658267, rs892961, rs566569, rs11650011, rs2411110	rs1656794

Table 4. Cont.

	Gene Symbol	Full Gene Name	AML SNPs	T2D SNPs
75	SGCG	Sarcoglycan Gamma	rs578196, rs501909, rs502068	rs9552911
76	SGCZ	Sarcoglycan Zeta	rs17608649, rs7826655, rs12547159, rs13278000	rs35753840, rs17294565
77	SHROOM3	Shroom Family Member 3	rs6848817, rs13151434, rs6810716, rs13105942, rs4241595, rs10050141, rs6854652	rs11723275, rs56281442
78	SLC39A11	Solute Carrier Family 39 Member 11	rs11077627, rs11077628, rs4530179, rs11658711	rs61736066
79	SYT10	Synaptotagmin 10	rs12227331, rs11052394	rs10844518, rs10844519
80	TMEM106B	Transmembrane Protein 106B	rs12537849, rs10237821, rs10269431, rs7794113	rs13237518
81	TMEM87B	Transmembrane Protein 87B	rs6713344, rs4848979, rs4848980	rs74677818
82	TTN	Titin	rs7604033, rs10497522, rs2291313, rs11902709, rs2291311, rs4894044, rs10497523, rs2054708, rs1484116, rs10171049, rs3754953, rs4471922, rs11895382, rs4894037, rs2291312, rs7600001	rs6715901
83	USP44	Ubiquitin-Specific Peptidase 44	rs3812813, rs10777699, rs2769444, rs7974458, rs10498964, rs301024, rs301003	rs2197973
84	XYLT1	Xylosyltransferase 1	rs4453460, rs4583225	rs551640889
85	ZFH3	Zinc Finger Homeobox 3	rs328398, rs328389, rs328317, rs328384, rs328395	rs6416749, rs1075855
86	ZNF800	Zinc Finger Protein 800	rs11563463, rs2285337, rs2285338, rs11563346, rs11563634	rs17866443

Table 5. AML- or T2D- specific SNPs that act as eQTLs on the 86 common AML/T2D susceptibility genes in a tissue-specific manner, as analyzed via the GTex portal [21] (May 2021).

AML-Specific			T2D-Specific		
SNP ID	Associated Gene	Affected Gene (s)	SNP ID	Associated Gene	Affected Gene (s)
Adipose, Muscle, Pancreas, Whole Blood					
1	rs1168446	AC093675.1, MERTK			MERTK (ad, pa, bl), TMEM87B (mu, bl)
2	rs4848980	TMEM87B			MERTK (pa, mu), TMEM87B (bl, ad)
3	rs5752972	ASCC2, MTMR3			MTMR3 (ad, bl, mu, pa)
4	rs11684321	MERTK			MERTK (pa, mu, ad, bl), TMEM87B (mu, ad, bl)
5	rs9625870	ASCC2, MTMR3			MTMR3 (ad, bl, pa)
6	rs4848979	TMEM87B			MERTK (pa, bl, mu, ad), TMEM87B (mu, pa, ad, bl)
7	rs1168446	AC093675.1, MERTK			MERTK (pa, mu, ad), TMEM87B (ad, pa, mu, bl)
Adipose, Muscle, Pancreas					
1	rs2769444	USP44	rs4382480	MFHAS1	FAM86B3P (ad, pa, mu), PRAG1 (ad), FAM85B (ad),
2	rs13274039	PRAG1, FAM86B3P			FAM86B3P (ad), FAM85B (ad)
3	rs301003	USP44			USP44 (pa, mu, ad)
4	rs301026	METAP2			USP44 (mu, pa, ad)
5	rs301024	USP44			USP44 (pa, ad)
6	rs301009	METAP2			USP44 (pa, mu, ad)

Table 5. Cont.

AML-Specific			T2D-Specific			
SNP ID	Associated Gene	Affected Gene (s)	SNP ID	Associated Gene	Affected Gene (s)	
Adipose, Muscle, Whole blood						
1	rs8139217	MTMR3, AC003681.1		rs7274168	CHMP4B	CHMP4B (bl, mu, ad)
2	rs737911	MTMR3, AC003681.1				MTMR3 (ad, bl, mu)
3	rs7285751	MTMR3, AC003681.1				MTMR3 (bl, mu, ad)
4	rs3788421	MTMR3, AC003681.1				MTMR3 (bl, mu, ad)
5	rs41158	HORMAD2-AS1, MTMR3, AC003681.1				MTMR3 (ad, bl, mu)
6	rs7284538	MTMR3, AC003681.1				MTMR3 (bl, ad, mu)
7	rs41170	HORMAD2-AS1, MTMR3, AC003681.1				MTMR3 (ad, bl, mu)
Adipose, Pancreas, Whole blood						
1	rs4261758	SPTBN1		rs34589210	AC093675.1, MERTK	MERTK (pa), TMEM87B (ad, bl)
2	rs4567941	AC093675.1				MERTK (pa, bl), TMEM87B (ad, pa, bl)
3	rs36605	MTMR3				MTMR3 (ad, bl, pa)
4	rs17039558	TDRP				EML6 (pa, ad, bl)
5	rs737904	MTMR3				MTMR3 (ad, bl, pa)
6	rs3811640	MERTK				MERTK (pa), TMEM87B (ad, bl)
7	rs6734445	SPTBN1				EML6 (pa, ad, bl)
8	rs36600	MTMR3				MTMR3(ad, bl, pa)
9	rs11904679	AC092839.1, SPTBN1				EML6 (pa, ad, bl)
10	rs6713344	TMEM87B				MERTK (pa, bl, ad), TMEM87B (ad, pa, bl)
Muscle, Pancreas, Whole blood						
1				rs13237518	TMEM106B	TMEM106B (bl, pa, mu)
Adipose, Muscle						
1	rs11563634	ZNF800		rs11723275	SHROOM3	SHROOM3 (mu, ad)
2	rs10937739	PPP2R2C				PPP2R2C (mu, ad)
3	rs2285338	ZNF800				ZNF800 (ad, mu)
4	rs11563346	ZNF800				ZNF800 (mu, ad)
5	rs4689465	PPP2R2C				PPP2R2C (ad, mu)
6	rs4689469	PPP2R2C				PPP2R2C (mu, ad)
Adipose, Pancreas						
1	rs11887259	MERTK		rs7841082	PRAG1, FAM86B3P	FAM86B3P (ad, pa), FAM85B (ad), PPP1R3B (pa)
2	rs6729826	SPTBN1				EML6 (ad)
3	rs4671956	AC092839.2, SPTBN1				EML6 (ad, pa)
4	rs4374383	MERTK				TMEM87B (ad), MERTK (pa, ad)
5	rs3811638	MERTK				TMEM87B (ad), MERTK (pa, ad)
6	rs2945230	PRAG1, FAM86B3P				FAM86B3P (ad, pa), FAM85B (ad)
7	rs13016942	SPTBN1				EML6 (ad, pa)
8	rs12104998	AC092839.1, SPTBN1				EML6 (ad, pa)
9	rs12105792	SPTBN1				EML6 (ad, pa)
10	rs1367295	AC092839.1, SPTBN1				EML6 (ad, pa)
11	rs11683409	MERTK				MERTK (ad, pa), TMEM87B (ad)
12	rs17344072	SPTBN1				EML6 (ad, pa)

Table 5. Cont.

AML-Specific			T2D-Specific		
SNP ID	Associated Gene	Affected Gene (s)	SNP ID	Associated Gene	Affected Gene (s)
Adipose, Liver					
1	rs4659444	DPPA2P2, HMG2N2			RPS6KA1 (li)
2	rs1359563	MLIP-AS1, MLIP			MLIP (ad, li)
3	rs12094989	DPPA2P2, RPS6KA1			RPS6KA1 (li, ad)
4	rs9637973	MLIP-AS1, MLIP			MLIP (li, ad)
5	rs1325831	MLIP-AS1, MLIP			MLIP (li, ad)
Adipose, Whole blood					
1	rs5997546	ASCC2, MTMR3			MTMR3 (ad)
2	rs5763688	MTMR3, AC003681.1			MTMR3 (ad, bl)
3	rs41159	HORMAD2-AS1, MTMR3, AC003681.1			MTMR3 (ad, bl)
4	rs634035	POLR1D			POLR1D (ad)
5	rs5763559	ASCC2, MTMR3			MTMR3 (ad, bl)
6	rs737909	MTMR3, AC003681.1			MTMR3 (ad, bl)
7	rs2051764	MTMR3			MTMR3 (bl)
8	rs667374	POLR1D			POLR1D (bl, ad)
Muscle, Whole blood					
1	rs382752	PCMI, ASAH1			ASAH1 (bl, mu)
Pancreas, Whole blood					
1			rs74677818	TMEM87B	TMEM87B (bl), MERTK (pa)
Adipose					
1	rs17821569	POLR1D		rs11201992	GRID1
2	rs12905649	PCSK6		rs56281442	SHROOM3
3	rs10883130	HPSE2		rs11201999	GRID1
4	rs12876263	POLR1D			GRID1 (ad)
5	rs898892	HPSE2			
6	rs7907389	HPSE2			
7	rs7337722	POLR1D			
8	rs737903	MTMR3			
9	rs10748739	HPSE2			
10	rs2980420	PRAG1, FAM86B3P			
11	rs650527	HPSE2			
12	rs7750294	MLIP-AS1, MLIP			MLIP (ad)
13	rs10883134	HPSE2			HPSE2 (ad)
14	rs2018085	HPSE2			HPSE2 (ad)
15	rs41164	HORMAD2-AS1, MTMR3, AC003681.1			MTMR3 (ad)
16	rs621644	HPSE2			HPSE2 (ad)
17	rs542610	POLR1D			POLR1D (ad)
18	rs489611	HPSE2			HPSE2 (ad)
Muscle					
1	rs4505896	PPP2R2C		rs11150745	RPTOR
					RPTOR (mu)
Pancreas					
1	rs9370050	PKHD1			PKHD1 (pa)
Liver					
1	rs12191362	MLIP-AS1, MLIP			MLIP (li)
2	rs16884633	MLIP-AS1, MLIP			MLIP (li)

Table 5. Cont.

AML-Specific			T2D-Specific		
SNP ID	Associated Gene	Affected Gene (s)	SNP ID	Associated Gene	Affected Gene (s)
Whole blood					
1	rs382964	PAM	rs115505614	GIN1	PAM (bl), PPIP5K2 (bl)
2	rs10179948	MERTK	rs35658696	PAM	PAM (bl), PPIP5K2 (bl)
3	rs382946	AC099487.2, PAM	rs75432112	AC011362.1	PAM (bl), PPIP5K2 (bl)
4	rs258132	PAM	rs9319382	AL136439.1, POLR1D	POLR1D (bl)
5	rs401114	PAM	rs610930	AUTS2	AUTS2 (bl)
6	rs442443	AC099487.2, PAM	rs7729395	PAM	PAM (bl), PPIP5K2 (bl)
7	rs462957	PAM			
8	rs6088343	CHMP4B, TPM3P2			CHMP4B (bl)
9	rs458256	PAM			PAM (bl), PPIP5K2 (bl)
10	rs451819	AC099487.2, PAM			PAM (bl)
11	rs17098576	GRK5			GRK5 (bl)
12	rs17692377	PCMI, ASAHI			ASAHI (bl)
13	rs10211152	MERTK			TMEM87B (bl), MERTK (bl)
14	rs12050009	POLR1D			POLR1D (bl)
15	rs11782529	PCMI, ASAHI			ASAHI (bl)
16	rs9551373	POLR1D			POLR1D (bl)
17	rs10095669	PRAG1, FAM86B3P			FAM86B3P (bl)
18	rs467186	PAM			PAM (bl)
19	rs6142044	PIGPP3, TPM3P2			CHMP4B (bl)
20	rs2657459	AC099487.2, PAM			PAM (bl), PPIP5K2 (bl)
21	rs438126	AC099487.2, PAM			PAM (bl), PPIP5K2 (bl)
22	rs647343	AC099487.2, PAM			PAM (bl), PPIP5K2 (bl)

ad: Adipose, bl: whole blood, li: liver, mu: muscle, pa: pancreas.

2.4. Pathway Analysis of the Proteins Encoded by the Common AML/T2D Susceptibility Genes

To investigate the possible involvement of the 86 common susceptibility genes in molecular networks correlated with both disorders, the developed gene/protein panel was further processed through the STRING and KEGG databases [25,26]. The following eGenes found to be affected by the five common susceptibility SNPs as well as by their proxies in disease-affected tissues were included in the analysis: *DHDDS* (Dehydrodichyl Diphosphate Synthase Subunit), *GATA3*, *METAP2*, *RP11-347H15.5*, *RPS6KA1*, *SYN2*, *TIMP4*. The corresponding protein–protein interaction (PPI) network is depicted in Figure 4A. Analysis revealed that numerous proteins of the above set are significantly involved in metabolic pathways, including pyrimidine, purine, choline metabolism, mTOR, AMPK, PI₃K-Akt and insulin signaling, as well as pathways deposited as related to AML (FDR < 0.05 for all) (Figure 4B and Table 6).

Differently colored nodes designate various genes/proteins involved in one or more pathways. Edges represent protein–protein associations—either known interactions, predicted interactions or other associations. All regulated pathways revealed in this analysis are included in Supplementary Table S3.

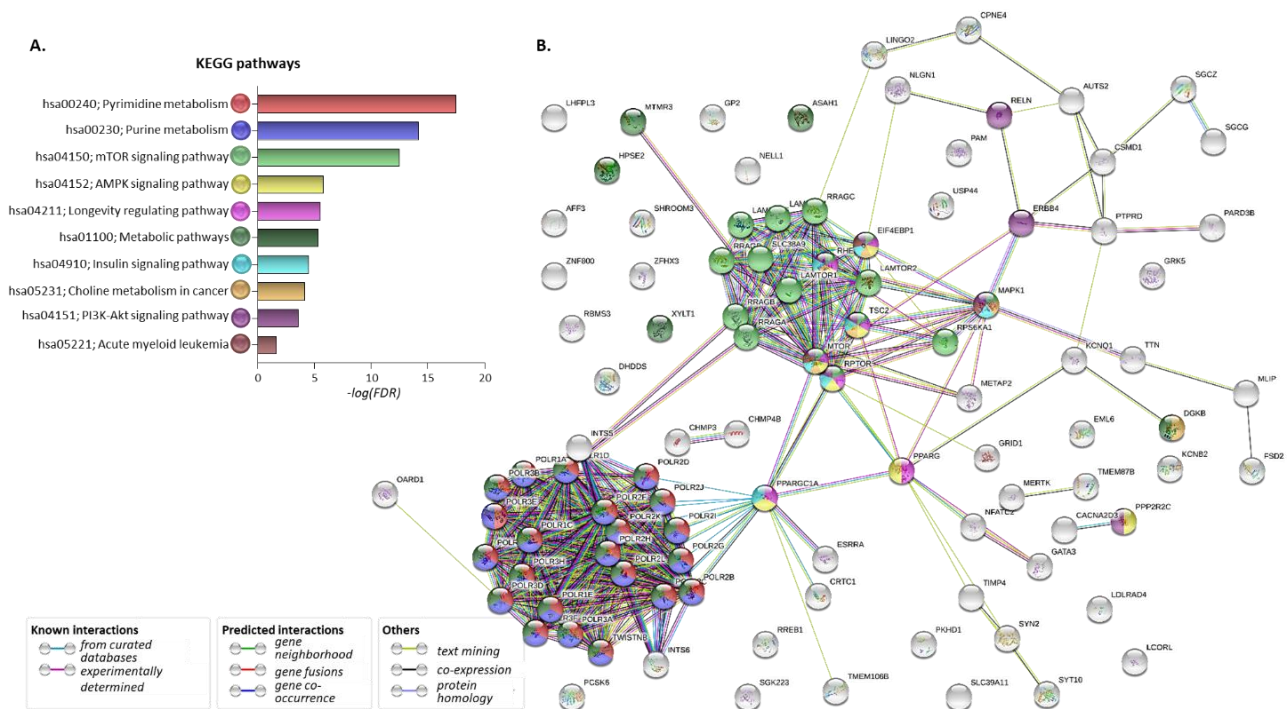


Figure 4. Pathways and protein–protein interactions regulated by the common AML/T2D-related genes. (A). Pathways enriched upon gene set analysis of 86 AML/T2D common susceptibility genes plus the seven eGenes affected by the five common AML/T2D susceptibility genes and their proxies, using KEGG database. (B). Protein–protein interaction (PPI) network developed upon processing the set in the STRING database. Different genes/proteins involved in different (one or more) pathways are designated by the differently colored nodes. Edges represent protein–protein associations—either known interactions, predicted interactions or other associations.

Table 6. Selected pathways significantly regulated by the set of 86 AML/T2D susceptibility genes plus seven eGenes affected by the five common AML/T2D susceptibility genes and their proxies, as analyzed upon processing in the STRING and KEGG databases [25,26]. Pathway IDs and description, number of susceptibility genes involved, number of background genes, their names as well as statistics (strength, FDR and log₁₀FDR) for each pathway are reported.

Term ID	Term Description	Observed Gene Count	Background Gene Count	Strength	FDR	log ₁₀ FDR	Matching Proteins in the Network
hsa00240	Pyrimidine metabolism	16	100	1.57	3.17 × 10 ⁻¹⁸	17.50	POLR2C, POLR2I, TWISTNB, POLR3B, POLR1A, POLR2D, POLR2J, POLR3E, POLR2G, POLR1D, POLR2L, POLR3C, POLR2K, POLR3H, POLR3A, POLR1C
hsa00230	Purine metabolism	16	173	1.33	6.30 × 10 ⁻¹⁵	14.20	POLR2C, POLR2I, TWISTNB, POLR3B, POLR1A, POLR2D, POLR2J, POLR3E, POLR2G, POLR1D, POLR2L, POLR3C, POLR2K, POLR3H, POLR3A, POLR1C
hsa04150	mTOR signaling pathway	14	148	1.34	3.30 × 10 ⁻¹³	12.48	MAPK1, TSC2, LAMTOR5, RHEB, RRAGB, LAMTOR1, RPTOR, EIF4EBP1, LAMTOR4, MTOR, LAMTOR2, RRAGD, RRAGC, RPS6KA1
hsa04152	AMPK signaling pathway	8	120	1.19	1.56 × 10 ⁻⁶	5.81	TSC2, RHEB, PPARGC1A, PPARG, RPTOR, PPP2R2C, EIF4EBP1, MTOR
hsa04211	Longevity regulating pathway	7	88	1.26	3.02 × 10 ⁻⁶	5.52	TSC2, RHEB, PPARGC1A, PPARG, RPTOR, EIF4EBP1, MTOR

Table 6. Cont.

Term ID	Term Description	Observed Gene Count	Background Gene Count	Strength	FDR	\log_{10} FDR	Matching Proteins in the Network
hsa01100	Metabolic pathways	20	1250	0.57	4.74×10^{-6}	5.32	POLR2C, POLR2I, TWISTNB, POLR3B, XYLT1, POLR1A, POLR2D, POLR2J, POLR2G, POLR1D, POLR2L, POLR3C, POLR2K, POLR3H, HPSE2, POLR3A, POLR1C, ASAH1, MTMR3, DGKB
hsa04910	Insulin signaling pathway	7	134	1.08	3.31×10^{-5}	4.48	MAPK1, TSC2, RHEB, PPARGC1A, RPTOR, EIF4EBP1, MTOR
hsa05231	Choline metabolism in cancer	6	98	1.15	6.93×10^{-5}	4.16	MAPK1, TSC2, RHEB, EIF4EBP1, MTOR, DGKB
hsa04151	PI ₃ K-Akt signaling pathway	9	348	0.77	2.60×10^{-3}	3.59	MAPK1, TSC2, RHEB, RPTOR, PPP2R2C, EIF4EBP1, ERBB4, MTOR, RELN
hsa05221	Acute myeloid leukemia	3	66	1.02	2.41×10^{-2}	1.62	MAPK1, EIF4EBP1, MTOR

2.5. Investigation of Aberrant mRNA Expression of T2D-Deregulated Genes in an AML Cohort

The second aim of the study was to investigate the possible deregulation of T2D-related metabolic mechanisms in AML patients. To this end, we selected a panel of genes previously reported to be deregulated in T2D patients [4] (*CAPN10*, *CDK5*, *CDKN2A*, *IGF2BP2*, *KCNQ1*, *THADA*, *TSPAN8*) and explored their mRNA levels in peripheral blood samples from AML- versus non-cancerous individuals utilizing RNAseq data and the TNMplot web tool [27]. Significantly increased mRNA levels of *CAPN10*, *CDK5*, *CDKN2A*, *IGF2BP2* and *THADA*, as well as significantly decreased levels of *KCNQ1* and *TSPAN8*, were found in 151 AML patients compared to 407 normal individuals tested (Mann-Whitney $p < 0.0004$ for all). The percentage (%) of AML samples that displayed up- or downregulated expression for each of the above genes, at each of the four quantile cut-off values (minimum, 1st quartile, median, 3rd quartile, maximum), as well as the specificity (the ratio of the number of AML samples to the sum of AML and non-cancerous samples over or below each given cut-off), are depicted in Figure 5.

To search for AML-specific SNPs on these deregulated genes, we used data obtained from the NHGRI-EBI Catalog of GWAS. It was found that rs10832134 (chromosomal location: 11:2481256), rs12576156 (11:2477588) and rs11523905 (11:2477029) variants lie in the *KCNQ1* ($p = 3 \times 10^{-15}$ for all), while the rest of the deregulated genes have not been identified to bear AML-related SNPs. Investigation for their proxies revealed three proxy SNPs (rs12574553, rs757092, rs7126330) for rs10832134 and five proxy SNPs (rs73419519, rs7937273, rs7928116, rs179395, rs7542142) for rs12576156, all of them in *KCNQ1*. No proxies were found for rs11523905 (data not shown). Out of these, the proxy SNP rs12574553 (allele C/T) consists of an eQTL for *KCNQ1*; the minor allele leads to the downregulation of mRNA levels in whole blood [21].

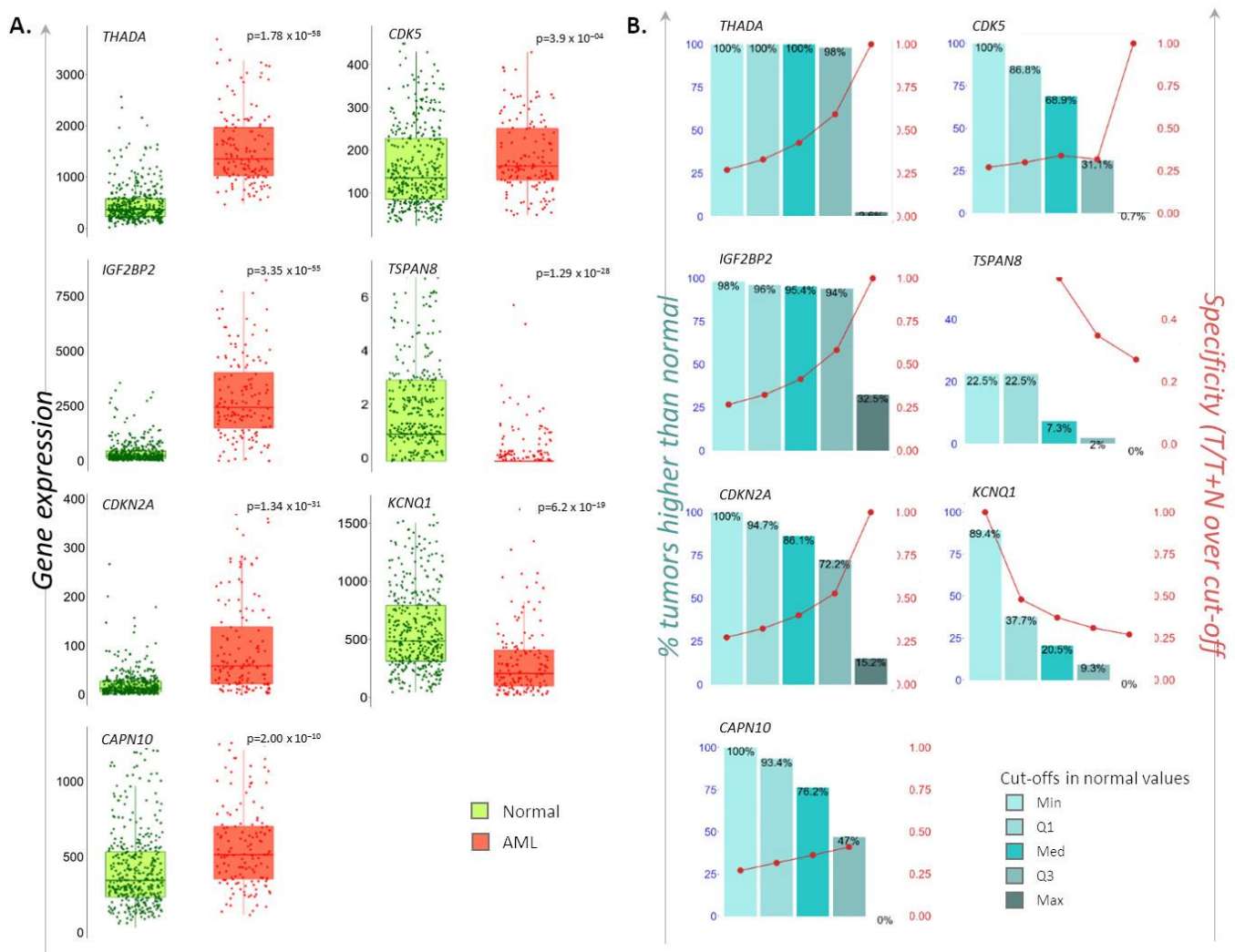


Figure 5. Differential expression levels of T2D-related genes in AML individuals. (A). Dot-plot/whisker bars depicting the differential mRNA levels of the *CAPN10*, *CDK5*, *CDKN2A*, *IGF2BP2*, *KCNQ1*, *THADA*, *TSPAN8* T2D susceptibility genes in AML patients. *p*-values of significance as obtained by Mann–Whitney test are reported. (B). Bar diagrams showing the: (i) percentage (%) of AML samples that possesses higher or lower of each gene-of-interest compared to non-cancerous samples, at each of the four quantile cut-off values (minimum, 1st quartile, median, 3rd quartile, maximum) (left y-axis), and (ii) specificity defined as the ratio of the number of AML samples to the sum of AML and non-cancerous samples over or below each given cut-off (right y-axis).

3. Discussion

Today, there is a well-accepted epidemiological link between T2D and cancer development [5]. However, in other types of human neoplasia, the association between T2D and hematological malignancies is less explored. Among them, AML represents one of the most intriguing morbidities for further investigation due to its increasing rates and relatively poor prognosis and response to treatment [10,28]. Accumulating clinical evidence connecting metabolic syndrome parameters (including BMI and T2D) to AML [9,11–16], together with corresponding in vitro data [17–19], highlights the need for investigation of the underlying mechanisms implicating genetic predisposition, which may regulate metabolic abnormalities.

In this study, we first aimed at the description of the possible common genetic background shared by the two disorders. Processing of the thousands of AML- and T2D-associated SNPs deposited in the GWAS NHGRI-EBI Catalog uncovered five SNPs that are significantly linked to both diseases (Table 1). Two of them (rs11709077, rs1801282) lie in

the *PPARG* gene, the first gene reproducibly associated with T2D [29,30]. The gene encodes for the PPAR- γ receptor, a molecular target of thiazolidinediones (insulin-sensitizing antidiabetic drugs); gene variants affecting its transcription levels in adipose tissue are associated with insulin sensitivity [29,30]. Although there are no data directly linking *PPARG* with AML, it is worth mentioning that the protein is implicated in the TGF-beta and mTOR signaling pathways, both associated with cancer development [31–33]. Our analyses also indicated that rs11709077 and rs1801282 on *PPARG* negatively affect the expression of *SYN2* (Synapsin II) in skeletal muscle and in whole blood (Table 2, Figure 1); however, there is not yet any evidence connecting *SYN2* with T2D or AML.

Another common SNP, which is a missense variant rs1801282, was found to negatively regulate the expression of the tissue inhibitor of metalloproteinases 4 (*TIMP4*) in visceral adipose tissue. The TIMP family has been associated with several cancers [34], but no information about its relation to T2D is available yet. Another interesting observation regards the negative impact of rs1801282 on *GATA3* in whole blood. *GATA3* is a transcription factor with a multi-faceted role in hematopoiesis [35], while related genetic and epigenetic aberrations are strongly associated with AML development, prognosis and response to therapy [36,37]. Regarding T2D, *GATA3* is considered an anti-adipogenic factor and a potential molecular therapeutic target for insulin resistance, through restoration of adipogenesis and amelioration of inflammation [38,39].

Rs6685701, located in the gene encoding for the ribosomal protein S6 kinase A1 (*RPS6KA1* or *P90S6K*), was found to be associated with its lower expression levels in visceral adipose tissue. The protein belongs to the family of serine/threonine kinases that govern various cellular processes, and it acts downstream of ERK (MAPK1/ERK2 and MAPK3/ERK1) signaling [33]. In murine models of T2D, *RPS6KA1* has been implicated in impaired glucose homeostasis in β -pancreatic, muscle and liver cells [40,41], which is improved upon sitagliptin (DPP-4 inhibitor; antidiabetic drug) administration [42]. Using an in vivo model of leukemia, *RPS6KA1* has been shown to promote the self-renewal of hematopoietic stem cells and disease progression through the regulation of the mTOR pathway [43]. More importantly, it was very recently reported that *RPS6KA1* may be a strong indicator of overall survival in AML patients, while aberrations in the miR-138-5p/*RPS6KA1* axis are associated with poor prognosis among patients [44].

The rs11108094 in *USP44* (ubiquitin-specific peptidase 44) was also recognized as a common susceptibility variant for AML and T2D, which acts as an eQTL downregulating the expression of *METAP2* (methionyl aminopeptidase 2) in subcutaneous and adipose tissue. The *USP44* protein is implicated in protein metabolism and ubiquitin-mediated proteasome-dependent proteolysis. More importantly, *METAP2* is involved in the metabolism of fat-soluble vitamins [33]. Its inhibition results in weight loss in obese rodents, dogs and humans and has been proposed as a therapeutic target against obesity [45]. On the other hand, *METAP2* inhibitors have been shown to induce apoptosis in leukemic cell lines [46], which renders them potent therapeutic agents also for leukemia. Lastly, the rs7929543 variant on the *AC118942.1* pseudogene was identified as an eQTL influencing the expression of the *RP11-347H15.5* pseudogene in visceral adipose tissue. The involvement of this deregulation in possible pathogenetic processes for both diseases might be part of the complex underlying genetic-molecular mechanisms.

To describe the network of genetic variants' inheritance more extensively, we developed a panel of 64 unique proxy SNPs associated with the five common AML/T2D ones (Table 2). Interestingly, these proxies are found to lie within and/or be eQTLs for the aforementioned genes (*PPARG*, *SYN2*, *TIMP4*, *GATA3*, *RPS6KA1*, *USP44*, *METAP2*, *AC118942.1*, *RP11-347H15.5*) in disease-target tissues. A new eGene added to the panel was *DHHS*, which is downregulated in whole blood by SNPs on *RP11-347H15.5*. The gene encodes for the dehydrolchyl diphosphate synthase subunit and is involved in pathways of protein metabolism and in N-glycan biosynthesis [33]. However, no direct data connecting the gene with neoplasias or diabetes have been reported to date.

Next, we identified a panel of 86 common AML/T2D susceptibility genes using the GWAS NHGRI-EBI Catalog (Figure 3). Several SNPs specific for each disease were found to impact the expression patterns of some of these common susceptibility genes in affected tissues, suggesting their possible functional involvement in disease development (Table 5). Pathway analysis revealed that the AML/T2D gene set regulates a series of metabolic pathways, with the highest significance observed for pyrimidine and purine metabolism. Although neither AML or T2D is purely a disorder of pyrimidine and/or purine metabolism, there are data supporting their implication in the development of each disease. The insulin effect on their regulation in diabetic liver is knowledge obtained decades ago [47,48]. Nevertheless, it was very recently described that the signatures of purine metabolites, including betaine metabolites, branched-chain amino acids, aromatic amino acids, acylglycine derivatives and nucleic acid metabolites, are associated with hyperglycemia or insulin resistance [49,50]. While there is no recent evidence regarding a possible role for purine and pyrimidine metabolites in leukemia, older studies support the notion that reciprocal alterations in the phenotype of specific enzymes may occur in leukemia cells [51,52].

Choline metabolism is another pathway that emerged through gene set enrichment analysis. Indeed, its upregulation in malignant transformation is well described [53], while the serum metabolomic signature of AML patients includes parameters of aberrant choline metabolism [54]. A group of metabolic pathways, including those of carbohydrates, lipids, nucleotides, amino acids, glycans, cofactors, vitamins, biosynthesis of terpenoids, polyketides and other secondary metabolites [25], as well as signaling pathways related to metabolic disturbances and the development of neoplasia and T2D, such as mTOR, AMPK, PI₃K-Akt and insulin signaling pathways, were also among the ontologies significantly regulated by the AML/T2D gene set. Analysis also revealed an association with a pathway category deposited as “Acute Myeloid Leukemia”, which refers to ERK, PI₃K and JAK-STAT signaling and transcription regulation pathways including mutated RUNX1 and the fusion genes AML1-ETO, PML-RARA and PLZF-RARA [33].

Finally, exploration through clinical datasets revealed that certain T2D-related genes, previously shown to be deregulated in T2D individuals [4], also exhibit deviated transcriptomic levels in AML patients. Expression levels of *THADA* (thyroid adenoma-associated protein), *IGF2BP2* (insulin-like growth factor 2 mRNA binding protein 2), *CDKN2A* (cyclin-dependent kinase inhibitor 2A) and *CDK5* (cyclin-dependent kinase 5) were upregulated, while levels of *KCNQ1* (potassium voltage-gated channel subfamily Q member 1) were downregulated in the peripheral blood of AML patients compared to normal subjects. *IGF2BP2*, *CDKN2A*, *CDK5* and *KCNQ1* are known to be implicated in the mass development, proliferation, and insulin secretory function of β -cells, and in metabolic processes in T2D-affected tissues [3,20,55,56]. As for *THADA*, despite its susceptibility to T2D, there are no data yet related to its involvement in the disease's pathogenesis and/or metabolic pathways [4]. However, chromosomal aberrations engaging this gene are observed in benign thyroid adenomas [57]. *CAPN10* (calpain 10) shows increased whereas *TSPAN8* (Tetraspanin 8) exhibits decreased mRNA levels in AML versus non-cancerous individuals, a trend opposite to what was observed in T2D versus healthy subjects. *CAPN10* plays important roles in the translocation of glucose transporter 4 (GLUT4), secretion of insulin and apoptotic processes in pancreatic cells [57], while *TSPAN8* has been described as a prognostic indicator for patients with certain solid tumors [58,59], but not for hematological malignancies.

In summary, this study provides, for the first time, evidence for a strong genetic network that is related to aberrations in metabolic processes and molecular pathways, shared between AML and T2D. Even though the metabolic vulnerability of AML cells and aberrant metabolic pathways observed in AML patients [54,60] have increasingly gained the attention of the research community, the genetic background leading to these metabolic disturbances had not yet been investigated. Data emerging from our study revealed that: (i) specific genetic variants (SNPs) associated with both AML and T2D, as well as their

co-inherited proxy SNPs, mostly specific for each disease rather than common, can alter the gene expression patterns in disease-target tissues; (ii) common susceptibility genes and genes with altered expression may be linked to the development of AML or T2D through common (such as *PPARG*) or different mechanisms (such as *GATA3*) and (iii) common susceptibility genes can regulate metabolic pathways, which may be implicated in the pathogenetic mechanisms leading to the development of the two disorders. It should be noted, however, that the study has certain limitations, including that it exclusively analyzed in silico data and the fact that other parameters affecting the gene expression, such as epigenetic mechanisms, were not explored. Moreover, in the case of certain genes and their SNPs, i.e., those of *PPARG* and *GATA3*, their specific implication in AML and/or T2D development is not well documented. Therefore, it is yet difficult to provide a plausible explanation regarding their possible impact as risk factors for AML in the context of T2D. Lastly, it needs to be clarified that, although some of the reported SNPs are associated with certain genes involved in AML (such as *RPS6KA1* and *METAP2*), the latter are not considered driver genes for AML initiation.

Despite these limitations, significant evidence emerging from this study can be further explored in future basic and clinical studies. For example, the common susceptibility genes revealed can be evaluated for their potential to serve as prognostic biomarkers of AML development in cohorts of T2D individuals. Moreover, in depth exploration of the described metabolic pathways and involved genes may lead to a better understanding of the pathogenetic basis of the increased risk for AML development observed in individuals with T2D. Finally, detailed investigation of the common therapeutic targets identified may suggest that repurposing of metabolic drugs (i.e., DPP-4 inhibitor targeting *RPS6KA1* or thiazolidinediones targeting *PPAR-γ*) could be exploited as novel therapeutic strategies to enhance the anti-leukemic armamentarium.

4. Materials and Methods

4.1. Study Design

Our study was performed in two axes. (A) Detection of common genetic variants and deregulated pathways in T2D and AML: We first created a panel of SNPs associated with AML or T2D, upon an in-depth search in the NHGRI-EBI Catalog of published GWAS [3], to detect common disease susceptibility genes. Their proxy SNPs were also detected using the LDLink web tool [24]. For the possible impact of the common susceptibility SNPs and their proxies on gene mRNA expression, a combined search in the Genotype-Tissue Expression (GTEx) project [21] and the Blood eQTL Browser [22] was performed. Moreover, a panel of mutual genes bearing common or disease (AML or T2D)-specific genes were processed through pathway analysis using the STRING (Search Tool for the Retrieval of Interacting Genes/Proteins) database [26], to reveal associated molecular networks and biological processes. (B) Investigation of possible deregulated expression of T2D susceptibility genes in AML cohorts: A panel of T2D susceptibility genes that were previously described to exert aberrant mRNA levels in diabetic patients was explored for their possible deregulated expression also in AML patients, using the TNMplot tool [27].

4.2. Development of the AML and T2D Susceptibility SNP Panels and Detection of Common SNPs

The panels of total susceptibility genes specific for AML and T2D were developed upon an in-depth search in the NHGRI-EBI GWAS Catalog [3]. All populations were considered for assessment. Common disease susceptibility genes were detected, generating Venn diagrams with the Draw-Venn-Diagrams online tool (<http://bioinformatics.psb.ugent.be/webtools/Venn/>) (May 2021). A genome-wide statistically significant p -value lower than or equal to 5×10^{-8} was applied to detect the SNPs that were significantly associated with the diseases. Data regarding the prevalence of the SNPs of interest in the general population were obtained from the gnomAD browser [61].

4.3. Detection of Proxy SNPs

Proxy SNPs of disease susceptibility SNPs of interest were detected utilizing the LDLink tool [24]. LDLink interactively explores proxy and putatively functional variants/SNPs for a query/tag variant (± 500 kilobases). The tool provides information about: (A) a squared correlation measure (R^2) of linkage disequilibrium (LD); proxy SNPs are considered those having $\geq 80\%$ possibility of coinheritance with the tag SNP, which equals to a R^2 value ≥ 0.8 , and (b) the combined recombination rate (cM/Mb) from HapMap; the recombination rate is the rate at which the association between the two loci is changed. It combines the genetic (cM) and physical positions (Mb) of the marker by an interactive plot.

4.4. Detection of Expression Quantitative Trait Loci (eQTLs)

Expression quantitative trait loci (eQTLs), which explain variations in mRNA expression levels, related to the SNPs of interest were explored utilizing the GTEx portal and the Blood eQTL Browser [21,22]. Analysis was focused on the expression patterns in the total target tissues of the two diseases (as per their availability in the databases). These included adipose tissue (subcutaneous, visceral), skeletal muscle, liver, pancreas and whole blood.

4.5. Pathway Analysis

Analysis through the STRING [26] and Kyoto Encyclopedia of Genes and Genomes (KEGG) [25] databases was performed to detect protein–protein interactions possibly regulated by a panel including: (i) proteins encoded by genes that bear disease susceptibility SNPs in both AML and T2D as well as (ii) proteins encoded by genes that are commonly affected by different AML-specific and T2D-specific SNPs. To filter significantly regulated pathways, a false discovery rate (FDR) < 0.05 was set as cut-off.

4.6. Investigation of the Expression Patterns of T2D-Deregulated Genes in AML Clinical Cohorts

To explore possible variations in the mRNA expression levels of previously described T2D-deregulated genes [4] in patients with AML, the TNMplot tool was used [27]. In more detail, analysis processed whole-exome sequencing data from 151 AML patients versus 407 non-cancerous individuals, available in the database. The tool compared the expression levels of each gene in the two groups using the Mann–Whitney non-parametric test, reporting the p -value of significance and the fold-change between groups. Other information included (a) the percentage (%) of AML samples that exerted up- or downregulated expression of query genes compared to non-cancerous samples, at each of the four quantile cut-off values (minimum, 1st quartile, median, 3rd quartile, maximum), and (b) the specificity, defined as the ratio of the number of AML samples to the sum of AML and non-cancerous samples over or below each given cut-off.

Supplementary Materials: The following are available online at <https://www.mdpi.com/article/10.3390/ijms22179322/s1>. Supplementary Table S1. Total SNPs associated with AML or T2D. Data obtained upon search in the NHGRI-EBI Catalog of GWAS [3] (May 2021). Supplementary Table S2. Total eQTLs affecting the 86 AML/T2D common susceptibility genes in adipose, skeletal muscle, liver, pancreas, and whole blood. Data obtained from the GTEx portal [21] (May 2021). Supplementary Table S3. Total KEGG pathways regulated by the 86 AML/T2D susceptibility genes and eGenes, as revealed upon analysis through STRING database [25,26] (May 2021). Supplementary Figure S1. Frequency of the five T2D/AML common SNPs in the general population. Bar diagrams depicting the number of carriers of each of the SNPs and the total number of individuals included in each age group. Details regarding their frequency in different populations and males or females are reported in the embedded table. Data were downloaded from <https://gnomad.broadinstitute.org/> (accessed on 11 August 2021).

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