EBioMedicine 73 (2021) 103642

Contents lists available at ScienceDirect

EBioMedicine

journal homepage: www.elsevier.com/locate/ebiom

Review Off balance: Interferons in COVID-19 lung infections

Milena Apetito Akamatsu, Júlia Tavares de Castro, Carolina Yumi Takano, Paulo Lee Ho*

Núcleo de Produção de Vacinas Bacterianas, Centro BioIndustrial, Instituto Butantan

ARTICLE INFO

Article History: Received 31 March 2021 Revised 14 September 2021 Accepted 7 October 2021 Available online xxx

Keywords: Interferons Cytokine Storms SARS-CoV-2 COVID-19 ΤΝFα

Introduction

Interferons (IFNs) are essential cytokines that mediate the host resistance response to viral infections. In the absence of IFNs or compromised cellular programs induced by IFNs, vertebrates may be susceptible to lethal viral infections. There are 21 human IFNs, classified in 3 types (16 Type-I: 12 IFN α s, IFN β , IFN ϵ , IFN κ , and IFN ω ; 1 Type-II, IFN γ ; and 4 Type-III, IFN λ 1, IFN λ 2, IFN λ 3, and IFN λ 4), involved in innate and adaptive immune responses against not only to viruses but also to other pathogens as well as to tumors and in tissue repair. In general, the production of distinct IFNs depends on the cell type and is stimulated during viral infection when the viral nucleic acid in the cytoplasm is detected by intracellular pattern recognition receptors (PRRs). Then, the produced and secreted IFNs will bind and activates heterodimeric receptors (IFNRs) on the cells in autocrine and paracrine mechanisms that initiate a complex cascade of signaling, culminating in transcription of hundreds of different Interferon-Stimulated Genes (ISGs) resulting in strong antiviral innate and adaptive immune responses [1-3].

Briefly, Type I and III IFNs bind to their heterodimeric receptors activating the JAK/STAT pathway, which in turn activate the heterotrimeric transcription factor complex ISGF3, which consists of phosphorylated STAT1/STAT2 and IRF9. However, unphosphorylated STAT molecules and IRF9-STAT2 homodimers have also been described [4]. Type II IFN also signals through the JAK/STAT pathway, resulting in the formation of phosphorylated STAT1 homodimers, also known as IFN γ -activated factor (GAF). Activated ISGF3 and GAF, resulting

* To whom correspondence should be addressed E-mail address: paulo.ho@butantan.gov.br (P.L. Ho).

ABSTRACT

Interferons are innate and adaptive cytokines involved in many biological responses, in particular, viral infections. With the final response the result of the balance of the different types of Interferons. Cytokine storms are physiological reactions observed in humans and animals in which the innate immune system causes an uncontrolled and excessive release of pro-inflammatory signaling molecules. The excessive and prolonged presence of these cytokines can cause tissue damage, multisystem organ failure and death. The role of Interferons in virus clearance, tissue damage and cytokine storms are discussed, in view of COVID-19 caused by Severe Acute Respiratory Syndrome Coronavirus 2 (SARS-CoV-2). The imbalance of Type I, Type II and Type III Interferons during a viral infection contribute to the clinical outcome, possibly together with other cytokines, in particular, TNF α , with clear implications for clinical interventions to restore their correct balance. © 2021 The Author(s). Published by Elsevier B.V. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/)

respectively from type I/III and type II IFN actions, translocate to the nucleus and bind IFN-stimulated response elements (ISRE) and gamma-activated sequences (GAS), respectively, in the promoter regions of ISGs (Figure 1). In this review, we will discuss the recent data showing the association of COVID19 severity with the correct balance between the IFN types and other cytokines for a favorable clinical outcome during infection by SARS-CoV-2.

Coronaviruses (CoVs) are viruses belonging to the family Coronaviridea, with a large positive-sense single-stranded RNA that cause diseases in humans and animals. The new SARS-CoV-2 was identified as a beta-coronavirus responsible for the pandemic COVID-19 (Coronavirus Disease 2019) [5]. Patients with severe COVID-19 present high concentrations of proinflammatory cytokines and chemokines in the plasma, massive infiltration of monocytes and neutrophils (Figure 2).

Types of Interferons (IFNs)

IFNs are components of the innate and adaptive immune systems. Type I IFNs are produced by many cell types including lymphocytes (NK cells, B-cells and T-cells), macrophages, fibroblasts, endothelial cells, osteoblasts, leukocytes, plasmacytoid and dendritic cells [6]. Type II IFN is produced predominantly by natural killer (NK) and natural killer T (NKT) cells as part of the innate immune response, and by CD4+ and CD8+ T cells of the adaptive immune cells [7]. While the main producer of Type III IFNs are type 2 myeloid dendritic cells, epithelial cells and hepatocytes [8,9], the receptors for Type I and Type II interferons are expressed ubiquitously on almost all nucleated cells and Type III interferon receptors are expressed on epithelial, dendritic cells and neutrophils. The antiviral effects of IFNs are dictated

https://doi.org/10.1016/j.ebiom.2021.103642

2352-3964/© 2021 The Author(s). Published by Elsevier B.V. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/)





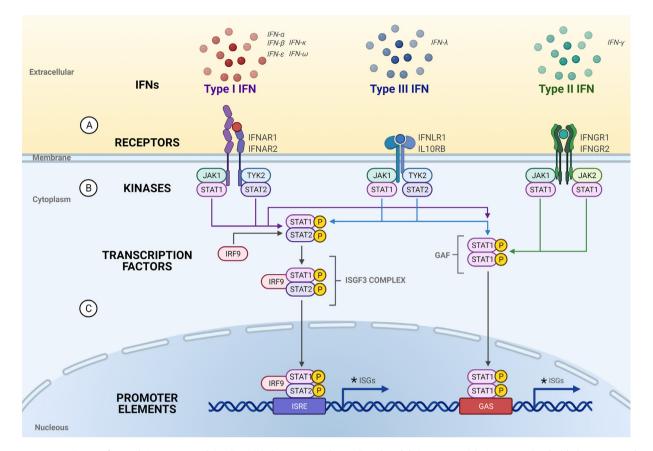


Figure 1. – Type I, II and III Interferons, their receptors and signalings. (A) The Type I, II and III IFN ligands and their receptors; (B) Kinases associated with the Type I, II and III receptors that mediate IFN signallings through receptor activation by ligand binding. Jak1 (Janus Kinase 1) interacts with STAT1 (Signal Transducer and Activator of Transcription 1) as well as to IFNRA1 (Interferon receptor alpha 1). Tyk2 (Tyrosine kinase 2) interacts to STAT2 as well as to IFNRA2. IFNRA1 forms a heterodimer with IFNRA2 that binds with Type I IFN; Jak1 and STAT1 also interacts to IFNRA1. IL-10RB (Interleukin-10 receptor B) also interacts with Tyk2 and STAT2. IFNLR1 forms a heterodimer with IL-10RB that binds to Type III IFNS. Jak1 and STAT1 also interacts to IFNCR1 and JAK2 and STAT1 also interacts to IFNCR1 forms a heterodimer with IL-10RB that binds to Type III IFNS. Jak1 and STAT1 also interacts to IFNCR1 and JAK2 and STAT1 also interacts with IFNGR2. IFNCR1 forms a heterodimer with IL-10RB (Interferon receptor B) also interacts to the nucleous and activate the transcription of genes with the GAS (gama interferon activation sites) promoters; STAT1 and STAT2 heterodimers binds to IRP9 (interferon regulatory factor 9) and the complex goes to the nucleous and activates the the transcription of genes with the ISRE (Interferon-stimulated responsive element) promoters. Some commom ISGs (trafd1, TNF receptor-associated factor -type zinc finger domain containing 1; parp 9, Poly(ADP-ribose) polymerase family member 9; parp12, Poly(ADP-ribose) polymerase family member 12; parp14, Poly(ADP-ribose) polymerase family member 14; nmi, N-myc and STAT1 interactor; epsti1, epithelial stromal interaction 1; eif2ak2, eukaryotic translation initiation factor 2 alpha kinase 2; irf1, interferon regulatory factor 8; treem67, transmembrane protein 67; treem140, transmembrane protein 140; treem173, transmembrane protein 173; dtx31, deltex E3 ubiquitin-protein ligase; samhd1, SAM (sterile alpha-motif) and HD (histidine-aspartate) do

by the presence of producer cells and responder cells with IFN receptors. For instance, Type III IFN receptors are restricted to epithelial cells in gastrointestinal, respiratory and reproductive tracts, they are considered as the first line of defense against viruses at these sites [10]. In contrast, all nucleated cells express Type I and II IFN receptors.

Types I and III IFNs in COVID-19

SARS-CoV-2 infects lung alveolar epithelial cells and these cells die due to the virus' cytopathic effect as viral replication proceeds. Recovery is achieved by proliferation and differentiation of neighboring progenitor epithelial cells. During viral infections, Type I and Type III IFNs are important mediators of the innate immune responses triggered by the recognition of pathogen-associated molecular patterns (PAMPs) by the PRRs of the cell [11]. Type I IFNs are more proinflammatory when compared to Type III IFNs [10]. Surprisingly, although both Type I and Type III IFNs have antiviral activities, mice infected with influenza virus have increased susceptibility to pneumonia caused by *Staphylococcus aureus* or *Streptococcus pneumoniae* that were associated with the increase in IFN λ production or

IFN λ treatment [12-15]. Similarly, IFN λ mRNAs were observed in bronchoalveolar fluid and naso-oropharyngeal samples of SARS-CoV-2 patients and increases in IFN λ mRNA expression were positively associated with increases in COVID-19 disease morbidity [16]. Interestingly, increases in IFN λ stimulate apoptosis and expression of tumor suppressor p53 that impairs proliferation, differentiation and repair of the lung epithelial cells, increasing disease severity, lung damage and susceptibility to bacterial superinfections (Figure 3F-G) [16,17]. Although Type I (IFN α and IFN β) also reduces lung epithelial cell proliferation after treatment during influenza, only IFN λ compromises lung epithelial tissue recovery [17]. Moreover, Type I IFN production and activity were highly suppressed in severe COVID-19 patients. These patients present a proinflammatory picture driven by nuclear factor- κ B (NF- κ B) and characterized by increased IL-6 and TNF α [18]. The reasons for this impaired production, signaling and activity of Type I IFN seems to be related, at least in part, to inborn errors associated to Type I IFN signaling cascade or production of autoantibodies, especially to IFN α or IFN ω [19-21]. In addition, it was also shown that the presence of autoantibodies against Type I IFN (IFN α and IFN ω) was proportionally higher in males and elderly with severe COVID-19 [20,21]. Altogether, these data suggest an important

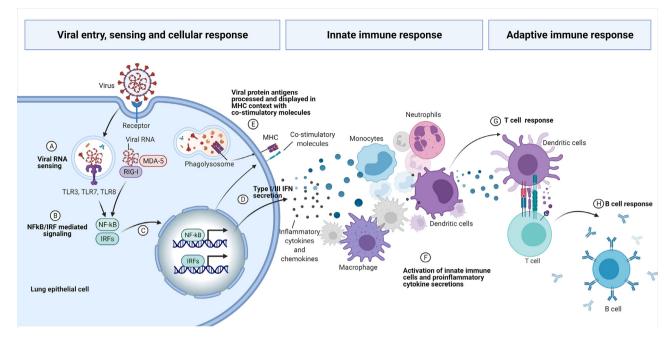


Figure 2. – Schematic view of the viral entry, sensing and host immune response. The RNA virus bind to the cellular receptor and is internalized. Pattern Recognition Receptors (PRRs) sense the viral RNA (A) and activate cellular signaling mediated by NF- κ B and IRFs transcription factors (B) to the nucleous (C), activating expression of specific genes, comprising Type I and Type III IFNs and proinflammatory cytokines (D). Viral protein antigens are also processed and displayed in MHC context with co-stimulatory molecules (E). The proinflammatory cytokines are responsible for the cellular innate response, with pronounced infiltration of monocytes and neutrophils (F). Activated dendritic cells present the antigens to the adaptive T cells (G) that will secrete cytokines to mature B cells to produce and secrete immunoglobulins (H) (Figure created by BioRender.com).

role of these IFNs in protection and susceptibility to severe pneumonia in COVID-19.

Viral proteins targeting IFN antiviral response

There are many clinical similarities between severe influenza and COVID-19. The COVID-19 clinical manifestations include profound and uncontrolled inflammatory response, massive cytokine release, diffuse alveolar damage and lung infiltration with inflammatory cells, most of them comprised of neutrophils. However, there are also striking differences, related to thrombosis and endothelial dysfunction in COVID-19, which appears to exceed that in influenza patients as observed from lung pathology of fatal cases [22,23].

Both influenza and SARS-CoV-2 have virulence factors targeting the IFN antiviral responses. Influenza hemagglutinins have been shown to facilitate ubiquitination and degradation of Type I IFN receptors, thus reducing the expression levels of Type I IFN receptors and impacting the strength of their responses and ISGs expressions [24,25]. Besides this, the non-structural protein 1 (NS1) inhibits the TRIM25, a RING-finger E3 ubiquitin ligase, blocking RIG-I-mediated innate immunity and suppression of host IFN response. Furthermore, NS1 also inhibits the production of Type I IFNs and downstream effector molecules of IFNs-mediated signal transduction processes by the inhibition of protein kinase RNA-activated (PKR) through the increased expression of host small non-coding RNAs (vault RNAs). As a result, the replication and production of virus particles are also increased [25,26]. NS1 may also affect the phosphorylation of IkB kinases (IKK) and decrease the NF- κ B complex in the nucleus and the gene expression mediated by NF-kB. The JAK-STAT pathways of IFNs are also impaired by NS1 that lowers the phosphorylation levels of STAT1, STAT2 and STAT3, preventing proper transcription of ISGs [27]. The transcription of genes is also affected by direct binding of NS1 to host double stranded DNA [28]. Therefore, NS1 is a master key to block the innate immune system, the transcription of IFNs and ISGs through many mechanisms but also of the adaptive immunity by modulating the dendritic cells capacities to induce the T cell responses [29,30]. However, besides NS1, other influenza proteins help to evade Type I IFN actions. PB1-F2 and PB2 influenza viral proteins also interferes with Type I IFN signaling pathway through interaction with MAVS in the mitochondria [31-33]. M2 protein from influenza also affects the generation of IFNs by blocking the host autophagy and TLRs pathways [34].

Although SARS-CoV-2 deregulates cell signaling pathways to generate a permissive environment for viral replication, mainly related to delayed hyperinflammation, generating weakened and delayed IFN responses [35], it was observed that the Spike protein (S) amplifies the activation of IFN β and the Envelope protein (E) stimulates the transcription of ISGs, in an opposite direction of creating a permissive milieu for SARS-CoV-2 [36]. The Membrane protein (M) and the Nucleocapsid (N) structural proteins inhibits the activation of IFN β or Type I IFN signaling pathway [36,37]. NSP1 inhibits STAT1 phosphorylation upon IFN α stimulation as well as the IFN β production in HEK293T cells [36,38]. In contrast, NSP2 accelerates the activation of IFN β [36]. NSP3, NSP12, NSP13, NSP14, ORF3, ORF6, ORF7 and ORF8 proteins inhibit Sendai virus-induced IFN β promoter activation, IFN α downstream signaling or NF- κ B-responsive promoter [35-39]. ORF9 protein targets the nuclear factor κ B (NF- κ B) essential modulator NEMO and interrupts its K63-linked polyubiquitination upon viral stimulation, thereby inhibiting the canonical IkB kinase alpha (IKK α)/ β / γ -NF- κ B signaling and subsequent IFN β production [40].

Thus, viral proteins in general have diverse roles in disruption of cellular programs to evade and to weaken the host immune response [41] including Influenza and SARS-CoV-2 proteins, most of them as negative effectors targeted to Type I IFN pathway, though positive effectors can also be observed, as in the case of Spike, Envelope and NSP2 viral proteins as discussed above.

Type II IFN in COVID-19

Type II IFN is produced predominantly by natural killer (NK) and natural killer T (NKT) cells as part of the innate immune response, and by CD4+ and CD8+ T cells of the adaptive immune cells [7]. Upon virus infection, IFN γ acts directly on CD8⁺ T cells to help them to

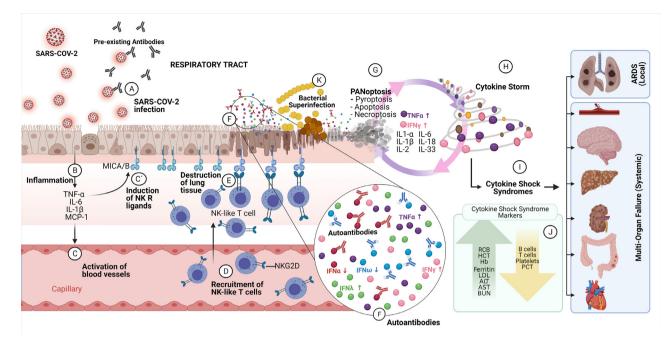


Figure 3. – Respiratory tract infection by SARS-CoV-2. Infection occurs at the epithelial cells of the respiratory tract and the infection may be inhibited by pre-existing cross-reactive antibodies resulted from previous infections with seasonal viruses (A). The infection causes inflammation in the respiratory tract with the secretion of proinflammatory cytokines (B), activation of the endothelial cells (C) and expression of NK receptor ligands (MICA/B, MHC class I chain related protein A or B) on respiratory epithilium (C). The activated endothelium promotes the infiltration of NK-like T cells expressing NK (Natural Killer) receptors (NKR) exemplified by NKG2D on the cell surface from the capillaries (D) to the respiratory tract epithelium. The infiltrating NK-like T cells binds to NKR receptor ligands (MICA/B) and induce TCR-independent killing of epithelium cells expressing the NKR ligands (E). In response to viral infections, epithelium cells secrete Type I or Type III IFNs. In severe cases of the diseases, the presence of autoantibodies against IFN α and IFN α (F) was observed and associated with higher morbidity, resulting in more viral infections. It was also observed an increase in the secretion of Type III IFN (IFN λ)(F). IFN λ impairs lung epithelial cell proliferation and tissue repair mediated by the expression of the tumor suppressor p53 gene and protein pathway. Cell death programs (PANoptosis) induced by cyto-kine storms, in special by IFN γ and TN α , perpetuates the local cytokine storms killing more epithelial cells in the respiratory tract (G) and the cytokine storms propagate (H) to other organs and tissues, provoking cytokine shock syndromes. Acute respiratory distress syndrome (ARDS) will be observed in the patient due to lung damage as well as multiorargan failures (I) due to systemic spread of the proinflammatory cytokines, in special of IFN γ and TN α . The cytokine shock syndromes can be identified by clinical markers as listed in the figure (J). Susceptibility to bacterial su

differentiate into cytotoxic T lymphocytes (CTL), which produce cytokines and effector molecules to restrict viral replication and kill virus-infected cells [42] as well as for the increase in CD4⁺ and CD8⁺ memory cells [43]. In general, type II IFN connects the innate immune response with the activation of the adaptive immune response [41].

The role of IFN γ during the infection of SARS-CoV-2 is beginning to be elucidated. It was shown that IFN γ is produced in high amount in patients with moderate to severe COVID-19 besides other proinflammatory cytokines. This Type II IFN is found in the serum of these patients and released in the culture medium when peripheral blood mononuclear cells (PBMCs) are infected with SARS-CoV-2 [44]. However, from the ten different cytokines that have their concentration increased in moderate to severe COVID-19 patients (IL-6, IL-18, $IFN\gamma$, IL-15, TNF α , IL-1 α , TNF β , IL-1 β , IL-33, IL-2), only the combination of IFN γ and TNF α induced inflammatory cell death characterized by pyroptosis, apoptosis and necroptosis in bone marrow-derived macrophages (BMDM), a process known as PANoptosis [44] (Figure 3A-J). Neither IFN γ nor TNF α alone or in combination with the other inflammatory cytokines were able to induce this response. Interestingly, combination of IFN γ and TNF α provoked a lethal cytokine shock in mouse model that was completely abolished by IFN γ and TNF α blockage by neutralizing antibodies [44]. Though these results associate the important role of IFN γ and TNF α combination in the cytokine storm and acute lung damage with patient mortality as observed in severe cases of COVID-19, recent results also associate recovery-like stage of patients with enhanced TNF α and IFN- γ gene expression [45]. Therefore, the role of these cytokines in COVID-19 has to be confirmed in future studies.

COVID-19 severity in male and older people is associated with exacerbated inflammation in the respiratory tract

COVID-19 mostly induces mild to moderate clinical symptoms in younger patients. However, SARS-CoV-2 causes higher morbidity and mortality in older patients, with exacerbated inflammation of their respiratory tracts [46,47]. Interestingly, older healthy individuals present a chronic low-grade sterile systemic inflammation (an inflammatory response not induced by a pathogen), a process known as inflamm-aging, with a higher baseline concentration of C-reactive protein (CRP) and some pro-inflammatory cytokines (IL-1 β , IL-6, IL8). The mechanisms that lead to pro- inflamm-aging are not completely understood but may involve impaired clearance of dead and dying cells with sustained inflammation, misfolded proteins, compromised gut barrier function, obesity and senescent cells which no longer divide but keep secreting pro-inflammatory cytokines, chemokines, growth factors and matrix metalloproteinases (MMPs), causing organ dysfunction during the aging process [48-51]. Interestingly, individuals in the same age group that do not present the inflamm-aging profile are healthier and live longer when compared with those with the inflamm-aging signs [48].

Although inflammation is essential to modulate both innate and adaptive immune response at the initial phases of immune stimulation, the inflammatory process is actively regulated by many processes, including the clearance of dead and dying cells and other mechanisms [49]. An excessive and non-resolved inflammation can inhibit antigen-specific immunity in vivo, as shown in the case of viral cancers [52]. It is also compelling that the inflamm-aging phenotype may contribute in exacerbating the hyperinflammation observed in older COVID-19 patients. The inflammation in the lungs of COVID-19 patients may involve the infiltration of monocytes, induction of Natural Killer Receptor (NKR) ligands (MICA/B) by cytokines produced in non-lymphoid cells [53], such as the pulmonary respiratory epithelial cells. These cells can be targeted and killed by NK-like T cells, possibly contributing to the pulmonary tissue damage in COVID-19 patients (Figure 3A-E) [47]. In contrast, NK cell production and proliferation in elderly is reduced and in elderly patients with COVID-19, the NK cells present in addition, an exhausted phenotype which affects the host defense against the SARS-CoV-2 acute infection in addition to the increased secretion of inflammatory cytokines [54,55].

Recent data showing that preexisting antibodies against seasonal human coronavirus able to cross-react with SARS-CoV-2 may also account for the clinical differences observed in children and adolescents versus elderly [56-58]. Children and adolescents are more recurrently infected by seasonal coronaviruses than elderly [58] and they present the highest amounts of cross-reactive antibodies between seasonal coronaviruses and SARS-CoV-2. Therefore, it is conceivable that sustained infection by seasonal coronavirus in children and adolescents induces protective cross-reactive antibodies and these preexisting antibodies influences the severity of COVID-19 by SARS-CoV-2 infection, in contrast to elderly. It is also possible that both mechanisms, inflammaging and preexisting antibodies influences the clinical outcome differences observed in children, adolescents and elderly (Figure 3A).

Males present a significantly higher COVID-19 mortality risk [59,60]. The plasma concentration of inflammatory cytokines and chemokines such as IL-8 and IL-18 are increased in males infected with SARS-CoV-2 when compared to females. The exacerbated systemic inflammation is associated with severe pulmonary pathology, which includes pronounced infiltration of monocytes and neutrophils [61-63]. Interestingly, it is well described that androgens drive the expression of Transmembrane protease serine 2 (TMPRSS2) in the lung and prostate. TMPRSS2 activates the spike protein by cleaving the S1/S2 and S2 sites of SARS-CoV-2 virus spike proteins. This cleavage step is necessary for the virus-host cell membrane fusion and cell entry mediated by the cellular receptor ACE2 that may account for some of the observed gender differences [64]. On the other hand, female patients have higher concentration of Type I IFN (IFN α) in their plasma that might also contribute to less disease severity [60]. Therefore, differences in Type I IFN expression may also account for distinct severities in COVID-19 or other viral pneumonias together with inflamm-aging condition, gender differences and pre-existing antibodies.

Future Directions

Altogether, these observations suggest several complex mechanisms SARS-CoV-2 induces for the diseases severities observed. Activation of NK-like T cells that bind to MICA/B and kill the MICA/B expressing cells is one possible mechanism. This response must be concerted with other mechanisms. Many recent studies have described that a fine balance between type I and type III IFN is critical for optimal protection and minimal host damage [16,17]. Deviation from this finely tuned balance can harm the patient through the unleashing of a cytokine storm. The mechanisms that may contribute to this balance could be the viral and host genetic backgrounds, previous infections leading to the presence of pre-existing viral antibodies, autoantibodies against Type I IFN (in particular, IFN α and IFN ω) or increased expression of IFN λ associated with poor lung epithelial cell/tissue repair processes [8-20]. Adding to this complex scenario, IFN γ and TNF α combination induced PANaptosis helps to perpetuate the cytokine storm and the disease progression with more lung epithelial cell death, tissue damage and increased susceptibility to bacterial superinfection [44]. Future therapeutic interventions have to consider these observations and interplays. It seems that binding of NK-like T cells to MICA/B presenting cells, IFN λ , IFN γ , TNF α and their associated signaling pathways should be targeted to increase tissue proliferation and repair, while decreasing damage to lung epithelial cell/tissue (for instance, interventions with neutralizing antibodies against Anti-TNF α , Anti-IFN γ , Anti-IFN λ or with JAK1/2 inhibitors, STAT1 inhibitors, NO inhibitor, PANoptosis inhibitors). Blockage of IFN γ and TNF α by neutralizing antibodies may increase survival and inhibit cell PANoptosis after SARS-CoV-2 infection or in other pathologies associated to cytokine storm. At the same time, treatment with Type I IFN (IFN α , IFN β IFN ω) might be effective in diminishing SARS-CoV-2, influenza and other respiratory viral infections. However, it is not clear how much we have to modulate/change the concentration of these molecules systemically for a proper clinical response and if the activation of these pathways can exacerbate the disease outcome since the SARS-CoV-2 cellular receptor ACE2 expression is increased in human nasal epithelia and lung tissue by Type I and II IFNs and also by Influenza infection [65]. All the possible interventions discussed above have to be monitored and provide qualitative and quantitative data for treatment decision and planning of patients with different ages, genders and co-morbilities. Furthermore, in light of pre-existing antibodies against the SARS-CoV-2, the presence of autoantibodies against Type I IFNs and also of inborn errors of Type I IFN that would affect the therapeutic response, it is important to generate information concerning the Type I IFN pathway integrity for a proper evaluation of the possible therapeutic intervention. At the current stage, the COVID-19 Treatment Guidelines Panel from National Institute of Health (NIH, USA) recommends against the use of interferons for the treatment of patients with severe or critical COVID-19, except in a clinical trial. There are also insufficient data to recommend either for or against the use of interferon beta for the treatment of early (i.e., <7 days from symptom onset) mild and moderate COVID-19 [66], though some recent trials showed some clinical benefits [67-69].

Outstanding Questions

The contributions of IFNs in viral infection responses is becoming clear. In SARS-CoV-2 infections, how can we restore the correct IFNs balances? What are the quantitative values for a proper clinical correction in the IFNs balances? What are the possible adjuvant therapies to be included for viral clearance with acceptable side effects in patients? How does TNF α affect IFN γ activities and vice-versa? Does TNF α interact with Type I or Type III IFNs in epithelial tissue proliferation and repair? Can the use of IFNs or influenza infection increase SARS-CoV-2 infections by the increase of its cellular receptor ACE2? Are these interplays occurring in the infections with the new strains of SARS-CoV-2?

Search Strategy

Data search and selection were identified through searches from PubMed with the following search terms: "COVID-19", "SARS-CoV-2", "Influenza", "Interferons", "Cytokine storms", "immune response". Only articles published in English between 2005 and 2021 were included.

Contributors

All the authors contributed in the discussion, writing, literature searches and revision of the manuscript. All the Figures were mentored and designed by PLH and MAA

Declaration of Competing Interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Acknowledgments

The authors would like to thank the financial support of the Brazilian financial agencies Fapesp (Grant 2020/07040-1 to PLH), CNPq (Grant 305430/2019-0 to PLH) and Fundação Butantan as well as all the comments received from the reviewers. We also declare that the funders had any role in paper design, data collection, data analysis, interpretation and writing of the paper.

References

- Walter MR. The Role of Structure in the. Biology of Interferon Signaling. Front. Immunol. 2020;11:606489. doi: 10.3389/fimmu.2020.606489.
- [2] Schoggins JW. Interferon-Stimulated Genes: What Do They All Do? Annu. Rev. Virol. 2019;6(1):567–84. doi: 10.1146/annurev-virology-092818-015756.
- [3] Onomoto K, Onoguchi K, Yoneyama M. Regulation of RIG-I-like receptor-mediated signaling: interaction between host and viral factors. Cell. Mol. Immunol. 2021;18:1–17. doi: 10.1038/s41423-020-00602-7.
- [4] Au-Yeung N, Horvath CM. Transcriptional and chromatin regulation in interferon and innate antiviral gene expression. Cytokine Growth Factor Rev 2018;44:11–7.
- [5] Monchatre-Leroy E, Boué F, Boucher J-M, Renault C, Moutou F, Ar Gouilh M, Umhang G. Identification of Alpha and Beta Coronavirus in Wildlife Species in France: Bats, Rodents, Rabbits, and Hedgehogs. Viruses 2017;9:364. doi: 10.3390/ v9120364.
- [6] Asselin-Paturel C, Trinchieri G. Production of type I interferons: plasmacytoid dendritic cells and beyond. J Exp Med 2005;202(4):461–5. doi: 10.1084/ jem.20051395.
- [7] Schoenborn JR, Wilson CB. Regulation of Interferon-γ During Innate and Adaptive Immune Responses. Regulation of interferon-gamma during innate and adaptive immune responses. Adv. Immunol. 2007;96:41–101. doi: 10.1016/S0065-2776 (07)96002-2.
- [8] Syedbasha M, Egli A. Interferon Lambda: Modulating Immunity in Infectious Diseases. Front Immunol. 2017;8:119. doi: 10.3389/fimmu.2017.00119.
- [9] Grajales-Reyes GE, Colonna M. Interferon responses in viral pneumonias. Science 2020;369(6504):626–7. doi: 10.1126/science.abd2208.
- [10] Galani IE, Triantafyllia V, Eleminiadou EE, Koltsida O, Stavropoulos A, Manioudaki M, Thanos D, Doyle SE, Kotenko SV, Thanopoulou K, Andreakos E. Interferonlambda Mediates Non-redundant Front-Line Antiviral Protection against Influenza Virus Infection without Compromising Host Fitness. Immunity 2017;46 (5):875–90 e6. doi: 10.1016/j.immuni.2017.04.025.
- [11] Kikkert M. Innate Immune Evasion by Human Respiratory RNA Viruses. J Innate Immun 2020;12(1):4–20. doi: 10.1159/000503030.
- [12] Lazear HM, Nice TJ, Diamond MS. Interferon-lambda: Immune Functions at Barrier Surfaces and Beyond. Immunity 2015;43(1):15–28. doi: 10.1016/j. immuni.2015.07.001.
- [13] Planet PJ, Parker D, Cohen TS, Smith H, Leon JD, Ryan C, Hammer TJ, Fierer N, Chen EI, Prince AS. Lambda Interferon Restructures the Nasal Microbiome and Increases Susceptibility to Staphylococcus aureus Superinfection. mBio 2016;7(1):e01915– e39. doi: 10.1128/mBio.01939-15.
- [14] Rich HE, McCourt CC, Zheng WQ, McHugh KJ, Robinson KM, Wang J, Alcorn JF. Interferon Lambda Inhibits Bacterial Uptake during Influenza Superinfection. Infect Immun 2019;87(5):e00114–9. doi: 10.1128/IAI.00114-19.
- [15] Broggi A, Ghosh S, Sposito B, Spreafico R, Balzarini F, Lo Cascio A, Clementi N, De Santis M, Mancini N, Granucci F, Zanoni I. Type III interferons disrupt the lung epithelial barrier upon viral recognition. Science 2020;369(6504):706–12. doi: 10.1126/science.abc3545.
- [16] Major J, Crotta S, Llorian M, McCabe TM, Gad HH, Priestnall SL, Hartmann R, Wack A. Type I and III interferons disrupt lung epithelial repair during recovery from viral infection. Science 2020;369(6504):712–7. doi: 10.1126/science.abc2061.
- [17] Hadjadj J, Yatim N, Barnabei L, Corneau A, Boussier J, Smith N, Péré H, Charbit B, Bondet V, Chenevier-Gobeaux C, Breillat P, Carlier N, Gauzit R, Morbieu C, Pène F, Marin N, Roche N, Szwebel TA, Merkling SH, Treluyer JM, Veyer D, Mouthon L, Blanc C, Tharaux PL, Rozenberg F, Fischer A, Duffy D, Rieux-Laucat F, Kernéis S, Terrier B. Impaired type I interferon activity and inflammatory responses in severe COVID-19 patients. Science 2020;369(6504):718–24. doi: 10.1126/science. abc6027.
- [18] Zhang Q, Bastard P, Liu Z, Le Pen J, Moncada-Velez M, Chen J, Ogishi M, Sabli IKD, Hodeib S, Korol C, Rosain J, Bilguvar K, Ye J, Bolze A, Bigio B, Yang R, Arias AA, Zhou Q, Zhang Y, Onodi F, Korniotis S, Karpf L, Philippot Q, Chbihi M, Bonnet-Madin L, Dorgham K, Smith N, Schneider WM, Razooky BS, Hoffmann HH, Michailidis E, Moens L, Han JE, Lorenzo L, Bizien L, Meade P, Neehus AL, Ugurbil AC, Corneau A, Kerner G, Zhang P, Rapaport F, Seeleuthner Y, Manry J, Masson C, Schmitt Y, Schlüter A, Le Voyer T, Khan T, Li J, Fellay J, Roussel L, Shahrooei M, Alosaimi MF, Mansouri D, Al-Saud H, Al-Mulla F, Almourfi F, Al-Muhsen SZ, Alsohime F, Al Turki S, Hasanato R, van de Beek D, Biondi A, Bettini LR, D'Angio' M, Bonfanti P,

Imberti L, Sottini A, Paghera S, Quiros-Roldan E, Rossi C, Oler AJ, Tompkins MF, Alba C, Vandernoot I, Goffard JC, Smits G, Migeotte I, Haerynck F, Soler-Palacin P, Martin-Nalda A, Colobran R, Morange PE, Keles S, Çölkesen F, Ozcelik T, Yasar KK, Senoglu S, SN Karabela, Rodríguez-Gallego C, Novelli G, Hraiech S, Tandjaoui-Lambiotte Y, Duval X, Laouénan C, COVID-STORM Clinicians, Snow AL, Dalgard CL, Milner JD, Vinh DC, Mogensen TH, Marr N, Spaan AN, Boisson B, Boisson-Dupuis S, Bustamante J, Puel A, Ciancanelli MJ, Meyts I, Maniatis T, Soumelis V, Amara A, Nussenzweig M, García-Sastre A, Krammer F, Pujol A, Duffy D, Lifton RP, Zhang SY, Gorochov G, Béziat V, Jouanguy E, Sancho-Shimizu V, Rice CM, Abel L, Notarangelo LD, Cobat A, Su HC, Casanova JL. COVID Clinicians; Imagine COVID Group; French COVID Cohort Study Group; CoV-Contact Cohort; Amsterdam UMC COVID-19 Biobank; COVID Human Genetic Effort; NIAID-USUHS/TAGC COVID Immunity Group. Inborn errors of type I IFN immunity in patients with lifethreatening COVID-19. Science 2020;370(6515):eabd4570. doi: 10.1126/science. abd4570.

- [19] Bastard P, Rosen LB, Zhang Q, Michailidis E, Hoffmann HH, Zhang Y, Dorgham K, Philippot Q, Rosain J, Béziat V, Manry J, Shaw E, Haljasmägi L, Peterson P, Lorenzo L, Bizien L, Trouillet-Assant S, Dobbs K, de Jesus AA, Belot A, Kallaste A, Catherinot E, Tandjaoui-Lambiotte Y, Le Pen J, Kerner G, Bigio B, Seeleuthner Y, Yang R, Bolze A, Spaan AN, Delmonte OM, Abers MS, Aiuti A, Casari G, Lampasona V, Piemonti L, Ciceri F, Bilguvar K, Lifton RP, Vasse M, Smadja DM, Migaud M, Hadjadj J, Terrier B, Duffy D, Quintana-Murci L, van de Beek D, Roussel L, Vinh DC, Tangye SG, Haerynck F, Dalmau D, Martinez-Picado J, Brodin P, Nussenzweig MC, Boisson-Dupuis S, Rodríguez-Gallego C, Vogt G, Mogensen TH, Oler AJ, Gu J, Burbelo PD, Cohen JI, Biondi A, Bettini LR, D'Angio M, Bonfanti P, Rossignol P, Mayaux J, Rieux-Laucat F, Husebye ES, Fusco F, Ursini MV, Imberti L, Sottini A, Paghera S, Quiros-Roldan E, Rossi C, Castagnoli R, Montagna D, Licari A, Marseglia GL, Duval X, J; Ghosn, Tsang JS, Goldbach-Mansky R, Kisand K, Lionakis MS, Puel A, Zhang SY, Holland SM, Gorochov G, Jouanguy E, Rice CM, Cobat A, Notarangelo LD, Abel L, Su HC, Casanova JL. HGID Lab; NIAID-USUHS Immune Response to COVID Group; COVID Clinicians; COVID-STORM Clinicians; Imagine COVID Group; French COVID Cohort Study Group; Milieu Intérieur Consortium; CoV-Contact Cohort; Amsterdam UMC COVID-19 Biobank; COVID Human Genetic Effort Autoantibodies against type I IFNs in patients with life-threatening COVID-19 Science 2020;370(6515): eabd4585. doi: 10.1126/science.abd4585
- [20] Beck DB, Aksentijevich I. Susceptibility to severe COVID-19. Science 2020;370 (6515):404–5. doi: 10.1126/science.abe7591.
- [21] Ackermann M, Verleden SE, Kuehnel M, Haverich A, Welte T, Laenger F, Vanstapel A, Werlein C, Stark H, Tzankov A, Li WW, Li VW, Mentzer SJ, Jonigk D. Pulmonary Vascular Endothelialitis, Thrombosis, and Angiogenesis in COVID-19. N Engl J Med 2020;383(2):120–8. doi: 10.1056/NEJMoa2015432.
- [22] Hartshorn KL Innate Immunity and Influenza A Virus Pathogenesis: Lessons for COVID-19. Front Cell Infect Microbiol 2020;10:563850. doi: 10.3389/ fcimb.2020.563850.
- [23] Xia C, Vijayan M, Pritzl CJ, Fuchs SY, McDermott AB, Hahm B. Hemagglutinin of Influenza A Virus Antagonizes Type I Interferon (IFN) Responses by Inducing Degradation of Type I IFN Receptor 1. J Virol 2015;90(5):2403–17. doi: 10.1128/ JVI.02749-15.
- [24] Chen X, Liu S, Goraya MU, Maarouf M, Huang S, Chen JL. Host Immune Response to Influenza A. Virus Infection. Front Immunol. 2018;9:320. doi: 10.3389/ fimmu.2018.00320.
- [25] Li Fang, Chen Yuhai, Zhang Zhaoyuan, Ouyang Jing, Wang Yi, Yan Ruoxiang, Huang Shile, Fu Gao George, Guo Guijie, Chen Ji-Long. Robust expression of vault RNAs induced by influenza A virus plays a critical role in suppression of PKRmediated innate immunity. Nucleic Acids Res 2015;43(21):10321–37. doi: 10.1093/nar/gkv1078.
- [26] Jia D, Rahbar R, Chan RW, Lee SM, Chan MC, Wang BX, Baker DP, Sun B, Peiris JS, Nicholls JM, Fish EN. Influenza virus non-structural protein 1 (NS1) disrupts interferon signaling. PLoS One 2010;5(11):e13927. doi: 10.1371/journal. pone.0013927.
- [27] Anastasina M, Le May N, Bugai A, Fu Y, Söderholm S, Gaelings L, Ohman T, Tynell J, Kyttänen S, Barboric M, Nyman TA, Matikainen S, Julkunen I, Butcher SJ, Egly JM, Kainov DE. Influenza virus NS1 protein binds cellular DNA to block transcription of antiviral genes. Biochim Biophys Acta 2016;1859(11):1440–8. doi: 10.1016/j. bbagrm.2016.09.005.
- [28] Fernandez-Sesma A, Marukian S, Ebersole BJ, Kaminski D, Park MS, Yuen T, Sealfon SC, García-Sastre A, Moran TM. Influenza virus evades innate and adaptive immunity via the NS1 protein. J Virol 2006;80(13):6295–304. doi: 10.1128/ JVI.02381-05.
- [29] Jiang H, Shen SM, Yin J, Zhang PP, Shi Y. Influenza virus non-structural protein 1 inhibits the production of interferon beta of alveolar epithelial cells upon the infection of influenza A H1N1. Mol Med Rep 2017;16(4):4553–60. doi: 10.3892/ mmr.2017.7138.
- [30] Varga ZT, Ramos I, Hai R, Schmolke M, García-Sastre A, Fernandez-Sesma A, Palese P. The influenza virus protein PB1-F2 inhibits the induction of type I interferon at the level of the MAVS adaptor protein. PLoS Pathog 2011;7(6):e1002067. doi: 10.1371/journal.ppat.1002067.
- [31] Iwai A, Shiozaki T, Kawai T, Akira S, Kawaoka Y, Takada A, Kida H, Miyazaki T. Influenza A virus polymerase inhibits type I interferon induction by binding to interferon beta promoter stimulator 1. J Biol Chem 2010;285(42):32064–74. doi: 10.1074/jbc.M110.112458.
- [32] Wang R, Zhu Y, Ren C, Yang S, Tian S, Chen H, Jin M, Zhou H. Influenza A virus protein PB1-F2 impairs innate immunity by inducing mitophagy. Autophagy 2020;11:1–16. doi: 10.1080/15548627.2020.1725375.
- [33] Beale R, Wise H, Stuart A, Ravenhill BJ, Digard P, Randow F. A LC3-interacting motif in the influenza A virus M2 protein is required to subvert autophagy and

maintain virion stability. Cell Host Microbe 2014;15(2):239-47. doi: 10.1016/j. chom.2014.01.006.

- [34] Suryawanshi RK, Koganti R, Agelidis A, Patil CD, Shukla D. Dysregulation of Cell Signaling by SARS-CoV-2. Trends Microbiol 2021;29(3):224–37. doi: 10.1016/j. tim.2020.12.007.
- [35] Lei X, Dong X, Ma R, Wang W, Xiao X, Tian Z, Wang C, Wang Y, Li L, Ren L, Guo F, Zhao Z, Zhou Z, Xiang Z, Wang J. Activation and evasion of type I interferon responses by SARS-CoV-2. Nat Commun 2020;11(1):3810. doi: 10.1038/s41467-020-17665-9.
- [36] Li JY, Liao CH, Wang Q, Tan YJ, Luo R, Qiu Y, Ge XY. The ORF6, ORF8 and nucleocapsid proteins of SARS-CoV-2 inhibit type I interferon signaling pathway. Virus Res 2020;286:198074. doi: 10.1016/j.virusres.2020.198074.
- [37] Xia H, Cao Z, Xie X, Zhang X, Chen JY, Wang H, Menachery VD, Rajsbaum R, Shi PY. Evasion of Type I Interferon by SARS-CoV-2. Cell Rep 2020;33(1):108234. doi: 10.1016/j.celrep.2020.108234.
- [38] Yuen CK, Lam JY, Wong WM, Mak LF, Wang X, Chu H, Cai JP, Jin DY, To KK, Chan JF, Yuen KY, Kok KH. SARS-CoV-2 nsp13, nsp14, nsp15 and orf6 function as potent interferon antagonists. Emerg Microbes Infect 2020;9(1):1418–28. doi: 10.1080/ 22221751.2020.1780953.
- [39] Wu J, Shi Y, Pan X, Wu S, Hou R, Zhang Y, Zhong T, Tang H, Du W, Wang L, Wo J, Mu J, Qiu Y, Yang K, Zhang LK, Ye BC, Qi N. (2021) SARS-CoV-2 ORF9b inhibits RIG-I-MAVS antiviral signaling by interrupting K63-linked ubiquitination of NEMO. Cell Rep. 2021 Feb 16;34(7):108761. doi: 10.1016/j.celrep.2021.108761.
- [40] Rojas JM, Alejo A, Martín V, Sevilla N. Viral pathogen-induced mechanisms to antagonize mammalian interferon (IFN) signaling pathway. Cell Mol Life Sci 2021;78(4):1423-44. doi: 10.1007/s00018-020-03671-z.
- [41] Whitmire JK, Tan JT, Whitton JL. Interferon-gamma acts directly on CD8+ T cells to increase their abundance during virus infection. J Exp Med 2005;201(7):1053–9. doi: 10.1084/jem.20041463.
- [42] Whitmire JK, Eam B, Benning N, Whitton JL. Direct interferon-gamma signaling dramatically enhances CD4+ and CD8+ T cell memory. J Immunol 2007;179 (2):1190–7. doi: 10.4049/jimmunol.179.2.1190.
- [43] Karki R, Sharma BR, Tuladhar S, Williams EP, Zalduondo L, Samir P, Zheng M, Sundaram B, Banoth B, Malireddi RKS, Schreiner P, Neale G, Vogel P, Webby R, Jonsson CB. Kanneganti TD Synergism of TNF-alpha and IFN-gamma Triggers Inflammatory Cell Death, Tissue Damage, and Mortality in SARS-CoV-2 Infection and Cytokine Shock Syndromes. Cell 2021;184(1):149–68 e17. doi: 10.1016/j. cell.2020.11.025.
- [44] Sadanandam A, Bopp T, Dixit S, Knapp DJHF, Emperumal CP, Vergidis P, Rajalingam K, Melcher A, Kannan N. A blood transcriptome-based analysis of disease progression, immune regulation, and symptoms in coronavirus-infected patients. Cell Death Discov 2020;6(1):141. doi: 10.1038/s41420-020-00376-x.
- [45] Ziegler CGK, Allon SJ, Nyquist SK, Mbano IM, Miao VN, Tzouanas CN, Cao Y, Yousif AS, Bals J, Hauser BM, Feldman J, Muus C, Wadsworth 2nd MH, Kazer SW, Hughes TK, Doran B, Gatter GJ, Vukovic M, Taliaferro F, Mead BE, Guo Z, Wang JP, Gras D, Plaisant M, Ansari M, Angelidis I, Adler H, Sucre JMS, Taylor CJ, Lin B, Waghray A, Mitsialis V, Dwyer DF, Buchheit KM, Boyce JA, Barrett NA, Laidlaw TM, Carroll SL, Colonna L, Tkachev V, Peterson CW, Yu A, Zheng HB, Gideon HP, Winchell CG, LJ in PL, Bingle CD, Snapper SB, Kropski JA, Theis FJ, Schiller HB, Zaragosi LE, Barbry P, Leslie A, Kiem HP, Flynn JL, Fortune SM, Berger B, Finberg RW, Kean LS, Garber M, Schmidt AG, Lingwood D, Shalek AK. Ordovas-Montanes J; HCA Lung Biological Network. Electronic address: lung-network@humancellatlas.org; HCA Lung Biological Network SARS-CoV-2 Receptor ACE2 Is an Interferon-Stimulated Gene in Human Airway Epithelial Cells and Is Detected in Specific Cell Subsets across Tissues Cell 2020;181(5):1016–35 e19. doi: 10.1016/j.cell.2020.04.035.
- [46] Merad M, Martin JC. Pathological inflammation in patients with COVID-19: a key role for monocytes and macrophages. Nat Rev Immunol 2020;20(6):355-62 Erratum, Nat Rev Immunol. 20(7):448. doi: 10.1038/s41577-020-0353-y. doi: 10.1038/s41577-020-0331-4.
- [47] Akbar AN, Gilroy DW. Aging immunity may exacerbate COVID-19. Science 2020;369(6501):256–7. doi: 10.1126/science.abb0762.
- [48] Franceschi C, Garagnani P, Vitale G, Capri M, Salvioli S. Inflammaging and 'Garbaging. Trends Endocrinol Metab 2017;28(3):199–212. doi: 10.1016/j. tem.2016.09.005.
- [49] De Maeyer RPH, van de Merwe RC, Louie R, Bracken OV, Devine OP, Goldstein DR, Uddin M, Akbar AN, Gilroy DW. Blocking elevated p38 MAPK restores efferocytosis and inflammatory resolution in the elderly. Nat Immunol 2020;21(6):615–25 Erratum: Nat Immunol. 2020 Jun;21(6):696. doi: 10.1038/s41590-020-0686-5. doi: 10.1038/s41590-020-0646-0.
- [50] Campisi J, Kapahi P, Lithgow GJ, Melov S, Newman JC, Verdin E. From discoveries in ageing research to therapeutics for healthy ageing. Nature 2019;571 (7764):183–92. doi: 10.1038/s41586-019-1365-2.
- [51] Furman D, Campisi J, Verdin E, Carrera-Bastos P, Targ S, Franceschi C, Ferrucci L, Gilroy DW, Fasano A, Miller GW, Miller AH, Mantovani A, Weyand CM, Barzilai N, Goronzy JJ, Rando TA, Effros RB, Lucia A, Kleinstreuer N, Slavich GM. Chronic inflammation in the etiology of disease across the life span. Nat Med 2019;25 (12):1822–32. doi: 10.1038/s41591-019-0675-0.
- [52] Ondondo BO. Fallen angels or risen apes? A tale of the intricate complexities of imbalanced immune responses in the pathogenesis and progression of immunemediated and viral cancers. Front Immunol 2014;5:90. doi: 10.3389/ fimmu.2014.00090.
- [53] Chauveau A, Tonnerre P, Pabois A, Gavlovsky PJ, Chatelais M, Coupel S, Charreau B. Endothelial cell activation and proliferation modulate NKG2D activity by

regulating MICA expression and shedding. J Innate Immun 2014;6(1):89-104. doi: 10.1159/000351605.

- [54] van Eeden C, Khan L, Osman MS. Cohen Tervaert JW (2020) Natural Killer Cell Dysfunction and Its Role in COVID-19. Int J Mol Sci 2021;21(17):6351. doi: 10.3390/ijms21176351.
- [55] Maucourant C, Filipovic I, Ponzetta A, Aleman S, Cornillet M, Hertwig L, Strunz B, Lentini A, Reinius B, Brownlie D, Cuapio A, Ask EH, Hull RM, Haroun-Izquierdo A, Schaffer M, Klingström J, Folkesson E, Buggert M, Sandberg JK, Eriksson LI, Rooyackers O, Ljunggren HG, Malmberg KJ, Michaëlsson J, Marquardt N, Hammer Q, Strålin K, Björkström NK. Karolinska COVID-19 Study Group (2020) Natural killer cell immunotypes related to COVID-19 disease severity. Sci Immunol 2021;5(50):eabd6832. doi: 10.1126/sciimmunol.abd6832.
- [56] Guthmiller JJ, Wilson PC. Remembering seasonal coronaviruses. Science 2020;370 (6522):1272–3. doi: 10.1126/science.abf4860.
- [57] MGH COVID-19 Collection & Processing Team Shrock E, Fujimura E, Kula T, Timms RT, Lee IH, Leng Y, Robinson ML, Sie BM, Li MZ, Chen Y, Logue J, Zuiani A, McCulloch D, Lelis FJN, Henson S, Monaco DR, Travers M, Habibi S, Clarke WA, Caturegli P, Laeyendecker O, Piechocka-Trocha A, Li JZ, Khatri A, Chu HY, Villani AC, Kays K, Goldberg MB, Hacohen N, Filbin MR, Yu XG, Walker BD, Wesemann DR, Larman HB, Lederer JA, Elledge SJ. Viral epitope profiling of COVID-19 patients reveals cross-reactivity and correlates of severity. Science 2020;370(6520):eabd4250. doi: 10.1126/science.abd4250.
- [58] Ng KW, Faulkner N, Cornish GH, Rosa A, Harvey R, Hussain S, Ulferts R, Earl C, Wrobel AG, Benton DJ, Roustan C, Bolland W, Thompson R, Agua-Doce A, Hobson P, Heaney J, Rickman H, Paraskevopoulou S, Houlihan CF, Thomson K, Sanchez E, Shin GY, Spyer MJ, Joshi D, O'Reilly N, Walker PA, Kjaer S, Riddell A, Moore C, Jebson BR, Wilkinson M, Marshall LR, Rosser EC, Radziszewska A, Peckham H, Ciurtin C, Wedderburn LR, Beale R, Swanton C, Gandhi S, Stockinger B, McCauley J, Gamblin SJ, McCoy LE, Cherepanov P, Nastouli E, Kassiotis G. Preexisting and de novo humoral immunity to SARS-CoV-2 in humans. Science 2020;370(6522):1339–43. doi: 10.1126/science.abe1107.
- [59] Scully EP, Haverfield J, Ursin RL, Tannenbaum C, Klein SL. Considering how biological sex impacts immune responses and COVID-19 outcomes. Nat Rev Immunol 2020;20(7):442–7. doi: 10.1038/s41577-020-0348-8.
- [60] T Takahashi, Ellingson MK, Wong P, Israelow B, Lucas C, Klein J, Silva J, Mao T, Oh JE, Tokuyama M, Lu P, Venkataraman A, Park A, Liu F, Meir A, Sun J, Wang EY, Casanovas-Massana A, Wyllie AL, Vogels CBF, Earnest R, Lapidus S, Ott IM, Moore AJ, Team Yale IMPACT Research, Shaw A, Fournier JB, Odio CD, Farhadian S, Dela Cruz C, Grubaugh ND, Schulz WL, Ring AM, Ko AI, Omer SB, Iwasaki A. Sex differences in immune responses that underlie COVID-19 disease outcomes. Nature 2020;588(7837):315–20. doi: 10.1038/s41586-020-2700-3.
- [61] C Lucas, Wong P, Klein J, Castro TBR, Silva J, Sundaram M, Ellingson MK, Mao T, Oh JE, Israelow B, Takahashi T, Tokuyama M, Lu P, Venkataraman A, Park A, Mohanty S, Wang H, Wyllie AL, Vogels CBF, Earnest R, Lapidus S, Ott IM, Moore AJ, Muenker MC, Fournier JB, Campbell M, Odio CD, Casanovas-Massana A, Team Yale IMPACT, Herbst R, Shaw AC, Medzhitov R, Schulz WL, Grubaugh ND, Dela Cruz C, Farhadian S, Ko AI, Omer SB, Iwasaki A. Longitudinal analyses reveal immunological misfiring in severe COVID-19. Nature 2020;584(7821):463–9. doi: 10.1038/s41586-020-2588-y.
- [62] Wang J, Li Q, Yin Y, Zhang Y, Cao Y, Lin X, Huang L, Hoffmann D, Lu M, Qiu Y. Excessive Neutrophils and Neutrophil Extracellular Traps in COVID-19. Front Immunol 2020;11:2063. doi: 10.3389/fimmu.2020.02063.
- [63] Nish S, Medzhitov R. Host defense pathways: role of redundancy and compensation in infectious disease phenotypes. Immunity 2011;34(5):629–36. doi: 10.1016/j.immuni.2011.05.009.
- [64] Mohamed MS, Moulin TC, Schiöth HB. Sex differences in COVID-19: the role of androgens in disease severity and progression. Endocrine 2021;71(1):3–8. doi: 10.1007/s12020-020-02536-6.
- [65] Pereda R, González D, Rivero HB, Rivero JC, Pérez A, Lopez LDR, Mezquia N, Venegas R, Betancourt JR, Domínguez RE, Nodarse H. Therapeutic Effectiveness of Interferon Alpha 2b Treatment for COVID-19 Patient Recovery. J Interferon Cytokine Res 2020;40(12):578–88. doi: 10.1089/jir.2020.0188.
- [66] Nacional Institutes of Health (NIH). COVID-19 Treatment Guidelines Interferons (Alfa, Beta) June 2015 Available from https://www.covid19treatmentguidelines. nih.gov/therapies/immunomodulators/interferons/ [Accessed 15th].
- [67] Alavi Darazam I, Shokouhi S, Pourhoseingholi MA, Naghibi Irvani SS, Mokhtari M, Shabani M, Amirdosara M, Torabinavid P, Golmohammadi M, Hashemi S, Azimi A, Jafarazadeh Maivan MH, Rezaei O, Zali A, Hajiesmaeili M, Shabanpour Dehbsneh H, Hoseyni Kusha A, Taleb Shoushtari M, Khalili N, Soleymaninia A, Gachkar L, Khoshkar A. Role of interferon therapy in severe COVID-19: the COVIFERON randomized controlled trial. Sci Rep 2021;11(1):8059. doi: 10.1038/s41598-021-86859-y.
- [68] Pandit A, Bhalani N, Bhushan BLS, Koradia P, Gargiya S, Bhomia V, Kansagra K. Efficacy and safety of pegylated interferon alfa-2b in moderate COVID-19: A phase II, randomized, controlled, open-label study. Int J Infect Dis 2021;105:516– 21. doi: 10.1016/j.ijid.2021.03.015.
- [69] Liu SY, Sanchez DJ, Aliyari R, Lu S, Cheng G. Systematic identification of type I and type II interferon-induced antiviral factors. Proc Natl Acad Sci U S A 2012;109 (11):4239–44. doi: 10.1073/pnas.1114981109.
- [70] Dai M, Xie T, Liao M, Zhang X, Feng M. Systematic identification of chicken type I, II and III interferon-stimulated genes. Vet Res 2020;51(1):70. doi: 10.1186/ s13567-020-00793-x.