DOI: 10.1111/cas.14852

REVIEW ARTICLE

Cancer Science Wiley

Revertant somatic mosaicism as a cause of cancer

Toshiya Inaba 💿 | Akiko Nagamachi

Department of Molecular Oncology and Leukemia Program Project, Research Institute for Radiation Biology and Medicine, Hiroshima University, Hiroshima, Japan

Correspondence Toshiya Inaba, 1-2-3 Kasumi, Minami-ku, Hiroshima 734-8553, Japan. Email: tinaba@hiroshima-u.ac.jp

Abstract

Revertant (somatic) mosaicism is a spontaneous correction of a causative mutation in patients with congenital diseases. A relatively frequent event, revertant mosaicism may bring favorable outcomes that ameliorate disorders, and is therefore called "natural gene therapy." However, it has been revealed recently that "overcorrection" of inherited bone marrow failure in patients with sterile alpha motif domain containing 9 (SAMD9)/9L syndromes by revertant mosaicism induces myelodysplastic syndrome (MDS) with monosomy 7 that occasionally proceeds to acute myelogenous leukemia (AML). In this review, we interpret very complex mechanisms underlying MDS/AML in patients with SAMD9/9L syndromes. This includes multiple myeloid tumor suppressors on the long arm of chromosome 7, all of which act in a haploinsufficient fashion, and a difference in sensitivity to interferon between cells carrying a mutation and revertants. Overcorrection of mutants by somatic mosaicism is likely a novel mechanism in carcinogenesis.

KEYWORDS

haploinsufficiency, revertant mosaicism, SAMD9/9L syndromes, somatic mutation, tumor suppressors

1 | INTRODUCTION

Somatic mutations, ranging from a single nucleotide mutation to the loss of chromosomes, will inevitably occur in mammals. The earlier a mutation occurs in postzygotic development, the larger the percentage of cells that will carry the mutation, resulting in "somatic mosaicism."¹ However, once development is complete, only mutations that occurred in long-lived cells, such as tissue stem cells, are retained indefinitely in a very low percentage of cells. The percentage increases when a cell gains an autonomous growth potential. This type of somatic mosaicism is called cancer. In addition, somatic mutations contribute to aging.²

Somatic mutations, however, do not always lead to unfavorable outcomes. For example, such mutations are indispensable for mammals in achieving diversity. More specifically, the cornerstones of acquired immunity are T and B lymphocytes that specifically target each of hundreds and thousands of harmful microorganisms. Needless to say, the mechanism of generating the variations in T/B lymphocytes depends heavily on somatic mutations, which also play critical roles in organs other than the immune system. For instance, the very high frequencies (up to 50%) of aneuploid hepatocytes are observed probably to adapt the exposure of countless xenobiotics.³ Even in the brain, where a vast majority of neurons cannot be renewed, mutations that occurred early in the development would remain present until old age, generating individual physiological diversity in brain function.⁴

Revertant (somatic) mosaicism, a term popularized by Jonkman et al,⁵ is one more favorable outcome of mutation, which is observed in patients carrying an inherited or de novo mutation that causes congenital disease. Since the first report of spontaneous reversion of Lesch-Nyhan mutations by *HPRT* gene rearrangement in 1988,⁶ hundreds of such cases have been reported. It has been revealed that in patients with congenital immunodeficiency, for example 30/272 (11%) cases of Wiskott–Aldrich syndrome,⁷ are ameliorated by a

This is an open access article under the terms of the Creative Commons Attribution-NonCommercial-NoDerivs License, which permits use and distribution in any medium, provided the original work is properly cited, the use is non-commercial and no modifications or adaptations are made. © 2021 The Authors. *Cancer Science* published by John Wiley & Sons Australia, Ltd on behalf of Japanese Cancer Association.

Wiley-Cancer Science

spontaneous additional somatic mutation that repairs or compensates the causal inherited mutation. Cases of revertant mosaicism have also been reported in various congenital diseases, including epidermolysis bullosa⁸ and Fanconi's anemia.⁹ Since this brings benefits to patients, revertant mosaicism is called "natural gene therapy." However, taking into consideration that mutations have both favorable and unfavorable aspects, revertant mosaicism might cause additional diseases, although these have not been reported until recently.

An unfavorable outcome by revertant mosaicism was eventually reported in patients with SAMD9/9L syndromes, a recently established category of inherited bone marrow failure (IBMF) syndromes.¹⁰ SAMD9/9L syndromes are caused by gain of function (g/f) mutations of either the *Samd9* or related *Samd9L* gene, which locates on the long arm of chromosome 7 (7q) in tandem. SAMD9/9L syndromes are characterized by the development of myelodysplastic syndromes (MDS) at extremely high frequency that sometimes progress to overt acute myeloid leukemia (AML). In the process of developing MDS, the region of 7q that contains the mutated *SAMD9/9L* gene is always lost, converting the genotype of bone marrow cells from *SAMD9/9L*^{+/mut} to *SAMD9/9L*^{+/-}. Thus, the elimination of the mutated *SAMD9/9L* gene by an "adaptation by aneuploidy" mechanism results in revertant mosaicism, which, however, brings on MDS/ AML, not an improvement in bone marrow failure.

In this review, we interpret the very complex mechanisms of how revertant mosaicism causes MDS/AML in patients with SAMD9/9L syndromes. This involves multiple myeloid tumor suppressors on 7q, all of which act in a haploinsufficient (h/i) fashion. Another important factor to promote myeloid malignancies is interferon (IFN). Finally, we will discuss whether this is a rare story of carcinogenesis or just the tip of the iceberg.

2 | TUMOR SUPPRESSORS ACTING IN A HAPLOINSUFFICIENT MANNER: ANOTHER PARADIGM OF ANTIONCOGENES

Monosomy 7 and the interstitial deletion of 7q [-7/del(7q)] are one of the most frequent (~15%) chromosomal abnormalities in adult patients with MDS and AML. -7/del(7q) is also frequently found in children with MDS and juvenile myelomonocytic leukemia (JMML). Intriguingly, as we discuss later, familial monosomy 7 syndrome, defined as bone marrow monosomy 7 occurring as the sole anomaly affecting more than two siblings, has been reported in 14 families (references in ref.¹¹), in which most patients are children or adolescents.

Investigating del(7q) cases provided researchers with an opportunity to identify the responsible myeloid tumor-suppressor gene(s). In the 1970s and 1980s, recessive tumor suppressors, such as *retinoblastoma* (*Rb*) and *tumor protein P53* (*TP53*) genes, were the focus of intensive research. These genes typically lose their function completely after the loss of one allele together with a loss of function (I/f) mutation on the remaining gene. Thus, a commonly deleted region (CDR) is an ideal guide to lead researchers to where the tumor suppressor gene is located. Once a CDR is identified, candidate genes within the CDR can be sequenced to find mutations. In spite of the enormous effort of laboratories worldwide using then-current techniques such as restriction fragment length polymorphism assessments, fluorescence in situ hybridization, and microsatellite surveys, the CDRs of MDS/AML patients with del(7q) identified by each laboratory did not overlap (reviewed in refs.^{12,13}). As a result, instead of narrowing down the location of the relevant CDRs, these were reported to be spread over the whole 7q region. The wide distribution of CDRs among del(7q)-myeloid malignancies was later confirmed by comparative genomic hybridization (CGH) microarray.¹⁴

The lack of CDRs in 7q suggests that myeloid tumor suppressor genes on 7q are totally different from the recessive-type classical tumor suppressors. Indeed, a relevant precedent exists: -5/del(5q). The deletion of -5/del(5q) is just a carbon copy of -7/del(7q) in that it is frequently detected in myeloid malignancies with broad and irregular borders of deletion, and has been explained by multiple genes that lose their myeloid tumor suppressor function by a one-allele loss of 5q.¹⁵ This suggests that multiple tumor suppressors that act in a haploinsufficient manner (h/i suppressors) are also involved in myeloid malignancies with -7/del(7q).

Although just the loss of one allele or I/f mutation of one gene encoding an h/i suppressor (= one hit) promotes cancer progression, little doubt exists that the effect is much smaller than the complete loss of the antitumor function of classical recessive tumor suppressors by two hits (eg, loss of one allele together with an I/f mutation in the remaining gene; Figure 1). If a one-allele loss of an h/i suppressor promotes carcinogenesis a great deal, the incidence of cancers will increase to levels that threaten the persistence of a species (eg, the very high incidence of cancers in patients with 13q- or Li-Fraumeni syndromes, who have a congenital one-allele loss of Rb or TP53 genes, respectively). Nevertheless, if h/i suppressor genes are located in a specific region of a chromosome, a deletion of this region (= one hit) causes the loss of multiple tumor suppressors, greatly promoting carcinogenesis. Localization of multiple h/i myeloid tumor suppressors to the specific region of chromosomes, such as 5q or 7q, can also explain why del(5q) or del(7q) is generally wide with no clear CDRs.

3 | H/I MYELOID TUMOR SUPPRESSORS IN 7q

The broad deletion regions with ambiguous borders of MDS/AML patients with del(7q) made it very difficult to identify tumor suppressor genes. Thus, it is not surprising that promising candidates have been identified only recently (Figure 2). Different strategies other than narrowing CDRs were necessary. One approach is to use microarray CGH to search for microdeletions in patients with myeloid malignancies that might be present on an apparently normal chromosome 7. By selecting JMML and JMML-like diseases, a common microdeletion spanning approximately 100 kb was identified in the

FIGURE 1 Two types of tumor suppressors. Classical recessive tumorsuppressor genes (upper panel), such as *RB*, *TP53*, *PTEN*, and *BRCA*. Cancer progression occurs only when all alleles on each gene lose their function by deletion, mutation or methylation. This is a rare event, but once it happens this would have a substantial effect on carcinogenesis. By contrast, tumor suppressor genes acting in a haploinsufficient (h/i) manner lose their function by lacking just one gene (lower panel). This will occasionally happen and damage is expected to be small Cancer Science - Wiley



• Tumor suppressors acting in a haploinsufficient (h/i) manner



FIGURE 2 Deletion of a chromosome region that contains many haploinsufficient (h/i) tumor suppressor genes would greatly increase cancer progression. Because considerable damage would be caused by even a partial deletion of the region, the deleted area in each patient would vary



7q21.3 sub-band that contains SAMD9/9L,¹⁶ details of which are described in the following sections. Similarly, from EVI1-dysregulated AML cell lines, two small microdeletions (0.39 and 1.33 Mb) in subband 7q36.1 were detected, the latter of which included the EZH2 gene^{17,18} that encodes a methyltransferase for lysine 27 of histone H3 (H3K27).¹⁹⁻²¹ It is well known that I/f mutations of EZH2 are frequently found (ca. 10%) in patients with MDS and related myeloproliferative neoplasms (MDS/MPN), as well as in patients with secondary AML.^{18,22,23} In addition, a focal deletion (8.8 Mb) at 7q35-36 encompassing the mixed-lineage leukemia protein (*MLL3*) gene (7q36.1) was isolated from a patient with relapsed AML showing a normal karyotype.²⁴ MLL3 possesses histone methyltransferase activity for lysine 4 of histone 3 (H3K4),^{25,26} and I/f mutations of *MLL3* have been identified in MDS and AML.^{27,28} The contribution of these four genes to the development of myeloid diseases has now been validated by gene targeting in mouse models.



FIGURE 3 Haploinsufficient (h/i) myeloid tumor suppressor genes on 7g

In addition, two more genes, cut like homeobox 1 (*CUX1*, 7q22.1) and Dedicator of cytokinesis 4 (*DOCK4*) were considered to be h/i myeloid tumor suppressors. *CUX1* encodes a homeobox transcription factor.^{29,30} Loss of one allele of this gene is frequently detected not only in myeloid tumors but also in uterine leiomyomas and breast cancers. Inactivating point mutations in one allele are also frequently found in cancers of the endometrium, large intestine, and lung.³¹ *DOCK4* (7q31.1) encoding a guanine exchange factor is disrupted in murine osteosarcoma cells,³² and its low expression has been linked to erythroid dysplasia.³³

4 | SAMD9/9L AS H/I MYELOID TUMOR SUPPRESSOR GENES

Human *SAMD9/9L* encode related cytosolic/endosomal proteins (60% amino acid identity). Interestingly, the distribution of these two genes in mammals is very odd. For example, (a) humans (and other higher primates), horses, and rats have both *SAMD9* and *SAMD9L*, (b) cows, sheep, and primitive primates (such as galagos) possess only *SAMD9*, and (c) cats, dogs, and mice have only *SAMD9L*.³⁴ This implies that the two gene products have common functions and can compensate for the biological functions of each other.

Using fibroblasts established from homo- and heterozygous *Samd9*L-deficient mice, it was revealed that Samd9L induce the homotypic fusion of primary/early endosomes to form sorting endosomes, which regulate endosomal trafficking including virus invasion and cytokine receptor metabolism.³⁵ Indeed, SAMD9/9L is a crucial factor for the defense against viruses. *Samd9L* gene was identified as IFN-inducible genes,³⁶ since the mouse *Samd9L* gene has IFN-responsive *cis* elements in the promoter, to which IFN regulatory factor 1 binds.³⁷ In addition, SAMD9/9L suppresses the replication of viruses, including Japanese encephalitis virus,³⁸ and serves as a barrier against cross-species poxvirus transmission.³⁹⁻⁴¹

Heterozygous (Samd9L^{+/-}) as well as homozygous (Samd9L^{-/-}) mice were found to develop MDS and die after 1.5 years.³⁵ Most Samd9L^{+/-} as well as Samd9L^{-/-} mice exhibited leukocytopenia and anemia with dysplasia in multiple hematopoietic lineages in their normal-to-hypercellular bone marrow. Competitive repopulation assays revealed that SAMD9L-deficiency confers a proliferative

advantage on hematopoietic stem/progenitor cells (HSPCs). Indeed, *SAMD9L*-deficient HSPCs possessed an enhanced sensitivity to cy-tokines, most likely due to disturbed metabolism of ligand-bound cytokine receptors.

5 | G/F MUTATIONS OF SAMD9/9L AS A CAUSE OF IBMF

In contrast to the involvement of SAMD9/9L-deficiencies in MDS/ myeloid leukemia in both the human and mouse, the g/f mutations of SAMD9/9L were identified in young IBMF patients, with or without additional nonhematopoietic symptoms (mainly degeneration of multiple organs), and were collectively defined as "SAMD9/9L syndromes."¹⁰ The entities initially reported were myelodysplasia, infection, growth restriction, adrenal hypoplasia, genital phenotypes, and enteropathy (MIRAGE) syndrome carrying SAMD9 mutations.^{42,43} and ataxia pancytopenia (AP) syndrome with SAMD9L mutations.⁴⁴⁻⁴⁶ The common symptom of these two syndromes is pancytopenia with hypocellular bone marrow in infancy that often requires transfusion but gradually improves over time. Degeneration of nonhematopoietic organs, such as the adrenal gland, testis/ovary, and cerebellum occurs. More recently, germline g/f mutations of the SAMD9/9L genes were identified at high frequencies in the cohorts of children and adolescents with IBMF⁴⁷ and isolated MDS at high frequencies.^{48,49} The latter cohorts include familial cases.

Mechanisms of how SAMD9/9L g/f mutations cause anemia or the degeneration of nonhematopoietic organs were analyzed by generating mice carrying a Samd9L mutation equivalent to the human SAMD9 mutation causative of MIRAGE syndrome.⁵⁰ These mice mimic the MIRAGE syndrome presenting with growth retardation, a short life, bone marrow failure, and multiorgan degeneration. In the erythroblasts of such mice, the endocytosis of transferrin and transferrin receptors is markedly slower than in normal erythroblasts, resulting in a decrease of iron uptake. Additionally, the internalization of c-Kit (the receptor for stem cell factor) in HSPCs is also delayed. In nonhematopoietic cells, in comparison, enhanced endocytosis of cytokine receptors, such as epidermal growth factor with activated lysosomes, degrades ligand-bound cytokine receptors, resulting in the downregulation of cytokine signals. In both surface receptors.

hematopoietic and nonhematopoietic cells, SAMD9/9L g/f mutants MDS with suppress cell growth and function through abnormal metabolism of HSPCs/-7.

6 | REVERTANT MOSAICISM AS A CAUSE OF MDS IN SAMD9/9L SYNDROME

Children with SAMD9/9L syndromes develop MDS with -7/del(7q) at extremely high frequencies that sometimes progresses to AML. The age of onset is mostly less than 5 years and, intriguingly, the mutated allele is always lost. This means the genotype of bone marrow cells changing from *SAMD9/9L*^{+/mut} to *SAMD9/9L*^{+/-} can "successfully" eliminate an IBMF-causative mutated *SAMD9/9L* gene (Figure 3). However, instead of an improvement in BMF by "revertant mosaicism" through a mechanism called "adaption by aneuploidy," patients developed new diseases, MDS/AML. Obviously, a loss of multiple h/i myeloid tumor suppressors on 7q, as mentioned above, was strongly involved in the development of myeloid malignancies, but it is most likely that two additional factors that are unique for patients with SAMD9/9L syndromes are also critical to promoting leukemogenesis.

One is the impaired proliferation potential of the surrounding bone marrow cells of patients with SAMD9/9L syndrome (namely, *SAMD9/9L*^{+/mut}; Figure 4). It has been reported that MDS cells from SAMD9/9L syndromes have few additional gene/chromosome alterations other than -7/del(7q),⁴⁹ in contrast to sporadic childhood MDS with -7/del(7q), which generally carries relevant gene mutations such as *GATA2* or those involved in the Ras pathway.⁵¹ Accordingly, it is assumed that HSPCs with -7/del(7q) (HSPC/-7) are susceptible to the development of MDS per se, but surrounding HSPCs suppress the expansion of HSPC/-7. When neighboring HSPCs are "weak" and unable to suppress HSPCs/-7, as is the case for the HSPCs of patients with SAMD9/9L syndrome, then HSPCs/-7 can develop into MDS without a long latency that allows additional mutation(s) to HSPCs/-7.

Gancer Science-Willey

The other factor is IFN. Patients with SAMD9/9L syndromes experience recurrent severe viral infections. The resulting elevation in IFN disturbs a balance between HSC self-renewal and differentiation, which is the fundamental regulatory mechanism of hematopoiesis.^{52,53} IFNγ signaling induces myeloid-biased HSC differentiation at the expense of self-renewal and differentiation to lymphoid and erythroid lineages. Because SAMD9/9L are IFN-responsive genes, the self-renewal of HSCs carrying a mutated SAMD9/9L will be further repressed by IFNy. Indeed, anemia and lymphocytopenia of patients with SAMD9/9L syndromes are worsened after episodes of viral infection. In addition, intraperitoneal injections of poly(I:C), an IFN inducer, into Samd9L^{+/mut} mice resulted in more profound anemia than Samd9L^{+/+} mice, while no significant hemoglobin reduction occurred in $Samd9L^{-/-}$ mice.⁵⁰ These data suggest that IFNy induced by recurrent viral infection facilitates clonal expansion of HSCs with -7/ del(7g) in the bone marrow of patients with SAMD9/9L.

This logic may extend to the pathogenesis of sporadic adult MDS. It is generally accepted that the accumulation of additional genetic and/or epigenetic alterations is required for HSPCs/-7 to develop MDS. This is assumed to be a reason why the great majority of MDS patients are >40 years of age. However, if the expansion potential of HSPCs/-7 is determined by the *relative* strength of the surrounding HSPCs, aging may also contribute to the development of MDS by "weakening" surrounding HSPCs to allow the expansion of HSPCs/-7. In addition, the elevation of IFN γ by viral infection or aberrant expression from bone marrow stroma cells could lead to a difference between HSPCs/-7 (SAMD9/9L^{+/-}) and surrounding cells (SAMD9/9L^{+/+}). Indeed, several lines of evidence revealed aberrant expression of IFN-inducible genes in MDS cells,⁵⁴ suggesting cross-talk of cytokines and IFN signaling contributes to the development of MDS.



FIGURE 4 A scheme of myelodysplastic syndrome (MDS) carrying -7/del(7q). In patients with SAMD9/9L syndromes, bone marrow cells with SAMD9/9L^{+/-} (revertants) show a high sensitivity to growth factors and a low sensitivity to (the suppressive effects of) interferon (IFN) γ . In addition, surrounding bone marrow cells (SAMD9/9L^{+/mut}) have a high sensitivity to IFN γ . As a result, the rapid expansion of a -7/del(7q) clone causes an "overcorrection," leading to MDS. This mechanism would be partially applied to sporadic MDS patients with -7/ del(7q) in old age

7 | DOES REVERTANT MOSAICISM CAUSE CANCER OTHER THAN MDS/AML IN SAMD9/9L SYNDROME?

Wiley-Cancer Science

In the case of SAMD9/9L syndrome, the accumulation of h/i myeloid tumor-suppressor genes on 7q "overcorrects" BMF, resulting in MDS/AML. Thus the answer to the question "Does revertant mosaicism cause cancers other than MDS/AML in SAMD9/9L syndromes?" most likely depends on whether h/i tumor-suppressor genes accumulate on a specific chromosome or not. Since nonrandom loss and/ or large deletions of chromosomes are frequently observed in a wide variety of cancers, we consider that SAMD9/9L syndromes are not alone in this regard.

DISCLOSURE

The authors have no conflict of interest.

ORCID

Toshiya Inaba D https://orcid.org/0000-0002-3455-6010

REFERENCES

- 1. Freed D, Stevens EL, Pevsner J. Somatic mosaicism in the human genome. *Genes*. 2014;5:1064-1094.
- Vijg J, Dong X. Pathogenic mechanisms of somatic mutation and genome mosaicism in aging. *Cell*. 2020;182:12-23.
- Duncan AW, Taylor MH, Hickey RD, et al. The ploidy conveyor of mature hepatocytes as a source of genetic variation. *Nature*. 2010;467:707-U793.
- Verheijen BM, Vermulst M, van Leeuwen FW. Somatic mutations in neurons during aging and neurodegeneration. *Acta Neuropathol.* 2018;135:811-826.
- Jonkman MF, Scheffer H, Stulp R, et al. Revertant mosaicism in epidermolysis bullosa caused by mitotic gene conversion. *Cell*. 1997;88:543-551.
- Yang TP, Stout JT, Konecki DS, Patel PI, Alford RL, Caskey CT. Spontaneous reversion of novel Lesch-Nyhan mutation by HPRT gene rearrangement. *Somat Cell Mol Genet*. 1988;14:293-303.
- Stewart DM, Candotti F, Nelson DL. The phenomenon of spontaneous genetic reversions in the Wiskott-Aldrich syndrome: a report of the workshop of the ESID Genetics Working Party at the XIIth Meeting of the European Society for Immunodeficiencies (ESID). Budapest, Hungary October 4–7, 2006. J Clin Immunol. 2007;27:634-639.
- 8. Lai-Cheong JE, McGrath JA, Uitto J. Revertant mosaicism in skin: natural gene therapy. *Trends Mol Med.* 2011;17:140-148.
- Kalb R, Neveling K, Nanda I, Schindler D, Hoehn H. Fanconi anemia: causes and consequences of genetic instability. *Genome Dyn*. 2006;1:218-242.
- Inaba T, Honda H, Matsui H. The enigma of monosomy 7. Blood. 2018;131:2891-2898.
- Gaitonde S, Boumendjel R, Angeles R, Rondelli D. Familial childhood monosomy 7 and associated myelodysplasia. J Pediatr Hematol Oncol. 2010;32:e236-e237.
- Honda H, Nagamachi A, Inaba T. -7/7q- syndrome in myeloidlineage hematopoietic malignancies: attempts to understand this complex disease entity. Oncogene. 2015;34:2413-2425.
- Todd R, Bia B, Johnson E, Jones C, Cotter F. Molecular characterization of a myelodysplasia-associated chromosome 7 inversion. Br J Haematol. 2001;113:143-152.

- Jerez A, Sugimoto Y, Makishima H, et al. Loss of heterozygosity in 7q myeloid disorders: clinical associations and genomic pathogenesis. *Blood.* 2012;119:6109-6117.
- 15. Ebert BL. Deletion 5q in myelodysplastic syndrome: a paradigm for the study of hemizygous deletions in cancer. *Leukemia*. 2009;23:1252-1256.
- Asou H, Matsui H, Ozaki Y, et al. Identification of a common microdeletion cluster in 7q21.3 subband among patients with myeloid leukemia and myelodysplastic syndrome. *Biochem Biophys Res Commun.* 2009;383:245-251.
- De Weer A, Poppe B, Vergult S, et al. Identification of two critically deleted regions within chromosome segment 7q35-q36 in EVI1 deregulated myeloid leukemia cell lines. *PLoS One.* 2010;5:e8676.
- Nikoloski G, Langemeijer SM, Kuiper RP, et al. Somatic mutations of the histone methyltransferase gene EZH2 in myelodysplastic syndromes. *Nat Genet*. 2010;42:665-667.
- 19. Cao R, Wang L, Wang H, et al. Role of histone H3 lysine 27 methylation in Polycomb-group silencing. *Science*. 2002;298:1039-1043.
- Czermin B, Melfi R, McCabe D, Seitz V, Imhof A, Pirrotta V. Drosophila enhancer of Zeste/ESC complexes have a histone H3 methyltransferase activity that marks chromosomal polycomb sites. *Cell.* 2002;111:185-196.
- Muller J, Hart CM, Francis NJ, et al. Histone methyltransferase activity of a Drosophila Polycomb group repressor complex. *Cell*. 2002;111:197-208.
- Bejar R, Stevenson K, Abdel-Wahab O, et al. Clinical effect of point mutations in myelodysplastic syndromes. N Engl J Med. 2011;364:2496-2506.
- 23. Ernst T, Chase AJ, Score J, et al. Inactivating mutations of the histone methyltransferase gene EZH2 in myeloid disorders. *Nat Genet*. 2010;42:722-726.
- 24. Chen C, Liu Y, Rappaport AR, et al. MLL3 is a haploinsufficient 7q tumor suppressor in acute myeloid leukemia. *Cancer Cell*. 2014;25:652-665.
- Ansari KI, Mandal SS. Mixed lineage leukemia: roles in gene expression, hormone signaling and mRNA processing. FEBS J. 2010;277:1790-1804.
- Herz HM, Hu D, Shilatifard A. Enhancer malfunction in cancer. *Mol Cell*. 2014;53:859-866.
- Kuhn MW, Radtke I, Bullinger L, et al. High-resolution genomic profiling of adult and pediatric core-binding factor acute myeloid leukemia reveals new recurrent genomic alterations. *Blood*. 2012;119:e67-75.
- Dolnik A, Engelmann JC, Scharfenberger-Schmeer M, et al. Commonly altered genomic regions in acute myeloid leukemia are enriched for somatic mutations involved in chromatin remodeling and splicing. *Blood.* 2012;120:e83-92.
- McNerney ME, Brown CD, Wang XY, et al. CUX1 is a haploinsufficient tumor suppressor gene on chromosome 7 frequently inactivated in acute myeloid leukemia. *Blood.* 2013;121:975-983.
- Ramdzan ZM, Nepveu A. CUX1, a haploinsufficient tumour suppressor gene overexpressed in advanced cancers. *Nat Rev Cancer*. 2014;14:673-682.
- 31. Wong CC, Martincorena I, Rust AG, et al. Inactivating CUX1 mutations promote tumorigenesis. *Nat Genet*. 2014;46:33-38.
- 32. Yajnik V, Paulding C, Sordella R, et al. DOCK4, a GTPase activator, is disrupted during tumorigenesis. *Cell*. 2003;112:673-684.
- Sundaravel S, Duggan R, Bhagat T, et al. Reduced DOCK4 expression leads to erythroid dysplasia in myelodysplastic syndromes. *Proc Natl Acad Sci USA*. 2015;112:E6359-6368.
- Yates A, Akanni WA-O, Amode MR, et al. Ensembl 2016. Nucleic Acids Res. 2016;44(D1):D710-D716.
- 35. Nagamachi A, Matsui H, Asou H, et al. Haploinsufficiency of SAMD9L, an endosome fusion facilitator, causes myeloid

malignancies in mice mimicking human diseases with monosomy 7. *Cancer Cell*. 2013;24:305-317.

- Pappas DJ, Coppola G, Gabatto PA, et al. Longitudinal systembased analysis of transcriptional responses to type I interferons. *Physiol Genomics*. 2009;38:362-371.
- Hershkovitz D, Gross Y, Nahum S, et al. Functional characterization of SAMD9, a protein deficient in normophosphatemic familial tumoral calcinosis. J Invest Dermatol. 2011;131:662-669.
- Zhang LK, Chai F, Li HY, Xiao G, Guo L. Identification of host proteins involved in Japanese encephalitis virus infection by quantitative proteomics analysis. J Proteome Res. 2013;12:2666-2678.
- Liu J, Wennier S, Zhang L, McFadden G. M062 is a host range factor essential for myxoma virus pathogenesis and functions as an antagonist of host SAMD9 in human cells. J Virol. 2011;85: 3270-3282.
- Nounamo B, Li Y, O'Byrne P, Kearney AM, Khan A, Liu J. An interaction domain in human SAMD9 is essential for myxoma virus hostrange determinant M062 antagonism of host anti-viral function. *Virology*. 2017;503:94-102.
- 41. Meng X, Zhang F, Yan B, et al. A paralogous pair of mammalian host restriction factors form a critical host barrier against poxvirus infection. *PLoS Pathog.* 2018;14:e1006884.
- Narumi S, Amano N, Ishii T, et al. SAMD9 mutations cause a novel multisystem disorder, MIRAGE syndrome, and are associated with loss of chromosome 7. Nat Genet. 2016;48:792-797.
- Buonocore F, Kuhnen P, Suntharalingham JP, et al. Somatic mutations and progressive monosomy modify SAMD9-related phenotypes in humans. J Clin Invest. 2017;127:1700-1713.
- Chen DH, Below JE, Shimamura A, et al. Ataxia-pancytopenia syndrome is caused by missense mutations in SAMD9L. Am J Hum Genet. 2016;98:1146-1158.
- 45. Tesi B, Davidsson J, Voss M, et al. Gain-of-function SAMD9L mutations cause a syndrome of cytopenia, immunodeficiency, MDS, and neurological symptoms. *Blood.* 2017;129:2266-2279.

 Gorcenco S, Komulainen-Ebrahim J, Nordborg K, et al. Ataxiapancytopenia syndrome with SAMD9L mutations. *Neurol Genet*. 2017;3:e183.

Cancer Science - Wiley

- Bluteau O, Sebert M, Leblanc T, et al. A landscape of germline mutations in a cohort of inherited bone marrow failure patients. *Blood*. 2018;131:717-732.
- Pastor VB, Sahoo SS, Boklan J, et al. Constitutional SAMD9L mutations cause familial myelodysplastic syndrome and transient monosomy 7. *Haematologica*. 2018;103:427-437.
- Schwartz JR, Ma J, Lamprecht T, et al. The genomic landscape of pediatric myelodysplastic syndromes. *Nat Commun.* 2017;8:1557.
- Nagamachi A, Kanai A, Nakamura M, et al. Multi-organ failure with abnormal receptor metabolism in mice mimicking Samd9/9L syndromes. J Clin Invest. 2021;131:e140147.
- Pastor V, Hirabayashi S, Karow A, et al. Mutational landscape in children with myelodysplastic syndromes is distinct from adults: specific somatic drivers and novel germline variants. *Leukemia*. 2017;31:759-762.
- 52. de Bruin AM, Voermans C, Nolte MA. Impact of interferon-gamma on hematopoiesis. *Blood.* 2014;124:2479-2486.
- 53. Morales-Mantilla DE, King KY. The role of interferon-gamma in hematopoietic stem cell development, homeostasis, and disease. *Curr Stem Cell Rep.* 2018;4:264-271.
- Pellagatti A, Cazzola M, Giagounidis AA, et al. Gene expression profiles of CD34+ cells in myelodysplastic syndromes: involvement of interferon-stimulated genes and correlation to FAB subtype and karyotype. *Blood*. 2006;108:337-345.

How to cite this article: Inaba T, Nagamachi A. Revertant somatic mosaicism as a cause of cancer. *Cancer Sci.* 2021;112:1383–1389. https://doi.org/10.1111/cas.14852