

**Original** Article

# Cytotoxic evaluation of different fractions of *Salvia chorassanica* Bunge on MCF-7 and DU 145 cell lines

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### Abstract

Because of antimicrobial, antioxidant, and anticancer potential, Salvia chorassanica Bunge (Lamiaceae) has been considered as a popular herb in Iranian traditional medicine. Previous studies have shown remarkable cytotoxic properties of the methanol, n-hexane and dichloromethane extract of S. chorassanica on human cervical cancer cells. To seek the therapeutic potentials of S. chorassanica, this study was undertaken to evaluate the cytotoxic activities of various extracts of this plant on human breast MCF-7 and prostate cancer DU 145 cells. The DU 145 cells were exposed to different concentrations of plant extracts (1-200 µg/ml). Cytotoxic activities were examined using alamarBlue<sup>®</sup> assay and apoptosis was assessed by acridine orange/propodium iodide double staining and evaluation of DNA fragmentation by flow cytometry. Our findings indicated that *n*-hexane and dichloromethane extracts had more cytotoxic activities against DU 145 and MCF-7 cell lines compared with other extracts (P<0.05). The acridine orange/propodium iodide staining showed apoptogenic properties of *n*-hexane and dichloromethane extracts which was consequently confirmed by flow cytometric histogram that exhibited an increase in sub-G1 peak in treated cells as compared with untreated cancer cell lines. Taken together, these observations demonstrated cytotoxic effects of S. chorassanica extracts on MCF-7 and DU 145 cell lines which is most likely exerted via apoptosis cell death. Therefore, further investigations on S. chorassanica extracts as potential chemotherapeutic agents are warranted.

Keywords: Salvia chorassanica; Lamiaceae; Cytotoxicity; Cell line

# **INTRODUCTION**

Cancer is one of the most dangerous multifactorial human disorders with high mortality and morbidity (1). Chemotherapy, radiotherapy. hormone therapy, immune therapy and surgery have been proposed effective modality for cancer treatment and prevention (2). In spite of the progression of cancer treatment approaches, it is still accounting as a prominent dreadful condition (3). Controlling cancer cell proliferation, reducing the risk of tumor progression, eradication improving malignancy have encouraged researchers to seek for novel

treatment options (4). In this regard, various natural product as therapeutic agents used in chemoprevention have been introduced through high throughput screenings (5). Many of the herbal natural compounds and purified secondary metabolites are used as nutriceuticals or dietary supplements (6). Herbal medicine maypropose better therapeutic approaches due to higher efficacy, minimal toxicity and lower cost of treatment (7). In addition, the whole natural extract represents the synergistic and antagonistic function of biochemicals compounds in the extract which improves the therapeutic efficacy (8).

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Naturally occurring substances exhibit pivotal role in elimination of malignant cells by modulation of apoptosis signaling (9,10). The programmed mode of cell death is an appreciate defense mechanism that exerted via self-destruction pathway and characterized with morphological and biochemical indicators such as DNA fragmentation, reducing of cell and membrane blebbing volume (11). Therefore, researchers have tried to discover innovative therapeutic agents accomplishing their cytotoxic effects through triggering of apoptosis in malignant cells (12-14). There are numerous reports on cytotoxic activities of some genus in Lamiaceae plants including Scutellaria (15), Lavandula (16) and Salvia (17). The genus Salvia (Lamiaceae) are traditional medicinal plants with unique biological properties which have been used due to their antimicrobial, antioxidant, and anti-cancer effects in folk medicine for many years (18).

The prominent constituents of this genus involve sesquiterpenoides, diterpenoides, triterpenes, essential oils and flavonoids (19,20). Salvia chorassanica is an Iranian endemic Salvia species which grows naturally in Khorasan Razavi province (18). There are a few literatures on biomedical efficacy of *S. chorassanica*.

In our two previous studies we have demonstrated the cytotoxic effect of *S. chorassanica* on leukemic and cervical cancer cells (19). Nevertheless, the comparative cytotoxic effects of different extracts of *S. chorassanica* on MCF-7 and DU 145 have not been reported previously. Thus, this study was planned to explore the cytotoxic effects of *S. chorassanica* on MCF-7 and DU 145 cell lines as two sex related common cancers in individuals.

#### MATERIALS AND METHODS

#### **Chemicals**

MCF-7 (breast cancer cells) and DU 145 (prostate cancer cells) were from National Cell Bank of Iran (NCBI). Roswell park memorial institute medium (RPMI)-1640 medium, fetal bovine serum (FBS), trypsinethylenediaminetetraacetic acid (trypsin-EDTA) and antibiotic (penicillin-streptomycin) were supplied by Gibco (USA). AlamarBlue<sup>®</sup> (resazurin) was from Sigma (Saint Louis, MO, USA). Acridine orange (AO) was procured from AppliChem (USA) and propodium iodide (PI) was obtained from Sigma (USA).

# **Plant materials**

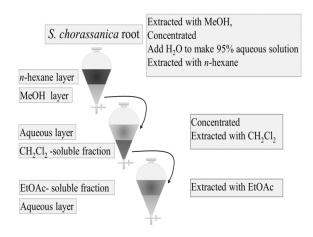
The roots of *S. chorassanica* were collected from Hosseinabad valley (2100 m height) in Pivejan, a village located 65 km south-west of Mashhad, Khorasan Razavi province, northeast of Iran. The plant was identified by Mr. M.R. Joharchi, from Ferdowsi University of Mashhad Herbarium (FUMH). Voucher specimen (No. 11289) was deposited in herbarium of Faculty of Pharmacy, Mashhad University of Medical Sciences, Iran.

#### **Plant extraction**

*n*-Hexane, dichloromethane (CH<sub>2</sub>Cl<sub>2</sub>), ethyl acetate (EtOAc) and water (H<sub>2</sub>O) extracts were prepared as previously described (21). Briefly, extracts were prepared after successive partitioning among different solvents started with crude dried methanol extract and stored at 4 °C until the use or analysis. A partitioning scheme of *S. chorassanica* methanol extract is presented in Fig. 1.

#### Sample preparation

To prepare the stock solutions (100 mg/ml), all extracts were dissolved in dimethyl sulfoxide (DMSO). The concentrations of 1-200  $\mu$ g/ml were then prepared by diluting these solutions with complete media, such that the final concentrations of DMSO did not exceed 0.5%.



**Fig. 1.** Extraction scheme of *Salvia chorassanica* using immiscible solvents.

#### Cells and culture

MCF-7 and DU 145 cells were obtained from the Pasteur Institute of Iran and grown in RPMI-1640 medium containing 10% FBS supplemented with 1% antibiotic (penicillin /streptomycin) at 37 °C in a humidified, 5%  $CO_2$  incubator.

#### Cytotoxic assay

The inhibitory effects of all prepared extracts of *S. chorassanica* on the proliferation of MCF-7 and DU 145 cancer cells were measured by alamarBlue<sup>®</sup> assay (22). Briefly, the cancer cells were cultured overnight at a concentration of  $10^4$  cells/well in 96-well plates. Then incubated with appropriate concentrations (1-200 µg/ml) of methanol, *n*-hexane, CH<sub>2</sub>Cl<sub>2</sub>, EtOAc and H<sub>2</sub>O extracts. The alamarBlue<sup>®</sup> assay was performed after 48 h incubation with extracts. Ten µl of alamarBlue<sup>®</sup> solution was added and incubated for 4 h, and eventually optical density was recorded at 570 and 600 nm using spectrophotometer (Epoch, USA).

#### Cellular morphology

MCF-7 and DU 145 cells  $(2 \times 10^5 \text{ cells/well})$ were grown in six-well culture plates in RPMI-1640 supplemented medium and incubated with cytotoxic extracts at a specified concentration for 24 h. After the desired time the cytomorphological changes were visualized under an inverted microscope (Bio Photonic, Brazil) and photographs were taken.

### Apoptosis assay

### Acridine orange/propidium iodide staining

The human MCF-7 and DU 145 cells (2  $\times$  10<sup>5</sup> cells/well) incubated with a defined concentrations of cytotoxic extracts for 24 h, then cells were trypsinized and stained with 10 µl of a dye mixture comprising 100 µg/ml PI and 100 µg/ml AO (23). The cells were then mounted on coverslips and observed under a fluorescence microscope.

# Apoptosis evaluation by detection of sub-G1 peak

The cultured MCF-7 and DU 145 cells were treated with a medium containing certain concentrations of cytotoxic extracts. Then the cells were washed twice and resuspended in PI solution containing 0.1% sodium citrate plus 0.1% Triton X-100 and incubated at 37 °C for 30 min.

The fluorescence of stained cells was evaluated using a FACScan laser flow cytometer (Becton Dickinson, NJ, USA), and the population of cells was calculated using the WinMDI 2.8 software.

# Statistical analysis

The results are expressed as the mean  $\pm$  SEM and experiments were carried out in triplicate. One-way ANOVA used for statistical analysis and the *P* values  $\leq 0.05$  was considered significant.

# RESULTS

#### Cytotoxicity of different extracts of S. chorassanica

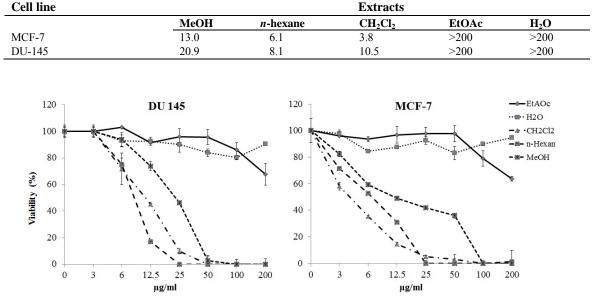
The data obtained from alamarBlue<sup>®</sup> assay indicated that the CH<sub>2</sub>Cl<sub>2</sub> extract had the most cytotoxic effect among other extracts of the *S. chorassanica* against MCF-7 cells (P<0.05), while DU 145 cells were more sensitive to the *n*-hexane extract in comparison with others (P<0.05) (Fig. 2).

IC<sub>50</sub> values for various concentrations of *S. chorassanica* extracts against MCF-7 and DU 145 cell lines are givenin Table 1. The IC<sub>50</sub> value of CH<sub>2</sub>Cl<sub>2</sub> extract against MCF-7 cells after 48 h was determined as 3.8 µg/ml and the IC<sub>50</sub> value of *n*-hexane extract against DU 145 cell line after 48 h was found to be 8.1 µg/ml.

# AO/PI staining and morphological characterization

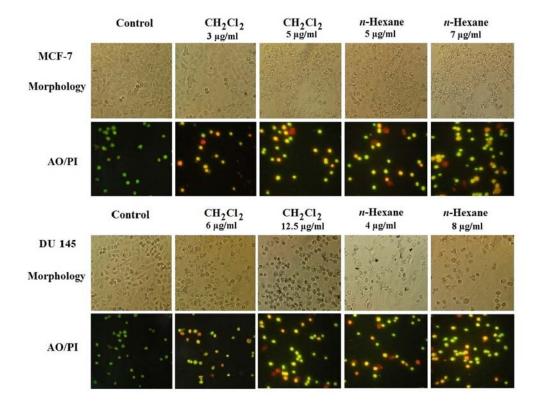
Apoptosis is a regulated mechanistic cell death identified by appearance of morphological and cellular features. Morphological alterations of MCF-7 and DU 145 cancer cells after incubation with efficient concentrations of the most cytotoxic extracts were identified using phase contrast inverted and fluorescent microscope.

As shown in Fig. 3 the untreated breast and prostate cancer cells exhibited polygonal shapes, while MCF-7 and DU 145 cells exposed to cytotoxic concentrations of CH<sub>2</sub>Cl<sub>2</sub> and *n*-hexane extracts represented cell shrinkage and loss of plasma membrane



**Table 1.**  $IC_{50}$  values (µg/ml) of various extracts of *Salvia chorassanica* in MCF-7 and DU 145 cell lines.

**Fig. 2.** The cytotoxic effect of various extracts obtained from *Salvia chorassanica*. MCF-7 and DU 145 cancer cells were incubated 48 h in the presence of various concentrations of MeOH, *n*-hexane,  $CH_2Cl_2$ , EtOAc, and  $H_2O$  extracts of *Salvia chorassanica*. Growth inhibitory effect was determined by alamarBlue<sup>®</sup> assay. Each point represents the mean  $\pm$  SEM of three replicates.



**Fig. 3.** Apoptotic morphological features under exposure with various concentrations of  $CH_2Cl_2$  and *n*-hexane extracts of *Salvia chorassanica* on MCF-7 and DU 145 cancer cells. Morphological visualization performed under inverted microscope and acridine orange/propodium iodide double staining was examined under fluorescent microscope.

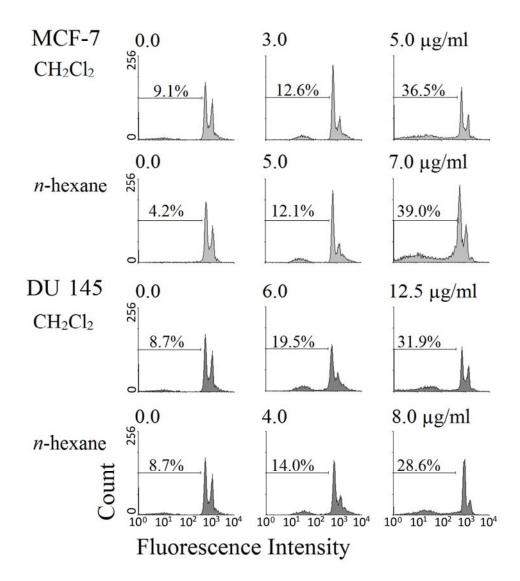
integrity as the most dominant characteristics of apoptosis induction.

To evaluate whether cytotoxic extracts can trigger apoptosis, the MCF-7 and DU 145 cells were double stained with AO/PI.

As illustratedin Fig. 3, AO/PI fluorescence double staining showed that MCF-7 and DU 145 cells treated with  $IC_{50}$  concentrations of *S. chorassanica* extracts were stained green to yellow and yellow to orange color which represents the involvement of apoptosis in cytotoxic activity of *S. chorassanica*.

# Sub-G1 peak and DNA fragmentation

The pro-apoptotic effect of *S. chorassanica* was evaluated by flow cytometric analysis of cells in hypotonic phosphate citrate buffer containing PI. As shown in Fig. 4 treatment of MCF-7 cells with  $CH_2Cl_2$  and *n*-hexane extracts of *S. chorassanica* significantly increased the sub-G1 peak, which was indicative of DNA fragmentation. In Fig. 4 flow cytometric histograms of prostate cancer cells treated with *n*-hexane and  $CH_2Cl_2$  for 48 h showed a concentration-dependent sub-G1 peak increment.



**Fig. 4.** Detection of apoptosis using flow cytometric analysis of cells stained with propodium iodide. MCF-7 and DU 145 cells remained untreated or treated with  $CH_2Cl_2$  and *n*-hexane extracts of *Salvia chorassanica*. As shown increasing sub-G1 peak in treated cells in comparison with control untreated cells demonstrated the involvement of apoptosis.

#### DISCUSSION

Breast cancer is the most serious type of malignancies in women which has the metastasis potential to other parts of the body and prostate cancer is the common leading cause of cancer-related deaths in men all over the world (24,25). Despite recent progression in cancer medicinal modalities, attempting to find alternative strategies to eradicate invasive cancer cells is going on (6). However, folk utilized natural extract medicine with therapeutic including antiproperties inflammatory, antioxidant and cytotoxic characteristic which have been isolated from terrestrial and marine resources. Many of the investigated substances play an important role in cancer cell elimination and have entered to clinical trials that means nature represents a promising alternative to enhance efficacy of cancer chemotherapy strategies (2).

The results of present study showed that S. chorassanica induce cytotoxicity against MCF-7 and DU 145 cell lines. n-Hexane and CH<sub>2</sub>Cl<sub>2</sub> extracts of S. chorassanica are among the active fractions with high cytotoxic effects against MCF-7 and DU 145 cancer cells while the methanol extract had moderate cytotoxicity and H<sub>2</sub>O, EtOAc extracts showed minimal effects. Different potency of the extracts can be associated with distinct constituents in each extracts. It should be noted that this finding is in accordance with our previous study. Previously we have indicated that CH<sub>2</sub>Cl<sub>2</sub> and n-hexane extracts of S. chorassanica have cytotoxic effect against HeLa cervical cancer cells with IC<sub>50</sub> value of 2.38 and 5.45 µg/ml for CH<sub>2</sub>Cl<sub>2</sub> and *n*-hexane extracts, respectively via induction of apoptosis (19). In addition, in a recent study we evaluated the cytotoxic and apoptosis inducing effect of S. chorassanica extracts and their active constituents such as taxodione, ferruginol and 6-hydroxysalvinolone against K-562 and HL-60 cancer cells and showed that CH<sub>2</sub>Cl<sub>2</sub> is the most cytotoxic extract and ferruginol is the most potent compounds among tested constituents Apoptotic resistance is (18).а kev complication that interfere with cancer therapeutic methods and therefore, recruitment of apoptosis signaling possess beneficial effects and improve the cytotoxic potential of phytochemicals (26).

The pro apoptotic activity of *S. chorassanica* extracts was evaluated using fluorescence microscopy. The morphological alterations such as cell shrinkage, membrane blebbing, increased cytoplasmic density, nuclear disintegration, activation of caspase cascade and formation of apoptotic bodies were considered as obvious apoptosis hallmarks (18).

The fluorescence micrographs showed that untreated cancer cells exhibited bright green fluorescence as the live cells which indicates intact membrane excluded PI, but permits uptake of AO. On the other hand, MCF-7 and DU-145 treated cells revealed more yelloworange fluorescence which demonstrated apoptotic cells could not exclude the dyes and stained with yellow-orange fluorescence.

Apoptosis inducing activity of S. *chorassanica* extracts was further assessed by flow cytometry using PI to evaluate cell membrane integrity and DNA fragmentation.

It should be notified that nuclear fragmentation produces small fragments of DNA that can be stained with a quantitative DNA-binding dye such as PI in a hypotonic phosphate-citrate buffer. It is proved that apoptosis cell death characterized by accumulation of a subpopulation of cells in sub-G1 area which is indicating endonuclease activation and subsequent leakage of DNA from the cells. Flow cytometric histogram of the S. chorassanica extract treated cells showed a sub-G1 peak in MCF-7 and DU-145 cells which exhibit the involvement of an apoptotic process in the cells. Collectively, apoptosis recruitment has pivotal role in cytotoxic mechanism of S. chorassanica extracts.

There are different reports on biomedical properties of *Salvia* genus (18). Xavier and colleagues examined the cytotoxic effect of *S. fruticosa* and *S. officinalis* on two human colorectal cancer cells and showed that these extracts can significantly inhibit colorectal cancer cell growth and induce apoptosis in both colorectal cell line (27). Tundis and coworkers have proven the growth inhibitory effect of extracts and isolated metabolites of S. leriifolia Benth. on amelanotic melanoma, colorectal adenocarcinoma, lung large cell malignant carcinoma, melanoma and hepatocellular carcinoma cells and indicated that the *n*-hexane and  $CH_2Cl_2$  extracts showed strongest cytotoxic activity against the melanoma cells and the EtOAc extract was the most active extract against lung large cell carcinoma cell line. Nevertheless, buchariol, a sesquiterpene obtained from CH<sub>2</sub>Cl<sub>2</sub> extracts and naringenin, a flavonoid isolated from the EtOAc extract showed remarkable cytotoxicity against melanoma and lung carcinoma (28). Recently, Zare Shahneh and colleagues reported that S. officinalis L. methanol extract could be cytotoxic against lymphoma and leukemiacells through apoptosis induction (29). However, there are rare information related with therapeutic efficacy of Iranian Salvia species and this study proved the cytotoxic and apoptogenic effects of various extracts of S. chorassanica on two frequent cancer cell lines including human breast carcinoma and prostate cancer cells.

# CONCLUSION

Taken together, the present study proved the cytotoxic properties of  $CH_2Cl_2$  and *n*-hexane extracts of *S. chorassanica* that proposed this popular plant for anticancer evaluations. There is need for further investigation and to explore the accurate mechanisms involved in apoptosis cell death and to provide more convincing evidence related to anticancer potential of *S. chorassanica* extract and its active compounds.

#### ACKNOWLEDGMENTS

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#### REFERENCES

1. Neergheen VS, Bahorun Th, Taylor EW, Jen LS, Aruoma OI. Targeting specific cell signaling transduction pathways by dietary and medicinal phytochemicals in cancer chemoprevention. Toxicology. 2010;278:229-41.

- Saunders FR, Wallace HM. On the natural chemoprevention of cancer. Plant Physiol Biochem. 2010;48:621-626.
- Boopathy NS, Kathiresan K. Anticancer drugs from marine flora: An overview. J Oncol. 2010;2010: ID 214186.
- Ganesh G, Abhishek T, Saurabh M, Sarada NC. Cytotoxic and apoptosis induction potential of *Mimusops elengi* L. in human cervical cancer (SiHa) cell line. J King Saud Univ Sci. 2014;26:333-337.
- Weli AM, AL-Hinai JRK, Al-Mjrafi JMA, Alnaaimi JRS, Hossain MA, Saeed S, *et al.* Effect of different polarities leaves crude extracts of Omani *Juniperus excelsa* on antioxidant, antimicrobial and cytotoxic activities and their biochemical screening. Asian Pac J Reprod. 2014;3:218-223.
- Nobili S, Lippi D, Witort E, Donnini M, Bausi L, Mini E, *et al.* Natural compounds for cancer treatment and prevention. Pharmacol Res. 2009;59:365-378.
- Kuppusamy P, Yusoff MM, Maniam GP, Govindan N. A case study regulation and functional mechanisms of cancer cells and control its activity using plants and their derivatives. J Pharm Res. 2013;6:884-892.
- Anter J, Fernández-bedmar Z, Villatoro-pulido M, Demyda-peyras S, Moreno-millán M, Alonsomoraga Á, *et al.* A pilot study on the DNA-protective, cytotoxic, and apoptosis-inducing properties of oliveleaf extracts. Mutat Res. 2011;723:165-170.
- Portt L, Norman G, Clapp C, Greenwood M, Greenwood MT. Anti-apoptosis and cell survival: a review. Biochim Biophys Acta. 2011;1813:238-259.
- Sun SY, Hail N, Lotan RJ. Apoptosis as a novel target for cancer chemoprevention. J Natl Cancer Inst. 2004;4:662-672.
- Thangam R, Sathuvan M, Poongodi A, Suresh V, Pazhanichamy K, Sivasubramanian S, *et al.* Activation of intrinsic apoptotic signaling pathway in cancer cells by *Cymbopogon citratus* polysaccharide fractions. Carbohydr Polym. 2014;107:138-150.
- 12. Felipe KB, Kviecinski MR, da Silva FO, Bücker NF, Farias MS, Porfirio Will Castro LSE, *et al.* Inhibition of tumor proliferation associated with cell cycle arrest caused by extract and fraction from *Casearia sylvestris* (Salicaceae). J Ethnopharmacol. 2014;155:1492-1499.
- Shojaee S, Parhiz H, Eshaghi A, Emami SA, Asili J, Mosaffa F. *In vitro* protective effects of *Scutellaria litwinowii* root extract against H<sub>2</sub>O<sub>2</sub>-induced DNA damage and cytotoxicity. J Complement Integr Med. 2014;11:121-127.
- 14. Ahmadi F, Mojarrab M, Ghazi-Khansari M, Hosseinzadeh L. A semipolar fraction of petroleum ether extract of *Artemisia aucheri* induces apoptosis and enhances the apoptotic response to doxorubicin in human neuroblastoma SKNMC cell line. Res Pharm Sci. 2015;10:335-344.

- Tayarani-Najaran Z, Mousavi SH, Asili J, Emami SA. Growth-inhibitory effect of *Scutellaria lindbergii* in human cancer cell lines. Food Chem Toxicol. 2010;48:599–604.
- 16. Tayarani-Najaran Z, Amiri A, Karimi G, Emami SA, Asili J, Mousavi SH. Comparative studies of cytotoxic and apoptotic properties of different extracts and the essential oil of *Lavandula angustifolia* on malignant and normal cells. Nutr Cancer. 2014;66:424-434.
- Tayarani-Najaran Z, Asili J, Aioubi E, Emami SA. Growth inhibition and apoptosis induction of *Salvia chloroleuca* on MCF-7 breast cancer cell line. Iran J Pharm Res. 2013;12:789-799.
- Tayarani-Najaran Z, Mousavi SH, Tajfard F, Asili J, Soltani S, Hatamipour M, *et al.* Cytotoxic and apoptogenic properties of three isolated diterpenoids from *Salvia chorassanica* through bioassay-guided fractionation. Food Chem Toxicol. 2013;57: 346-351.
- Parsaee H, Asili J, Mousavi SH, Soofi H, Emami SA, Tayarani-Najaran Z. Apoptosis induction of *Salvia chorassanica* root extract on human cervical cancer cell line. Iran J Pharm Res. 2013;12:75-83.
- Kharazian N. Flavonoid constituents in some of endemic *Salvia* L. (Lamiaceae) species in Iran. Res Pharm Sci. 2012;7:S752.
- 21. Boozari M, Mohammadi A, Asilia A, Emami SA, Tayarani-Najaran Z. Growth inhibition and apoptosis induction by *Scutellaria pinnatifida* A. Ham. on HL-60 and K562 leukemic cell lines. Environ Toxicol Pharmacol. 2015; 39:307-312.
- 22. O'Brien J, Wilson I, Orton T, Pognan F. Investigation of the Alamar Blue (resazurin) fluorescent dye for the assessment of mammalian cell cytotoxicity. Eur J Biochem. 2000;267: 5421-5426.
- Cury-Boaventura MF, Pomp C, Curi R. Comparative toxicity of oleic acid and linoleic acid on Jurkat cells. Clin Nutr. 2004;23:721-732.
- 24. Angelova S, Gospodinova Z, Krasteva M, Antov G, Lozanov V, Bozhanov S, *et al.* Antitumor activity of Bulgarian herb *Tribulus terrestris* L. on human breast cancer cells. J BioSci Biotech. 2013;2:25-32.
- 25. Chen P, Shih Y, Huang H, Cheng H. Diosgenin, a steroidal saponin, inhibits migration and invasion of human prostate cancer PC-3 cells by reducing matrix metalloproteinases expression. PLoS One. 2011;6:e20164.
- 26. Mousavi Sh, Tayrani-najaran Z, Hersey P. Apoptosis: from signalling pathways to therapeutic tools. Iran J Basic Med Sci. 2008;11:121-142.
- 27. Xavier CPR, Lima CL, Ferreira MF, Pereira-wilson C. Salvia fruticosa, Salvia officinalis, and rosmarinic acid induce apoptosis and inhibit proliferation of human colorectal cell lines: The role in MAPK/ERK pathway. Nutr Cancer. 2009;61: 564-571.
- Tundis R, Loizzo MR, Menichini F, Bonesi M, Colica C, Menichini F. *In vitro* cytotoxic activity of extracts and isolated constituents of *Salvia leriifolia*

Benth. against a panel of human cancer cell lines. Chem Biodivers. 2011;8:1152-1162.

29. Zare Shahneh F, Valiyari S, Baradaran B, Abdolalizadeh J, Bandehagh A, Azadmehr A, *et al.* Inhibitory and cytotoxic activities of *Salvia officinalis* L. extract on human lymphoma and leukemia cells by induction of apoptosis. Adv Pharm Bull. 2013;3:51-55.