




# Draft Genome Sequences of Three *Salmonella enterica* Serovar 4,[5],12:i:– Strains and One *S. enterica* Serovar Typhimurium Strain, Isolated in Brazil

Ana Isabela Lopes Sales,<sup>a</sup>  Guilherme Paier Milanez,<sup>b</sup> Leandro C. Nascimento,<sup>c</sup> Camila Pinheiro do Carmo,<sup>b</sup> Fernanda Luz Paulino da Costa,<sup>b</sup> Gonçalo Amarante Guimarães Pereira,<sup>b</sup> Roberto Martinez,<sup>d</sup> Marcelo Brocchi<sup>b</sup>

<sup>a</sup>Departamento de Biologia Celular, Molecular e Bioagentes Patogênicos, Faculdade de Medicina de Ribeirão Preto, Universidade de São Paulo (USP), Ribeirão Preto, SP, Brazil

<sup>b</sup>Departamento de Genética, Evolução Microbiologia e Imunologia, Instituto de Biologia, Universidade Estadual de Campinas (UNICAMP), Campinas, SP, Brazil

<sup>c</sup>Laboratório de Tecnologias de Alto Desempenho (LaCTAD), Universidade Estadual de Campinas (UNICAMP), Campinas, SP, Brazil

<sup>d</sup>Departamento de Clínica Médica, Faculdade de Medicina de Ribeirão Preto, Universidade de São Paulo (USP), Ribeirão Preto, SP, Brazil

**ABSTRACT** Draft genomes of three *Salmonella enterica* 4,[5],12:i:– (STi) strains isolated from human infections were obtained using Illumina sequencing. They were negative for the *fljBA* operon but positive for *hin*, and k-mer analyses revealed their identity as *S. enterica* 4,[5],12:i:– 08-1736 and *S. Typhimurium*. A draft *S. Typhimurium* sequence is described for comparison.

*Salmonella enterica*, an important human and animal pathogen (1, 2), is often associated with antibiotic resistance (3). This species is classified into different serotypes according to its somatic, flagellar, and, when expressed, capsular antigens (4). It is classified as either typhoid *Salmonella* (TS) or nontyphoid *Salmonella* (NTS) based on the serovar and host specificity. *S. enterica* subsp. *enterica* serovar Typhimurium (antigenic formula *S. enterica* I 4,[5],12:i:1,2) is among the most frequently identified NTS serovars isolated from human infections (2). Characteristics of this serovar include the nonconcomitant expression of two flagellar antigens (flagellar phases I and II), encoded by the *fliC* and *fljB* genes, respectively (5). Expression of *fliC* is negatively regulated by the *fljA* gene product, which forms an operon with *fljB* (6–8). The promoter of the *fljBA* operon extends into the *hin* sequence, a DNA fragment that is able to invert its orientation. Therefore, the type of flagellar antigen expressed is governed by the orientation of *hin* (5). Since the last century, *S. Typhimurium* variants unable to express flagellar phase II antigen have been isolated at an increased rate, becoming emergent an *S. enterica* serotype (9). *S. enterica* 4,[5],12:i:– originated from *S. Typhimurium* through the deletion of the *fljBA* operon. Different clones are distinguished based on the molecular events responsible for the *fljBA* deletion (9). Here, we describe the draft genome of three strains of *S. enterica* 4,[5],12:i:– (607STi, 633STi, and 691STi) and one of *S. Typhimurium* (662ST), all isolated in Brazil from human infections. Genomic DNA was prepared using the Wizard R genomic DNA purification kit (Promega, USA) according to manufacturer's instructions. Genomic DNA integrity was assessed by electrophoresis on 1% agarose gel. Library preparation was performed using the Nextera paired-end sample preparation kit and TruSeq DNA PCR-free LT sample prep kit and processed in a HiSeq 2500 system (Illumina, San Diego, CA). Genome assembly was performed with Velvet (10) and annotation with RAST (11).

The number of contigs was 30 or 36, and the mean scaffold length was 134,767 to 163,767 bp. The GC content of all strains was 52.2%. Bacterial typing was performed

Received 7 May 2018 Accepted 7 May 2018 Published 5 July 2018

**Citation** Sales AIL, Milanez GP, Nascimento LC, do Carmo CP, da Costa FLP, Pereira GAG, Martinez R, Brocchi M. 2018. Draft genome sequences of three *Salmonella enterica* serovar 4,[5],12:i:– strains and one *S. enterica* serovar Typhimurium strain, isolated in Brazil. *Genome Announc* 6:e00488-18. <https://doi.org/10.1128/genomeA.00488-18>.

**Copyright** © 2018 Sales et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Ana Isabela Lopes Sales, [belasales@gmail.com](mailto:belasales@gmail.com), or Marcelo Brocchi, [mbrocchi@unicamp.br](mailto:mbrocchi@unicamp.br).

with MLST 1.7, a platform for multilocus sequence typing (MLST) from assembled genomes (12). All strains were typed as sequence type 19 (ST-19), although the *S. Typhimurium* strain did not match perfectly for the *thra* locus due to a 1-base substitution. The ST-19 type is common for *S. Typhimurium*.

The prediction of prokaryotic species by k-mer genomic comparisons using Kmer-Finder 2.0 (13) indicated high scores for *S. enterica* 4,[5],12:i:– strain 08-1736, a 4,[5],12:i:– strain, and other *S. Typhimurium* strains. Scores for other *S. enterica* serovars were lower. BLAST analyses indicated an absence of the *fljBA* operon in the *S. enterica* 4,[5],12:i:– strains, though the *hin* sequence was present. The *S. Typhimurium* strain was positive for both sequences.

In conclusion, the three sequenced 4,[5],12:i:– strains presented high identity to *S. enterica* 4,[5],12:i:– strain 08-1736 and *S. Typhimurium*. Further analyses are under way to characterize this *S. Typhimurium* variant serovar in Brazil.

**Accession number(s).** This whole-genome shotgun project has been deposited in GenBank under the accession no. [QAWG00000000](#), [QAWH00000000](#), [QAWI00000000](#), and [QAWJ00000000](#). The versions described in this paper are the first versions, QAWG01000000, QAWH01000000, QAWI01000000, and QAWJ01000000.

## ACKNOWLEDGMENTS

This work was supported by grants from Fundação de Amparo à Pesquisa do Estado de São Paulo (FAPESP 2003/00501-8). A.I.L.S. was a fellow of the Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq 142048/2003-8). M.B. is a fellow of the Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq 309800/2015-3).

## REFERENCES

- Andrews-Polymeris HL, Bäumlér AJ, McCormick BA, Fang FC. 2010. Taming the elephant: *Salmonella* biology, pathogenesis, and prevention. *Infect Immun* 78:2356–2369. <https://doi.org/10.1128/IAI.00096-10>.
- Crump JA, Sjölund-Karlsson M, Gordon MA, Parry CM. 2015. Epidemiology, clinical presentation, laboratory diagnosis, antimicrobial resistance, and antimicrobial management of invasive *Salmonella* infections. *Clin Microbiol Rev* 28:901–937. <https://doi.org/10.1128/CMR.00002-15>.
- Fernandes SA, Camargo CH, Francisco GR, Bueno MFC, Garcia DO, Doi Y, Casas MRT. 2017. Prevalence of extended-spectrum  $\beta$ -lactamases CTX-M-8 and CTX-M-2-producing *Salmonella* serotypes from clinical and nonhuman isolates in Brazil. *Microb Drug Resist* 23:580–589. <https://doi.org/10.1089/mdr.2016.0085>.
- Agbaje M, Begum RH, Oyekunle MA, Ojo OE, Adenubi OT. 2011. Evolution of *Salmonella* nomenclature: a critical note. *Folia Microbiol* 56:497–503. <https://doi.org/10.1007/s12223-011-0075-4>.
- McQuiston JR, Fields PI, Tauxe RV, Logsdon JM, Jr. 2008. Do *Salmonella* carry spare tyres? *Trends Microbiol* 16:142–148. <https://doi.org/10.1016/j.tim.2008.01.009>.
- Bonifield HR, Hughes KT. 2003. Flagellar phase variation in *Salmonella enterica* is mediated by a posttranscriptional control mechanism. *J Bacteriol* 185:3567–3574. <https://doi.org/10.1128/JB.185.12.3567-3574.2003>.
- Aldridge P, Gnerer J, Karlinsey JE, Hughes KT. 2006. Transcriptional and translational control of the *Salmonella fliC* gene. *J Bacteriol* 188:4487–4496. <https://doi.org/10.1128/JB.00094-06>.
- Yamamoto S, Kutsukake K. 2006. FliA-mediated posttranscriptional control of phase 1 flagellin expression in flagellar phase variation of *Salmonella enterica* serovar Typhimurium. *J Bacteriol* 188:958–967. <https://doi.org/10.1128/JB.188.3.958-967.2006>.
- Switt AIM, Soyer Y, Warnick LD, Wiedmann M. 2009. Emergence, distribution, and molecular and phenotypic characteristics of *Salmonella enterica* serotype 4,5,12:i:–. *Foodborne Pathog Dis* 6:407–415. <https://doi.org/10.1089/fpd.2008.0213>.
- Zerbino DR, Birney E. 2008. Velvet: algorithms for de novo short read assembly using de Bruijn graphs. *Genome Res* 18:821–829. <https://doi.org/10.1101/gr.074492.107>.
- Overbeek R, Olson R, Pusch GD, Olsen GJ, Davis JJ, Disz T, Edwards RA, Gerdes S, Parrello B, Shukla M, Vonstein V, Wattam AR, Xia F, Stevens R. 2014. The SEED and the Rapid Annotation of microbial genomes using Subsystems Technology (RAST). *Nucleic Acids Res* 42:D206–D214. <https://doi.org/10.1093/nar/gkt1226>.
- Larsen MV, Cosentino S, Rasmussen S, Friis C, Hasman H, Marvig RL, Jelsbak L, Sicheritz-Pontén T, Ussery DW, Aarestrup FM, Lund O. 2012. Multilocus sequence typing of total-genome-sequenced bacteria. *J Clin Microbiol* 50:1355–1361. <https://doi.org/10.1128/JCM.06094-11>.
- Larsen MV, Cosentino S, Lukjancenko O, Saputra D, Rasmussen S, Hasman H, Sicheritz-Pontén T, Aarestrup FM, Ussery DW, Lund O. 2014. Benchmarking of methods for genomic taxonomy. *J Clin Microbiol* 52:1529–1539. <https://doi.org/10.1128/JCM.02981-13>.