

# Regulation of PI3K signaling in cancer metabolism and PI3K-targeting therapy

# Beinan Han<sup>1</sup>, Xiaorong Lin<sup>2</sup>, Hai Hu<sup>3</sup>

<sup>1</sup>Department of Oncology, Sun Yat-sen Memorial Hospital, Sun Yat-sen University, Guangzhou, China; <sup>2</sup>Diagnosis and Treatment Center of Breast Diseases, Shantou Central Hospital, Shantou, China; <sup>3</sup>The Cancer Hospital of the University of Chinese Academy of Sciences (Zhejiang Cancer Hospital), Institute of Basic Medicine and Cancer (IBMC), Chinese Academy of Sciences, Hangzhou, China

*Contributions:* (I) Conception and design: B Han, H Hu; (II) Administrative support: X Lin, H Hu; (III) Provision of study materials or patients: B Han, X Lin; (IV) Collection and assembly of data: B Han; (V) Data analysis and interpretation: B Han, X Lin; (VI) Manuscript writing: All authors; (VII) Final approval of manuscript: All authors.

*Correspondence to:* Hai Hu, PhD. The Cancer Hospital of the University of Chinese Academy of Sciences (Zhejiang Cancer Hospital), Institute of Basic Medicine and Cancer (IBMC), Chinese Academy of Sciences, Hangzhou 310022, China. Email: huhai@zjcc.org.cn; Xiaorong Lin, MD. Diagnosis and Treatment Center of Breast Diseases, Shantou Central Hospital, Shantou 515000, China. Email: clarelynn\_lin@163.com.

Abstract: The phosphatidylinositol-3-kinase (PI3K) signaling plays a key role in various cellular functions and is frequently activated in cancer, making it an attractive therapeutic target. The PI3K signaling pathway influencing glucose metabolism, lipid synthesis, nucleotide production, and protein synthesis, all of which contribute to cancer cell proliferation and survival. It enhances glucose uptake through the activation of glucose transporters and glycolysis, while also promoting lipid synthesis via downstream factors like mTORC1. This pathway boosts nucleotide synthesis by regulating transcription factors like MYC, activating key enzymes for purine and pyrimidine production. Additionally, due to its essential role in cancer cell growth, the PI3K pathway is a key target for anticancer therapies. However, treatment using PI3K inhibitors alone has limitations, including drug resistance and significant side effects such as hyperglycemia, fatigue, and liver dysfunction. Clinical trials have led to the development of isoform-specific PI3K inhibitors to reduce toxicity. Combining PI3K inhibitors with other treatments, such as hormone therapy or surgery, may improve efficacy and minimize side effects. Further research is needed to fully understand the mechanisms of PI3K inhibitors and improve individualized treatment approaches. In this review, we introduce the characteristic of three classes of PI3Ks, discuss the regulation of cancer metabolism including the control of glucose uptake, glycolysis, de novo lipid synthesis, nucleotide synthesis and protein synthesis, and review the current statuses of different PI3K inhibitors therapy.

Keywords: Phosphatidylinositol-3-kinase (PI3K); cancer; metabolism; target therapy; PI3K inhibitor

Received: 15 June 2024; Accepted: 11 October 2024; Published online: 29 October 2024. doi: 10.21037/tbcr-24-29 **View this article at:** https://dx.doi.org/10.21037/tbcr-24-29

## Introduction

The phosphatidylinositol-3-kinase (PI3K) signaling pathway is a crucial intracellular mechanism that regulates various cellular functions, including growth, metabolism, movement, survival, and angiogenesis. The activation of the PI3K/AKT/mTOR pathway is crucial in driving tumor progression, as well as other physiological and pathophysiological functions. After PI3K activation, PI3K phosphorylates phosphatidylinositol-4,5-bisphosphate (PIP2) to form phosphatidylinositol-3,4,5-trisphosphate (PIP3), leading to the translocation and activation of protein kinase B (AKT) on the cell membrane. AKT belongs to the serine/threonine-specific protein kinase family and exists in three main isoforms: Akt1, Akt2, and Akt3. AKT can phosphorylate various intracellular substrates such as

glycogen synthase kinase 3 beta (GSK-3 $\beta$ ), Bcl-2-associated death promoter (BAD), and forkhead box O (FOXO) transcription factors, thereby regulating multiple biological functions of cells including metabolism, proliferation, survival, and metastasis.

Under physiological conditions, the activation of the PI3K/AKT signaling pathway begins with the binding of growth factors, hormones, and other extracellular signals to receptor tyrosine kinases (RTKs), regulating organismal metabolism. This pathway plays a crucial role in maintaining normal cell growth, proliferation, and survival. However, dysregulation of the PI3K/AKT pathway is common in various cancers, leading to metabolic reprogramming of cells. This results in the overactivation of cellular processes that enhance cell survival, growth, and metabolism, promoting tumorigenesis to meet the synthetic demands of tumor cells.

The PI3K/AKT signaling pathway plays a critical role in regulating cellular metabolism by various mechanisms, including direct modulation of metabolism-associated proteins, indirect regulation of metabolism through transcription factors, acute metabolic changes mediated by phosphorylation of metabolic enzymes, and longterm metabolic effects through the regulation of gene expression programs. These regulatory mechanisms enable the PI3K/AKT pathway to exert key roles in cell growth, proliferation, and survival.

Therefore, targeting the PI3K/AKT signaling pathway has become an attractive strategy for cancer treatment. This is because the pathway plays crucial roles in cell growth, metabolism, and survival. In cancer cells, the PI3K/AKT pathway is often aberrantly activated, leading to abnormal proliferation and survival. By inhibiting this pathway, it is possible to reprogram the metabolic processes of cancer cells, inducing apoptosis (programmed cell death) or senescence (loss of cellular function), thereby preventing further proliferation and spread of cancer cells.

Numerous inhibitors have been developed to target the PI3K signaling pathway, with some approved for clinical therapy, leading to improvements in the survival rates of cancer patients. However, there are still numerous issues related to side effects and drug resistance that need to be addressed. Despite some successes in clinical use, PI3K inhibitors are still plagued by concerns regarding side effects and drug resistance. Side effects may include hyperglycemia, rash, fatigue, among others, while resistance issues may lead to decreased drug efficacy. Therefore, systematic exploration of the mechanisms of action of PI3K inhibitors and the formulation of individualized treatment decisions are crucial for further enhancing the clinical effectiveness of PI3K inhibitors.

#### **Overview of the PI3K**

PI3K is a group of enzymes associated with the plasma membrane, primarily involved in regulating essential biological processes such as cell growth, survival, proliferation, and metabolism. Its function involves the synthesis of PIP3 in signaling pathways, thereby activating a cascade of downstream signaling molecules, including AKT and mammalian target of rapamycin (mTOR), to influence cellular physiology and pathology (1).

PI3K can be divided into three main classes: I, II, and III, each composed of specific subunits, including a regulatory subunit and a catalytic subunit. These classes have distinct substrate specificities and functions, regulating cellular signaling and metabolic pathways differently (2).

The PI3K pathway plays a critical role in regulating tumor cell metabolism, particularly in glycolysis and fatty acid synthesis. It promotes glucose uptake and utilization, enhancing energy storage by influencing AKT and mTOR signaling. Additionally, the PI3K pathway stimulates cell growth and proliferation through the activation of AKT and mTOR. Studies have shown that this pathway is not only vital for the growth of normal cells but is also aberrantly activated in various cancers, driving tumor cell growth and survival, and closely associated with tumor invasion and metastasis. Moreover, the PI3K pathway integrates signals from multiple growth factors and cytokines, allowing cells to adapt to environmental changes and promote growth and survival. In cancer treatment, the abnormal activation of the PI3K pathway is often linked to treatment resistance. Understanding the modern mechanisms of this pathway can help develop new therapeutic strategies to overcome resistance. Furthermore, the PI3K pathway interacts with other signaling pathways in the tumor microenvironment, such as the MAPK and Wnt pathways, influencing tumor growth, metastasis, and response to treatment. Therefore, PI3K has become an important target in cancer therapy, and inhibitors targeting this pathway have been extensively researched and used in clinical trials, offering new hope for cancer patients' treatment (3).

# Class I PI3Ks

Class I PI3Ks are a crucial group of cellular signaling enzymes involved in regulating key biological processes

such as cell growth, survival, proliferation, and metabolism. Class I PI3Ks include PI3Ka, PI3Kβ, PI3Kγ, and PI3Kδ, with their catalytic subunits encoded by PIK3CA, PIK3CB, PIK3CG, or PIK3CD genes. They catalyze the conversion of PIP2 on the cell membrane to PIP3. Class I PI3Ks are further divided into two subclasses based on their structure, substrate specificity, and regulation: IA and IB. Class IA PI3Ks are activated by RTKs and consist of a regulatory subunit (p85) and a catalytic subunit (p110) (4). The p85 subunit has five variants (p85a, p55a, p50a, p85β, and p55 $\gamma$ ), while p110 has three isoforms (p110 $\alpha$ , p110 $\beta$ , and p1108). Class IA PI3Ks can bind to intracellular signals from proteins like Rho, SHP1, and protein kinase C, and activate downstream signaling molecules through the p110 subunit (5). Class IB PI3Ks include a catalytic subunit  $(p110\gamma)$  and a regulatory subunit (p101 or p84) and are typically activated by G protein-coupled receptor (GPCR) Gβγ subunits. In summary, Class I PI3Ks are critical regulators of cell growth, metabolism, and survival. Class IA and IB subtypes play distinct roles in cellular signaling pathways, and their dysregulation is associated with various diseases, making them important targets for therapeutic research.

#### Class II PI3Ks

Class II PI3Ks are a class of relatively complex enzymes characterized prominently by their C-terminal C2 domain. This domain lacks critical aspartic acid residues necessary for binding Ca2+, indicating that Class II PI3Ks can interact with lipids independently of Ca2+. Class II PI3Ks comprise three subtypes: PI3K-C2 $\alpha$ , PI3K-C2 $\beta$ , and PI3K-C2 $\gamma$ . PI3K-C2 $\alpha$  and PI3K-C2 $\beta$  are widely expressed, whereas PI3K-C2 $\alpha$  is expressed predominantly in the liver, breast, prostate, salivary glands, and exocrine pancreas. They can synthesize two distinct products, phosphatidylinositol-3-phosphate (PI(3)P) and phosphatidylinositol-3,4-bisphosphate (PI(3,4) P2), depending on substrate specificity (6).

Compared to Class I PI3Ks, Class II PI3Ks have been relatively overlooked in research, especially in the field of cancer. However, recent studies suggest their potential significant roles in cancer development and progression. Class II PI3Ks primarily regulate intracellular dynamics and membrane trafficking rather than functioning in signal transduction. Specifically, PI3K-C2 $\alpha$  regulates cancer cell death and mitosis by controlling spindle stability. In contrast, PI3K-C2 $\beta$  is involved in cancer cell migration and invasion, potentially influencing proliferation through the regulation of cyclin B1 expression. However, our understanding of the role of PI3K-C2 $\gamma$  in cancer is still limited (7).

Due to their regulation of crucial membrane-related processes such as endocytosis, lysosomal activity, and cell adhesion, Class II PI3Ks may play either promotive or inhibitory roles in tumor growth, invasion, and metastasis. Therefore, gaining a deeper understanding of the functions and regulatory mechanisms of Class II PI3Ks is essential for developing more effective cancer treatment strategies. Future research should further explore the roles of Class II PI3Ks in the tumor microenvironment and the development of targeted drugs against these enzymes, aiming to provide more personalized and efficient treatment options for cancer patients (8).

## Class III PI3Ks

Class III PI3Ks are heterodimeric enzymes consisting of a regulatory subunit (VPS15) and a catalytic subunit (VPS34), which have been found to be involved in the regulation of autophagy. VPS34 phosphorylates phosphatidylinositol (PI) to generate PI(3)P. VPS34, the smallest PI3K catalytic core, forms two tetrameric complexes—complex I and complex II, which play roles in autophagy and endocytosis, respectively. Complex I is involved in the recruitment of the endoplasmic reticulum (ER), crucial for the formation and elongation of autophagosomes, while complex II regulates endosome maturation and promotes fusion between autophagosomes and late endosomes/lysosomes.

Research on VPS34 in cancer has revealed its role in promoting cell survival and proliferation through inducing autophagy. For instance, studies have shown that VPS34 expression correlates with the oncogenic activity of human breast cancer cells. VPS34 enhances the binding of PKC- $\delta$ to p62, leading to its phosphorylation at serine 349, thereby positively reinforcing Nrf2-dependent transcription of oncogenes and promoting tumor growth. Additionally, inhibitors of VPS34 significantly enhance the sensitivity of breast cancer cells to tyrosine kinase inhibitors by suppressing autophagy. Therefore, VPS34 represents a promising therapeutic target for combination therapies in breast cancer (9).

## Key regulation of cancer metabolism

Metabolic reprogramming, including lipid dysfunctions, is a key characteristic of cancer cells. It leads to changes

#### Page 4 of 15

in metabolic enzymes and pathways that impact the development, progression, and metastasis of cancer. The activity of upstream oncogenes of PI3K is increased in cancer cells, resulting in altered nutrient transporters and metabolic enzymes. This alteration in metabolism meets the anabolic needs of abnormal cells and contributes to cancer progression.

# Control of glucose metabolism (glucose uptake and glycolysis)

One of the most common metabolic characteristics that distinguish tumor cells from normal cells is altered glucose metabolism. This metabolic feature is known as aerobic glycolysis, also referred to as the Warburg effect. Aerobic glycolysis refers to the tendency of tumor cells to convert glucose to lactate through glycolysis, even in the presence of sufficient oxygen, rather than producing energy through mitochondrial oxidative phosphorylation. This process is characterized by increased glucose uptake and lactate production; tumor cells exhibit a higher rate of glucose uptake and predominantly convert glucose to lactate, even in an oxygen-rich environment (10). Glycolysis not only generates adenosine triphosphate (ATP) but also provides metabolic intermediates necessary for biosynthetic processes, including the synthesis of proteins, lipids, and nucleotides, which are essential for cell growth and proliferation (11). Aerobic glycolysis also helps maintain redox balance within the cell by regenerating NAD+ through the primary conversion of pyruvate to lactate.

The PI3K/AKT pathway promotes glycolysis through various mechanisms, including increasing glucose uptake, activating key glycolytic enzymes, regulating the expression of glycolysis-related genes, and modulating the utilization of metabolic products (12). This pathway plays a critical role in cellular energy metabolism and growth, with a particularly significant impact in tumor cells, enabling them to grow rapidly even under adverse conditions (13).

The PI3K/AKT pathway plays a significant role in regulating anabolic metabolism through directly phosphorylating mechanisms. The pathway directly regulates glucose uptake and glycolysis through glucose transporters (GLUTs), with AKT promoting glucose uptake via both GLUT1 and GLUT4 (14-17). AKT promotes the translocation of the glucose transporter GLUT4 by phosphorylation, facilitating its movement from intracellular vesicles to the cell membrane, thereby increasing cellular glucose uptake (18). Additionally, upon AKT activation, FoxO1 and FoxO3a are phosphorylated and translocated from the nucleus to the cytoplasm, leading to the suppression of thioredoxin-interacting protein (TXNIP) expression. TXNIP is a protein that promotes the endocytosis of GLUT1 and GLUT4, thereby inhibiting glucose uptake (19,20). Furthermore, the AKT signaling pathway enhances the metabolic activation of glucose by directly phosphorylating and activating hexokinase 2 (HK2), a key enzyme in the glycolytic pathway. HK2 catalyzes the conversion of glucose to glucose-6-phosphate, which is the first step in glycolysis and a critical rate-controlling step. HK2 is highly expressed in many cancer cells and associates with the outer mitochondrial membrane, contributing to the high glycolytic activity of cancer cells. AKT phosphorylates HK2 directly, increasing its enzymatic activity and mitochondrial localization. This process facilitates HK2 to more efficiently perform its catalytic function within the cell, thereby enhancing glycolytic efficiency (21,22).

PI3K activation has also been found to promotes glycolytic flux in an AKT-independent manner, except for PI3K-initiated AKT activation. Aldolase is a key glycolytic enzyme responsible for catalyzing the breakdown of fructose-1,6-bisphosphate (F1,6BP) into glyceraldehyde-3-phosphate (G3P) and dihydroxyacetone phosphate (DHAP). This step is the fourth in the glycolytic pathway and is crucial because it splits the six-carbon molecule into two three-carbon molecules, allowing the metabolic process to continue. Stimulation of PI3K by growth factors and insulin prompts the release of the glycolytic enzyme aldolase A bound to filamentous actin, and a subsequent rise in aldolase activity, which leads to heightened glycolytic flux (23). Increased levels of aldolase A in cancer are linked to adverse patient prognosis. Inhibition of aldolase A has been demonstrated to reduce tumor growth in xenograft models (24). Therefore, inhibiting the PI3K pathway to reduce the activity of aldolase A and consequently suppress tumor growth may represent a promising therapeutic approach.

Furthermore, the PI3K/AKT pathway indirectly regulates glycolysis and other metabolic pathways through downstream transcription factors such as FOXO, HIF1 (hypoxia-inducible factor 1), MYC, and ATF4 (activating transcription factor 4) (25-27). These transcription factors oversee the regulation of gene expression related to glucose transporter production and glycolytic enzyme activity (28).

Those studies indicate that PI3K signaling is crucial in facilitating glucose uptake and glycolysis, in both normal and cancerous cells. In the context of oncogenic PI3K

signaling, the aforementioned regulatory mechanisms collectively contribute to the persistent activation of aerobic glycolysis. This metabolic trait, commonly observed in cancer cells, enables metabolic intermediates to be utilized by metabolic pathways branching off glycolysis, ultimately contributing to the biosynthesis of cellular macromolecules.

# Control of de novo lipid synthesis

The PI3K/AKT signaling pathway is essential for the regulation of various cellular processes, including cell survival, growth, and metabolism. One of the metabolic processes regulated by this pathway is de novo lipid synthesis. Lipid synthesis is necessary for membrane biogenesis, signal transduction, and energy storage, and plays a crucial role in regulating normal cellular function. Nevertheless, irregularities in lipid synthesis have been associated with the development of cancer (29). The production of fatty acids and sterols from cytosolic acetyl-CoA through de novo lipid synthesis can be initiated in two ways: either from citrate with the involvement of ATP citrate lyase (ACLY), or from acetate via acetyl-CoA synthetase. Extensive research has demonstrated that PI3K signaling can activate multiple enzymes involved in lipid synthesis and promote this process through various mechanisms, including transcriptional regulation and post-translational modifications (30,31).

By directly phosphorylating ACLY, the PI3K/AKT pathway can activate *de novo* lipid synthesis, leading to heightened production of cytosolic acetyl-CoA, which is then utilized in the synthesis of fatty acids and sterols (32,33). In addition, this pathway can also contribute to protein acetylation reactions. Studies have shown that the AKT-ACLY axis plays a pivotal role in tumor growth and histone acetylation (34). ACLY is frequently often observed to be upregulated in different human cancer types, and inhibiting this enzyme has demonstrated efficacy in decreasing cancer cell proliferation in both laboratory and animal models, indicating its promise as a therapeutic target for cancer treatment (35).

The activation of the PI3K/AKT pathway can also promote the synthesis of fatty acids and cholesterol by activating the sterol regulatory element-binding protein (SREBP) family of transcription factors, which regulate the expression of genes related to lipid biosynthesis. Studies have shown that tumors can increase glucose uptake through the PI3K/AKT/mTOR signaling pathway, thereby driving the N-glycosylation of SREBP cleavageactivating protein (SCAP), which in turn activates SREBP. The N-glycosylation of SCAP is crucial for its transport and activation. The precursor protein of SREBP is bound to the ER membrane in an inactive form. Its activation is inhibited by the ER-resident protein Insig, which prevents the translocation and nuclear activation of SREBP. For SREBP to become an active transcription factor, it needs to be transported to the Golgi apparatus for proteolytic cleavage. Once cleaved, it can then enter the nucleus to regulate the expression of target genes. Insig binds to SCAP, and glucose-mediated N-glycosylation promotes the dissociation of SCAP from Insig, forming the SCAP/ SREBP complex and facilitating its transport from the ER to the Golgi apparatus. In the Golgi, the precursor protein of SREBP is proteolytically cleaved, and the active form of SREBP translocates to the nucleus. Once in the nucleus, it binds to the sterol regulatory elements (SRE) of target gene promoters, upregulating the transcription of genes involved in lipid biosynthesis. These genes encode enzymes such as acetyl-CoA carboxylase and fatty acid synthase for fatty acid synthesis, as well as HMG-CoA reductase for cholesterol synthesis (36,37). SREBP family induce the expression of a wide range of enzymes involved in the synthesis of sterols and fatty acids, including SLC25A1, ACLY, ACSS2, ACC, FASN and SCD1 (38). In addition to promoting the processing and nuclear translocation of SREBP, the PI3K/AKT/mTORC1 signaling pathway can also activate SREBP through other pathways, resulting in the upregulation of lipogenic gene expression and increased de novo lipid synthesis. For example, SREBP can be phosphorylated by synthase kinase-3 (GSK3) at either Ser-73 or Arg-96, facilitating its dissociation from the SREBP-1c-SCAP complex and subsequent GSK-3-dependent proteasomal degradation via the ubiquitin ligase pathway. And AKT can inhibit glycogen GSK3, thereby enhancing the stability of active SREBP and preventing its degradation via ubiquitination (39-42). Hence, AKT signaling can potentially boost SREBP processing by activating mTORC1 and maintain the stability of activated SREBP by inhibiting GSK3 activity. Moreover, recent studies have unveiled a collaborative role of MYC with SREBP in fostering lipogenesis and promoting cancer progression (43,44).

In addition, mTORC1 affects lipid synthesis through the phosphorylation and activation of S6 kinase 1 (S6K1). S6K1 is one of the downstream effector proteins of mTORC1, playing a critical role in protein synthesis, cell growth, and metabolic regulation. S6K1 enhances ribosomal translation activity by phosphorylating and regulating multiple translation initiation and post-translational

#### Page 6 of 15

modification factors, leading to increased synthesis and activity of enzymes involved in lipid synthesis. Specifically, activated S6K1 can directly phosphorylate SR protein kinase 2 (SRPK2), enabling its regulatory role in RNA splicing within the nucleus, impacting the transcription and translation levels of genes related to lipid synthesis, thus influencing lipid biosynthesis and metabolism (45). Moreover, numerous *de novo* lipid synthesis enzymes have been found to be upregulated in diverse cancer types, encompassing SREBPs, SRPK2, and the lipogenic enzymes activated by SREBP family. These constituents continue to be regarded as promising targets for cancer therapy (46,47).

# Control of nucleotide synthesis

Nucleotides play a crucial role in the synthesis of nucleic acids and the facilitation of diverse cellular processes, comprising purines and pyrimidines. In cancer cells, there's a significant increase in the generation of new nucleotides, vital for driving their rapid growth and proliferation—a departure from the behavior of healthy cells (48).

Various metabolic pathways play crucial roles in nucleotide synthesis, including the pentose phosphate pathway (PPP), which provides ribose-5-phosphate necessary for nucleotide synthesis. Serine and glycine are essential precursors for pyrimidine and purine synthesis, participating in one-carbon unit transfer. One-carbon metabolism, which includes the folate cycle and methionine cycle, supplies one-carbon units required for the synthesis of purines and pyrimidines. Glutamine is converted into glutamate and ammonia in cells and further participates in the tricarboxylic acid (TCA) cycle, providing the nitrogen and carbon skeletons needed for nucleotide synthesis. The TCA cycle provides precursor molecules for nucleotide synthesis, such as oxaloacetate, which can be converted to aspartate and contribute to pyrimidine and purine synthesis (49,50). Therefore, it's evident that AKT signaling regulates nucleotide synthesis through multiple mechanisms, thereby impacting metabolism.

The activation of the PI3K/AKT/mTORC1 signaling pathway can enhance the metabolic flux of glucose carbon into the PPP, facilitating the production of ribose essential for nucleotide synthesis (51). The PI3K/ AKT/mTORC1 pathway enhances the metabolic flux of the oxidative PPP by upregulating the expression of its key enzyme, glucose-6-phosphate dehydrogenase (G6PD). G6PD catalyzes the conversion of glucose-6phosphate to 6-phosphogluconolactone while reducing NADP+ to NADPH. This reaction is the first step in the oxidative PPP and serves as the rate-controlling step for this pathway. Glucose-6-phosphate gains entry into the oxidative arm of the PPP through the activity of glucose-6-phosphate dehydrogenase, resulting in the generation of ribose-5-phosphate and bypassing glycolysis (52,53). Additionally, research has shown that AKT can directly activate transketolase (TKT), a key enzyme in the nonoxidative phase of the PPP (54). Evidence shows that the PI3K/AKT signal crosstalks with PPP branching metabolic pathways to promote PPP metabolism by stabilizing G6PD protein and inhibiting G6PD E3 ligase TIRM21, while PPP metabolites reinforce AKT activation and promote cancer metabolic reprogramming (51). In breast cancer cells, metabolic enzymes in both oxidative and nonoxidative PPP are upregulated, therefore, inhibiting them can reduces tumor proliferation; After treatment with PI3K inhibitors, the metabolic flux through the non-oxidative PPP in breast cancer cells is significantly reduced (55,56), indicating a greater dependency of tumors on this metabolic pathway (57). Hence, investigating PPP metabolic enzymes or targeting PI3K holds considerable significance and can offer more tailored and individualized approaches to cancer treatment.

AKT can regulate nucleotide synthesis by modulating the transcription of MYC. Additionally, MYC directly regulates the expression of enzymes involved in the pyrimidine and purine synthesis pathways. For example, phosphoribosyl pyrophosphate (PRPP) is essential for nucleotide synthesis, and MYC controls the synthesis of PRPP by regulating phosphoribosyl pyrophosphate synthase 2 (PRPS2) at both the transcriptional and translational levels (58-60). Moreover, glutamine is a crucial primary nitrogen donor for synthesizing pyrimidine and purine bases. MYC can promote both the synthesis and uptake pathways of glutamine. MYC regulates the transcription of several key enzymes involved in glutamine synthesis, such as glutamatecysteine ligase (GCLC), glutamate kinase (GK), and glutamate dehydrogenase (GDH). MYC also upregulates the expression of transporters SLC1A5 and LAT1, thereby increasing the rate of cellular glutamine uptake (61,62).

The PI3K/AKT/mTORC1 signaling pathway influences nucleotide synthesis through various mechanisms, including the regulation of both pyrimidine and purine synthesis. In pyrimidine synthesis, growth factor signaling activates the PI3K/AKT pathway, subsequently activating mTORC1. mTORC1 then phosphorylates and activates the primary rate-limiting enzyme in the pyrimidine synthesis pathway, thereby promoting pyrimidine synthesis (63). In addition to pyrimidine synthesis, mTORC1 triggers purine resynthesis through transcriptional mechanisms. Moreover, mTORC1 activates ATF4, increasing the expression of enzymes involved in serine synthesis and its conversion to formate, providing one-carbon units for purine ring formation. Through these mechanisms, the PI3K/AKT/ mTORC1 signaling pathway coordinates nucleotide cellular metabolism to ensure an adequate supply of pyrimidines and purines, supporting nucleotide synthesis and the rapid proliferation and growth of cells (64).

Therefore, the PI3K/AKT/mTORC1 signaling network is pivotal in governing *de novo* nucleotide synthesis by its capacity to regulate both post-translational and transcriptional mechanisms. This is accomplished through the activation of key enzymes involved in both pyrimidine and purine synthesis pathways, including CAD (cytosolic carbamoyl-phosphate synthetase II, aspartate transcarbamylase and dihydroorotase), and through the induction of specific metabolic enzymes and the injection of metabolites into purine synthesis pathways. These discoveries underscore the significance of mTORC1 as a central figure in cellular metabolism and propose that directing interventions toward this pathway could present a promising therapeutic approach for addressing specific diseases, such as cancer, that are characterized by dysregulated nucleotide synthesis.

## Control of protein synthesis

Protein synthesis is a complex and resource-intensive process, and the PI3K/AKT/mTORC1 signaling pathway plays a crucial regulatory role in this process. mTORC1 regulates protein synthesis by phosphorylating multiple downstream substrates. Two key downstream substrates are S6K1 and eIF4E binding protein (4E-BP) (65). mTORC1 phosphorylates and activates S6K1, which further promotes protein synthmTORC1 activates S6K1 by directly phosphorylating it at multiple sites (such as T389). Activated S6K1 further phosphorylates ribosomal protein S6 and the translation initiation factor eIF4B, thereby enhancing the initiation of translation and the ribosome biogenesis function, promoting protein synthesis. For 4E-BP1, in its unphosphorylated state, it binds to eIF4E and inhibits the initiation of translation. Once phosphorylated, 4E-BP1 releases eIF4E, allowing it to bind with eIF4G, forming the active translation initiation complex eIF4F (66). In addition to 4E-BP, the phosphorylation of La-related protein 1 (LARP1) downstream of mTORC1 is also involved in

selectively inducing the translation of 5'-TOP mRNAs (67). Both AKT and S6K1 can phosphorylate LARP1, facilitating its dissociation from the 5' UTRs and thus relieving the inhibition on translation, promoting 5' cap-dependent translation initiation (68). Additionally, S6K1 activates eIF4B, a positive regulator of translation initiation, and facilitates the degradation of the eIF4A inhibitor PDCD4, further promoting the initiation of translation.

Additionally, the PI3K/AKT/mTORC1 signaling pathway regulates protein synthesis by enhancing rRNA transcription. mTORC1, through its downstream effectors like S6K1, phosphorylates and activates nucleolar transcription factors such as UBF and transcription initiation factor 1A (TIF-1A). TIF-1A and UBF work together to promote the binding and activation of RNA polymerase I at the rDNA promoter (69). This upregulates the transcription of rRNA, promoting ribosome biogenesis. Through these mechanisms, the cell's capacity for protein synthesis is significantly enhanced. This pathway plays a crucial role in regulating proliferation and cell growth.

#### **Targeting PI3K signal therapy in cancer**

# Pan-PI3K inhibitors

Pan-PI3K inhibitors are primarily designed to target the p110 subunits of class IA PI3Ks, as these subunits are closely associated with the development and progression of tumors. Several small-molecule pan-PI3K inhibitors, such as buparlisib (BKM120), pictilisib (GDC-0941) and pilaralisib (XL147), have been developed as potential cancer therapies.

The thienopyrimidine derivative GDC-0941 was the first pan-class I PI3K inhibitor to undergo clinical trials. It exhibits potent inhibitory activity against p110 $\alpha$  and p110 $\delta$  enzymes, as well as inhibiting p110 $\beta$  and p110 $\gamma$  at nanomolar concentrations in kinase assays. Research has shown that GDC-0941 exhibits significant anti-tumor activity in patients, mouse xenograft models, and *in vitro* models, whether used alone or in combination with other therapeutic methods (70-72).

BKM120 (buparlisib) is a potent inhibitor of class I PI3Ks, which targets the p110- $\alpha/\beta/\delta/\gamma$  isoforms with IC50 values of 5, 27, 7, and 14 nM, respectively, in cell-free assays. It has exhibited significant anti-cancer effects across various solid cancer models (73). At tolerated doses, buparlisib displays substantial antitumor activity in human tumor xenograft models and possesses favorable oral

bioavailability *in vivo* (74). In a phase II study (EPOC1303) involving patients with advanced esophageal squamous cell carcinoma (ESCC), including those previously treated for the disease, BKM120 demonstrated promising efficacy (75). Additionally, a derivative of BKM120, known as PQR309, has been developed. This compound differentiates the microtubule inhibitory activity of BKM120 from its PI3K inhibitory activity, providing enhanced safety and flexibility for combination therapies (76). The phase I clinical trial of PQR309 (NCT01940133) has shown promising efficacy and PI3K inhibition, and a phase II clinical trial is currently in progress (77).

XL147 is a class I PI3K inhibitor that exhibits high specificity towards class I PI3Ks ( $\alpha$ ,  $\beta$ ,  $\gamma$ , and  $\delta$ ). In vitro assays have demonstrated the effective inhibition of PIP3 formation in cell membranes by XL147, along with the phosphorylation of downstream kinases such as AKT, p70S6K (the 70 kDa ribosomal protein S6 kinase), and ribosomal protein S6 in various tumor cell lines. This inhibition leads to a broad spectrum of potencies in impeding cell proliferation. Following oral administration, XL147 has shown to sustain inhibition of Akt, p70S6K, and S6 phosphorylation in multiple human xenograft models for at least 24 hours. Subsequent administration of XL147 has resulted in significant suppression of tumor growth with manageable toxicity profiles (78). In a phase I study involving patients with advanced solid tumors, the maximum tolerated dose of pilaralisib was determined to be 600 mg once daily. The administration of pilaralisib tablets was associated with favorable safety outcomes and exhibited preliminary antitumor efficacy (79). Furthermore, pilaralisib was evaluated in a phase II study involving patients with advanced or recurrent endometrial carcinoma, demonstrating a favorable safety profile and limited antitumor activity in this patient cohort (80). Clinical trials have indicated that combining XL147 with other therapeutic modalities enhances its antitumor efficacy compared to its monotherapy use (81).

# PI3K isoform-specific inhibitors

PI3K inhibitors with specificity for certain isoforms have been shown to have fewer toxic side effects and offtarget toxicity than pan-inhibitors. Consequently, isotypespecific inhibitors are considered more efficacious and can be administered at higher doses. Alpelisib (BYL-719), inavolisib (GDC-0077), idelalisib (GS-1101/CAL-101), GDC-0326 and taselisib (GDC-0032) are exemplars of isoform-specific inhibitors.

The first oral specific inhibitor targeting class I p110 $\alpha$ PI3K is BYL-719 (alpelisib) (82). Clinical trial findings from a phase I study of alpelisib showcased its efficacy as a monotherapy in managing patients with solid tumors harboring PI3KCA mutations (83). In a subsequent Phase 1b clinical trial, the combination of BYL-719 and fulvestrant demonstrated a manageable safety profile among patients with estrogen receptor-positive advanced breast cancer, irrespective of PIK3CA alterations. Initial data indicate a potentially enhanced clinical activity of this combination in tumors with PIK3CA alterations compared to those lacking such mutations (84). Furthermore, in the SOLAR-1 trial, the efficacy of alpelisib combined with fulvestrant was assessed in patients both with and without PIK3CA mutations. The findings revealed a significant improvement in progression-free survival (PFS) with the addition of alpelisib to fulvestrant in patients with PIK3CAmutated, hormone receptor-positive (HR+), human epidermal growth factor receptor 2-negative (HER2-) advanced breast cancer (85,86).

A novel PI3Ka inhibitor, inavolisib, has been discovered in recent years. PIK3CA mutations occur in approximately 40% of HR+ breast cancer cases globally, making targeted therapies for these mutations clinically significant. Inavolisib has emerged as a promising treatment for PIK3CA-mutated HR+/HER2- advanced breast cancer. It selectively induces degradation of the mutant  $p110\alpha$  protein, the catalytic subunit of PI3Ka, with a selectivity for PI3Ka that is 300 times greater than for other PI3K subunits (87). Inavolisib has advanced to phase III clinical trials (NCT04191499), showing significant efficacy in combination therapies for endocrine-resistant populations with PIK3CA mutations, with good safety and tolerability profiles. It offers a new and more effective treatment option for patients with PIK3CAmutated breast cancer and is poised to become a first-line therapy for this patient population (88).

Idelalisib was granted Food and Drug Administration (FDA) approval as the initial PI3K inhibitor for managing relapsed/refractory chronic lymphocytic leukemia (R/R CLL) patients (89). A randomized, double-blind, placebocontrolled phase 3 trial was conducted to assess the efficacy of combining idelalisib with bendamustine and rituximab in relapsed or refractory chronic lymphocytic leukemia patients. The findings indicated that this combination therapy led to enhanced progression-free survival compared to bendamustine plus rituximab monotherapy. Nonetheless, the idelalisib-treated group exhibited a heightened risk of

infections (90). Another phase III clinical trial compared the outcomes of idelalisib plus rituximab versus placebo plus rituximab in relapsed CLL patients, revealing that the idelalisib and rituximab combination therapy significantly improved both progression-free survival and overall survival compared to rituximab monotherapy (91).

In addition, GDC-0326, another p110 $\alpha$ -selective inhibitor, has been identified (92,93). It has demonstrated promising efficacy in a mouse model of pancreatic neuroendocrine tumors (PanNETs), showing advantages in reducing angiogenesis and decreasing lymph node metastasis (94). Taselisib is also a selective PI3K inhibitor designed to target p110 $\alpha$  rather than p110 $\beta$  (95). However, it also exhibits inhibitory effects on the p110 $\delta$  and p110 $\gamma$  isoforms, and this off-target inhibition may be a key factor contributing to its suboptimal performance in clinical trials (96).

# Dual PI3K and mTOR inhibitors

Inhibitors targeting both the PI3K and mTOR signaling pathways, known as PI3K/mTOR inhibitors, have been developed. Despite lacking FDA approval for cancer treatment currently, they are anticipated to yield improved therapeutic outcomes by concurrently suppressing both signaling pathways. This is because PI3K and mTOR share a structurally similar p110 subunit. The current pan-PI3K/ mTOR inhibitors available for clinical use include apitolisib, gedatolisib, SF1126, voxtalisib, omipalisib, samotolisib, bimiralisib, paxalisib, and GSK1059615 (97-104). Although PI3K/mTOR inhibitors exhibit lower specificity compared to isotype-specific inhibitors, they have shown promise in the treatment of various types of tumors.

Apitolisib, also referred to as GDC-0980, serves as a potent inhibitor targeting class I PI3K, displaying notable potency against the p110- $\alpha/\beta/\delta/\gamma$  isoforms with IC50 values of 5, 27, 7, and 14 nM, respectively, as observed in cell-free assays. Beyond its primary impact on PI3K, apitolisib also engages with the mTOR signaling pathway. Preclinical studies have revealed apitolisib's capacity to induce cytotoxicity and apoptosis in glioma cell lines in a dose- and time-dependent manner (105). In a randomized open-label phase II trial, the safety and efficacy of apitolisib were compared with everolimus in patients diagnosed with metastatic renal cell carcinoma. Results indicated that apitolisib exhibited inferior efficacy compared to everolimus, likely attributed to multiple on-target adverse events stemming from the complete blockade of PI3K/ mTOR signaling (106).

Several dual PI3K/mTOR inhibitors have been studied in clinical trials to evaluate their efficacy and pharmacokinetic properties. Among them, gedatolisib (PKI-587) is an effective dual inhibitor targeting PI3K $\alpha/\gamma$  and mTOR, demonstrating high efficacy in cell-free assays with IC50 values of 0.4 nM, 5.4 nM, and 1.6 nM for PI3K $\alpha/\gamma$  and mTOR, respectively (107). SF1126, developed by SignalRx, is a small molecule conjugate selectively inhibiting all PI3K class IA isoforms as well as other members of the PI3K superfamily, including DNA-PK and mTOR. Voxtalisib (XL765), developed by Sanofi, is also an effective inhibitor targeting the PI3K p110y, DNA-PK, and mTOR pathways. Omipalisib (GSK458) is a selective, pan-PI3K ATP-competitive inhibitor that also targets the mTOR pathway, with a Ki for the catalytic p110α subunit in the subnanomolar range. These dual PI3K/mTOR inhibitors have demonstrated promising therapeutic effects in research studies.

# Adverse reactions and management of toxicity of PI3K inhibitors

At present, various PI3K inhibitors have been developed, and they have shown good efficacy in experimental models or clinical trials. However, the toxicity caused by the ontarget and off-target effects of PI3K inhibitors presents a major challenge, impacting the effectiveness of treatment. For example, in pan-PI3K inhibitors, common adverse effects include hyperglycemia with BKM120, rash with XL147, and neutropenia with GDC-0941. Additionally, pan-PI3K inhibitors are associated with neuropsychiatric effects (confusion, depression, anxiety), hepatotoxicity, and diarrhea. Among PI3K p110a isoform inhibitors, BYL719's common adverse effect is hyperglycemia, while GDC-0032 commonly causes rash, along with diarrhea and pneumonitis. For dual PI3K and mTOR inhibitors, voxtalisib may cause stomatitis, SF1126 frequently leads to hyperglycemia, and GSK1059615 often causes immunosuppression (108,109).

Managing the toxicity of PI3K inhibitors requires a comprehensive approach that includes considering the drug dosage, adjusting treatment regimens, and symptomatic management. First, it is advisable to choose more selective PI3K inhibitors. For instance, early pan-PI3K inhibitors, due to their broad targeting and higher toxicity, did not gain wide clinical use. In contrast, alpelisib, with its  $\alpha$ -isoform specificity, has demonstrated significant antitumor activity with manageable toxicity in HR+, HER2– advanced breast

## Page 10 of 15

cancer patients, making it a preferred option. Additionally, close monitoring of patient safety data and symptomatic management of adverse effects is essential. Appropriate symptomatic treatments, such as using insulin to control hyperglycemia, topical corticosteroids for skin issues, and antidiarrheal agents for diarrhea, can help reduce adverse events and prolong treatment duration. Furthermore, besides PIK3CA mutations, other potential biomarkers that may affect the efficacy of PI3K inhibitors in advanced breast cancer warrant further clinical investigation. Identifying these markers could support the development of combination therapies to simultaneously target multiple pathways, helping to prevent the development of resistance (110).

# Conclusions

The PI3K signaling pathway plays a critical role in tumor metabolism, involving regulation in various aspects including glucose metabolism, lipid synthesis, nucleotide metabolism, and protein synthesis. This pathway enhances the uptake and utilization of glucose, augmenting the dependency of tumor cells on glucose. This process primarily occurs through increased expression of glucose transporters, activation of glycolysis pathways, and promoting glucose conversion to pyruvate. Additionally, the PI3K pathway promotes lipid synthesis pathways by activating downstream factors like mTORC1, leading to increased lipid synthesis and accumulation, thereby facilitating the growth and proliferation of tumor cells. Moreover, PI3K signaling enhances nucleotide synthesis by regulating the activity of transcription factors such as MYC, which involves pathways for both pyrimidine and purine synthesis. This process entails the activation of several key enzymes like PRPS2 and thymidylate synthase. Furthermore, the PI3K pathway boosts protein synthesis by activating downstream factors like mTORC1, facilitating translation initiation, and increasing translation rates. This supports the rapid proliferation of tumor cells by meeting the demand for protein synthesis. Overall, the PI3K signaling pathway orchestrates a metabolic reprogramming in cancer cells, favoring anabolic processes such as glycolysis, lipogenesis, nucleotide biosynthesis, and protein synthesis to sustain their rapid proliferation and survival.

Targeting the PI3K signaling pathway is highly appealing for anticancer therapy because of its essential role in cell growth, proliferation, and survival. Clinical trials with dual PI3K/mTOR and pan-PI3K inhibitors have reported high rates of side effects, prompting the creation of isoformspecific PI3K inhibitors with better specificity and lower toxicity.

However, the efficacy of using PI3K inhibitors alone for treatment is limited, which is associated with drug resistance, a narrow therapeutic window, and medication side effects. Therefore, combining them with other treatment modalities, such as hormone therapy, surgery and other anticancer drugs, may enhance effectiveness and reduce side effects. Common adverse reactions include hyperglycemia, vomiting, fatigue, rash, nausea, loss of appetite, diarrhea and abnormal liver function. It is also important to closely monitor medication side effects and provide corresponding symptomatic treatment when using PI3K inhibitors.

To date, the precise mechanism of action of PI3K inhibitors remains incompletely established, necessitating further extensive and systematic exploration. This endeavor will facilitate the monitoring of therapeutic efficacy during PI3K inhibition therapy, optimize the management of adverse effects, and enable the development of more individualized treatment regimens.

# **Acknowledgments**

*Funding*: This work was supported by the Natural Science Foundation of China (82002823), and the Program from Guangdong Basic and Applied Basic Research Foundation (2022A1515010168).

# Footnote

*Peer Review File:* Available at https://tbcr.amegroups.org/ article/view/10.21037/tbcr-24-29/prf

*Conflicts of Interest:* All authors have completed the ICMJE uniform disclosure form (available at https://tbcr. amegroups.org/article/view/10.21037/tbcr-24-29/coif). B.H., X.L., H.H. were supported by the Natural Science Foundation of China (82002823), and the Program from Guangdong Basic and Applied Basic Research Foundation (2022A1515010168). The authors have no other conflicts of interest to declare.

*Ethical Statement:* The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are

appropriately investigated and resolved.

*Open Access Statement:* This is an Open Access article distributed in accordance with the Creative Commons Attribution-NonCommercial-NoDerivs 4.0 International License (CC BY-NC-ND 4.0), which permits the non-commercial replication and distribution of the article with the strict proviso that no changes or edits are made and the original work is properly cited (including links to both the formal publication through the relevant DOI and the license). See: https://creativecommons.org/licenses/by-nc-nd/4.0/.

# References

- Katso R, Okkenhaug K, Ahmadi K, et al. Cellular function of phosphoinositide 3-kinases: implications for development, homeostasis, and cancer. Annu Rev Cell Dev Biol 2001;17:615-75.
- Bilanges B, Posor Y, Vanhaesebroeck B. PI3K isoforms in cell signalling and vesicle trafficking. Nat Rev Mol Cell Biol 2019;20:515-34.
- Thibault B, Ramos-Delgado F, Guillermet-Guibert J. Targeting Class I-II-III PI3Ks in Cancer Therapy: Recent Advances in Tumor Biology and Preclinical Research. Cancers (Basel) 2023;15:784.
- 4. Burke JE, Williams RL. Synergy in activating class I PI3Ks. Trends Biochem Sci 2015;40:88-100.
- Aytenfisu TY, Campbell HM, Chakrabarti M, et al. Class I PI3K Biology. Curr Top Microbiol Immunol 2022;436:3-49.
- Heng EYZ, Maffucci T. An Overview of Class II Phosphoinositide 3-Kinases. Curr Top Microbiol Immunol 2022;436:51-68.
- Gulluni F, De Santis MC, Margaria JP, et al. Class II PI3K Functions in Cell Biology and Disease. Trends Cell Biol 2019;29:339-59.
- Margaria JP, Ratto E, Gozzelino L, et al. Class II PI3Ks at the Intersection between Signal Transduction and Membrane Trafficking. Biomolecules 2019;9:104.
- Devereaux K, Dall'Armi C, Alcazar-Roman A, et al. Regulation of mammalian autophagy by class II and III PI 3-kinases through PI3P synthesis. PLoS One 2013;8:e76405.
- Warburg O, Wind F, Negelein E. THE METABOLISM OF TUMORS IN THE BODY. J Gen Physiol 1927;8:519-30.
- 11. Lunt SY, Vander Heiden MG. Aerobic glycolysis: meeting the metabolic requirements of cell proliferation. Annu Rev

Cell Dev Biol 2011;27:441-64.

- Xie Y, Shi X, Sheng K, et al. PI3K/Akt signaling transduction pathway, erythropoiesis and glycolysis in hypoxia (Review). Mol Med Rep 2019;19:783-91.
- Buzzai M, Bauer DE, Jones RG, et al. The glucose dependence of Akt-transformed cells can be reversed by pharmacologic activation of fatty acid beta-oxidation. Oncogene 2005;24:4165-73.
- Mueckler M, Thorens B. The SLC2 (GLUT) family of membrane transporters. Mol Aspects Med 2013;34:121-38.
- 15. Arponen M, Jalava N, Widjaja N, et al. Glucose transporters GLUT1, GLUT3, and GLUT4 have different effects on osteoblast proliferation and metabolism. Front Physiol 2022;13:1035516.
- Jiang T, Zhou ML, Fan J. Inhibition of GLUT-1 expression and the PI3K/Akt pathway to enhance the chemosensitivity of laryngeal carcinoma cells in vitro. Onco Targets Ther 2018;11:7865-72.
- 17. Sharma M, Dey CS. AKT ISOFORMS-AS160-GLUT4: The defining axis of insulin resistance. Rev Endocr Metab Disord 2021;22:973-86.
- Nozaki S, Takeda T, Kitaura T, et al. Akt2 regulates Rac1 activity in the insulin-dependent signaling pathway leading to GLUT4 translocation to the plasma membrane in skeletal muscle cells. Cell Signal 2013;25:1361-71.
- Albers PH, Pedersen AJ, Birk JB, et al. Human muscle fiber type-specific insulin signaling: impact of obesity and type 2 diabetes. Diabetes 2015;64:485-97.
- Sullivan WJ, Mullen PJ, Schmid EW, et al. Extracellular Matrix Remodeling Regulates Glucose Metabolism through TXNIP Destabilization. Cell 2018;175:117-132.e21.
- 21. Wei Q, Ren Y, Zheng X, et al. Ginsenoside Rg3 and sorafenib combination therapy relieves the hepatocellular carcinomaprogression through regulating the HK2mediated glycolysis and PI3K/Akt signaling pathway. Bioengineered 2022;13:13919-28.
- 22. Zhang T, Zhu X, Wu H, et al. Targeting the ROS/PI3K/ AKT/HIF-1α/HK2 axis of breast cancer cells: Combined administration of Polydatin and 2-Deoxy-d-glucose. J Cell Mol Med 2019;23:3711-23.
- Hu H, Juvekar A, Lyssiotis CA, et al. Phosphoinositide 3-Kinase Regulates Glycolysis through Mobilization of Aldolase from the Actin Cytoskeleton. Cell 2016;164:433-46.
- 24. Niu Y, Lin Z, Wan A, et al. Loss-of-Function Genetic Screening Identifies Aldolase A as an Essential Driver for Liver Cancer Cell Growth Under Hypoxia. Hepatology

# Page 12 of 15

2021;74:1461-79.

- Lee S, Dong HH. FoxO integration of insulin signaling with glucose and lipid metabolism. J Endocrinol 2017;233:R67-79.
- Semenza GL. Hypoxia-inducible factors: coupling glucose metabolism and redox regulation with induction of the breast cancer stem cell phenotype. EMBO J 2017;36:252-9.
- 27. Abdel-Wahab AF, Mahmoud W, Al-Harizy RM. Targeting glucose metabolism to suppress cancer progression: prospective of anti-glycolytic cancer therapy. Pharmacol Res 2019;150:104511.
- Tameire F, Verginadis II, Leli NM, et al. ATF4 couples MYC-dependent translational activity to bioenergetic demands during tumour progression. Nat Cell Biol 2019;21:889-99.
- 29. Beloribi-Djefaflia S, Vasseur S, Guillaumond F. Lipid metabolic reprogramming in cancer cells. Oncogenesis 2016;5:e189.
- Ricoult SJ, Yecies JL, Ben-Sahra I, et al. Oncogenic PI3K and K-Ras stimulate de novo lipid synthesis through mTORC1 and SREBP. Oncogene 2016;35:1250-60.
- Liu G, Wang N, Zhang C, et al. Fructose-1,6-Bisphosphate Aldolase B Depletion Promotes Hepatocellular Carcinogenesis Through Activating Insulin Receptor Signaling and Lipogenesis. Hepatology 2021;74:3037-55.
- 32. Icard P, Wu Z, Fournel L, et al. ATP citrate lyase: A central metabolic enzyme in cancer. Cancer Lett 2020;471:125-34.
- 33. Wei X, Shi J, Lin Q, et al. Corrigendum: Targeting ACLY Attenuates Tumor Growth and Acquired Cisplatin Resistance in Ovarian Cancer by Inhibiting the PI3K-AKT Pathway and Activating the AMPK-ROS Pathway. Front Oncol 2021;11:742374.
- Lee JV, Carrer A, Shah S, et al. Akt-dependent metabolic reprogramming regulates tumor cell histone acetylation. Cell Metab 2014;20:306-19.
- Gu L, Zhu Y, Lin X, et al. The IKKβ-USP30-ACLY Axis Controls Lipogenesis and Tumorigenesis. Hepatology 2021;73:160-74.
- Su L, Zhou L, Chen FJ, et al. Cideb controls sterolregulated ER export of SREBP/SCAP by promoting cargo loading at ER exit sites. EMBO J 2019;38:e100156.
- Shimano H, Sato R. SREBP-regulated lipid metabolism: convergent physiology - divergent pathophysiology. Nat Rev Endocrinol 2017;13:710-30.
- 38. Yi J, Zhu J, Wu J, et al. Oncogenic activation of PI3K-

AKT-mTOR signaling suppresses ferroptosis via SREBP-mediated lipogenesis. Proc Natl Acad Sci U S A 2020;117:31189-97.

- Raghow R, Dong Q, Elam MB. Phosphorylation dependent proteostasis of sterol regulatory element binding proteins. Biochim Biophys Acta Mol Cell Biol Lipids 2019;1864:1145-56.
- 40. Lewis CA, Griffiths B, Santos CR, et al. Regulation of the SREBP transcription factors by mTORC1. Biochem Soc Trans 2011;39:495-9.
- Dong Q, Giorgianni F, Beranova-Giorgianni S, et al. Glycogen synthase kinase-3-mediated phosphorylation of serine 73 targets sterol response element binding protein-1c (SREBP-1c) for proteasomal degradation. Biosci Rep 2015;36:e00284.
- 42. Frame S, Cohen P, Biondi RM. A common phosphate binding site explains the unique substrate specificity of GSK3 and its inactivation by phosphorylation. Mol Cell 2001;7:1321-7.
- Guo D, Bell EH, Mischel P, et al. Targeting SREBP-1driven lipid metabolism to treat cancer. Curr Pharm Des 2014;20:2619-26.
- Wu Y, Chen K, Liu X, et al. Srebp-1 Interacts with c-Myc to Enhance Somatic Cell Reprogramming. Stem Cells 2016;34:83-92.
- Lee G, Zheng Y, Cho S, et al. Post-transcriptional Regulation of De Novo Lipogenesis by mTORC1-S6K1-SRPK2 Signaling. Cell 2017;171:1545-1558.e18.
- 46. Cheng C, Geng F, Li Z, et al. Ammonia stimulates SCAP/Insig dissociation and SREBP-1 activation to promote lipogenesis and tumour growth. Nat Metab 2022;4:575-88.
- Geng F, Cheng X, Wu X, et al. Inhibition of SOAT1 Suppresses Glioblastoma Growth via Blocking SREBP-1-Mediated Lipogenesis. Clin Cancer Res 2016;22:5337-48.
- Feng X, Ma D, Zhao J, et al. UHMK1 promotes gastric cancer progression through reprogramming nucleotide metabolism. EMBO J 2020;39:e102541.
- Lane AN, Fan TW. Regulation of mammalian nucleotide metabolism and biosynthesis. Nucleic Acids Res 2015;43:2466-85.
- Ali ES, Lipońska A, O'Hara BP, et al. The mTORC1-SLC4A7 axis stimulates bicarbonate import to enhance de novo nucleotide synthesis. Mol Cell 2022;82:3284-3298.e7.
- 51. Cheng J, Huang Y, Zhang X, et al. TRIM21 and PHLDA3 negatively regulate the crosstalk between the PI3K/AKT pathway and PPP metabolism. Nat Commun

2020;11:1880.

- 52. Poulain L, Sujobert P, Zylbersztejn F, et al. High mTORC1 activity drives glycolysis addiction and sensitivity to G6PD inhibition in acute myeloid leukemia cells. Leukemia 2017;31:2326-35.
- Düvel K, Yecies JL, Menon S, et al. Activation of a metabolic gene regulatory network downstream of mTOR complex 1. Mol Cell 2010;39:171-83.
- Saha A, Connelly S, Jiang J, et al. Akt phosphorylation and regulation of transketolase is a nodal point for amino acid control of purine synthesis. Mol Cell 2014;55:264-76.
- 55. Mishra R, Yuan L, Patel H, et al. Phosphoinositide 3-Kinase (PI3K) Reactive Oxygen Species (ROS)-Activated Prodrug in Combination with Anthracycline Impairs PI3K Signaling, Increases DNA Damage Response and Reduces Breast Cancer Cell Growth. Int J Mol Sci 2021;22:2088.
- 56. Juvekar A, Hu H, Yadegarynia S, et al. Phosphoinositide 3-kinase inhibitors induce DNA damage through nucleoside depletion. Proc Natl Acad Sci U S A 2016;113:E4338-47.
- 57. Patra KC, Hay N. The pentose phosphate pathway and cancer. Trends Biochem Sci 2014;39:347-54.
- Mannava S, Grachtchouk V, Wheeler LJ, et al. Direct role of nucleotide metabolism in C-MYC-dependent proliferation of melanoma cells. Cell Cycle 2008;7:2392-400.
- 59. Dejure FR, Eilers M. MYC and tumor metabolism: chicken and egg. EMBO J 2017;36:3409-20.
- 60. Huang F, Huffman KE, Wang Z, et al. Guanosine triphosphate links MYC-dependent metabolic and ribosome programs in small-cell lung cancer. J Clin Invest 2021;131:e139929.
- 61. Zhao X, Petrashen AP, Sanders JA, et al. SLC1A5 glutamine transporter is a target of MYC and mediates reduced mTORC1 signaling and increased fatty acid oxidation in long-lived Myc hypomorphic mice. Aging Cell 2019;18:e12947.
- Amaya ML, Inguva A, Pei S, et al. The STAT3-MYC axis promotes survival of leukemia stem cells by regulating SLC1A5 and oxidative phosphorylation. Blood 2022;139:584-96.
- Robitaille AM, Christen S, Shimobayashi M, et al. Quantitative phosphoproteomics reveal mTORC1 activates de novo pyrimidine synthesis. Science 2013;339:1320-3.
- Ben-Sahra I, Hoxhaj G, Ricoult SJH, et al. mTORC1 induces purine synthesis through control of the mitochondrial tetrahydrofolate cycle. Science 2016;351:728-33.

- 65. Morita M, Gravel SP, Chénard V, et al. mTORC1 controls mitochondrial activity and biogenesis through 4E-BP-dependent translational regulation. Cell Metab 2013;18:698-711.
- 66. Böhm R, Imseng S, Jakob RP, et al. The dynamic mechanism of 4E-BP1 recognition and phosphorylation by mTORC1. Mol Cell 2021;81:2403-2416.e5.
- 67. Jia JJ, Lahr RM, Solgaard MT, et al. mTORC1 promotes TOP mRNA translation through sitespecific phosphorylation of LARP1. Nucleic Acids Res 2021;49:3461-89.
- 68. Hong S, Freeberg MA, Han T, et al. LARP1 functions as a molecular switch for mTORC1-mediated translation of an essential class of mRNAs. Elife 2017;6:e25237.
- Iadevaia V, Zhang Z, Jan E, et al. mTOR signaling regulates the processing of pre-rRNA in human cells. Nucleic Acids Res 2012;40:2527-39.
- 70. Shapiro GI, LoRusso P, Kwak E, et al. Phase Ib study of the MEK inhibitor cobimetinib (GDC-0973) in combination with the PI3K inhibitor pictilisib (GDC-0941) in patients with advanced solid tumors. Invest New Drugs 2020;38:419-32.
- 71. Schöffski P, Cresta S, Mayer IA, et al. A phase Ib study of pictilisib (GDC-0941) in combination with paclitaxel, with and without bevacizumab or trastuzumab, and with letrozole in advanced breast cancer. Breast Cancer Res 2018;20:109.
- 72. Mehraj U, Wani NA, Hamid A, et al. Adapalene inhibits the growth of triple-negative breast cancer cells by S-phase arrest and potentiates the antitumor efficacy of GDC-0941. Front Pharmacol 2022;13:958443.
- 73. Hwang Y, Kim HC, Shin EJ. BKM120 alters the migration of doublecortin-positive cells in the dentate gyrus of mice. Pharmacol Res 2022;179:106226.
- 74. Peng X, Zhang S, Jiao W, et al. Hydroxychloroquine synergizes with the PI3K inhibitor BKM120 to exhibit antitumor efficacy independent of autophagy. J Exp Clin Cancer Res 2021;40:374.
- 75. Kojima T, Kato K, Hara H, et al. Phase II study of BKM120 in patients with advanced esophageal squamous cell carcinoma (EPOC1303). Esophagus 2022;19:702-10.
- 76. Bohnacker T, Prota AE, Beaufils F, et al. Deconvolution of Buparlisib's mechanism of action defines specific PI3K and tubulin inhibitors for therapeutic intervention. Nat Commun 2017;8:14683.
- 77. Wicki A, Brown N, Xyrafas A, et al. First-in human, phase 1, dose-escalation pharmacokinetic and pharmacodynamic study of the oral dual PI3K and mTORC1/2 inhibitor

# Page 14 of 15

PQR309 in patients with advanced solid tumors (SAKK 67/13). Eur J Cancer 2018;96:6-16.

- 78. Foster P, Yamaguchi K, Hsu PP, et al. The Selective PI3K Inhibitor XL147 (SAR245408) Inhibits Tumor Growth and Survival and Potentiates the Activity of Chemotherapeutic Agents in Preclinical Tumor Models. Mol Cancer Ther 2015;14:931-40.
- 79. Edelman G, Rodon J, Lager J, et al. Phase I Trial of a Tablet Formulation of Pilaralisib, a Pan-Class I PI3K Inhibitor, in Patients with Advanced Solid Tumors. Oncologist 2018;23:401-e38.
- Matulonis U, Vergote I, Backes F, et al. Phase II study of the PI3K inhibitor pilaralisib (SAR245408; XL147) in patients with advanced or recurrent endometrial carcinoma. Gynecol Oncol 2015;136:246-53.
- 81. Soria JC, LoRusso P, Bahleda R, et al. Phase I doseescalation study of pilaralisib (SAR245408, XL147), a panclass I PI3K inhibitor, in combination with erlotinib in patients with solid tumors. Oncologist 2015;20:245-6.
- Furet P, Guagnano V, Fairhurst RA, et al. Discovery of NVP-BYL719 a potent and selective phosphatidylinositol-3 kinase alpha inhibitor selected for clinical evaluation. Bioorg Med Chem Lett 2013;23:3741-8.
- Juric D, Rodon J, Tabernero J, et al. Phosphatidylinositol 3-Kinase α-Selective Inhibition With Alpelisib (BYL719) in PIK3CA-Altered Solid Tumors: Results From the Firstin-Human Study. J Clin Oncol 2018;36:1291-9.
- Juric D, Janku F, Rodón J, et al. Alpelisib Plus Fulvestrant in PIK3CA-Altered and PIK3CA-Wild-Type Estrogen Receptor-Positive Advanced Breast Cancer: A Phase 1b Clinical Trial. JAMA Oncol 2019;5:e184475.
- 85. Rugo HS, André F, Yamashita T, et al. Time course and management of key adverse events during the randomized phase III SOLAR-1 study of PI3K inhibitor alpelisib plus fulvestrant in patients with HR-positive advanced breast cancer. Ann Oncol 2020;31:1001-10.
- 86. André F, Ciruelos EM, Juric D, et al. Alpelisib plus fulvestrant for PIK3CA-mutated, hormone receptorpositive, human epidermal growth factor receptor-2negative advanced breast cancer: final overall survival results from SOLAR-1. Ann Oncol 2021;32:208-17.
- Hanan EJ, Braun MG, Heald RA, et al. Discovery of GDC-0077 (Inavolisib), a Highly Selective Inhibitor and Degrader of Mutant PI3Kα. J Med Chem 2022;65:16589-621.
- Juric D, Kalinsky K, Turner NC, et al. First-line inavolisib/placebo + palbociclib + fulvestrant (Inavo/ Pbo+Palbo+Fulv) in patients (pts) with PIK3CA-mutated,

hormone receptor-positive, HER2-negative locally advanced/metastatic breast cancer who relapsed during/ within 12 months (mo) of adjuvant endocrine therapy completion: INAVO120 Phase III randomized trial additional analyses. J Clin Oncol 2024;42:1003.

- Gordon MJ, Huang J, Chan RJ, et al. Medical comorbidities in patients with chronic lymphocytic leukaemia treated with idelalisib: analysis of two large randomised clinical trials. Br J Haematol 2021;192:720-8.
- 90. Zelenetz AD, Barrientos JC, Brown JR, et al. Idelalisib or placebo in combination with bendamustine and rituximab in patients with relapsed or refractory chronic lymphocytic leukaemia: interim results from a phase 3, randomised, double-blind, placebo-controlled trial. Lancet Oncol 2017;18:297-311.
- 91. Sharman JP, Coutre SE, Furman RR, et al. Final Results of a Randomized, Phase III Study of Rituximab With or Without Idelalisib Followed by Open-Label Idelalisib in Patients With Relapsed Chronic Lymphocytic Leukemia. J Clin Oncol 2019;37:1391-402.
- 92. Heffron TP, Heald RA, Ndubaku C, et al. The Rational Design of Selective Benzoxazepin Inhibitors of the α-Isoform of Phosphoinositide 3-Kinase Culminating in the Identification of (S)-2-((2-(1-Isopropyl-1H-1,2,4triazol-5-yl)-5,6-dihydrobenzo[f]imidazo[1,2-d][1,4] oxazepin-9-yl)oxy)propanamide (GDC-0326). J Med Chem 2016;59:985-1002.
- Soler A, Figueiredo AM, Castel P, et al. Therapeutic Benefit of Selective Inhibition of p110α PI3-Kinase in Pancreatic Neuroendocrine Tumors. Clin Cancer Res 2016;22:5805-17.
- 94. Freitas de Sousa FJ, Nunes Azevedo FF, Santos de Oliveira FL, et al. Quantum biochemistry description of PI3Kα enzyme bound to selective inhibitors. J Biomol Struct Dyn 2024;42:9283-93.
- 95. Ndubaku CO, Heffron TP, Staben ST, et al. Discovery of 2-{3-[2-(1-isopropyl-3-methyl-1H-1,2-4-triazol-5-yl)-5,6-dihydrobenzo[f]imidazo[1,2-d][1,4]oxazepin-9-yl]-1H-pyrazol-1-yl}-2-methylpropanamide (GDC-0032): a β-sparing phosphoinositide 3-kinase inhibitor with high unbound exposure and robust in vivo antitumor activity. J Med Chem 2013;56:4597-610.
- 96. Dent S, Cortés J, Im YH, et al. Phase III randomized study of taselisib or placebo with fulvestrant in estrogen receptor-positive, PIK3CA-mutant, HER2-negative, advanced breast cancer: the SANDPIPER trial. Ann Oncol 2021;32:197-207.
- 97. Shah N, Mohammad AS, Saralkar P, et al. Investigational

chemotherapy and novel pharmacokinetic mechanisms for the treatment of breast cancer brain metastases. Pharmacol Res 2018;132:47-68.

- Shor RE, Dai J, Lee SY, et al. The PI3K/mTOR inhibitor Gedatolisib eliminates dormant breast cancer cells in organotypic culture, but fails to prevent metastasis in preclinical settings. Mol Oncol 2022;16:130-47.
- Singh AR, Joshi S, Burgoyne AM, et al. Single Agent and Synergistic Activity of the "First-in-Class" Dual PI3K/ BRD4 Inhibitor SF1126 with Sorafenib in Hepatocellular Carcinoma. Mol Cancer Ther 2016;15:2553-62.
- 100. Munakata W, Tobinai K. Clinical development of voxtalisib: a pan-PI3K/mTOR inhibitor. Lancet Haematol 2018;5:e134-5.
- 101. Álvarez RM, García AB, Riesco-Fagundo C, et al. Omipalisib inspired macrocycles as dual PI3K/mTOR inhibitors. Eur J Med Chem 2021;211:113109.
- 102.Zhao YY, Wu DM, He M, et al. Samotolisib Attenuates Acute Liver Injury Through Inhibiting Caspase-11-Mediated Pyroptosis Via Regulating E3 Ubiquitin Ligase Nedd4. Front Pharmacol 2021;12:726198.
- 103. Collins GP, Eyre TA, Schmitz-Rohmer D, et al. A Phase II Study to Assess the Safety and Efficacy of the Dual mTORC1/2 and PI3K Inhibitor Bimiralisib (PQR309) in Relapsed, Refractory Lymphoma. Hemasphere 2021;5:e656.
- 104.Jonchere B, Williams J, Zindy F, et al. Combination of Ribociclib with BET-Bromodomain and PI3K/mTOR Inhibitors for Medulloblastoma Treatment In Vitro and In

#### doi: 10.21037/tbcr-24-29

**Cite this article as:** Han B, Lin X, Hu H. Regulation of PI3K signaling in cancer metabolism and PI3K-targeting therapy. Transl Breast Cancer Res 2024;5:33.

Vivo. Mol Cancer Ther 2023;22:37-51.

- 105.Omeljaniuk WJ, Krętowski R, Ratajczak-Wrona W, et al. Novel Dual PI3K/mTOR Inhibitor, Apitolisib (GDC-0980), Inhibits Growth and Induces Apoptosis in Human Glioblastoma Cells. Int J Mol Sci 2021;22:11511.
- 106. Powles T, Lackner MR, Oudard S, et al. Randomized Open-Label Phase II Trial of Apitolisib (GDC-0980), a Novel Inhibitor of the PI3K/Mammalian Target of Rapamycin Pathway, Versus Everolimus in Patients With Metastatic Renal Cell Carcinoma. J Clin Oncol 2016;34:1660-8.
- 107. Venkatesan AM, Dehnhardt CM, Delos Santos E, et al. Bis(morpholino-1,3,5-triazine) derivatives: potent adenosine 5'-triphosphate competitive phosphatidylinositol-3-kinase/mammalian target of rapamycin inhibitors: discovery of compound 26 (PKI-587), a highly efficacious dual inhibitor. J Med Chem 2010;53:2636-45.
- 108. Drullinsky PR, Hurvitz SA. Mechanistic basis for PI3K inhibitor antitumor activity and adverse reactions in advanced breast cancer. Breast Cancer Res Treat 2020;181:233-48.
- 109. Sabbah DA, Hajjo R, Bardaweel SK, et al. Targeting the PI3K/AKT signaling pathway in anticancer research: a recent update on inhibitor design and clinical trials (2020-2023). Expert Opin Ther Pat 2024;34:141-58.
- 110. Nunnery SE, Mayer IA. Management of toxicity to isoform α-specific PI3K inhibitors. Ann Oncol 2019;30 Suppl 10:x21-6.