RESEARCH

Open Access



GCKR common functional polymorphisms are associated with metabolic syndrome and its components: a 10-year retrospective cohort study in Iranian adults

Asiyeh Sadat Zahedi, Mahdi Akbarzadeh, Bahareh Sedaghati-Khayat, Atefeh Seyedhamzehzadeh and Maryam S. Daneshpour^{*}

Abstract

Background: Previous studies reported that common functional variants (rs780093, rs780094, and rs1260326) in the glucokinase regulator gene (GCKR) were associated with metabolic syndrome despite the simultaneous association with the favorable and unfavorable metabolic syndrome components. We decided to evaluate these findings in a cohort study with a large sample size of Iranian adult subjects, to our knowledge for the first time. We investigated the association of the GCKR variants with incident MetS in mean follow-up times for nearly 10 years.

Methods: Analysis of this retrospective cohort study was performed among 5666 participants of the Tehran Cardiometabolic Genetics Study (TCGS) at 19–88 years at baseline. Linear and logistic regression analyses were used to investigate the metabolic syndrome (JIS criteria) association and its components with rs780093, rs780094, and rs1260326 in an additive genetic model. Cox regression was carried out to peruse variants' association with the incidence of metabolic syndrome in the TCGS cohort study.

Results: In the current study, we have consistently replicated the association of the GCKR SNPs with higher triglyceride and lower fasting blood sugar levels (p < 0.05) in Iranian adults. The CT genotype of the variants was associated with lower HDL-C levels. The proportional Cox adjusted model regression resulted that TT carriers of rs780094, rs780093, and rs1260326 were associated with 20%, 23%, and 21% excess risk metabolic syndrome incidence, respectively (p < 0.05).

Conclusions: Elevated triglyceride levels had the strongest association with GCKR selected variants among the metabolic syndrome components. Despite the association of these variants with decreased fasting blood sugar levels, T alleles of the variants were associated with metabolic syndrome incidence; so whether individuals are T allele carriers of the common functional variants, they have a risk factor for the future incidence of metabolic syndrome.

Keywords: Metabolic syndrome, Triglyceride, Single nucleotide polymorphisms, GCKR

Background

The metabolic syndrome (MetS) represents a combination of metabolic abnormalities that increase the risk of developing Type 2 diabetes mellitus by fivefold, cardiovascular diseases (CVDs) by twofold, and the risk of all-cause mortality by 1.5-fold [1, 2]. These metabolic abnormalities include insulin resistance, elevated

*Correspondence: daneshpour@sbmu.ac.ir Cellular and Molecular Research Center, Research Institute for Endocrine Sciences, Shahid Beheshti University of Medical Sciences, POBox: 19195-4763, Tehran, Iran



© The Author(s) 2021. This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativeco mmons.org/licenses/by/4.0/. The Creative Commons Public Domain Dedication waiver (http://creativecommons.org/publicdomain/ $\frac{1}{2}$ zero/1.0/) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

blood pressure, dyslipidemia, and central obesity [3]. So, early diagnosis of metabolic syndrome and evaluation of its predisposing factors can be as valuable as the study of diabetes biomarkers [4, 5] and strong biomarkers of cardiovascular disease [6-8] in the prevention and treatment of two disorders. The pathophysiological mechanism that is explicating MetS remains unclear, and maybe more than one. It is noticeable that lipid profile and glucose metabolism change due to central obesity and insulin resistance, and this change leads to the development of MetS [9]. The average worldwide prevalence of metabolic syndrome is 31% [10]. In comparison, according to the International Diabetes Federation (IDF) criteria, the prevalence of this disorder in Iranian adults estimate 37.4% [11] and, based on a meta-analysis of published data during 2000-2016, is higher in some ethnic groups (Bushehr 57.8%) [12]. Therefore, it is crucial to consider the factors that contribute to metabolic syndrome as a health threat. MetS is a multifactorial disease, and besides environmental factors, genetic studies have revealed susceptible loci in associations with MetS [13].

In the liver and the Langerhans islets' beta-cells, glycolytic enzyme glucokinase (GCK) contributes to glycogen synthesis regulation and gluconeogenesis as a primary glucose sensor [14]. In the first step of glycolysis, GCK is responsible for glucose phosphorylation; thus, GCK carries out a pivotal role in maintaining blood glucose homeostasis [15]. Glucokinase regulatory protein (GKRP) regulates glycolysis by inhibiting GCK enzymatic activity at low glucose concentrations. When glucose concentration rises, the GCK/GKRP complex disconnects so that more GCK are available, leading to increased hepatic glucose utilization [16]. GCK activity rises through overexpression of the glucokinase regulator gene (GCKR) in the liver, leading to decreased glucose and increased triglyceride concentrations [17].

Glucokinase regulator gene (GCKR) locates on chromosome 2p23.3-p23.2 and containing 19 exons, encodes GKRP (68 kDa, 625 amino acids) [18, 19]. Genome-wide association studies (GWAS) and multiple candidate gene studies have reported that GCKR genetic variation is associated with metabolic parameters such as triglyceride (TG) levels [20-26], fasting blood sugar (FBS) [21-23, 26–28], and insulin [27, 28] or metabolic disorders like type 2 diabetes (T2DM) [27, 29], dyslipidemia (high triglyceride and low high-density lipoprotein cholesterol levels) [23, 30]. Common functional variants, rs780094, rs780093, and rs1260326, are broadest studied genetic variants of the GCKR gene [21, 24, 26, 29, 30]. The T allele of mentioned SNPs is associated with FBS concentration reduction, insulin level, lower insulin resistance, and T2DM prevalence. Otherwise, the associations with higher fasting serum triglyceride concentration in several populations were reported [21–24, 26, 29, 30]. This opposite effect on both TG and insulin traits did not affect the association between the T-allele of the variants and MetS in some previous findings [30, 31].

Association between common polymorphisms of the *GCKR* gene and MetS investigated in a few studies among the Iranian population; Most of them performed during childhood. None of which focused on the associations between the *GCKR* variants and MetS components. Earlier in a cross-sectional study, we confirmed the association of rs780094 and rs1260326 with an increased risk of developing metabolic syndrome and its components, published in Persian [32].

In the present study, a retrospective cohort approach, we investigated the association of *GCKR* variants (rs780093, rs780094, rs1260326) with MetS incidence and its components in new cases MetS by utilizing data of the Tehran Cardiometabolic Genetic Study (TCGS).

Methods

Study population

Subjects of this retrospective cohort study were selected from TCGS participants [33]. TCGS is an ongoing genetic study that is a part of the Tehran Lipid and Glucose Study (TLGS). This cohort study estimates the prevalence of non-communicable disease risk factors in a sample of Tehran's (capital of Iran) 13th district residents [34]. The first survey of the TLGS was initiated from 1999 to 2001 on 15,005 individuals aged 3 years, and subjects were genotyped and followed up to identify recently developed diseases every 3 years. In the current study, we excluded individuals with prevalent MetS at baseline. A total of 5666 adults 19-88 aged years who had complete demographic, anthropometric, biochemical, and genotype baseline data were included in the statistical analyses. All subjects' written consent was obtained, and the Research Institute for Endocrine Sciences (affiliated to Shahid Beheshti University of Medical Sciences, Tehran, Iran) approved this study.

Measurements

At each survey of TLGS, participants signed a consent form. Information for age, sex, and history of using medication for diabetes, hypertension, smoking, and lipid disorders collected with a standardized questionnaire. Anthropometric measurements were recorded using standard protocols; waist circumference (WC) was measured and recorded to the nearest 0.1 cm using a non-stretch tape meter. Systolic blood pressure (SBP), diastolic blood pressure (DBP) were measured as described previously [34]. A blood sample was drawn after 12–14 h overnight fasting. Samples were centrifuged within 30–45 min of collection, and the sera were used for biochemical measurements. FBS, TG, and highdensity lipoprotein cholesterol (HDL-C) were measured by the enzymatic colorimetric method, using commercial kits (Pars Azmoon, Tehran, Iran); also, coefficients of variation (CV) for HDL-C and triglyceride measurements were below 5%.

Genetic analysis

To select SNPs in *GCKR*, we searched for variants whose function was identified and proven. In previous studies, the function of rs1260326 is described as reducing the inhibitory properties of the GKRP, which leads to increased glucokinase activity in the liver [35, 36]. The rs780093 and rs780094 are in strong linkage disequilibrium with this functional variant in different populations [37]; therefore, we investigated these three variants' effects on metabolic syndrome in the Iranian population.

Genomic samples were extracted from the buffy coat using the standard Proteinase K, the salting-out method. DNA samples were genotyped with HumanOmniExpress-24-v1-0 bead chips (containing 649,932 SNP loci with an average mean distance of 4 kb) at the deCODE genetics company (Iceland) according to the manufacturer's specifications (Illumina Inc., San Diego, CA, USA). Quality control procedures were performed by the PLINK program (V 1.07) and R statistic (V 3.2), and the genotyping data of *GCKR* polymorphisms (rs780094, rs1260326, and 780093) were used for present association analysis.

Study outcomes

Outcomes were the components of MetS as continuous and categorized traits [30], and the prospective outcome was incident MetS. The MetS were defined According to the most recent Joint Interim Statement (JIS) of the International Diabetes Federation Task Force on Epidemiology and Prevention [32]. Individuals were considered with metabolic syndrome if they had at least three of the following metabolic components in at least one survey of TCGS: central obesity (waist circumference \geq 90 cm for both genders based on the Guidelines of the Iranian National Committee for Obesity [38]); elevated TG (fasting serum $TG \ge 150 \text{ mg/dl}$ and drug treatment); reduced HDL-c (fasting serum HDL-c < 40 mg/dl in men and < 50 mg/dl in women and drug treatment); elevated blood pressure (SBP \geq 130 mmHg or DBP \geq 85 mmHg or taking hypertension medication); high fasting glucose (FBS \geq 100 mg/dl or taking diabetes medication).

Statistical analysis

SPSS software version 21.0 (SPSS Inc. Chicago, IL, USA) was used for most statistical analyses. Only we calculated pairwise linkage disequilibrium (LD) using

the "genetics" package in R. The baseline characteristics were presented as means \pm standard deviation (SD) for continuous traits or as number (percentage) for categorical traits. Hence, serum triglyceride levels were not normally distributed, so their natural logarithm (Ln) was taken before analysis. *P*-value < 0.05 was considered significant. Chi-square test was used to investigate deviations of observed genotype frequencies from those predicted by the Hardy–Weinberg equation and for determining the frequency of genotypes in metabolic trait groups.

Chi-square test and logistic regression were performed to test the association between the MetS and its components as categorized traits and the three variants. To reduced heterogeneity of differing lengths of followup in odds ratios and considered the duration of followup of censored observations, we estimated the hazard ratio and 95% confidence interval of the incidence of MetS. Two Cox models were performed; model 1 unadjusted model, and model 2 was adjusted for age (baseline for Mets and without MetS), gender, and smoking. We defined time to event for participants with MetS as the mid-time between the date of the follow-up visit when the Mets were diagnosed for the first time and the most recent follow-up visit before the diagnosis, and for participants without MetS, the time between baseline to the last follow-up. The effects of the GCKR polymorphisms on the MetS components as quantitative traits were evaluated first by analyzing variance, ANOVA followed by posthoc Tukey's test, and then utilizing linear regression models. In examining the association between the SNPs and MetS components (categorized or quantitative) for individuals with MetS, the first occurrence of MetS in the follow-up period was considered the baseline.

Results

Population characteristics

The population's average age was 39.2 years in the baseline, and 57.4% were female. During the follow-up period of about ten years, two thousand eight hundred eightyfour subjects (50.9%) developed MetS in at least one survey of the present study. Analysis of the distribution of MetS components demonstrated that low-level HDL-C was the most common component of metabolic syndrome, which observes in 69.8% of subjects. Increased levels of fasting glucose had the lowest frequency (22.3%) (Table 1).

Genotypes characteristics

Genotype frequencies of three polymorphisms followed the Hardy–Weinberg equilibrium (p > 0.01). The minor

	Total	rs780094			rs780093			rs1260326		
		CC (n = 2543)	CT (n=4169)	TT (n = 1879)	CC (n=2509)	CT (n = 4180)	TT (n = 1902)	CC (n=2478)	CT (n = 4174)	TT (n = 1939)
With MetS	50.9%	28.1%	49%	23%*	28.2%	48%	23.8%*	27.4%	49.0%	23.6%*
Without MetS	49.1%	32.5%	47.1%	20.4%*	31.6%	48.2%	20.2%*	31.4%	47.7%	20.9%*
Male	42.6%	30.9%	47.8%	21.2%	30.8%	47.5%	21.6%	30.3%	48.0%	21.8%
Female	57.4%	29.7%	48.0%	22.2%	29.3%	48.4%	22.3%	28.8%	48.5%	22.7%
Smoking never	83%	30.6%	47.9%	21.5%	30.3%	48.0%	21.7%	29.7%	48.4%	21.9%
Ex-smoker and smoker	17%	28.0%	48.9%	23.1%	28.0%	49.0%	23.0%	27.8%	48.2%	23.9%
Age (years)	39土16	39土17	39土16	40土17	39土17	39土16	40土17	39土17	39土16	39.5 土 16.6
WC (cm)	85±21 (53.3%) ^a	85 土 21	85 土 21	85 土 22	85 土 22	85 土 2 1	85 土 22	85 土 22	85 ± 21	85.4±21.5
HDL-C (mg/dl)	42土14 (69.8%) ^b	43土14	42 土 14	42 土 15	43土14	42 土 14	42土14	43土14	42土14	42土14
TG (mg/dl)	137土86 (41.8%) ^c	125 土 75	139±88	$150 \pm 91^{+}$	126土81	137±84	$152 \pm 92^{+}$	125 土 74	138±88	$151 \pm 92^{+}$
FBS (mg/dl)	89±25 (22.3%) ^d	90 土 26	89 土 23	88 土 27	90±27	89±23	89±27	90±27	89.4±23.1	88.8±26.5
SBP (mmHg)	110土26(28.8%) ^e	110土26	110土25	111土27	110土26	110土25	111土26	110土26	110土26	111.1 土 26
DBP (mmHg)	73±17 (28.8%) ^e	72 土 17	73 土 17	73 土 18	72±18	73 土 17	73±18	72 土 18	72 土 17	73 土 17
From the 6th row onwards, data are presented as mean $\pm\text{SD}$, data are presented as	mean ± SD								

if participants
haracteristics o
Baseline c
Table 1

 a Frequency of subjects with high waist circumference: waist circumference \geq 90 cm for both genders

 $^{\rm b}\,$ Frequency of subjects low HDL-C: HDL-C < 40 mg/dl in men and < 50 mg/dl in women and drug treatment

 $^{\rm c}\,$ Frequency of subjects high TG: TG \ge 150 mg/dl and drug treatment

 $^{\rm d}\,$ Frequency of subjects high FBS: FBS \ge 100 mg/dl and drug treatment

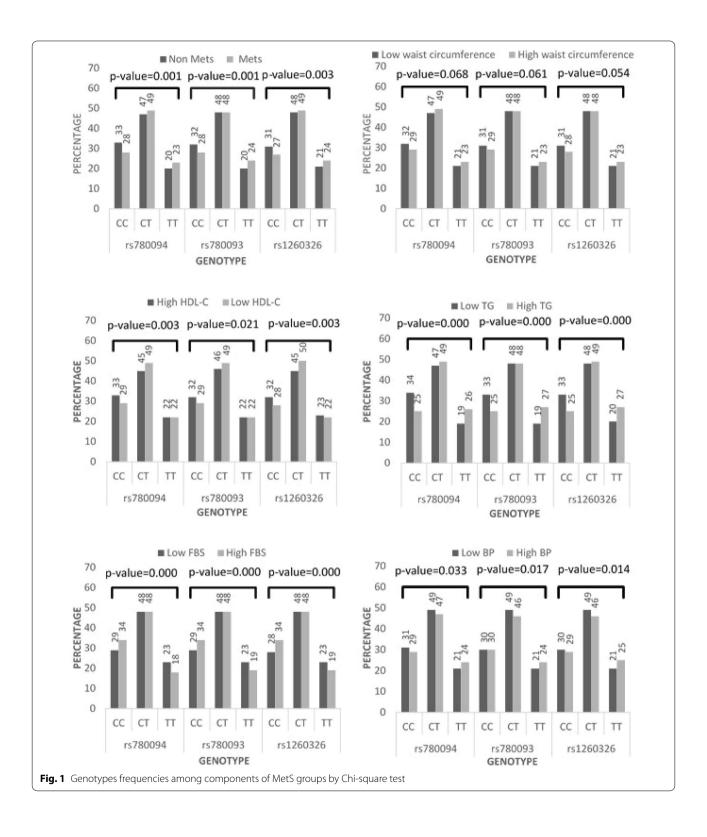
 $^{\rm e}~$ Frequency of subjects high blood pressure: SBP \ge 130 mmHg and DBP \ge 85 mmHg and drug treatment

 $^{\ast}\,$ Frequency differences of genotypes between MetS and without MetS, P < 0.05 $\,$ $^{\dagger}\,$ TT vs. CC, P < 0.05 and TT vs CT, P < 0.05

T.

allele frequencies (MAF) were 45.8%, 46.1%, and 46.4% for the rs780094, rs780093, and rs1260326 alleles. The three SNPs are in strong LD in our population (D' = 0.972

for rs780094 and rs780093, D' = 0.964 for rs1260326 and rs780094, and D' = 0.924 for rs1260326 and rs780093).



The T alleles of the common variants of *GCKR* were frequently found in the MetS affected by non-MetS (Table 1 and Fig. 1). Results of the logistic regression analysis confirmed the association between T alleles and MetS. Adjusted odds ratios for age, gender, and smoking were 1.28 (95% CI 1.08 to 1.53), 1.34 (95% CI 1.13 to 1.6), and 1.28 (95% CI 1.08 to 1.53) (Table 2) respectively; so the homozygotes for T were at higher risk of MetS incidence.

Cox regression analysis

The mean follow-up period was nearly 10 years. In Cox unadjusted model, hazard ratios were significantly higher with each additional copy of the T allele in rs780094 and rs1260326. Also, in rs780093, TT carriers had a higher incidence of MetS than CC carriers (Table 3, model 1). After adjusting for age, sex, and smoking, TT carriers of rs780094, rs780093, and rs1260326 were associated with

Table 2 Association of GCKR SNPs with MetS and qualitative MetS components in an additive	genetic model

Categorical traits	rs780094	OR (CI) model 1	Adjusted OR (CI) model 2	rs780093	OR (CI) model 1	Adjusted OR (CI) model 2	rs1260326	OR (CI) model 1	Adjusted OR (CI) model 2
Metabolic syndrome	CC (refer- ence)	1	1	CC (refer- ence)	1	1	CC (refer- ence)	1	1
	CT	1.2 (1.06– 1.36)*	1.22 (1.05– 1.41)*	CT	1.11 (0.99– 1.26)	1.14 (0.99– 1.32)	СТ	1.17 (1.04– 1.33)*	1.19 (1.03– 1.38)*
	TT	1.3 (1.12– 1.51)*	1.28 (1.08– 1.53)*	TT	1.32 (1.14– 1.53)*	1.34 (1.13–1.6)*	TT	1.29 (1.11– 1.49)*	1.28 (1.08– 1.53)*
High waist circumfer-	CC (refer- ence)	1	1	CC (refer- ence)	1	1	CC (refer- ence)	1	1
ence	CT	1.12 (0.99– 1.27)	1.09 (0.95– 1.25)	CT	1.08 (0.95– 1.22)	1.08 (0.94– 1.24)	CT	1.1 (0.97– 1.25)	1.08 (0.94– 1.24)
	Π	1.18 (1.02– 1.38)*	1.13 (0.96– 1.34)	TT	1.2 (1.03– 1.4)*	1.17 (0.99– 1.39)	TT	1.2 (1.03– 1.4)*	1.17 (0.99– 1.38)
Low HDL-C	CC (refer- ence)	1	1	CC (refer- ence)	1	1	CC (refer- ence)	1	1
	CT	1.26 (1.1–1.44)*	1.25 (1.09– 1.43)*	CT	1.21 (1.06– 1.38)*	1.2 (1.04– 1.37)*	CT	1.26 (1.1–1.44)*	1.25 (1.09– 1.43)*
	TT	1.11 (0.94–1.3)	1.09 (0.93– 1.28)	TT	1.12 (0.95– 1.31)	1.1 (0.93– 1.29)	TT	1.12 (0.95– 1.31)	1.08 (0.91– 1.27)
High TG	CC (refer- ence)	1	1	CC (refer- ence)	1	1	CC (refer- ence)	1	1
	CT	1.36 (1.19– 1.55)*	1.38 (1.2–1.58)*	CT	1.3 (1.14– 1.48)*	1.33 (1.16– 1.53)*	CT	1.36 (1.2–1.56)*	1.39 (1.21– 1.6)*
	TT	1.78 (1.52– 2.08)*	1.78 (1.5–2.1)*	TT	1.8 (1.54– 2.1)*	1.83 (1.55– 2.16)*	TT	1.84 (1.57– 2.14)*	1.86 (1.57– 2.19)*
High FBS	CC (refer- ence)	1	1	CC (refer- ence)	1	1	CC (refer- ence)	1	1
	CT	0.85 (0.73– 0.98)*	0.82 (0.71– 0.96)*	CT	0.84 (0.73– 0.97)*	0.83 (0.71– 0.97)*	СТ	0.83 (0.71– 0.96)*	0.8 (0.68– 0.94)*
	TT	0.68 (0.57– 0.82)*	0.64 (0.52– 0.77)*	TT	0.7 (0.58– 0.84)*	0.65 (0.53– 0.79)*	TT	0.67 (0.56–0.8)*	0.63 (0.52– 0.77)*
High blood pressure	CC (refer- ence)	1	1	CC (refer- ence)	1	1	CC (refer- ence)	1	1
	CT	0.99 (0.87– 1.14)	0.96 (0.83– 1.13)	CT	0.96 (0.83–1.1)	0.95 (0.81–1.1)	CT	0.96 (0.84– 1.11)	0.93 (0.8–1.09)
	TT	1.20 (1.02– 1.41)*	1.16 (0.97–1.4)	TT	1.19 (1.01–1.4)*	1.16 (0.97– 1.39)	ТТ	1.2 (1.02– 1.41)*	1.18 (0.98– 1.42)

High TG: TG \geq 150 mg/dl and drug treatment; low HDL-C: HDL-C < 40 mg/dl in men and < 50 mg/dl in women and drug treatment; high blood pressure: SBP \geq 130 mmHg and/or DBP \geq 85 mmHg and drug treatment; high FBS: FBS \geq 100 mg/dl and drug treatment; high waist circumference: waist circumference \geq 90 cm for both genders

Cl confidence interval, OR odds ratio

Model 1: unadjusted

Model 2: adjusted for age, gender, and smoking

* *P* < 0.0

Table 3 Hazard ratios for metabolic syndrome by GCKR SNPs in an additive genetic model

Variants	Genotype	HR (CI) model 1	Adjusted HR (CI) model 2
rs780094	CC (reference)	1	1
	СТ	1.14 (1.04–1.24)*	1.15 (1.05–1.26)*
	TT	1.22 (1.1–1.353)*	1.2 (1.08–1.34)*
rs780093	CC (reference)	1	1
	СТ	1.08 (0.99–1.18)	1.13 (1.03–1.23)*
	TT	1.23 (1.11–1.36)*	1.23 (1.11–1.37)*
rs1260326	CC (reference)	1	1
	СТ	1.11 (1.02–1.21)*	1.14 (1.04–1.25)*
	TT	1.21(1.09–1.34)*	1.21 (1.09–1.35)*

Model 1: unadjusted

Model 2: adjusted for age, gender, and smoking

Cl confidence interval, HR hazard ratio

* P<0.05

20%, 23%, and 21% excess risk MetS incidence, respectively (Table 3, model 2).

The values of odds ratios and hazard ratios showed that the strongest association, among the three variants, was observed between TT carriers of rs780093 and the incidence of metabolic syndrome.

Associations with metabolic syndrome components

In baseline characteristics (first occurrence of MetS for patients with MetS), a significant difference in mean TG was observed in all genotype groups; T allele carriers of the *GCKR* variants had higher mean TG (Table 1).

Chi-square test showed that in subjects with a high TG, high blood pressure, and low HDL-C, T homozygotes of the three SNPs were more frequent. However, in people with high FBS, C homozygotes people were significantly more prevalent (Fig. 1). The first and second logic models' results determined that, with and without adjustment, C allele carriers, T allele carriers, and CT carriers in all SNPs were associated with high FBS, high TG, and low HDL-C levels, respectively. In model 1, associations were observed between high waist circumference and high blood pressure with TT carriers of the three SNPs; this association was lost after adjusting in the second model (Table 2).

The findings of the linear regression showed after age, sex, and smoking adjustment, the TT genotypes of rs780094, rs780093 and rs1260326 were associated with increased TG levels (effect per allele respectively: 0.11 mg/dl, 0.12 mg/dl and 0.12 mg/dl, p < 0.000) and reduced FBS levels (effect per allele in all SNPs: -0.03 mg/dl, p < 0.01) (Table 4).

Discussion

In the present retrospective cohort study, the strongest association of MetS components with the common functional *GCKR* variants was observed between elevated TG levels and T alleles. T alleles of the variants also were associated with MetS incidence over 10 years.

We consistently replicated previous GWAS results that reported the T allele carriers of selected variants had significant associations with lower FBS and higher TG levels among MetS components [22, 23, 25, 39–42]. Among Iranian studies, two studies in children and adolescents (CASPIAN III) have previously demonstrated the association between the rs780094 risk allele and higher triglyceride levels [43, 44]. The correlation of rs1260326 and rs780094 with these MetS components has also been confirmed in our previous cross-sectional study in adults [32]. In 74 patients with non-alcoholic fatty liver disease from Tabriz city in the north-west of Iran, the lack of association between the *GCKR* rs780094 genotypes

Table 4 Association of GCKR SNPs with quantitative components o	of MetS in an additive genetic model
---	--------------------------------------

Quantitative	rs780094			rs780093			rs1260326		
traits	Unstandardized coefficients ^a	Standardized coefficients ^a	P-value	Unstandardized coefficients ^a	Standardized coefficients ^a	P-value	Unstandardized coefficients ^a	Standardized coefficients ^a	P-value
	B(SE)	В		B(SE)	β		B(SE)	β	
Waist circumfer- ence	- 0.16 (0.33)	- 0.01	0.624	- 0.01 (0.33)	0.00	0.979	0 (0.33)	0.00	0.998
HDL-C	- 0.35 (0.24)	- 0.02	0.135	- 0.34 (0.24)	- 0.02	0.15	- 0.28 (0.24)	- 0.01	0.243
FBS	- 0.95 (0.4)	- 0.03	0.017*	- 0.88 (0.4)	- 0.03	0.026*	- 0.9 (0.4)	- 0.03	0.023*
TG (natural log)	0.08 (0.01)	0.11	0.000*	0.08 (0.01)	0.12	0.000*	0.09 (0.01)	0.12	0.000*
SBP	0.07 (0.38)	0.00	0.845	0.12 (0.38)	0.00	0.758	0.12 (0.38)	0.00	0.762
DBP	0.21 (0.28)	0.01	0.465	0.3 (0.28)	0.01	0.288	0.32 (0.28)	0.01	0.251

^a In age, gender, and smoking-adjusted linear regression model

* P<0.05

and lipid profile was observed. This finding was probably attributed to the limited sample size of this study [45].

As described above in the current study, the common variants of *GCKR* were concurrently associated with both desired and undesired MetS components and MetS, like the findings of some previous studies [22, 30]. In some other investigations, the authors suggested that the T allele's declining effect on the FBS levels has contributed to a lack of association between the *GCKR* variants and MetS [23, 46]. The obtained regression coefficient (β) values in the current study demonstrated a stronger association of the *GCKR* variants with TG than FBS. Besides, the adjusted odds ratio of the MetS components for the development of MetS (IDF criteria) in a previous study in the TLGS population, from which subjects of the present study were derived, showed that triglyceride had the highest power among other MetS components [47].

Based on our findings, CT genotypes of the *GCKR* SNPs were associated with lower HDL-C levels. In a cohort study (Atherosclerosis Risk in Communities) among white participants, the T-allele of rs780094 was associated with higher HDL-C levels. While in black participants of this study, no association was found with HDL-C [30]. This disparity in findings may be attributed more to different ethnicities than any other cause. As in another study where the participants were Taiwanese Adolescents, T-allele carriers of rs780094 were more highly represented among participants with low HDL-C levels [31].

The expression of GCKR produces glucokinase regulatory protein (GKRP), inhibits the enzyme GCK in competition with glucose substrate, and plays a vital role in regulating glucose storage and disposal. The GKRP itself is activated by fructose 6-phosphate binding and inactivated by fructose 1-phosphate binding. The variants investigated in this study can affect the regulation of protein by fructose 6-phosphate; for instance, based on the results of a study, GKRP from the expression of mutation in rs1260326 less regulated by fructose in the physiological range of fructose-6-phosphate concentrations, which leads to a reduction in inhibition of GCK. This reduction increases the rate of glycolysis and, therefore, glucose uptake by the liver. Enhanced glycolytic flux leads to increased triglyceride levels [35]. GKRP inhibits GCK and also stabilizes this enzyme; at low glucose concentrations. GK/GKRP complex locates in the nucleus of liver hepatocytes. As glucose levels increase, GCK enters the cytoplasm, and GKRP protects GCK against degradation by cytoplasmic proteases. Therefore, structure and function changes affect glycolysis and lipolysis regulation [30, 35, 48]. Thus defects in the mentioned pathways can lead to metabolic syndrome [49].

The minor allele frequency (MAF) of the selected GCKR SNPs was different from the other Iranian population study [43]. This difference could be Because of the current study's greater sample size. The SNPs' minor allele frequency in our study was similar to those in East Asia and Europe [37]. The LD pattern of 3 SNPs varies in different Hap Map populations [30]. In the TCGS population, rs780093, rs1260326, and rs780094 were in strong LD. Based on obtained β and OR, rs780093, which, based on our knowledge, had not yet been studied in the Iranian population, slightly stronger than other variants in LD, was associated with MetS incidence. The present cohort study's strengths included using large sample size and selecting new cases of MetS, which attenuated the likelihood of any behavior changes due to awareness of metabolic abnormalities.

Conclusions

We replicated associations of the common *GCKR* variants with metabolic syndrome components, including fasting blood sugar and triglyceride levels in Iranian adults. In the TCGS study participants, rs780094, rs780093, and rs1260326 T alleles were also associated with increased metabolic syndrome incidence. Given that in the present study, three polymorphisms were in linkage disequilibrium; therefore, whether individuals are T allele carriers, they have a risk factor for the future incidence of MetS.

Abbreviations

CI: Confidence interval; CV: Coefficients of variation; CVDs: Cardiovascular diseases; DBP: Diastolic blood pressure; FBS: Fasting blood sugar; GCK: Glycolytic enzyme glucokinase; *GCKR*: Glucokinase regulator gene; GKRP: Glucokinase regulatory protein; GWAS: Genome-wide association studies; HDL-C: High-density lipoprotein cholesterol; HR: Hazard ratio; IDF: International Diabetes Federation; JIS: Joint Interim Statement; LN: Natural logarithm; MetS: Metabolic syndrome; OR: Odds ratios; SBP: Systolic blood pressure; SD: Standard deviation; SNPs: Single nucleotide polymorphisms; T2DM: Type 2 diabetes; TCGS: Tehran Cardiometabolic Genetics Study; TG: Triglyceride; TLGS: Tehran Lipid and Glucose Study; WC: Waist circumference.

Acknowledgements

The authors would like to thank the participants in the TCGS project. This study was financially and scientifically supported by the Research Institute of Endocrine Sciences, Shahid Beheshti University of Medical Sciences, I.R. Iran (Grant No. 12228). We also express our gratitude for supporting the deCODE genetic company (Reykjavik, Iceland).

Authors' contributions

BS and MD, study design. AZ, AS, and MA, literature review, data analysis, interpretation, and manuscript preparation. MA and AZ, data collection, and analysis. MD and AZ, manuscript review, critical appraisal, and specialist advice. All authors read and approved the final manuscript.

Funding

This study was supported in part by Grant No. 12228 from Shahid Beheshti University of Medical Sciences.

Availability of data and materials

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Ethics approval and consent to participate

Written informed consent was obtained from all participants. The research protocol was approved by the ethics committee of the Research Institute for Endocrine Sciences, Shahid Beheshti University of Medical Sciences, Tehran, Iran.

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

Received: 27 December 2020 Accepted: 8 February 2021 Published online: 18 February 2021

References

- Mottillo S, Filion KB, Genest J, Joseph L, Pilote L, Poirier P, Rinfret S, Schiffrin EL, Eisenberg MJ. The metabolic syndrome and cardiovascular risk: a systematic review and meta-analysis. J Am Coll Cardiol. 2010;56:1113–32.
- Nsiah K, Shang VO, Boateng KA, Mensah FO. Prevalence of metabolic syndrome in type 2 diabetes mellitus patients. Int J Appl Basic Med Res. 2015;5:133–8.
- 3. Reaven G. Metabolic syndrome: pathophysiology and implications for management of cardiovascular disease. Circulation. 2002;106:286–8.
- Giglio RV, Lo Sasso B, Agnello L, Bivona G, Maniscalco R, Ligi D, Mannello F, Ciaccio M. Recent updates and advances in the use of glycated albumin for the diagnosis and monitoring of diabetes and renal, cerebro- and cardio-metabolic diseases. J Clin Med. 2020;9(11):3634.
- Bellia C, Zaninotto M, Cosma C, Agnello L, Bivona G, Marinova M, Lo Sasso B, Plebani M, Ciaccio M. Clinical usefulness of glycated albumin in the diagnosis of diabetes: results from an Italian study. Clin Biochem. 2018;54:68–72.
- Bivona G, Agnello L, Bellia C, Lo Sasso B, Ciaccio M. Diagnostic and prognostic value of H-FABP in acute coronary syndrome: still evidence to bring. Clin Biochem. 2018;58:1–4.
- Zinellu A, Sotgia S, Porcu P, Casu MA, Bivona G, Chessa R, Deiana L, Carru C. Carotid restenosis is associated with plasma ADMA concentrations in carotid endarterectomy patients. Clin Chem Lab Med. 2011;49:897–901.
- Agnello L, Bellia C, Scazzone C, Bivona G, Iacolino G, Gambino CM, Muratore M, Lo Sasso B, Ciaccio M. Establishing the 99(th) percentile for high sensitivity cardiac troponin I in healthy blood donors from Southern Italy. Biochem Med. 2019;29:020901.
- 9. Kahn R, Buse J, Ferrannini E, Stern M. The metabolic syndrome: time for a critical appraisal. Diabetologia. 2005;48:1684–99.
- 10. Engin AB, Engin A. Obesity and lipotoxicity. Cham: Springer; 2017.
- Delavari A, Forouzanfar MH, Alikhani S, Sharifian A, Kelishadi R. First nationwide study of the prevalence of the metabolic syndrome and optimal cutoff points of waist circumference in the Middle East: the national survey of risk factors for noncommunicable diseases of Iran. Diabetes Care. 2009;32:1092–7.
- Farmanfarma KK, Kaykhaei MA, Adineh HA, Mohammadi M, Dabiri S, Ansari-moghaddam A. Prevalence of metabolic syndrome in Iran: a metaanalysis of 69 studies. Diabetes Metab Syndr Clin Res Rev. 2019;13:792–9.
- 13. Fall T, Ingelsson E. Genome-wide association studies of obesity and metabolic syndrome. Mol Cell Endocrinol. 2014;382:740–57.
- Matschinsky FM, Glaser B, Magnuson MA. Pancreatic beta-cell glucokinase: closing the gap between theoretical concepts and experimental realities. Diabetes. 1998;47:307–15.
- Matschinsky FM. A lesson in metabolic regulation inspired by the glucokinase glucose sensor paradigm. Diabetes. 1996;45:223–41.
- 16. Van Schaftingen E. A protein from rat liver confers to glucokinase the property of being antagonistically regulated by fructose 6-phosphate and fructose 1-phosphate. Eur J Biochem. 1989;179:179–84.

- O'Doherty RM, Lehman DL, Télémaque-Potts S, Newgard CB. Metabolic impact of glucokinase overexpression in liver: lowering of blood glucose in fed rats is accompanied by hyperlipidemia. Diabetes. 1999;48:2022–7.
- Veiga-da-Cunha M, Delplanque J, Gillain A, Bonthron D, Boutin P, Van Schaftingen E, Froguel P. Mutations in the glucokinase regulatory protein gene in 2p23 in obese French caucasians. Diabetologia. 2003;46:704–11.
- Warner J, Leek J, Intody S, Markham A, Bonthron D. Human glucokinase regulatory protein (GCKR): cDNA and genomic cloning, complete primary structure, and chromosomal localization. Mamm Genome. 1995;6:532–6.
- Benyamin B, Middelberg RP, Lind PA, Valle AM, Gordon S, Nyholt DR, et al. GWAS of butyrylcholinesterase activity identifies four novel loci, independent effects within BCHE and secondary associations with metabolic risk factors. Hum Mol Genet. 2011;20:4504–14.
- Kitamoto A, Kitamoto T, Nakamura T, Ogawa Y, Yoneda M, Hyogo H, et al. Association of polymorphisms in GCKR and TRIB1 with nonalcoholic fatty liver disease and metabolic syndrome traits. Endocr J. 2014. https://doi.org/10.1507/endocrj.EJ14-0052.
- Kraja AT, Vaidya D, Pankow JS, Goodarzi MO, Assimes TL, Kullo IJ, et al. A bivariate genome-wide approach to metabolic syndrome: STAMPEED consortium. Diabetes. 2011;60:1329–39.
- Kristiansson K, Perola M, Tikkanen E, Kettunen J, Surakka I, Havulinna AS, et al. Genome-wide screen for metabolic syndrome susceptibility Loci reveals strong lipid gene contribution but no evidence for common genetic basis for clustering of metabolic syndrome traits. Circ Cardiovasc Genet. 2012;5:242–9.
- 24. Orho-Melander M, Melander O, Guiducci C, Perez-Martinez P, Corella D, Roos C, et al. Common missense variant in the glucokinase regulatory protein gene is associated with increased plasma triglyceride and C-reactive protein but lower fasting glucose concentrations. Diabetes. 2008;57:3112–21.
- Saxena R, Voight BF, Lyssenko V, Burtt NP, de Bakker PI, Chen H, et al. Genome-wide association analysis identifies loci for type 2 diabetes and triglyceride levels. Science. 2007;316:1331–6.
- Kim OY, Kwak SY, Lim H, Shin MJ. Genotype effects of glucokinase regulator on lipid profiles and glycemic status are modified by circulating calcium levels: results from the Korean Genome and Epidemiology Study. Nutr Res. 2018;60:96–105.
- 27. Dupuis J, Langenberg C, Prokopenko I, Saxena R, Soranzo N, Jackson AU, et al. New genetic loci implicated in fasting glucose homeostasis and their impact on type 2 diabetes risk. Nat Genet. 2010;42:105.
- Ramos E, Chen G, Shriner D, Doumatey A, Gerry N, Herbert A, et al. Replication of genome-wide association studies (GWAS) loci for fasting plasma glucose in African-Americans. Diabetologia. 2011;54:783–8.
- Vaxillaire M, Cavalcanti-Proença C, Dechaume A, Tichet J, Marre M, Balkau B, Froguel P. The common P446L polymorphism in GCKR inversely modulates fasting glucose and triglyceride levels and reduces type 2 diabetes risk in the DESIR prospective general French population. Diabetes. 2008;57:2253–7.
- Bi M, Kao WHL, Boerwinkle E, Hoogeveen RC, Rasmussen-Torvik LJ, Astor BC, North KE, Coresh J, Köttgen A. Association of rs780094 in GCKR with metabolic traits and incident diabetes and cardiovascular disease: the ARIC study. PLoS ONE. 2010;5(7):e11690.
- Chang H-W, Lin F-H, Li P-F, Huang C-L, Chu N-F, Su S-C, et al. Association between a glucokinase regulator genetic variant and metabolic syndrome in Taiwanese adolescents. Genet Test Mol Biomark. 2016;20:137–42.
- Zahedi AS, Sedaghati-Khayat B, Behnami S, Azizi F, Daneshpour MS. Associations of common polymorphisms in GCKR with metabolic syndrome. Tehran Univ Med J TUMS Publ. 2018;76:459–68.
- Daneshpour MS, Fallah M-S, Sedaghati-Khayat B, Guity K, Khalili D, Hedayati M, et al. Rationale and design of a genetic study on cardiometabolic risk factors: protocol for the Tehran Cardiometabolic Genetic Study (TCGS). JMIR Res Protoc. 2017;6:e28.
- 34. Azizi F, Ghanbarian A, Momenan AA, Hadaegh F, Mirmiran P, Hedayati M, Mehrabi Y, Zahedi-Asl S. Prevention of non-communicable disease in a population in nutrition transition: Tehran lipid and glucose study phase II. Trials. 2009;10:5.
- Beer NL, Tribble ND, McCulloch LJ, Roos C, Johnson PR, Orho-Melander M, Gloyn AL. The P446L variant in GCKR associated with fasting plasma glucose and triglyceride levels exerts its effect through increased glucokinase activity in liver. Hum Mol Genet. 2009;18:4081–8.

- Rees MG, Wincovitch S, Schultz J, Waterstradt R, Beer NL, Baltrusch S, Collins FS, Gloyn AL. Cellular characterisation of the GCKR P446L variant associated with type 2 diabetes risk. Diabetologia. 2012;55:114–22.
- Auton A, Abecasis GR, Altshuler DM, Durbin RM, Abecasis GR, Bentley DR, et al. A global reference for human genetic variation. Nature. 2015;526:68–74.
- Azizi F, Khalili D, Aghajani H, Esteghamati A, Hosseinpanah F, Delavari A et al. Appropriate waist circumference cut-off points among Iranian adults: the first report of the Iranian National Committee of Obesity. 2010.
- Kamatani Y, Matsuda K, Okada Y, Kubo M, Hosono N, Daigo Y, Nakamura Y, Kamatani N. Genome-wide association study of hematological and biochemical traits in a Japanese population. Nat Genet. 2010;42:210.
- Kathiresan S, Willer CJ, Peloso GM, Demissie S, Musunuru K, Schadt EE, et al. Common variants at 30 loci contribute to polygenic dyslipidemia. Nat Genet. 2009;41:56–65.
- Oh S-W, Lee J-E, Shin E, Kwon H, Choe EK, Choi S-Y, Rhee H, Choi SH. Genome-wide association study of metabolic syndrome in Korean populations. PLoS ONE. 2020;15:e0227357.
- Surakka I, Horikoshi M, Mägi R, Sarin A-P, Mahajan A, Lagou V, et al. The impact of low-frequency and rare variants on lipid levels. Nat Genet. 2015;47:589–97.
- Hovsepian S, Javanmard SH, Mansourian M, Tajadini M, Hashemipour M, Kelishadi R. Interaction of GCKR, MLXIPL and FADS genes polymorphisms with obesity in the occurrence of childhood metabolic syndrome. Middle East J Fam Med. 2018;99:1–9.
- 44. Hovsepian S, Javanmard SH, Mansourian M, Hashemipour M, Tajadini M, Kelishadi R. Lipid regulatory genes polymorphism in children with and

without obesity and cardiometabolic risk factors: the CASPIAN-III study. J Res Med Sci. 2018;23:11.

- Mohammadi S, Farajnia S, Shadmand M, Mohseni F, Baghban R. Association of rs780094 polymorphism of glucokinase regulatory protein with non-alcoholic fatty liver disease. BMC Res Notes. 2020;13:26.
- 46. Kong X, Zhang X, Xing X, Zhang B, Hong J, Yang W. The association of type 2 diabetes loci identified in genome-wide association studies with metabolic syndrome and its components in a Chinese population with type 2 diabetes. PLoS ONE. 2015;10:e0143607.
- Heidari Z, Hosseinpanah F, Mehrabi Y, Safarkhani M, Azizi F. Predictive power of the components of metabolic syndrome in its development: a 6.5-year follow-up in the Tehran lipid and glucose study (TLGS). Eur J Clin Nutr. 2010;64:1207–14.
- 48. Mohás M, Kisfali P, Járomi L, Maász A, Fehér E, Csöngei V, et al. GCKR gene functional variants in type 2 diabetes and metabolic syndrome: do the rare variants associate with increased carotid intima-media thickness? Cardiovasc Diabetol. 2010;9:79.
- Nirala NK, Rahman M, Walls SM, Singh A, Zhu LJ, Bamba T, et al. Survival response to increased ceramide involves metabolic adaptation through novel regulators of glycolysis and lipolysis. PLoS Genet. 2013;9:e1003556.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

At BMC, research is always in progress.

Learn more biomedcentral.com/submissions

