



Case Control Study

Histological difference in ligament flavum between degenerative lumbar canal stenosis and non-stenotic group: A prospective, comparative study

Mantu Jain, Mukund Sable, Amit Purushottam Tirpude, Rabi Narayan Sahu, Sudeep Kumar Samanta, Gurudip Das

Specialty type: Orthopedics

Provenance and peer review:

Invited article; Externally peer reviewed.

Peer-review model: Single blind

Peer-review report's scientific quality classification

Grade A (Excellent): 0

Grade B (Very good): B, B

Grade C (Good): 0

Grade D (Fair): 0

Grade E (Poor): 0

P-Reviewer: Elgafy H, United States; Vahedi P, Iran

Received: January 17, 2022

Peer-review started: January 17, 2022

First decision: February 21, 2022

Revised: March 2, 2022

Accepted: August 24, 2022

Article in press: August 24, 2022

Published online: September 18, 2022



Mantu Jain, Sudeep Kumar Samanta, Gurudip Das, Department of Orthopaedics, All India Institute of Medical Sciences Bhubaneswar, Bhubaneswar 751019, Odisha, India

Mukund Sable, Department of Pathology, All India Institute of Medical Sciences Bhubaneswar, Bhubaneswar 751019, Odisha, India

Amit Purushottam Tirpude, Department of Anatomy, All India Institute of Medical Sciences Raipur, Raipur 492009, Chattisgarh, India

Rabi Narayan Sahu, Department of Neurosurgery, All India Institute of Medical Sciences Bhubaneswar, Bhubaneswar 751019, Odisha, India

Corresponding author: Mantu Jain, MD, Doctor, Surgeon, Department of Orthopaedics, All India Institute of Medical Sciences Bhubaneswar, Bhubaneswar 751019, Odisha, India. montu_jn@yahoo.com

Abstract

BACKGROUND

Ligament flavum (LF) hypertrophy is the main etiopathogenesis of lumbar canal stenosis (LCS). The purely elastic LF undergoes a morphological adaptation including a reduction in the elastic fibers and a consequent increase in the collagen content, fibrosis, cicatrization, and calcification. However, the morphometric analysis can delineate the LF in patients with LCS from those without LCS, which would help in better understanding LCS pathogenesis.

AIM

To compare the histopathological changes in LF between the degenerative LCS and non-stenotic (non-LCS) group.

METHODS

The present prospective study was conducted in 82 patients who were divided into two groups, namely LCS and non-LCS. Demographic details of the patients such as duration of symptoms, level of involvement, and number of segments were recorded. The LF obtained from both groups was histopathologically examined for the fibrosis score, elastic fiber degeneration, calcification, and

chondroid metaplasia. Morphometrical details included a change in elastin and collagen percentages, elastin/collagen ratio, elastic fiber fragmentation, and ligamentocyte numbers. All parameters were compared between the two groups by using the independent t test, Chi-square test, and Pearson's correlation test.

RESULTS

Out of 82 cases, 74 were analysed, 34 in LCS and 40 in non-LCS group. The mean $\pm$ SD age of presentation in LCS and non-LCS group was 49.2 ± 8.9 and 43.1 ± 14.3 respectively. The LCS group ($n = 34$) exhibited significant differences in fibrosis ($P = 0.002$), elastic fiber degeneration ($P = 0.01$), % elastic fragmentation (66.5 ± 16.3 vs 29.5 ± 16.9), % elastic content (26.9 ± 6.7 vs 34.7 ± 8.4), % collagen content (63.6 ± 10.4 vs 54.9 ± 6.4), reduction of elastic/collagen (0.4 ± 0.1 vs 0.6 ± 0.1), and ligamentocyte number (39.1 ± 19.1 vs 53.5 ± 26.9) as compared to non-LCS group ($n = 40$). The calcification ($P = 0.08$) and Pearson's correlation between duration and loss of elastin was not significant. The difference in LF morphology is consistent in patient's ≥ 40 years of age among the groups as found in subgroup analysis. Similarly in the patients < 40 and > 40 in the non-LCS group.

CONCLUSION

LF is vital in the pathogenesis of LCS. The purely elastic LF undergoes a morphological adaptation that includes a reduction in the elastic fibers with a consequent increase in the collagen content, fibrosis, cicatrization, and calcification. The present study provides a detailed morphometric analysis to semiquantitatively delineate the LF changes in patients with LCS from those in patients without LCS.

Key Words: Spinal stenosis; Lumbar spine; Ligamentum flavum; histopathology; Morphometry

©The Author(s) 2022. Published by Baishideng Publishing Group Inc. All rights reserved.

Core Tip: The ligament flavum (LF) is vital in the pathogenesis of lumbar canal stenosis (LCS). The purely elastic LF undergoes a morphological adaptation that includes a reduction in the elastic fibers with a consequent increase in the collagen content, fibrosis, cicatrization, and calcification. The present study provides a detailed morphometric analysis to semiquantitatively delineate the LF changes in patients with LCS from those in patients without LCS.

Citation: Jain M, Sable M, Tirpude AP, Sahu RN, Samanta SK, Das G. Histological difference in ligament flavum between degenerative lumbar canal stenosis and non-stenotic group: A prospective, comparative study. *World J Orthop* 2022; 13(9): 791-801

URL: <https://www.wjgnet.com/2218-5836/full/v13/i9/791.htm>

DOI: <https://dx.doi.org/10.5312/wjo.v13.i9.791>

INTRODUCTION

Lumbar canal stenosis (LCS) is a common spinal disorder that affects elderly patients, leading to lower back pain, leg pain, and neurogenic claudication, which rarely ends in paresis[1,2]. Because the ligamentum flavum (LF) covers most of the posterior and lateral part of the lumbar spinal canal, the hypertrophied facets and LF hypertrophy (LFH) are responsible for LCS despite the disc complex, contributing to spinal canal narrowing[2,3]. Thus, morphological and histological changes in developing lumbar spinal canal encroachment must be studied[4,5]. With aging of the global population, a paradigm shift toward symptomatic LCS that requires surgical treatment is observed[6].

LF contains the purest form of elastic tissue among ligaments. These elastic fibers decrease with age and are replaced by collagen fibers[2]. The causes of LFH are multifactorial, including the activity levels, age, and mechanical stress[7,8]. Based on transmission electron microscopy findings, Postacchini *et al*[9] concluded that the reduced elasticity might cause bulging of the LF into the spinal canal even in the standing position.

Studies have exhibited a qualitative transformation in the dynamics of the LF components with degeneration[1,6,10-12]. The LFH exhibited loss of elastic fibers, increased content of collagen fibers, and chondrometaplasia, leading to calcification. A few studies have suggested this association of calcification[10], whereas two other studies have suggested a decrease in the elastin/collagen ratio[2,4]. Electron microscopy has revealed fragmentation and changes in the quality of elastic fibers.

Therefore, the present study attempted to explain the LF structure histologically in a semi-quantitative manner by using advanced imaging software.

MATERIALS AND METHODS

The present prospective study was conducted following the Helsinki Declaration principles and approved by the local institutional review board (T/IM/18-19/43 dated 04/01/2019). Valid written informed consent was obtained from all participants.

Study population

The present study was conducted in 74 adult patients undergoing lumbar spine surgery between January 2019 and March 2020 in the departments of orthopedics or neurosurgery in a tertiary care center. Patients with characteristic clinical and radiological findings of spinal stenosis were grouped as the LCS study group. Simultaneously, patients with lumbar disc herniation, infective etiology (tubercular or non-tubercular), trauma, and malignancy constituted the non-LCS study group. Patients with prior lumbar surgery were excluded from the study. Clinical details such as age, sex, duration of symptoms, and level of involvement were recorded.

LF samples were obtained from 82 patients who underwent decompressive laminectomy in piecemeal by using a Kerrison's rongeur and sent for histopathological assay in neutral buffer formalin. Of these, eight samples were excluded due to sufficient tissue availability. Thus, the final analysis was conducted in 74 patients.

Histology

The harvested LF components were kept in a solution containing 10% neutral buffered formalin for 24 h. The tissues were processed overnight on an automatic tissue processor (Leica Biosystems Ltd.) and embedded in paraffin. Multiple 4- μ m thick sections were cut. All sections were stained with haematoxylin and eosin (H & E), Verhoef-Van-Gieson stains (VVG), Masson trichrome stain (MTS), reticulin, and Von-kossa stains.

Light microscopy

The histological evaluation was performed by two experts who were blinded to the nature of the groups. Light microscopy was used to examine LF sections for elastic degeneration, fibrosis, metaplasia (chondroid or osteoid), hemorrhage, and calcification. Elastic degeneration was graded depending upon the percentage of elastic fibers exhibiting degenerative changes and elastin fiber fragmentation (1+: 0%–33%, 2+: 33%–66%, and 3+: > 67% of elastic fibers). Fibrosis was graded as per the criterion described previously[2].

Image acquisition and morphometric analysis

Digital images were obtained using a commercial imaging system (ZEN blue edition on ZEISS Scope A.1 microscope) at 200X magnification for H & E, VVG, and MTS stains in the tagged image file format. For each case, at least two areas from different LF regions were captured randomly. All images were imported in Image J software java windows-64 application (ImageJ bundled with 64-bit Java 1.8.0_172) for morphometric analysis.

The morphological analysis of the digital images of different stains was performed for elastin content (VVG), collagen content (MTS), and the number of ligamentocytes (H & E). The tool was used to estimate the percentage of collagen and elastic fibers, elastic to collagen fiber ratio, number of ligamentocytes, and elastic fiber width. Each parameter was estimated three times, and two independent observers recorded the average.

Statistical analysis

Continuous data are presented as mean $\pm$ standard deviation (SD). Differences in the mean between the groups were tested using the independent t test. The Chi-square test was used to compare the categorical variables between the two groups. Correlation between various morphological parameters and duration of symptoms was determined using the Pearson's correlation test for the stenotic group. Subgroup analysis was also performed for patients aged ≥ 40 years and for patients aged < 40 and ≥ 40 years in the non-LCS group. All differences associated with a chance probability of ≤ 0.05 were considered. Data were analyzed using the IBM Statistical Package for Social Sciences ver. 17 (SPSS Inc., Chicago, IL, United States).

RESULTS

Clinical features

Of the 74 cases, 34 cases constitute the LCS group and 40 cases constituted the non-LCS group (Figure 1). The mean $\pm$ SD age of presentation in the LCS and non-LCS groups was 49.2 ± 8.9 and 43.1 ± 14.3 years, respectively. The percentage of patients in the stenotic group aged > 40 years was 91%, whereas that in the non-stenotic group was 57.5%. This difference was statistically significant ($P = 0.001$) (Fischer exact test). Although the non-stenotic group included permanent lumbar disc herniation, other pathologies were also observed (Table 1). Single-level involvement was observed in majority of cases. L4-5 involvement was observed in 43.5% patients, followed by L3-4 and L5-S1 involvement (23% patients each). The remaining patients exhibited involvement in L1-2 and L2-3 levels.

Histological differences between LCS and non-LCS groups

The histological differences in elastin fibers, collagen content, and ligamentocytes were compared between the two groups. The LCS group exhibited higher elastic degeneration and fibrosis than the non-LCS group ($P = 0.01$ and 0.002 , respectively) (Table 2; Figures 2-4). On the other hand, the extent of calcification, chondroid metaplasia, and hemorrhage was statistically nonsignificant between the groups.

LF exhibited a significant reduction in the elastin content in the LCS group ($P < 0.0001$, independent t test) and an increase in the collagen content ($P < 0.0001$, independent t test) compared with those in the non-LCS group. The elastin/collagen ratio, width of elastic fibers, and ligamentocyte number were also significantly lower in the LCS group than in the non-LCS group (Table 3).

Correlation of clinicopathological features between the LCS and non-LCS groups

The Pearson's correlation test indicated a moderate correlation between decrease in the number of ligamentocytes and age in both groups (non-LCS-R: -0.52 , $P < 0.001$ and LCS-R: -0.578 , $P < 0.001$). All other morphological changes were statistically nonsignificant. The duration of symptoms in the LCS group was not significantly correlated with morphological changes in both groups.

The ligamentocyte number was moderately correlated with the elastin content in the LCS group (R: -0.450 , $P = 0.008$), (Figure 5). On the other hand, the collagen content was moderately correlated with the elastic content in the non-LCS group (R: -0.504 , $P = 0.001$). The LCS group exhibited an inverse correlation between the elastin and collagen contents. However, this difference was statistically nonsignificant.

Subgroup analysis

Subgroup analysis exhibited a statistically nonsignificant difference between the patients aged < 40 years and those aged > 40 years in the LCS group. Additionally, the difference between the LCS and non-LCS groups in the percentage of patients aged > 40 years was statistically significant (Table 4).

DISCUSSION

The LF, which envelops the spinal canal, is a highly elastic structure that contains four times pure elastin than collagen[6]. If the LF surrounding the spinal canal becomes hypertrophic, it will compress the dural sac containing the cauda equina or the nerve root. Elsberg[13] first reported a case of LFH causing sciatica. Several clinical studies have reported that LFH is the primary pathology in LCS[1,2,4,6-10]. Although surgical excision is the only therapeutic management for patients with LFH in LCS, a deeper understanding of the pathophysiology can encourage future nonsurgical or prophylactic treatment modalities. Thus, the present study attempted to study the pathological changes in LF and compare the LCS and non-LCS study groups.

Etiology

Sairyo *et al*[1] had proposed that LFH occurs due to degenerative changes with aging process, and also due to increased mechanical stress occurring in instability. Wang *et al*[14] have experimentally demonstrated increased motion in lumbar spine induced LFH. Chuang *et al*[15] have found that age-related LFH occurred due to activation of the Akt and MAPK (apoptotic) pathways. The authors also postulated that hypertrophy is initiated in all subjects after the second decade of life. Zaki *et al*[16] also found that older individuals had some loss and rupture of elastic fibres with abnormal collagen, increase in vascularity and ossification particularly in the lumbar region as compared to the thoracic and cervical spine. Postacchini *et al*[9] observed that although older individuals with disc herniation exhibited some elastic fiber loss, the stenotic group of similar age exhibited more collagen and chondroid metaplasia and were strikingly different. However, the authors noted that there was no difference was observed in stenotic changes related to age and listhesis (degenerative), implying that instability does not accelerate hypertrophic changes[9]. The present study also exhibited no LCS-

Table 1 Demographic profile of the groups

Categories		LSS (n = 34)	Non-LSS (n = 40)	P value
Age in years, mean (SD)		49.2 (8.9)	43.1 (14.3)	
Sex in %		M:F – 59:41	M:F – 60:40	
Duration in weeks, mean (SD)		44.1 (11.6)	9.4 (8.1)	
Level	One	30 (88.2)	32 (80)	0.553
	Two	3 (8.8)	7 (17.5)	
	Three	1 (2.9)	1 (2.5)	
Diagnosis		Lumbar canal stenosis	PIVD – 18	
			Trauma – 8	
			Potts spine – 7	
			Tumour – 5	
			Epidural abscess (non-TB) – 2	

Table 2 Histological differences between the groups

Variable	Histological features	Stenotic (n = 34)	Non stenotic (n = 40)	P value (Chi square test)
Elastin degeneration	1+ and 2+	9 (26.5)	22 (55)	0.010
	3+	25 (73.5)	18 (45)	
Fibrosis	1+ and 2+	10	17	0.002
	3+	24	13	
Calcification	Absent	28	38	0.081
	Present	6	2	
Chondroid metaplasia	Absent	22	26	0.979
	Present	12	14	
Haemorrhage	Absent	17	23	0.519
	Present	17	17	

induced morphological changes in the non-LCS group of similar age. Similarly, no difference was observed in the non-LCS group with age < 40 years, whereas a statistically significant difference was observed from age > 40 years.

Levels and age

Sairyo *et al*[1] exhibited that LF thickness increased with age; however, the changes with age exhibited spinal level dependence. The increment at L4/5 and L3/4 levels was more extensive than that at L2/3 and L5/S1 levels. Similar changes in magnetic resonance imaging (MRI) were also reported by Kolte *et al*[7]. Okuda *et al*[10] exhibited that thickening was correlated to calcification, which was prime in LCS. A positive correlation was also observed by authors between calcification and clinical scoring (Japanese orthopedic association scores). The present study did not measure the thickness, either grossly, histologically, or radiologically. The present understanding of LCS has evolved from a pure 'static compression' (dependent on width) to a more "dynamic compression" that arises from the imbalance in LF components rather than actual width. Altun *et al*[2] hypothesized that the loss of elasticity is the contributing factor for LF infolding, leading to spinal canal narrowing[2].

Schröder *et al*[12] also studied 41 Ligaments in 21 patients and reported single-level stenosis in five patients, bi-segmental stenosis in 24 patients, and stenosis on three levels in 12 patients. Additionally, Hulmani *et al*[4] exhibited more double-level stenosis in their series. However, the present study exhibited more single-level involvement ($n = 32$) than double-level ($n = 7$) or multilevel ($n = 1$) involvement. This may be due to the higher age group cohort in the study by Hulmani *et al*[4] compared to that in the present study (72 vs 49) due to the preferential selection. The present study exhibited that L4-5 was the most common involvement, followed by L3-4. Sairyo *et al*[1] proposed that high mechanical stress might be responsible for the preferential increase in thickness at the L4-5 level.

Table 3 Morphometric differences between the groups

Factor		mean	SD	t	df	P value
% Elastin fragmentation	NS (40)	29.50	16.9	-9.51	71	< 0.0001
	S (34)	66.47	16.3			
Collagen content (area %)	NS (40)	54.94	6.41	-4.38	53.2	< 0.0001
	S (34)	63.59	10.36			
Elastic content (area %)	NS (40)	34.73	8.36	4.36	72	< 0.0001
	S (34)	26.94	6.71			
Elastin/Collagen ratio	NS (40)	0.63	0.14	6.01	71.27	< 0.0001
	S (34)	0.43	0.13			
Width of elastin fibers	NS (39)	4.37	1.07	5.44	71	< 0.0001
	S (34)	3.27	0.54			
Ligamentocyte numbers	NS (40)	53.45	26.85	2.6	72	< 0.0001
	S (34)	39.11	19.05			

Table 4 Subgroup analysis among groups > 40 years

Factor		mean	SD	t	df	P value
% Elastin fragmentation	NS (25)	31.80	18.123	-7.4	43.93	< 0.0001
	S (31)	67.10	16.96			
Collagen content (area %)	NS (25)	55.12	7.62	-3.4	53.19	0.001
	S (31)	63.58	10.75			
Elastic content (area %)	NS (25)	34.898	7.92	4.20	46.6	< 0.0001
	S (31)	26.57	6.60			
Elastin/Collagen ratio	NS (25)	0.63	0.14	5.54	50.09	< 0.0001
	S (31)	0.43	0.13			
Width of elastin fibers	NS (25)	3.75	0.98	4.42	31.94	< 0.0001
	S (31)	2.77	0.49			
Ligamentocyte numbers	NS (25)	46.56	22.4.0	1.88	41.03	0.66
	S (31)	36.612	15.41			

Histology

Okuda *et al*[10] and Elsberg *et al*[13] observed that nearly all ligaments were calcified in LCS. Calcium deposition within the ligament significantly aggravated the symptoms, and this process increased with age. The increase in the formation of calcium crystals is a significant factor for LF thickening. Okuda *et al* [10] observed that the mean age of patients with calcification and those without calcification was 74 ± 2.0 and 68 ± 1.4 years, respectively. Therefore, patients with calcification were significantly older than those in the LCS group. No reactive granulomatous tissue formation was noticed in the calcification focus. Other researchers have reiterated a smooth transition between calcific zones and surroundings[10,17]. Okuda *et al*[10] reported focal and dispersion-type calcification in their patients with LCS and correlated calcification with a low clinical score.

Altun *et al*[2], Hulmani *et al*[4], and Reyes-Sánchez *et al*[18] exhibited contrasting findings regarding calcification. No statistical difference was observed in calcification between the LCS group and the lumbar disc herniation (LDH) group in these studies.

Schröder *et al*[12] exhibited calcification of all the ligaments, and the patients also exhibited relevant fibrosis with decrease in the elastic/collagenous fiber ratio. Additionally, Sairyo *et al*[1] reported that the LCS group exhibited increased LF thickness and fibrosis with reduced elastic fibers. These transformations were more predominant along the dorsal side than those along the middle of the dural side. Sato *et al* also found that dorsal side is affected 30 more than the dural side[19] Peng *et al*[20] revealed that the dorsal fibers of the LF were subjected to higher stress than the dural fibers that have a fluid-filled tube,

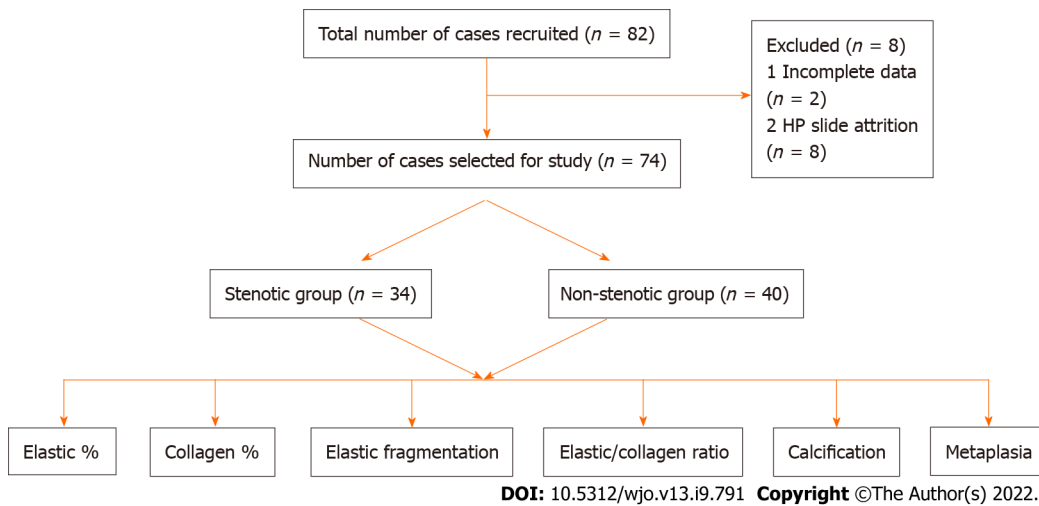


Figure 1 Flow diagram of patients.

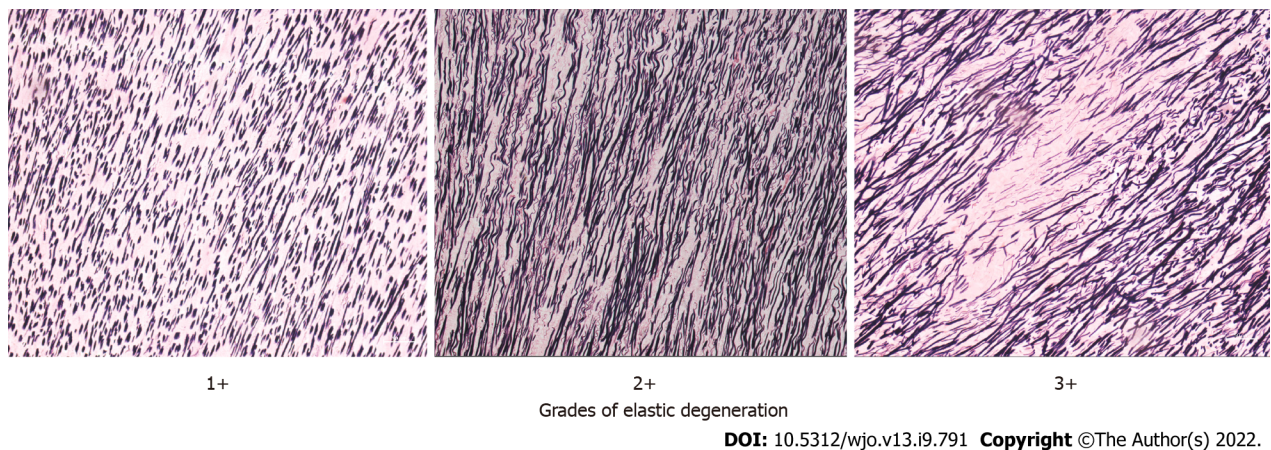


Figure 2 The three grades of elastic degeneration.

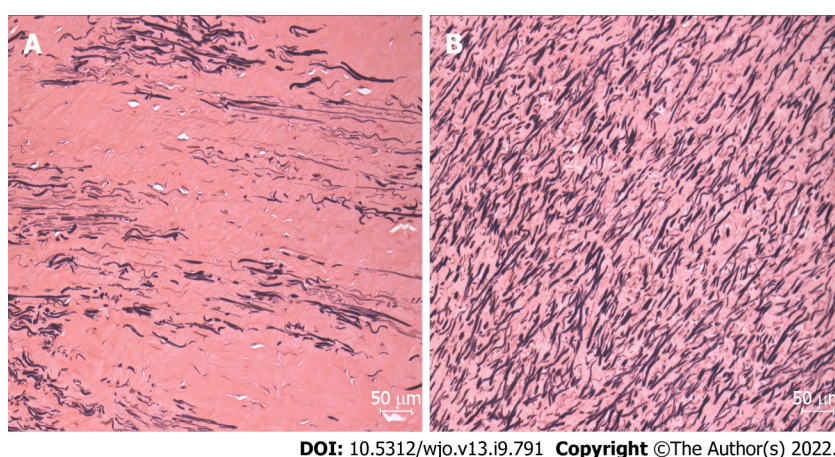
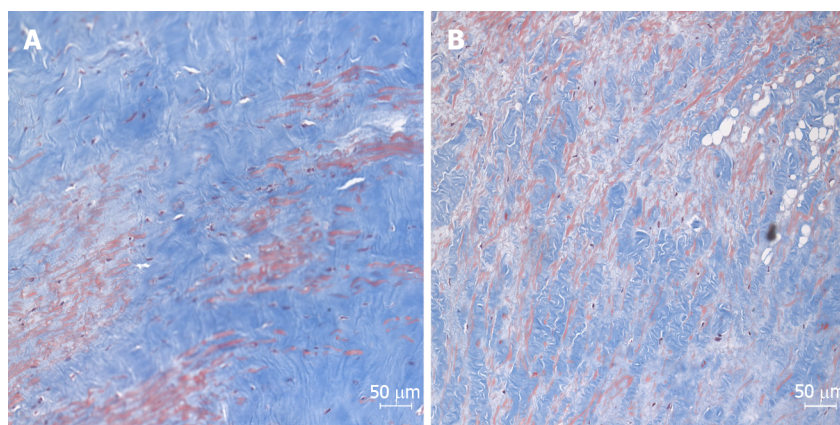


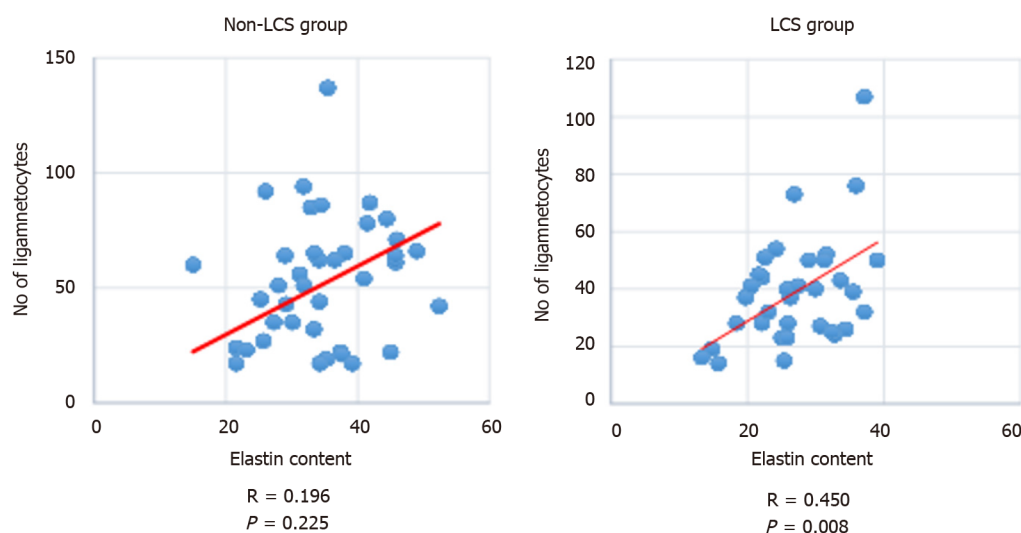
Figure 3 Von Geisson's stain showing the difference between lumbar canal stenosis & Non-lumbar canal stenosis in the number of elastic fibers. A: Lumbar canal stenosis; B: Non-lumbar canal stenosis.

which keeps it smooth. Hamdan *et al*[11] exhibited that the LCS changes were more in the central portion than in the attachments. Yabe *et al*[21] reported a severe reduction in elastic fibres on the dorsal hypertrophied LF. We did not differentiate among the sides as the stenotic effect was due to in-toto changes in LF. Moreover, the LF was removed piecemeal in most of our cases. Hence, such differences



DOI: 10.5312/wjo.v13.i9.791 Copyright ©The Author(s) 2022.

Figure 4 Masson's Trichrome stain showing the difference in the number of collagen fibers. A: Lumbar canal stenosis; B: Non-lumbar canal stenosis.



DOI: 10.5312/wjo.v13.i9.791 Copyright ©The Author(s) 2022.

Figure 5 Scatter plot showing the correlation between the elastic fibers and ligamentocyte numbers in the groups. LCS: Lumbar canal stenosis.

were not observed in the present study.

Fuertes *et al*[22] were first to comment on the anatomical and fiber arrangements on different layers. The authors realized that aging and repetitive stress altered the elastic fiber organization, causing disarray, derangement, and even complete transformation with no elastic fibers. In the present study, morphologic changes such as diameter irregularity, orientation turbulence, and extent of fragmentation were observed in elastic fibers. A similar finding was observed though electron microscopy in the study by Hulmani *et al*[4]. The image software was used to perform the morphometric study based on light microscopy. The present study did not exhibit significant ganglionic cystic changes or chondroid metaplasia despite substantial fibrosis. The present study noted a statistically significant reduction in the elastin percentage, increase in collagen, and reduction in the elastin/collagen ratio. This finding is in contrast to that of other studies. Additionally, a significant change was observed in the elastic fibers (4.37 ± 1.07 vs 3.27 ± 0.54 ; $P < 0.001$), fragmentation, and decrease in the number of ligamentocytes. Altun *et al*[2] reported that elastic fiber reduction or collagen increase was not significant, except for calcification. Okuda *et al*[10] first described elastic fiber degeneration, and it was accompanied by a proliferation of collagen fibers among elastic fibers. Additionally, Hulmani *et al*[4] reported findings such as ganglion-like cystic lesion, mucinous degeneration, and vascularization that were confirmed through electron microscopy. Reyes-Sánchez *et al*[18] exhibited more cystic degeneration, fibrillar appearance, and hypercellularity in the degenerative listhetic group than in the degenerative stenotic group. These results could be caused by instability rather than a degenerative disease in the spine. Schröder *et al*[12] noted that the parallel arrangement of LF elastic fibers was lost in degenerative LCS. This finding is concurrent with that of the present study and the study by Altun *et al*[2].

Significant fibrosis was observed in the LCS group in the present study[2,4]. This finding is in contrast to that of Hulmani *et al*[4] or Altun *et al*[2], and Cheung *et al*[23] exhibited a positive correlation between fibrosis and LFH in the LCS group. On the other hand, the developmental stenotic group exhibited paradoxically less fibrosis[23]. Okuda *et al*[10] also exhibited graded fibrosis in their LCS patients. However, the correlation with the clinical symptoms of patients was not significant. Additionally, the authors exhibited a large number of chondroid cells in patients with spondylolisthesis [10]. These findings are similar to those of Fukuyama *et al*[8], who postulated that unstable lumbar spine accelerates LF degeneration and chondrometaplasia.

LFH etiology is multifactorial. The morphological transformation that includes reduction in the elastin/collagen ratio, degeneration, fragmentation of the residual elastic fibers, fibrosis, cicatrization, and calcification leads to a loss in elasticity that can infold into the spinal canal, causing narrowing. Future studies can evaluate the correlation between symptom duration and progression of specific changes.

Several inflammatory cytokines have been studied which are responsible for the growth and reproduction and some of these plays' crucial role in inflammatory response and progressive LF fibrosis [24]. However, we have not studied any such markers.

Limitation

The present study exhibited that morphometric findings can be studied satisfactorily even in the absence of an electron microscope, which can be reproduced even in less sophisticated setups. However, it could not differentiate between the dorsal and dural aspects as we removed the LF piecemeal and not as a whole. The central and peripheral parts could not be segregated, which could allow more in-depth understanding, particularly of chondroid metaplasia. Clinical scoring, occupational activity, and MRI measurement were also ignored to keep the study simple. Additionally, gene expression was not studied in the present study.

CONCLUSION

The LCS and non-LCS groups differ in clinical parameters, mainly symptom duration. Histopathologically, the two groups exhibited significant differences in elastin degeneration, fragmentation, elastic/collagen ratio, fibrosis, and number of ligamentocytes. However, calcification was not significant between the groups.

ARTICLE HIGHLIGHTS

Research background

Ligament flavum (LF) hypertrophy is the main etiopathogenesis of lumbar canal stenosis (LCS). The purely elastic LF undergoes a morphological adaptation including a reduction in the elastic fibers and a consequent increase in the collagen content, fibrosis, cicatrization, and calcification. However, the morphometric analysis can delineate the LF in patients with LCS from those without LCS, which would help in better understanding LCS pathogenesis.

Research motivation

The research is motivated due to high footfall of these patient on Orthopedic outpatient department. An interdepartmental meeting was made to analyze these patients and funds were provided by the institute.

Research objectives

To compare the histopathological changes in LF between the degenerative LCS and non-stenotic (non-LCS) group.

Research methods

The present prospective study was conducted in 82 patients who were divided into two groups, namely LCS and non-LCS. Demographic details of the patients such as duration of symptoms, level of involvement, and number of segments were recorded. The LF obtained from both groups was histopathologically examined for the fibrosis score, elastic fiber degeneration, calcification, and chondroid metaplasia. Morphometrical details included a change in elastin and collagen percentages, elastin/collagen ratio, elastic fiber fragmentation, and ligamentocyte numbers. All parameters were compared between the two groups by using the independent *t* test, Chi-square test, and Pearson's correlation test.

Research results

Of the total, we selected 74 patients. The number of patients in the LCS and non-LCS groups was 34 and 40, respectively. The mean $\pm$ standard deviation of age of presentation in the LCS and non-LCS groups was 49.2 ± 8.9 and 43.1 ± 14.3 years, respectively. The difference in fibrosis ($P = 0.002$), elastic fiber degeneration ($P = 0.01$), elastic fragmentation percentage ($66.5\% \pm 16.3\%$ vs $29.5\% \pm 16.9\%$), elastic content percentage ($26.9\% \pm 6.7\%$ vs $34.7\% \pm 8.4\%$), collagen content percentage ($63.6\% \pm 10.4\%$ vs $54.9\% \pm 6.4\%$), reduction of elastic/collagen ratio (0.4 ± 0.1 vs 0.6 ± 0.1), and ligamentocyte number (39.1 ± 19.1 vs 53.5 ± 26.9) between the LCS and non-LCS groups was statistically significant. The difference in calcification ($P = 0.08$) and Pearson's correlation between duration and loss of elastin was statistically nonsignificant. Subgroup analysis exhibited a consistent difference in LF morphology in patients aged ≥ 40 years between the two groups. A similar finding was observed in patients aged < 40 and > 40 years in the non-LCS group.

Research conclusions

The quality change in elastin fibers and an increase in the collagen content and fibrosis cause loss of elasticity in LF, contributing to LCS pathogenesis. However, calcification did not play a significant role in LCS pathogenesis.

Research perspectives

The study compares the histopathological changes in LF between the degenerative LCS and non-stenotic (non-LCS) group.

FOOTNOTES

Author contributions: Jain M, Sable B, Tirpude AP, and Sahu RN conceived the idea; Jain M got the ethical clearance and grant for the study; Jain M, Sahu RN, and Das G collected the sample; Sable M and Tirpude AP performed the histological analysis; The data were compiled by Jain M and Samanta SK; Sable M and Jain M performed the statistical analysis; Jain M, Samanta SK, and Das G wrote the manuscript, whereas the other authors provided critical inputs; All authors have read and agree to the content of the manuscript.

Supported by Institutional research grant (AIIMS Bhubaneswar).

Institutional review board statement: The study was reviewed and approved by the Institutional ethics committee of AIIMS Bhubaneswar (T/IM/18-19/43 dated 04/01/2019).

Informed consent statement: All patients gave informed consent.

Conflict-of-interest statement: There is no conflict of interest.

Data sharing statement: Participants gave informed consent for data sharing.

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <https://creativecommons.org/licenses/by-nc/4.0/>

Country/Territory of origin: India

ORCID number: Mantu Jain 0000-0003-3848-4277; Mukund Sable 0000-0002-5364-1464; Amit Purushottam Tirpude 0000-0003-0223-2270; Rabi Narayan Sahu 0000-0003-1429-2123; Sudeep Kumar Samanta 0000-0003-3543-4485; Gurudip Das 0000-0001-7887-5526.

S-Editor: Liu JH

L-Editor: A

P-Editor: Liu JH

REFERENCES

- 1 Sairyo K, Biyani A, Goel V, Leaman D, Booth R Jr, Thomas J, Gehling D, Vishnubhotla L, Long R, Ebraheim N. Pathomechanism of ligamentum flavum hypertrophy: a multidisciplinary investigation based on clinical, biomechanical, histologic, and biologic assessments. *Spine (Phila Pa 1976)* 2005; **30**: 2649-2656 [PMID: 16319751 DOI: 10.1097/01.brs.0000188117.77657.ee]
- 2 Altun I, Yüksel KZ. Histopathological Analysis of Ligamentum Flavum in Lumbar Spinal Stenosis and Disc Herniation.

- Asian Spine J* 2017; **11**: 71-74 [PMID: 28243372 DOI: 10.4184/asj.2017.11.1.71]
- 3 **Hansson T**, Suzuki N, Hebelka H, Gaulitz A. The narrowing of the lumbar spinal canal during loaded MRI: the effects of the disc and ligamentum flavum. *Eur Spine J* 2009; **18**: 679-686 [PMID: 19277726 DOI: 10.1007/s00586-009-0919-7]
 - 4 **Hulmani D**, Garg B, Mehta N, Mridha AR, Nag TC, Farooque K. Morphological Changes in the Ligamentum Flavum in Degenerative Lumbar Canal Stenosis: A Prospective, Comparative Study. *Asian Spine J* 2020; **14**: 773-781 [PMID: 32429020 DOI: 10.31616/asj.2020.0041]
 - 5 **Giles LG**, Kaveri MJ. Some osseous and soft tissue causes of human intervertebral canal (foramen) stenosis. *J Rheumatol* 1990; **17**: 1474-1481 [PMID: 2273488]
 - 6 **Zhong ZM**, Zha DS, Xiao WD, Wu SH, Wu Q, Zhang Y, Liu FQ, Chen JT. Hypertrophy of ligamentum flavum in lumbar spine stenosis associated with the increased expression of connective tissue growth factor. *J Orthop Res* 2011; **29**: 1592-1597 [PMID: 21484860 DOI: 10.1002/jor.21431]
 - 7 **Kolte VS**, Khambatta S, Ambiyev MV. Thickness of the ligamentum flavum: correlation with age and its asymmetry-an magnetic resonance imaging study. *Asian Spine J* 2015; **9**: 245-253 [PMID: 25901237 DOI: 10.4184/asj.2015.9.2.245]
 - 8 **Fukuyama S**, Nakamura T, Ikeda T, Takagi K. The effect of mechanical stress on hypertrophy of the lumbar ligamentum flavum. *J Spinal Disord* 1995; **8**: 126-130 [PMID: 7606119]
 - 9 **Postacchini F**, Gumina S, Cinotti G, Perugia D, DeMartino C. Ligamenta flava in lumbar disc herniation and spinal stenosis. Light and electron microscopic morphology. *Spine (Phila Pa 1976)* 1994; **19**: 917-922 [PMID: 8009349 DOI: 10.1097/00007632-199404150-00009]
 - 10 **Okuda T**, Baba I, Fujimoto Y, Tanaka N, Sumida T, Manabe H, Hayashi Y, Ochi M. The pathology of ligamentum flavum in degenerative lumbar disease. *Spine (Phila Pa 1976)* 2004; **29**: 1689-1697 [PMID: 15284518 DOI: 10.1097/01.brs.0000132510.25378.8c]
 - 11 **Hamdan TA**, Jbara KK, Hatem HA. Histological Changes of Ligamenta Flava in Lumbar Disc Herniation and Spinal Canal Stenosis. *Basrah J Surg* 2005; **11**: 1-14 Available from: <https://www.semanticscholar.org/paper/HISTOLOGICAL-CHANGES-OF-LIGAMENTA-FLAVA-IN-LUMBAR-Hatem-Jbara/9df6335c9351393e307b723bf70dea3a7980d2c2>
 - 12 **Schröder PK**, Grob D, Rahn BA, Cordey J, Dvorak J. Histology of the ligamentum flavum in patients with degenerative lumbar spinal stenosis. *Eur Spine J* 1999; **8**: 323-328 [PMID: 10483836 DOI: 10.1007/s005860050181]
 - 13 **Elsberg C**. Experiences in spinal surgery: observations upon 60 Laminectomies for spinal disease. *Surg Gynecol Obs* 1913; **16**: 117-132
 - 14 **Wang B**, Gao C, Zhang P, Sun W, Zhang J, Gao J. The increased motion of lumbar induces ligamentum flavum hypertrophy in a rat model. *BMC Musculoskelet Disord* 2021; **22**: 334 [PMID: 33823825 DOI: 10.1186/s12891-021-04203-x]
 - 15 **Chuang HC**, Tsai KL, Tsai KJ, Tu TY, Shyong YJ, Jou IM, Hsu CC, Shih SS, Liu YF, Lin CL. Oxidative stress mediates age-related hypertrophy of ligamentum flavum by inducing inflammation, fibrosis, and apoptosis through activating Akt and MAPK pathways. *Aging (Albany NY)* 2020; **12**: 24168-24183 [PMID: 33223505 DOI: 10.18632/aging.104105]
 - 16 **Zaki SM**. Study of the human ligamentum flavum in old age: a histological and morphometric study. *Folia Morphol (Warsz)* 2014; **73**: 492-499 [PMID: 25448909 DOI: 10.5603/FM.2014.0074]
 - 17 **Baba H**, Maezawa Y, Furusawa N, Imura S, Tomita K. The role of calcium deposition in the ligamentum flavum causing a cauda equina syndrome and lumbar radiculopathy. *Paraplegia* 1995; **33**: 219-223 [PMID: 7609980 DOI: 10.1038/sc.1995.49]
 - 18 **Reyes-Sánchez A**, García-Ramos CL, Deras-Barrientos CM, Alpizar-Aguirre A, Rosales-Olivarez LM, Pichardo-Bahena R. Ligamentum flavum in lumbar spinal stenosis, disc herniation and degenerative spondylolisthesis. An histopathological description. *Acta Ortop Mex* 2019; **33**: 308-313 [PMID: 32253853]
 - 19 **Sato N**, Higashino K, Sakai T, Terai T, Goel VK, Biyani A, Ebraheim N, Takata Y, Hayashi F, Yamashita K, Morimoto M, Manabe H, Sairyo K. Role of Epiligament in Ligamentum Flavum Hypertrophy in Patients with Lumbar Spinal Canal Stenosis: a Pilot Study. *J Med Invest* 2018; **65**: 85-89 [PMID: 29593200 DOI: 10.2152/jmi.65.85]
 - 20 **Peng YX**, Zheng ZY, Wang Md WG, Liu L, Chen Md F, Xu Md HT, Zhang ZM. Relationship between the location of ligamentum flavum hypertrophy and its stress in finite element analysis. *Orthop Surg* 2020; **12**: 974-982 [PMID: 32489000 DOI: 10.1111/os.12675]
 - 21 **Yabe Y**, Hagiwara Y, Tsuchiya M, Honda M, Hatori K, Sonofuchi K, Kanazawa K, Koide M, Sekiguchi T, Itaya N, Itoi E. Decreased elastic fibers and increased proteoglycans in the ligamentum flavum of patients with lumbar spinal canal stenosis. *J Orthop Res* 2016; **34**: 1241-1247 [PMID: 26679090 DOI: 10.1002/jor.23130]
 - 22 **Viejo-Fuertes D**, Liguoro D, Rivel J, Midy D, Guerin J. Morphologic and histologic study of the ligamentum flavum in the thoraco-lumbar region. *Surg Radiol Anat* 1998; **20**: 171-176 [PMID: 9706675 DOI: 10.1007/BF01628891]
 - 23 **Cheung PWH**, Tam V, Leung VYL, Samartzis D, Cheung KM, Luk KD, Cheung JPY. The paradoxical relationship between ligamentum flavum hypertrophy and developmental lumbar spinal stenosis. *Scoliosis Spinal Disord* 2016; **11**: 26 [PMID: 27635416 DOI: 10.1186/s13013-016-0088-5]
 - 24 **Sun C**, Zhang H, Wang X, Liu X. Ligamentum flavum fibrosis and hypertrophy: Molecular pathways, cellular mechanisms, and future directions. *FASEB J* 2020; **34**: 9854-9868 [PMID: 32608536 DOI: 10.1096/fj.202000635R]



Published by **Baishideng Publishing Group Inc**
7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA

Telephone: +1-925-3991568

E-mail: bpgoffice@wjgnet.com

Help Desk: <https://www.f6publishing.com/helpdesk>

<https://www.wjgnet.com>

