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EDITED BY

Anamaria Brozovic,
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REVIEWED BY

Sutapa Mukherjee,
Chittaranjan National Cancer Institute,
India
Aling Shen,
Fujian University of Traditional Chinese
Medicine, China
Xiaoqun Ye,
Second Affiliated Hospital of
Nanchang University, China

*CORRESPONDENCE

Ke Xie
mei97@sina.com
Zhisen Shen
szs7216@163.com

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Overcoming therapeutic resistance to platinum-based drugs by targeting Epithelial–Mesenchymal transition

Xirui Duan¹, Maochao Luo², Jian Li³, Zhisen Shen^{4*}
and Ke Xie^{1*}

¹Department of Oncology, Sichuan Academy of Medical Sciences and Sichuan Provincial People's Hospital, School of Medicine, University of Electronic Science and Technology of China, Chengdu, China, ²State Key Laboratory of Biotherapy and Cancer Center, West China Hospital, and Collaborative Innovation Center for Biotherapy, Chengdu, China, ³West China School of Basic Medical Sciences and Forensic Medicine, Sichuan University, Chengdu, China, ⁴Department of Otorhinolaryngology and Head and Neck Surgery, The Affiliated Lihuli Hospital, Ningbo University, Ningbo, China

Platinum-based drugs (PBDs), including cisplatin, carboplatin, and oxaliplatin, have been widely used in clinical practice as mainstay treatments for various types of cancer. Although there is firm evidence of notable achievements with PBDs in the management of cancers, the acquisition of resistance to these agents is still a major challenge to efforts at cure. The introduction of the epithelial-mesenchymal transition (EMT) concept, a critical process during embryonic morphogenesis and carcinoma progression, has offered a mechanistic explanation for the phenotypic switch of cancer cells upon PBD exposure. Accumulating evidence has suggested that carcinoma cells can enter a resistant state *via* induction of the EMT. In this review, we discussed the underlying mechanism of PBD-induced EMT and the current understanding of its role in cancer drug resistance, with emphasis on how this novel knowledge can be exploited to overcome PBD resistance *via* EMT-targeted compounds, especially those under clinical trials.

KEYWORDS

platinum-based drugs, epithelial-mesenchymal transition, drug resistance, targeted therapy, cancer stem cells

Introduction

Metallodrugs play essential roles in anticancer therapy, where platinum-based drugs (PBDs) are most widely used. More than 40 years ago, the United States Food and Drug Administration (FDA) approved the platinum (Pt) compound, cisplatin, to treat ovarian cancer, bladder cancer, and metastatic testicular cancer (1). With the wide application of

PBDs, these drugs have become the first-line agents in anticancer therapy for other types of malignancies (2). Despite the prospective effect, resistance to PBDs has become the main reason for chemotherapy failure in clinical treatments.

Numerous studies have demonstrated that the epithelial-mesenchymal transition (EMT) is a primary cause of PBD resistance in cancer cells. The EMT is a process in which cells lose the epithelial marker E-cadherin and express the mesenchymal marker vimentin. It is involved in embryogenesis, tissue morphogenesis, gastrulation, and wound repair (3). Multiple studies have revealed that cancerous tissue has both epithelial and mesenchymal characteristics (4). During the EMT process, the connection between cells is weakened, cell motility is enhanced, and the synthesis of multidrug-resistant proteins is increased, which makes the tumor tissue show the characteristics of high invasiveness and metastasis and high drug resistance. Cancer stem cells (CSCs), a minor population of cancer cells, are regulated by EMT, and these cells play an important role in invasion, metastasis, drug resistance, and phenotypic cell switching (5). With the current deeper exploration of oncology and cancer-related problems, researchers have found that the EMT is closely related to drug resistance, tumor stemness and aggressiveness, and metastasis (Figure 1) (6).

Development of the EMT phenotype is one of the fundamental reasons that tumor cells transform from complete

sensitivity to extreme resistance during PBD chemotherapy. In this review, we attempt to synthetically illustrate the underlying mechanism of EMT-induced resistance to metallo drugs. In addition, we highlight novel, potentially effective strategies that combine targeted molecular inhibitors to obstruct the EMT or inhibit EMT-related cellular changes to overcome drug resistance and improve the antitumor effects of Pt compounds (Table 1 and 2).

Mechanisms involved in PBD-induced EMT

When cancer cells receive signals from tumor-associated reactive stroma (e.g., Wnt, TGF- β , and Notch signaling), the expression of EMT-inducible transcription factors is elevated and this can activate the EMT (37). When treating cancer patients with chemotherapeutic drugs, some cancer cells develop drug resistance, and the resistance of cancer cells is negatively correlated with patient survival. Moreover, recent findings have shown that drug-resistant tumor cells tend to undergo EMT (38). This section summarizes developments in the last five years regarding the signaling pathways associated with the induction of EMT by PBDs (Figure 2). Other scholars have demonstrated the effects of mitotic kinases such as Nek2 and Mps1 (TTK) on EMT, focusing on AuroraA, AuroraB, Bub1

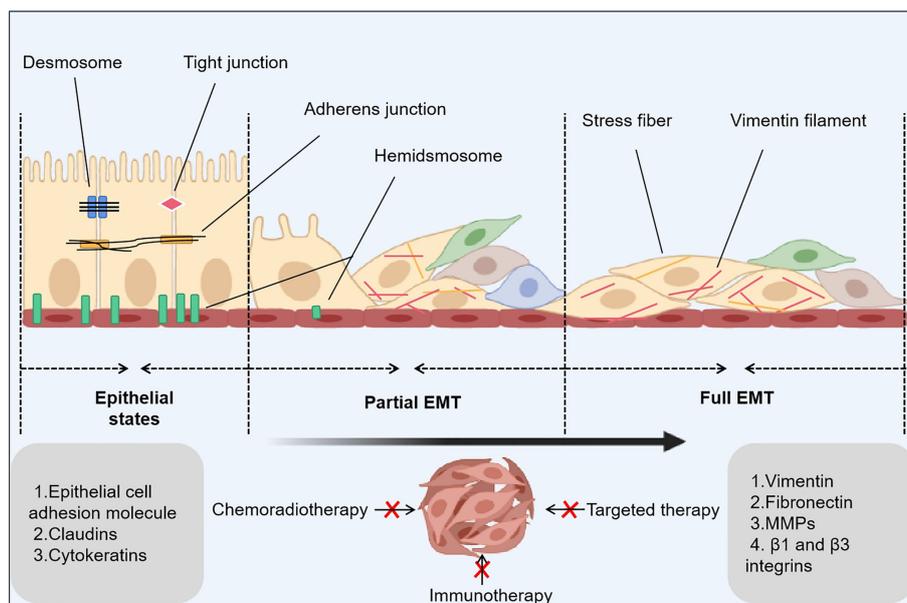


FIGURE 1 EMT-related principles underlying cancer drug resistance. The epithelial-mesenchymal transition (EMT) is a dynamic process that consists of three states: an epithelial state with epithelial phenotypes, a hybrid state with epithelial phenotypes and mesenchymal phenotypes, and a mesenchymal state with mesenchymal phenotypes. In the latter two states, a small number of cancer stem cells and drug-tolerant persister cells occur to inhibit anticancer treatments.

TABLE 1 Classes of synthetic drugs targeting epithelial-mesenchymal transition.

Drug	Mechanism of action	Clinical trial for	Ref
Thymoquinone	β -catenin inhibitor	Bladder cancer	(7)
Luteolin	Notch-1 inhibitor	Gastric cancer	(8)
Curcumin	β -catenin inhibitor	Breast cancer	(9)
	Snail inhibitor	Oral cancer	
	CXCR4 inhibitor	Colorectal cancer	
	Hedgehog inhibitor	Pancreatic cancer	
	TGF- β inhibitor	Hepatoma	
		retinal pigment epithelial cancer	
	ERK inhibitor	Hepatocellular carcinoma	
	NF- κ B inhibitor	Breast cancer	
	STAT3 inhibitor	Melanoma	
	ERK1/2 inhibitor	Bladder cancer	
Glucosamine	Smad2/3 inhibitor	Thyroid cancer	(10)
	NF- κ B inhibitor	Breast cancer	
Ginsenosides	NF- κ B inhibitor	Breast cancer	(11)
Combretastatin	Akt inhibitor	Thyroid papillary carcinoma	(12)
Oxyresveratrol	Snail inhibitor	Colon cancer	(13)
N-arachidonoyl dopamine	ERK1/2 inhibitor	Breast cancer	(14)
Kaempferol	Snail inhibitor	NSCLC	(15)
		Breast cancer	
Doxycycline	Snail1 inhibitor	Breast cancer	(16)
	twist1 inhibitor		
	ZEB1 inhibitor		
	Slug inhibitor		
Resveratrol	Hedgehog inhibitor	Gastric Cancer	(17)
Genistein	Twist1 inhibitor		
Pantoprazole	β -catenin inhibitor		
Astragaloside	TGF- β 1 inhibitor		
Kaempferol	Akt inhibitor	Cervical cancer	(18)
Niclosamide	β -catenin inhibitor	Colon cancer	(19)
Lipoic acid	TGF- β inhibitor	Breast cancer	(20)
Piperlongumine	Snail1 inhibitor	Epithelial cancer	(21)
	Twist1 inhibitor		
Manganese-12 acetate	β -catenin inhibitor	Breast cancer	(22)
	Akt inhibitor		
Shikonin	Akt inhibitor	Glioblastoma	(23)
Actinomycin V	Snail inhibitor	Breast cancer	(24)
	Slug inhibitor		
Hydroxytyrosol	β -catenin inhibitor	Breast cancer	(25)
	Smad2/3 inhibitor		
Antrodia camphorata	Akt inhibitor	Ovarian cancer	(26)
	β -catenin inhibitor	Colon cancer	
Quercetin	β -catenin inhibitor	Colon cancer	(27)

TABLE 2 Classes of natural drugs targeting epithelial-mesenchymal transition.

Drug	Mechanism of action	Clinical trial for	Ref
Sulforaphane	ERK5 activator	Lung cancer	(28)
Nitidine chloride	JAK2/STAT3 inhibitor	Glioblastoma	(29)
Eribulin	TGF-β/Smad2/3 inhibitor	Triple-negative breast cancer	(30)
Ursolic acid	TGF-β1/smad3 inhibitor	Renal fibrosis	(31)
Curcumin	TGF-β/Smad inhibitor	Hepatic fibrosis	(32)
Celastrol	TGF-β1/smad2/3 inhibitor	Pulmonary Fibrosis	(33)
Baicalin	ERK inhibitor	Hepatocellular Carcinoma	(34)
Isoviolanthin	TGF-β/Smad inhibitor	Hepatocellular Carcinoma	(35)
Astragaloside IV	Akt/GSK-3β/β-catenin inhibitor	Hepatocellular Carcinoma	(36)

and Hec1 (high expression in cancer) as potential targets for cancer therapeutic intervention through their impact on EMT. The established relationships and interactions between these and other mitotic kinases are highlighted, together with the impact of classical signaling pathways and long RNAs on EMT. Recent studies have found that microbial metabolism can increase the intake of 5-fluorouracil (5FU) to counteract the drug resistance

in CRC by modulating the FoxO3-FOXM1 axis to alter drug transporter sensitivity. The researchers also found that this process involved changes in the expression of EMT mediators (39). There have been few studies on the relationship between EMT and drug transporters, and regulating drug transporters (drug import/export pumps) is expected to become a new approach and research direction for anti-EMT treatment.

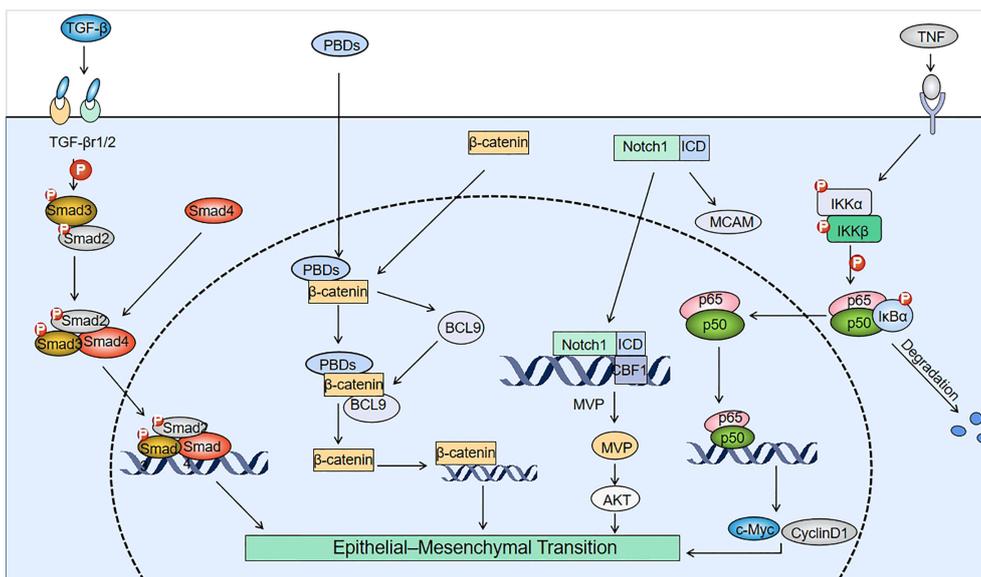


FIGURE 2 Mechanisms involved in PBD-induced EMT. TGF-β interacts with TGF-β receptors (TGF-βr1/2) to induce phosphorylation of Smad2/3, which binds to Smad4 to form a Smad2/3/4 complex. After platinum-based drugs (PBDs) enter tumor cells, they bind to the N-terminus of β-catenin and upregulate BCL9. BCL9 can form a complex with β-catenin to create positive feedback that promotes β-catenin expression, and induces the epithelial-mesenchymal transition (EMT). The pathways for Notch1 to induce EMT include: promoting melanoma cell adhesion molecule (MCAM) expression and binding of the Notch1 internal domain (ICD) to the CBF1 site of the major vault protein (MVP) gene to upregulate MVP. The binding of tumor necrosis factor (TNF) to membrane surface receptors drives IκB kinase (IKK) to phosphorylate IκB. Degradation of IκB forms the p53/p50 dimeric complex that binds to EMT-related genes.

Wnt

The Wnt signaling pathway is a complex regulatory network that includes the Wnt/ β -catenin signaling pathway, the Wnt/PCP pathway (planner-cell polarity pathway), and the Wnt/Ca²⁺ pathway (40). PBDs induce EMT mainly by activating the classical Wnt signaling pathway. The expression of c-MYB, a proto-oncogene, is increased in drug-resistant cells, which directly induces EMT by promoting miR-21 expression to increase the mesenchymal marker zinc finger E-box binding homeobox 1 (ZEB1) and decrease E-cadherin expression. By activating the Wnt pathway, c-MYB can also inhibit β -catenin phosphorylation and thus indirectly induce EMT (41). It has been found that in non-small cell lung cancer (NSCLC), PBDs can increase intracellular B-cell CLL/lymphoma 9 (BCL9) expression and form the β -catenin/BCL9 complex by selectively binding to the N-terminal structural domain of β -catenin. This binding enhances the transcriptional activity of Wnt signaling and promotes the transcription of β -catenin, thus increasing nuclear translocation and inducing EMT (42). It has also been shown that overexpression of BMAL1 (the central positive loop element that initiates circadian oscillations) and the PDZ-binding motif (TAZ) can induce the EMT by promoting nuclear expression of β -catenin in chemo-resistant colorectal cancer (CRC) cells (38, 43). Ectopic expression of disheveled (DVL) was effective in activating the Wnt/ β -catenin signaling pathway even in the absence of Wnt ligand stimulation. Indeed, the forkhead box M1 (FOXO1) induction of Snail expression, metastasis, and chemoresistance requires DVL2, whereas FOXO1 does not alter DVL2 expression (44).

In addition, miR-28-5p expression was also decreased in drug-resistant CRC cells, thereby attenuating the inhibitory effect on downstream structure-specific recognition protein-1 (SSRP1) and increasing its expression to promote EMT by activating the Wnt/ β -catenin pathway (45). The expression of miR-302 was reduced in drug-resistant cells, which directly targeted and upregulated ATPase family AAA domain containing 2 (ATAD2). This alteration significantly downregulated the tumor suppressor APC and upregulated nuclear-catenin *via* activation of the Wnt/ β -catenin signaling pathway. The result was a decrease in expression of E-cadherin and increase in expression of vimentin (46). In drug-resistant NSCLC cells, miR-192 and miR-662 were overexpressed. MiR-192 may activate the Wnt pathway by silencing its atypical inhibitors (e.g., NOTUM) and upregulating FOXO1 expression. In parallel, miR-662 may enhance Wnt by upregulating the forkhead box N3 (FOXN3), forkhead box protein 1 (FOXP1), and receptor tyrosine kinase-like orphan receptor 1 (ROR1), and glioma-associated oncogene protein-2 (GLI2) signaling pathways and thus promote EMT. Similarly, miR-192 and miR-662 could upregulate the expression of the lncRNA, CASC9, and promote EMT in gastric cancer cells (47). In addition, BCL9 expression was also increased in drug-resistant

NSCLC cells, activating the EMT mechanism by stimulating nuclear translocation of β -catenin (48).

TGF- β

Transforming growth factor- β (TGF- β) is a multifunctional cytokine in mammals consisting of three isoforms, TGF- β 1, TGF- β 2, and TGF- β 3. TGF- β binds to the TGF- β receptor type 1 complex (TGF β -R1 and TGF β -R2), leading to phosphorylation of the Smad2/3 protein in the cytoplasm. Smad2/3 interacts with Smad4 to form the Smad2/3/4 complex, which is translocated to the nucleus where it activates transcription of TGF- β -responsive downstream genes. This leads to activation of the signaling cascade that induces EMT (49). Cisplatin has been shown to increase IL-6 release and TGF- β expression in cancer-associated fibroblasts. IL-6 blocked apoptosis during inflammation, which could also protect cancer cells from apoptosis and chemotherapeutic agents. In addition, IL-6 enhanced the TGF- β -induced EMT in NSCLC. MiRNA-17, 20a, and 20b were expressed at low levels in cancer cells, and could also activate the TGF- β pathway (50). In the following section, we will describe the detailed mechanism of platinum-induced activation of the TGF- β signaling pathway.

Following cisplatin treatment of lung cancer cells, the C-X-C motif chemokine ligands/CXC chemokine receptor (CXCLs/CXCR2) signaling pathway was altered in resistant cells, resulting in increased expression of CXCR2-associated chemokines (CXCL1, CXCL2, and CXCL5), increased neutrophil infiltration and concomitant upregulation of the immunosuppressive factors, TGF- β and arginase (Arg-1). These factors induced neutrophil polarization toward the N2 type, which has a significantly reduced tumor-killing capacity. These cells then produced more TGF- β and Arg-1, suppressing the antitumor immune response, and thereby inducing EMT and promoting tumor progression (51). In drug-resistant SCLC cells, epithelial splicing regulatory protein 1 (ESRP1) expression was significantly downregulated, which increased the proportion of full-length CARM1 (CARM1FL) by regulating the selective splicing of coactivator associated arginine methyltransferase 1 (CARM1). This inhibited arginine methylation of Smad7, activated the TGF- β /Smad pathway, and increased Smad3 phosphorylation that promoted the EMT (52). Increased expression of CD24 in drug-resistant ovarian cancer cells led to enhanced TGF- β signaling. This TGF- β signaling cascade enabled the activation of downstream PI3k/Akt and MAPK signaling pathways, which further increased Snail expression and decreased E-cadherin expression, leading to the development of EMT (53). In patients with nasopharyngeal carcinoma who were treated with cisplatin, overexpression of miR-449b led to the degradation of TGF- β 1 mRNA, resulting in

a decrease in TGF- β 1 expression. This led to downregulation of miR-34c, which directly induced SRY-box transcription factor 2 (SOX2) upregulation and promoted EMT (54). Bone morphogenetic protein 4 (BMP4), a member of the TGF- β superfamily, was overexpressed in drug-resistant hepatocellular carcinoma (HCC). This significantly reduced the expression of Bax proapoptotic protein and caspase-3 and significantly enhanced the expression of Bcl-2 antiapoptotic protein. BMP4 induced cyclin B1 and cyclin-dependent kinase 1 (CDK1) expression in HCC cells, thereby promoting cell cycle progression from G2 to M and resisting the oxaliplatin-derived G2/M blockade. BMP4 also promoted EMT by increasing the activity of matrix metalloproteinases (MMPs), upregulating vimentin expression, and downregulating E-cadherin (55). In drug-resistant oral squamous cell carcinoma (OSCC), circANKS1B was overexpressed and attenuated the direct inhibitory effect of miR-515-5p on TGF-1 by sequestering miR-515-5p, leading to activation of the TGF- β pathway. TGF- β pathway activation increased N-cadherin expression and decreased E-cadherin expression, resulting in the development of EMT (56). Notably, esophageal adenocarcinoma (EAC) cells were capable of producing and secreting large amounts of active TGF- β under the high therapeutic stress of chemotherapy, thereby inducing EMT. The reversal of EMT in EAC cells could not be achieved by short-term drug abstinence, suggesting that the drug dormancy period (i.e., intermittent radiotherapy) was insufficient to prevent radiotherapy-induced EMT. The addition of the TGF-neutralizing antibody fresolimumab during the second week of radiotherapy, however, was able to block the EMT in EAC cells and improve therapeutic efficacy (57).

Notch

The NOTCH pathway has been highly conserved during evolution and is involved in controlling cell proliferation and inhibiting apoptosis (58). In mammals, the NOTCH pathway has four receptors (NOTCH1, 2, 3, and 4) and five ligands (JAG1 and 2, DLL1, 3, and 4), all of which are type 2 transmembrane proteins (59). Following receptor-ligand binding in the NOTCH pathway, the γ -secretase complex releases the intracellular structural domain of the NOTCH receptor, which translocates into the nucleus and induces expression of target genes, such as the HES/HEY family (60). A large body of evidence suggests that the NOTCH pathway is involved in the induction of EMT in normal and tumor tissues. In drug-resistant head and neck squamous cell carcinoma (HNSCC), increased NOTCH4 expression and resulted in specific upregulation of the downstream gene Hes-related family basic helix-loop-helix transcription factor with YRPW motif 1 (HEY1) expression, without affecting other NOTCH downstream genes. This resulted in reduced E-cadherin expression and increased

expression of vimentin, fibronectin, Twist-related protein 1 (TWIST1), and SOX2, which induced EMT. HEY1 inhibition also inversely reduced NOTCH4 expression in HNSCC (60). In CR cells, TAZ overexpression increased the activation of the downstream signaling molecule Notch1 (38). In addition, Notch1 expression was increased in resistant cells after carboplatin treatment of triple-negative breast cancer (TNBC). On the one hand, Notch1 could promote melanoma cell adhesion molecule (MCAM) expression through direct activation of the MCAM promoter, thereby inducing EMT (61). On the other hand, the Notch1 intracellular structural domain (ICD) bound to the CBF1 binding site on the MVP (major vault protein) promoter, thereby upregulating the expression of MVP and activating the AKT pathway to promote EMT (62). In addition, in high-grade serous ovarian cancer, the Notch3 pathway was activated, with increased expression of downstream SUSD2. This, in turn, induced the EMT by reducing E-cadherin expression through increased epithelial cell adhesion molecule (EpCAM) expression (63). Low concentrations of cisplatin (DDP, CDDP) could induce EMT in osteosarcoma cells *via* the Notch signaling pathway by promoting the expression of NOTCH2 and its target gene HEY1 (64).

NF- κ B

NF- κ B proteins commonly form homo/heterodimers with p65 and p50 and are inactivated in the cytoplasm by binding to the inhibitory protein I κ B to form a trimeric complex (65). When the upstream signaling factor, tumor necrosis factor (TNF), binds to its cell membrane surface receptor, the receptor conformation changes and signals IKK (I κ B kinase) kinase to phosphorylate I κ B and dissociate it from the trimer. The NF- κ B dimer then exposes the nuclear localization sequence (NLS) and rapidly moves from the cytoplasm into the nucleus, where it binds to specific sequences on the nuclear DNA and promotes the transcription of related genes, such as cyclinD1, c-Myc, matrix metalloproteinase-9 (MMP-9), and vascular endothelial growth factor (VEGF), which are EMT-related proteins (66). Studies have shown that aberrant activation of NF- κ B could induce the development of EMT in drug-resistant cells. The expression of EGFR was increased in drug-resistant head and neck squamous cell carcinoma (HNSCC), resulting in increased IKK β expression and activation of the downstream NF- κ B signaling pathway. NF- κ B increased downstream signal transducer and activator of transcription 3 (STAT3) by promoting IL-6 expression, which induced overexpression of N-cadherin and decreased E-cadherin expression, promoting EMT (67). In CRC cells, increased DJ-1 expression promoted EMT by activating the NF- κ B/Snail signaling pathway and increasing Snail protein expression (48). In triple-negative breast cancer (TNBC) cells,

cisplatin-induced activation of the ERK1/2-p90RSK signaling pathway led to NF- κ B activation, which promoted the expression of Snail Twist and ZEB-1 and induced EMT (68).

Hedgehog

The hedgehog signaling pathway is important for cellular self-renewal, tissue maintenance, and cell regeneration. The GLI family is a transcription factor for hedgehog signaling that includes glioma-associated oncogene homolog 1 (GLI1), GLI2, and GLI3. GLI1 usually acts as a potent target activator, while GLI2 and 3 have dual functions as repressors or activators depending on the situation (69). In cisplatin-resistant breast cancer cells, elevated expression of ubiquitin-specific peptidase 37 (USP37) upregulated the expression of purmorphamine (PM). Expression of the Hh targets (Smo and gli1) and the cell proliferation marker Ki-67 were also elevated through the hedgehog signaling pathway, resulting in significant upregulation of Snail1, N-cadherin, and vimentin, downregulation of E-cadherin, and induction of EMT (70). In oral squamous cell carcinoma (OSCC), drug-resistant cells enhanced CSC-associated features and increased CD10 expression, resulting in increased expression of Smo and gli1, which activated the hedgehog signaling pathway and induced EMT (71). In addition, cisplatin induced the expression of prostaglandin E2 (PGE2) in osteoclast cells, which promoted GLI1 expression by activating integrin β 1. GLI1, in turn, promoted expression of B cell-specific Moloney murine leukemia virus integration site 1 (BMI1) by activating the hedgehog signaling pathway, thereby altering the microenvironment and inducing CSC-like features that promoted EMT (69).

PI3K/AKT/mTOR

PI3K (phosphatidylinositol kinase) is a dimer consisting of the regulatory subunit p85 and the catalytic subunit p110. When it binds to growth factor receptors (EGFR), it alters the protein structure of Akt and activates it, promoting or inhibiting the activity of a range of downstream substrates such as the apoptosis-associated proteins Bad and caspase9 by phosphorylation, thereby regulating cell proliferation, differentiation, apoptosis and migration phenotypes. In addition, Akt also activated IKK, which interacted with the NF- κ B pathway. In this section, we describe the molecular changes that activate the PI3K/AKT signaling pathway in drug-resistant cells from different cancers.

After cisplatin treatment of non-small cell lung cancer (NSCLC), the expression of FOXC2 was increased, inactivating the pro-apoptotic factor GSK-3 β by activating AKT. This leads

to stabilization and nuclear localization of Snail, increasing the expression of the mesenchymal markers vimentin and N-cadherin, and decreasing the expression of the epithelial marker E-cadherin, thereby promoting EMT (48, 72). Increased expression of *FAM83D*, a mitosis-related gene, activated the AKT/mTOR signaling pathway, resulting in increased phosphorylation of p70s6k, downregulation of the epithelial markers E-cadherin and α -catenin, and upregulation of the mesenchymal markers N-cadherin and vimentin, thereby inducing EMT (73). Increased expression of *PAX6* activated the downstream PI3K/AKT signaling pathway by directly binding to the promoter region of *ZEB2* and upregulating its expression (74). Similarly, increased expression of the lncRNA, *UCA1* induced EMT by activating the AKT/mTOR pathway (75).

In nasopharyngeal carcinoma cells, cisplatin treatment caused specific overexpression of miR-205-5p in drug-resistant cells, which promoted NPC cell proliferation and decreased PTEN (a classical tumor suppressor) expression by targeting it through phosphorylation of the PI3K/AKT signaling pathway. Low expression of PTEN induced the absence or reduction of epithelial markers (E-cadherin) and increased expression of mesenchymal markers (vimentin and N-cadherin) through the regulation of Snail/Slug, thereby promoting EMT and increasing the migration and invasive capacity of NPC cells. MiR-205-5p promoted cell metastasis by upregulating MMP-9 and MMP-2, which caused ECM degradation (76). Furthermore, increased expression of miR-BART22 increased MYH9 expression through activation of the PI3K/AKT/c-Jun axis. MYH9 then induced GSK-3 β degradation and mediated nuclear translocation of β -catenin, thereby stimulating the EMT machinery (48). The PI3K/Akt pathway was activated in drug-resistant gastric cancer cells, leading to an increase in downstream Rac1 activity, which further activated the JNK pathway and promoted self-renewal of GA CSCs and EMT (EMT may be a mechanism for generating CSCs). At the same time, Rac1 led to increased expression of N-cadherin and Slug. In GA cells, inhibition of any component of the PI3K/Akt-Rac1/JNK axis could block expression of the EMT transcription factor, Slug, and inhibit EMT (77).

Long non-coding RNAs

Long non-coding RNAs (lncRNAs) are defined as RNA transcripts longer than 200 nucleotides that do not encode polypeptides. Studies have shown that lncRNAs and miRNAs play a role in promoting the continuing proliferation of tumor cells and drug resistance (78); therefore, they are considered key regulators of cancer pathways and are widely studied as biomarkers of disease (79). One of the important aspects is the impact of miRNA and miRNA-lncRNA competition on tumor EMT progression.

MiR-574-3p has a tumor-suppressive effect on gastric cancer cells. In cisplatin-resistant cancer cells, miR-574-3p expression decreased and increased the expression of ZEB1 protein by attenuating the direct inhibition of ZEB1 protein, which in turn inhibited the transcription of E-cadherin, reduced its expression, increased the expression of vimentin, and induced EMT (80). In addition, miR-492 overexpression promoted CSCs by directly targeting the 3' UTR of DNMT3B and inhibiting its expression, thereby inducing EMT (81). MiR-17 expression was also increased, which promoted EMT by downregulating DEDD (82). In ovarian cancer (OC) cells, decreased expression of miR-1294 allowed an increase in IGF1-R expression, which induced the EMT (48). The expression of miR-363 was also decreased, promoting EMT by attenuating the direct inhibitory effect on Snail (83, 84). In contrast, the expression of miR-149-3p was increased, promoting EMT by downregulating TIMP2 and CDKN1A expression (85). Notably, in OC cells, there is a miR-374b-5p-FOXP1 feedback loop in which miR-374b-5p can negatively regulate FOXP1 by binding to the 3'UTR of FOXP1, which can also repress miR-374b-5p transcription. In poor prognosis cells, the expression of miR-374b-5p was downregulated, while FOXP1 expression was upregulated, thereby inducing EMT (86). The competition between miRNA and lncRNA is also closely related to EMT. lncRNA-ATB is a ceRNA that is a regulator of the TGF β -ZEB1/ZEB2 axis. As a transcriptional target of TGF β , lncRNA-ATB competes with the miR-200 family, which eliminates EMT, resulting in up-regulation of ZEB1/ZEB2 expression. Our results are similar to those in other studies. lncRNA ZFAS1 was amplified at the genomic DNA level and competitively bound to miR-150, inhibiting ZEB1, MMP14 and MMP16 gene expression (87). STAT3-activated lncRNA HOXD-AS1 inhibited SOX4 mRNA expression by competing with miR-130a-3p to promote HCC lymph node metastasis (88).

EMT-mediated chemotherapeutic resistance

EMT is characterized by the loss of tightly knit epithelial cells and transdifferentiation into mesenchymal cells. In recent years, meta-analysis of existing research data about the connection between gene expression and curative cancer effects showed that EMT-related genes were closely linked to resistance to certain kinds of treatment. For instance, an analysis of data from the clinical samples of breast cancer patients demonstrated that resistance to chemotherapy was an important cause of gene regulation of stromal cells, and the expression of these genes was mediated by EMT program activation (89). In addition, seventy-six EMT marker genes were used to explore gene expression and the effect of PI3K

inhibitors in non-small cell lung carcinoma (NSCLC), and to predict resistance to these agents and assess prognosis (90). Gene changes can induce the generation of specific cell types as well as changes in phenotype, EMT-related cells, such as partial EMT cells (named hybrid EMT cells), drug-tolerant persister cells, and CSCs, which have a crucial function in creating drug resistance (Table 3). Next, we will look at how these cells are related to EMT and how to deal with PBD resistance (Figure 3).

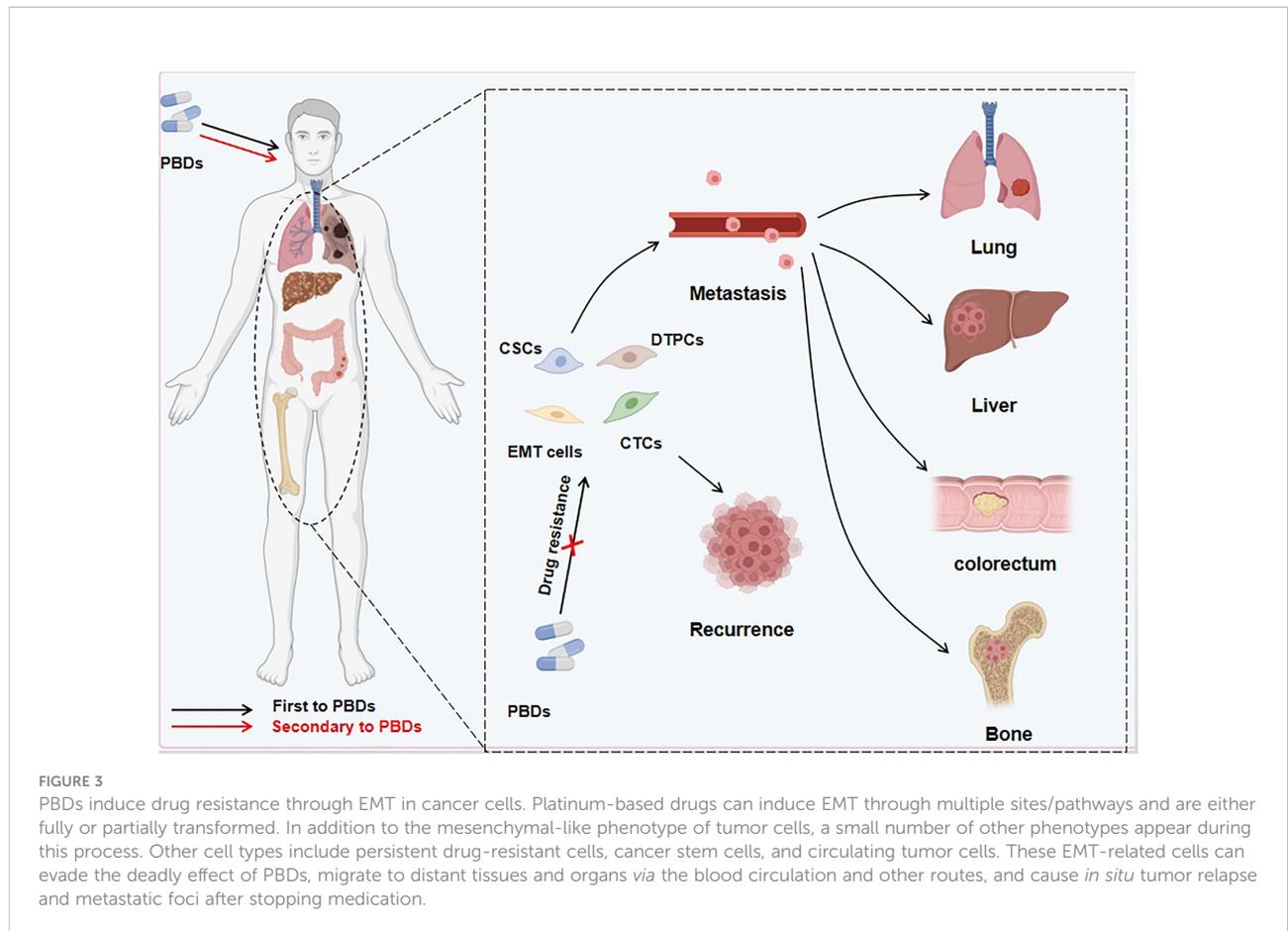
Partial EMT cells

Under normal physiological conditions, the EMT program is involved in embryonic development and wound repair, but tumor cells often enhance their invasive and metastatic capabilities by activating EMT, and especially to mount a resistance to chemotherapy drugs that ensures their survival (108). The EMT program involves the loss of junctional tightness between epithelial cells and their gradual differentiation into mesenchymal cells. Because mesenchymal cells have motility and are capable of degrading extracellular matrix (ECM) proteins, these cells can invade the surrounding tissues and metastasize to distal sites (109). Therefore, EMT is considered a sign that the tumor is progressing to a higher stage. Researchers have found that there is an intermediate state in the EMT process, in which cells have both epithelial and mesenchymal phenotypes. These are called partial EMT cells, and experimental and clinical results have shown that such cells are more malignant and drug-resistant than completely differentiated EMT cells (110).

Analysis of clinical data showed that most cancer rarely undergoes adequate transformation (3, 111). Non-small-cell lung cancers (NSCLC) patients were usually treated with gefitinib and erlotinib, which are epidermal growth factor receptor (EGFR) and tyrosine kinase inhibitors (112). However, EGFR resistance was usually accompanied by a partial EMT phenotype with co-expression of vimentin and E-cadherin. Similar evidence emerged from another clinical trial, where the epithelial cell subset and the mesenchymal cell subset were more sensitive to cisplatin, paclitaxel, and salinomycin than partial EMT cells (high CD44, high CD24, low EpCAM) in oral squamous cell carcinoma (OSCC) (113). These results indicate that, to some extent, partial EMT cells may have a greater chance of developing drug resistance. To verify the correctness of this view that partial EMT cells are more likely to develop drug resistance than completed EMT cells, we selected docetaxel-resistant PC-3 with complete EMT and DU145 with partial EMT cell lines for comparative experiments. PC-3 appeared to undergo apoptotic death and DU145 was resistant to treatment with salinomycin (114). Similarly, this hypothesis was validated by clinical results in which cancer cells altered their metabolic program to meet changing nutrition and energy requirements. This metabolic change began when tumor cells

TABLE 3 Phenotypic differences of CSCs, DTP cells and pEMT cells.

Type of cell	Markers	Type of cancer	Ref
CSCs	CD44 and CD133	Breast cancer	(91)
	CD44	Gastric cancer	(92)
	CD51	Prostate cancer	(93)
	CD133	Lung cancer	(94)
	CD133, nestin, and A2B5	Glioblastoma	(95)
	CD34	AML	(96)
DTP cells	ABC B5	Melanoma	(97)
	CD133		(98)
	CD133	Gastric cancer	(99)
	CD271	Osteosarcoma	(100)
	ALDHs	Brain cancer	(101)
pEMT cells	L1CAM	Colorectal cancer	(102)
	CD106, CD61 and CD51	Squamous cell carcinoma	(103)
	S100A6	Breast cancer	(104)
	NRF2	Non-small-cell lung carcinoma, Bladder cancer	(105)
	VEGF	Esophageal squamous cell carcinoma	(106)
	Cathepsin B	Salivary adenoid cystic carcinoma	(107)



started the EMT program and was most robust in the partial EMT state. Autophagy is divided into protective autophagy and lethal autophagy, and the condition is enhanced in the partial EMT state. During chemotherapy or the loss of nutrients or growth factors, the protective autophagy activity in tumor cells was enhanced by AMPK expression to ensure lysosomal function (115, 116). This conclusion was confirmed in clinical studies as well as by *in vitro* research. For example, inhibiting protective autophagy promoted the release of arginase 1 (Arg1) from the liver, which promoted arginine synthesis and stimulated cancer cell growth (117). In clinical practice, enhanced autophagy promoted the occurrence and development of autochthonous pancreatic cancer (118). In addition, other experiments have shown that the presence of an epithelial cell adhesion molecule (EpCAM) on the surface of epithelial cells could induce the production of multidrug-resistant proteins (MDRPs) (119); however, cells that have completed the EMT lose this phenotype, leading to a reduction in the production of MDRPs. Although we know that partial EMT plays an important role in tumor resistance, an interesting phenomenon provides us with new ideas about the function of partial EMT. In the whole EMT lineage, partial EMT cells located at intermediate sites showed greater plasticity than fully transitioned EMT cells (120). We hypothesized that different degrees of EMT and different physical and chemical properties caused by differences in phenotype and morphological structure had different degrees of influence on tumor invasiveness, metastasis, and drug resistance.

Cancer stem cells

CSCs have been considered an important factor in the sensitivity and tolerance to chemotherapy. Clinical practice demonstrated that CSCs existed in various tumor tissues, and the proportion of CSCs was significantly higher than that of the tumor before chemotherapy (121). However, the mechanism of how CSCs are produced is still unclear. With the introduction of the EMT, it is easy to determine whether CSCs and EMT are always found together in pathological tissue and to study the interaction between EMT and CSCs, because EMT induces and maintains CSCs and CSC phenotypes.

One of the possible explanations for the secreted proteins of carcinoma cells is that EMT leads to autocrine signaling loops. Multiple studies have revealed that autocrine signaling loops had the necessary activity to induce and maintain the characteristics of stem cells. EMT activation promoted the work of autocrine signaling loops to maintain the stem cell phenotype *via* the TGF-Smad and Wnt-catenin pathways in the secretome, which was secreted by transformed HMLER human mammary epithelial cells (122). Non-CSCs also had the plasticity necessary to promote cancer cell dedifferentiation, and this process depended on the

occurrence of EMT (123). For example, some secreted extracellular vesicles and macromolecules such as microRNAs and cytokines, were released by EMT to induce differentiated tumor cells to dedifferentiate into CSCs (124). Intracellular signaling pathways and cytokines participated in the EMT process to generate the CSC phenotype. The tumor-suppressor protein, p53, is restrained by the expression of Snai1, which is linked to the tumorigenic ability of cancer cells *via* the Snai1-histone deacetylase 1 (HDAC1)-p53 complex. However, this ability could be uncovered by the downregulation of Tp53, which encoded p53 (125). EMT was first discovered during embryonic development and required TGF- β pathway activation (126). Researchers also found that TGF- β , IL-6, and hepatocyte growth factor (HGF) could induce the EMT process and enhance the conversion of the differentiated cell phenotype to CSC characteristics (127, 128). In addition, it has been found that excessive EMT activation led to maximal tumor-initiating ability and chemotherapy resistance (129). A partial EMT program may contribute to the frequent and substantial production of CSCs. In a mouse model of breast cancer, CSCs had different phenotypes, called epithelial-like CSCs and mesenchymal-like CSCs. These cells had a greater ability to seed cancer cells to distant tissues (130, 131). Moreover, the combination of epithelial and mesenchymal phenotypes of CSCs enhanced the progression of tumor drug resistance and metastasis (132). Although the relationship between EMT and CSCs provided a partial explanation, the hypothesis that the partial EMT could acquire more stem phenotypes than the complete EMT, and induce CSCs that were related to specific cell types and epigenetic abnormalities deserves further exploration.

CSCs have some physical and biochemical properties that influence sensitivity to clinical treatments, including chemotherapy, radiotherapy, molecular targeting, and immunotherapy. The first important feature is the ability to self-update, which optimizes tumorigenesis and differentiation in multiple types of cells, including drug-resistant cells. Recent research discovered that many signaling pathways, such as Wnt and Notch, engaged in self-renewal activation. For example, in breast cancer (133), PBDs induced EMT *via* the Notch pathway, and RO4929097, a Notch1 inhibitor, could repress N-cadherin and CD44 to produce better patient outcomes (134). In the tumor microenvironment (TME) after Notch pathway activation, the CSCs were 'self-updated' compared to the non-EMT environment. Signaling through Notch and PI3K/AKT/mTOR stimulated quiescent CSCs to progress into the S-phase of the cell cycle (135, 136) and tumor tissue exhibited higher resistance to radiotherapy and chemotherapy in the presence of active CSCs. The body of preclinical and clinical observations shows that radio-chemotherapy effectively eliminated most non-CSC cells but did not appreciably reduce the number of CSCs (137–139), which was why CSCs could continue to enter the G0 phase and remain there for a long period. These dormant cancer cells were not affected by the drugs, and CSC resistance to

targeted treatment and immunotherapy was significantly higher than that of non-CSCs (140).

Drug-tolerant persister cells

Research has determined that the entry into drug-tolerant persister (DTP) states of cancer cells is an adaptive response to chemotherapy with PBDs (141). The emergence of this state is currently subject to several hypotheses, including that DTP cancer cells are already present in tumor tissue and are selected for proliferation after drug treatment (142). Another likely reason is that the production of DTP cancer cells is induced by activation of cell reprogramming rather than through survival of the fittest by drug selection (143). This transformation process has been widely observed in clinical research and is most commonly considered to be EMT. For example, cancer tissues had abundant DTP cancer cells when treatment activated EMT (144). Therefore, the close relationship between DTP cancer cells and EMT was an essential factor in the multidrug resistance of these cells to tumor therapy.

DTP cancer cells have some characteristic phenotypes that help them evade the killing effects of drugs, such as reversible biological capacity, slow cell cycling, and proliferative activity (143, 145, 146). For example, a slowly cycling cell population that expressed high levels of the histone H3K4 demethylase, JARID1B, was preserved after chemotherapy in melanoma (147). In addition, several researchers reported that cancer cells with DTPs exhibited a stem-like phenotype to reduce drug influence; thus, EMT programs contributed to inducing and maintaining the CSC phenotype. This DTP phenotype allowed cancer cells to acquire plasticity, pluripotency, self-renewal capacity, and transferability, and to enter the G0 phase or slow-cycling state, which protected them from cytotoxic agents (148, 149). In addition, the activation of the Wnt/ β -catenin pathway was found in the DTP cancer cells (150). These results also indirectly proved that the emergence of DTPs was closely related to the EMT process. Osimertinib, an EGFR tyrosine kinase inhibitor, upregulated the AXL gene-inducing EMT program. AXL bound to its ligand (GAS6) and promoted formation of DTP cancer cells by increasing the expression of DNA repair proteins to reduce DNA damage (151, 152). Mesenchymal EGFR mutant cancers survived initial EGFR inhibitors because fibroblast growth factor receptor 1 (FGFR1) was expressed on the cell surface (153). MiR-99b was activated when tumor cells underwent EMT reprogramming, and as FGFR3 was a target of miR-99b, it participated in the induction of DTP cancer cells (153, 154). In addition to FGFR1 and FGFR3, studies have shown that insulin-like growth factor 1 (IGF-1) and the lipid hydroperoxidase, GPX4, could activate the PI3K/AKT and

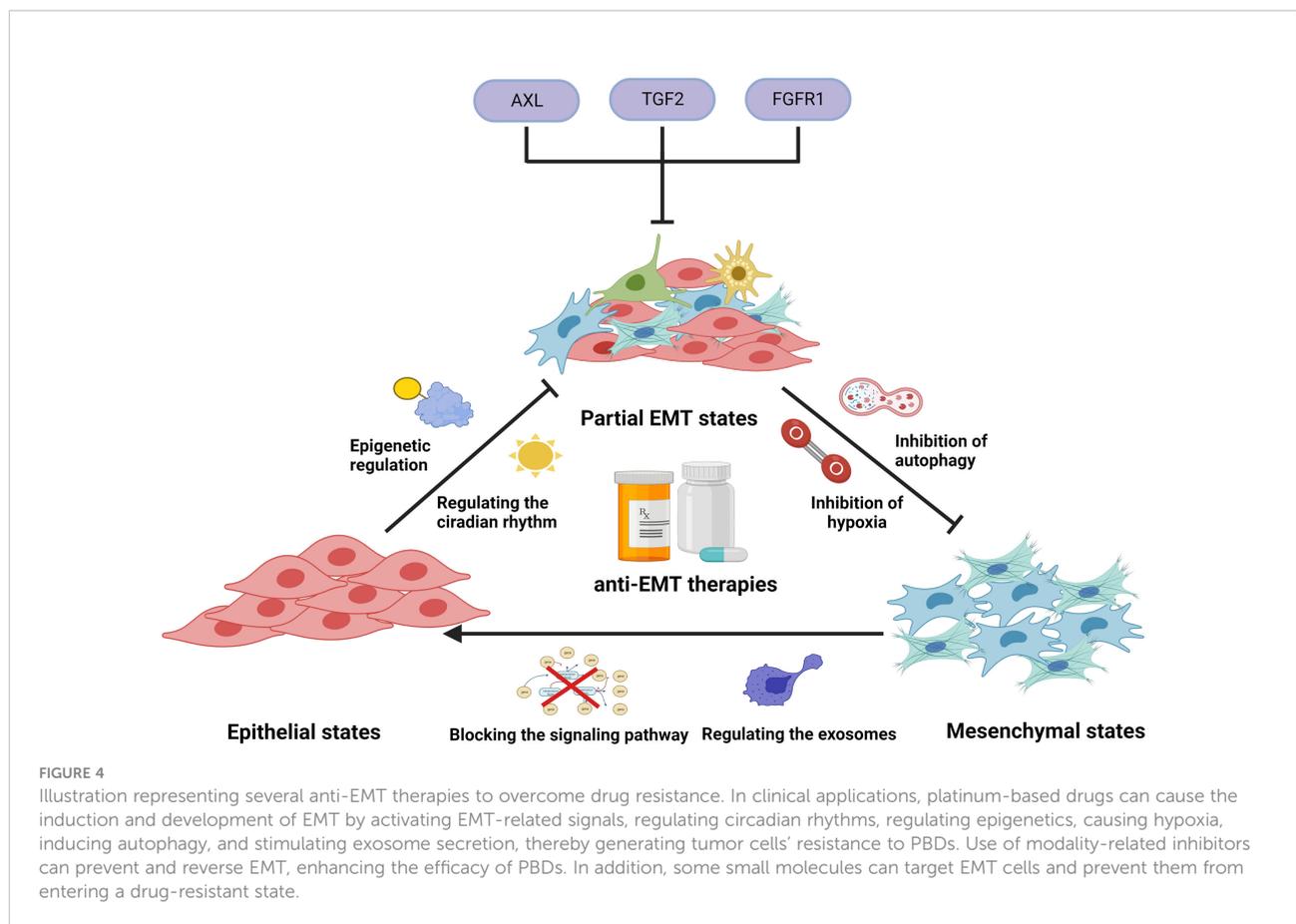
ERK pathways to induce the EMT program, and IGF-1R phosphorylation in cancer cells led to the production of DTPs (145). TGF β 2 linked the EMT program to fatty acid metabolism in an acidic environment (155). In the EMT process, the epithelial cells transformed into mesenchymal cells, gaining motility and losing cell-cell adhesion, making them more capable of entering the circulatory system. Recent research has shown that persistent subsets of proliferative cells in CTCs—cycling persister cells (CPCs)—not only survived cancer drugs but also maintained their ability to grow and multiply with continuing drug treatment. Moreover, single-cell RNA sequencing and metabolic analysis demonstrated that CPCs enhanced antioxidant gene capacity by increasing glutathione metabolism. It was also found that CPCs were dependent on fatty acid-based metabolism rather than glucose-based metabolism (156). Therefore, CTCs with EMT activation are more likely to transform into CPCs, resulting in drug resistance, distant recurrent lesions, and metastasis.

Targeting the EMT for cancer therapy

It has been proven that in the development of EMT, cancer cells generated new phenotypes, enabling them to survive PBD treatment. To eliminate drug resistance caused by EMT, different strategies to target the EMT program, such as preventing EMT initiation, targeting partial EMT cells, and reversing the EMT process, have been pursued (Figure 4).

Preventing EMT initiation

Tumor cells develop EMT after chemotherapy due to alterations in signaling pathways, circadian rhythms, and epigenetic regulation, making the tumor cells more malignant and eventually drug-resistant (157). For this reason, blocking the induction of EMT in tumor cells is crucial. Chemotherapy results in abnormal activation or inhibition of signaling pathways in tumor cells, and these abnormal signaling pathways can lead to EMT. We have previously described in detail the signaling pathways involved in platinum-induced EMT, including Wnt, TGF- β , Notch, NF- κ B, and hedgehog. Blocking any of the links in the signaling pathway of drug-induced EMT can affect the development of EMT in tumor cells to varying degrees. It was shown that the CXCL/CXCR2 axis was activated in resistant cells after cisplatin treatment, allowing CXCR2 to be highly expressed in human lung cancer tissues. Overexpression of CXCR2 promoted EMT by activating the p38/ERK MAPK pathway. SB225002, a selective CXCR2 inhibitor, blocked CXCR2 expression, thus preventing EMT



formation (51). In addition, hematopoietic PBX interacting protein (HPIP) was overexpressed in high-grade primary ovarian tumors, which increased Snail stability by activating the PI3K/AKT pathway and inhibiting the expression of E-cadherin by phosphorylating glycogen synthase kinase-3 β (GSK-3 β), thereby inducing EMT. Therefore, it is reasonable to attempt to inhibit the formation of EMT by knocking down HPIP using PI3K and AKT inhibitors (158). Reducing the expression of Snail could also have an inhibitory effect on the formation of EMT (159).

Epigenetic regulation plays an important role in cancer progression. Ten-eleven translocation 1 (TET1), an important DNA demethylase, was found to be overexpressed in cisplatin-resistant ovarian cancer cells and could induce partial EMT by increasing vimentin expression through demethylation of the vimentin promoter. Thus, we could inhibit EMT formation by downregulating TET1 expression using TET1 siRNA (160). Another study found that cisplatin-resistant ovarian cancer cells had increased expression of histone deacetylase (HDAC), which resulted in histone hypoacetylation and suppression of many genes, leading to EMT induction and subsequent PBD

resistance. Consequently, the HDAC inhibitor sodium butyrate (Nabu) could be used to restore histone acetylation, allowing tumor cells to upregulate E-cadherin expression and thus inhibit EMT formation. In addition, the combination of Nabu and cisplatin enhanced the toxicity of cisplatin on tumor cells (161).

Hypoxic conditions can occur because of rapid tumor growth and lack of blood vessel formation. Hypoxia drives adaptive changes that can lead to malignant tumor transformation. In NSCLC, hypoxia significantly upregulated antizyme inhibitor 2 (AZIN2) expression by increasing the binding of HIF-1 α and AZIN2 promoters, resulting in decreased E-cadherin expression and increased N-cadherin and vimentin expression, thereby promoting EMT (162). Therefore, maintaining oxygenation at the appropriate level could be a viable strategy for preventing EMT initiation in tumor cells. In treating cisplatin-resistant gastric cancer with Danggui-Sayuk-Ga-Osuyu-Saenggang-tang (DSGOST), an accumulation of GFP-LC3 puncta was induced, which promoted the release of exosomes through activation of autophagy, thereby inducing EMT. Consequently, the activation of DSGOST-mediated EMT markers, including N-

cadherin, Snail, Slug, vimentin, β -catenin, p-Smad2, and p-Smad3, could be blocked by inhibiting autophagy, which in turn inhibited EMT formation (163).

Traditional Chinese medicine (TCM) research on the EMT process is only in the initial stages, but we can still identify new potential drugs and strategies for inhibiting the formation of EMT from existing TCM extracts and monomers. As a prime example of the value of TCM, curcumin often appears in various prescriptions. In recent years, studies have shown that curcumin inhibited TGF- β -induced EMT through the PPAR γ pathway. Other researchers found that curcumin could effectively inhibit the NF- κ B pathway to reduce the expression of EMT-related genes (164–166). In addition, ginsenoside Rg3, triptolide, and resveratrol had similar effects (167, 168). Therefore, analyzing the TCM extracts for active compounds is a worthy effort that could lead to new drugs for translation into clinical treatments.

Targeting partial EMT cells.

The intermediate state on the way to complete EMT is called partial EMT, the essential feature of which is the simultaneous expression of E-cadherin and vimentin. Multiple studies have shown that cancer cells with coexpression of E-cadherin and vimentin led to poorer prognosis compared with those that expressed E-cadherin or vimentin alone, or neither (169). Therefore, when the tumor tissue presents a partial EMT state and is highly resistant to multiple drugs, the elimination of some or all of them could increase the therapeutic effect and prolong the survival of patients to a certain extent. In addition to these EMT markers, a number of other markers were also found associated with partial EMT, such as integrin beta 4 (ITGB4), interrogating the grainyhead-like 2 (GRHL2), ferroptosis suppressor protein 1 (FSP1), and ZEB1. These markers were also associated with shorter overall survival, poor relapse-free and disease-free survival (132, 170, 171). Whole-genome CRISPR screening revealed that mesenchymal EGFR mutant non-small cell lung cancers highly express FGFR1, which promotes DTPs. Combining EGFR and FGFR inhibitors could block the development of persistent resistance and drug-tolerant cell survival (153). Some molecules, such as recombinant AXL receptor tyrosine kinase (AXL), TGF2, and FGFR1, could be targeted to inhibit cells in EMT states from becoming DTPs, which could be a novel approach for correcting EMT-induced drug resistance. In addition, the cancer cells of the EMT process released proinflammatory cytokines, inducing drug resistance. For example, comprised of chemokine ligand 21 (CCL21) stimulated the AKT/GSK3 β /Snail pathway, which promoted the expression of multidrug resistance proteins such as P-glycoprotein1 (172). Therefore, we speculated that preventing partial EMT cells from secreting small molecules might inhibit the production of drug-resistant proteins. In addition, it is also

interesting to see whether the protective molecules secreted by these cells play a role in immune cell aggregation to engulf chemotherapeutic drugs and prevent immune escape.

Reversing the EMT process.

The EMT program is positively correlated with poor cancer prognosis. Therefore, taking the reversal of EMT as the starting point for resistance to cancer should increase the success rate of chemotherapy. For example, the ionophore antibiotic salinomycin upregulates the E-cadherin gene while reducing the expression of vimentin in CD133+ colorectal cancer cells and restoring cancer cell sensitivity to chemotherapy. This research also found that CD133 was a target of salinomycin (173). Therefore, combining salinomycin and PBDs could reverse the drug-induced EMT process and specifically target CD133+ CSCs. The specific signaling pathways play a role in both the induction of EMT and the drug resistance induced by EMT; thus, turning off the activated pathways should reverse EMT and eliminate drug resistance (174). A combination of cisplatin and BEZ235, an ATP-competitive dual inhibitor of PI3K and mTOR, could reverse the EMT program, induce apoptosis, and decrease the number of resistant cells (175).

Mesenchymal stem cells secrete exosomes that increase the mRNA levels of CK-19 and E-cadherin and decrease the expression of vimentin. In addition, exosomes reduce phosphorylation of the upregulated proteins, vimentin, TGF- β 1, and Smad2 (176). Further studies suggest that miRNAs play an essential role in this process. For example, exosomes derived from mesenchymal cells can transfer miR-182-5p and miR-23a-3p into EMT cells and target Ikbk and ubiquitin specific peptidase 5 (Usp5) to repress IKK β ubiquitination, leading to the inhibition of NF- κ B signaling and the reversal of EMT (177). The secretion by mesenchymal cells of specific molecules that reverse the EMT process could partly explain the presence of partial EMT. It might be possible to enhance the secretion of these substances and the formation of exosomes to eliminate tumor tissue resistance to chemotherapy.

Conclusions and perspectives

Research in the field of EMT has evolved from promoting normal development and wound repair to having a significant and complex impact on tumor resistance. On the one hand, EMT can induce the generation of cells with different phenotypes, which are resistant to PDBs to different degrees. On the other hand, the EMT process can induce tumor cells to produce multidrug-resistant proteins to evade the killing effect of drugs. It has been recognized that the induction of EMT required the activation of multiple signaling pathways, offering

several opportunities for therapeutic intervention to inhibit the occurrence and development of the EMT process, and even to reverse it. However, the combination of EMT inhibitors and PBDs has not achieved the desired effect in clinical practice. This may be related to the synergistic effect of multiple signaling pathways, the constantly changing tumor microenvironment, or stress-induced metabolic changes in the tumor. New research findings together with advances in technology could better promote the development of new drugs so that PBDs combined with radiotherapy could be precisely targeted to ensure a more substantial inhibitory effect on the EMT process with fewer toxic side effects. Moreover, the application of new technology and bioinformatics methods has led to a deeper understanding of EMT, with the discovery of new proteins and new pathways, checkpoints and bypasses, which play important roles in resistance to chemotherapeutic drugs. Current research on EMT has been focused on identifying and neutralizing the EMT-related proteins involved in drug resistance. Highlights of the recent research include the effects of CSCs, CTCs, and DTPs on tumor recurrence, metastasis, and drug resistance, which have promoted an in-depth exploration of the relationship between these cells' metabolism and EMT. Another fruitful area of research has been the effects of the extracts and individual components of traditional Chinese medicines on EMT. The results of these studies are expected to be the keys to breaking the bottleneck of EMT-induced tumor resistance. In the coming decades, research on EMT and the mechanism of drug resistance will deepen our understanding of how to solve the resistance to PBDs and reveal ways to Kibbe make these agents safer and more effective in clinical practice.

References

- Rottenberg S, Disler C, Perego P. The rediscovery of platinum-based cancer therapy. *Nat Rev Cancer* (2021) 21:37–50. doi: 10.1038/s41568-020-00308-y
- Giacomini I, Ragazzi E, Pasut G, Montopoli M. The pentose phosphate pathway and its involvement in cisplatin resistance. *Int J Mol Sci* (2020) 21(3):937. doi: 10.3390/ijms21030937
- Nieto MA, Huang RY, Jackson RA, Thiery JP. EMT: 2016. *Cell* (2016) 166:21–45. doi: 10.1016/j.cell.2016.06.028
- Burger GA, Danen EHJ, Beltman JB. Deciphering epithelial-mesenchymal transition regulatory networks in cancer through computational approaches. *Front Oncol* (2017) 7:162. doi: 10.3389/fonc.2017.00162
- Bjerkvig R, Tysnes BB, Aboody KS, Najbauer J, Terzis AJ. Opinion: the origin of the cancer stem cell: current controversies and new insights. *Nat Rev Cancer* (2005) 5:899–904. doi: 10.1038/nrc1740
- Krebs AM, Mitschke J, Lasierra Losada M, Schmalhofer O, Boerries M, Busch H, et al. The EMT-activator Zeb1 is a key factor for cell plasticity and promotes metastasis in pancreatic cancer. *Nat Cell Biol* (2017) 19:518–29. doi: 10.1038/ncb3513
- Zhang M, Du H, Wang L, Yue Y, Zhang P, Huang Z, et al. Thymoquinone suppresses invasion and metastasis in bladder cancer cells by reversing EMT through the wnt/ β -catenin signaling pathway. *Chem Biol Interact* (2020) 320:109022. doi: 10.1016/j.cbi.2020.109022
- Hussain Y, Cui JH, Khan H, Aschner M, Batiha GE, Jeandet P. Luteolin and cancer metastasis suppression: focus on the role of epithelial to mesenchymal transition. *Med Oncol* (2021) 38:66. doi: 10.1007/s12032-021-01508-8
- Bahrami A, Majeed M, Sahebkar A. Curcumin: a potent agent to reverse epithelial-to-mesenchymal transition. *Cell Oncol (Dordr)* (2019) 42:405–21. doi: 10.1007/s13402-019-00442-2
- Valinezhad Sani F, Palizban A, Mosaffa F, Jamialahmadi K. Glucosamine attenuates drug resistance in mitoxantrone-resistance breast cancer cells. *J Pharm Pharmacol* (2021) 73:922–7. doi: 10.1093/jpp/rgaa032
- Wang YS, Li H, Li Y, Zhang S, Jin YH. (20S)G-Rh2 inhibits NF- κ B regulated epithelial-mesenchymal transition by targeting annexin A2. *Biomolecules* (2020) 10(4):528. doi: 10.3390/biom10040528
- Sherbet GV. Suppression of angiogenesis and tumour progression by combretastatin and derivatives. *Cancer Lett* (2017) 403:289–95. doi: 10.1016/j.canlet.2017.06.032
- Lin TA, Lin WS, Chou YC, Nagabhushanam K, Ho CT, Pan MH. Oxyresveratrol inhibits human colon cancer cell migration through regulating epithelial-mesenchymal transition and microRNA. *Food Funct* (2021) 12:9658–68. doi: 10.1039/D1FO01920A
- Bandyopadhyaya S, Akimov MG, Verma R, Sharma A, Sharma D, Kundu GC, et al. N-arachidonoyl dopamine inhibits epithelial-mesenchymal transition of breast cancer cells through ERK signaling and decreasing the cellular cholesterol. *J Biochem Mol Toxicol* (2021) 35:e22693. doi: 10.1002/jbt.22693
- Lee GA, Choi KC, Hwang KA. Kaempferol, a phytoestrogen, suppressed triclosan-induced epithelial-mesenchymal transition and metastatic-related behaviors of MCF-7 breast cancer cells. *Environ Toxicol Pharmacol* (2017) 49:48–57. doi: 10.1016/j.etap.2016.11.016

Author contributions

XD and ML wrote the manuscript. JL made the figures. ZS and KX revised the manuscript. All authors contributed to the article and approved the submitted version.

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Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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16. Zhang L, Xu L, Zhang F, Vlashi E. Doxycycline inhibits the cancer stem cell phenotype and epithelial-to-mesenchymal transition in breast cancer. *Cell Cycle* (2017) 16:737–45. doi: 10.1080/15384101.2016.1241929
17. Kozak J, Forma A, Czezelewski M, Kozyra P, Sitarz E, Radzikowska-Büchner E, et al. Inhibition or reversal of the epithelial-mesenchymal transition in gastric cancer: Pharmacological approaches. *Int J Mol Sci* (2020) 22(1):277. doi: 10.3390/ijms22010277
18. Imran M, Rauf A, Shah ZA, Saeed F, Imran A, Arshad MU, et al. Chemopreventive and therapeutic effect of the dietary flavonoid kaempferol: A comprehensive review. *Phytother Res* (2019) 33:263–75. doi: 10.1002/ptr.6227
19. Oh HC, Shim JK, Park J, Lee JH, Choi RJ, Kim NH, et al. Combined effects of niclosamide and temozolomide against human glioblastoma tumorspheres. *J Cancer Res Clin Oncol* (2020) 146:2817–28. doi: 10.1007/s00432-020-03330-7
20. Farhat D, Lincet H. Lipoic acid a multi-level molecular inhibitor of tumorigenesis. *Biochim Biophys Acta Rev Cancer* (2020) 1873:188317. doi: 10.1016/j.bbcan.2019.188317
21. Park MJ, Lee DE, Shim MK, Jang EH, Lee JK, Jeong SY, et al. Piperlongumine inhibits TGF- β -induced epithelial-to-mesenchymal transition by modulating the expression of e-cadherin. *Snail1 Twist1 Eur J Pharmacol* (2017) 812:243–9. doi: 10.1016/j.ejphar.2017.07.036
22. Ju H, Li Y, Xing X, Miao X, Feng Y, Ren Y, et al. Manganese-12 acetate suppresses the migration, invasion, and epithelial-mesenchymal transition by inhibiting wnt/ β -catenin and PI3K/AKT signaling pathways in breast cancer cells. *Thorac Cancer* (2018) 9:353–9. doi: 10.1111/1759-7714.12584
23. Matias D, Balça-Silva J, Dubois LG, Pontes B, Ferrer VP, Rosário L, et al. Dual treatment with shikonin and temozolomide reduces glioblastoma tumor growth, migration and glial-to-mesenchymal transition. *Cell Oncol (Dordr)* (2017) 40:247–61. doi: 10.1007/s13402-017-0320-1
24. Lin S, Zhang C, Liu F, Ma J, Jia F, Han Z, et al. Actinomycin V inhibits migration and invasion via suppressing Snail/Slug-mediated epithelial-mesenchymal transition progression in human breast cancer MDA-MB-231 cells *In vitro*. *Mar Drugs* (2019) 17(5):305. doi: 10.3390/md17050305
25. Cruz-Lozano M, González-González A, Marchal JA, Muñoz-Muela E, Molina MP, Cara FE, et al. Hydroxytyrosol inhibits cancer stem cells and the metastatic capacity of triple-negative breast cancer cell lines by the simultaneous targeting of epithelial-to-mesenchymal transition, wnt/ β -catenin and TGF β signaling pathways. *Eur J Nutr* (2019) 58:3207–19. doi: 10.1007/s00394-018-1864-1
26. Hseu YC, Chao YH, Lin KY, Way TD, Lin HY, Thiyagarajan V, et al. Antrodia camphorata inhibits metastasis and epithelial-to-mesenchymal transition via the modulation of claudin-1 and wnt/ β -catenin signaling pathways in human colon cancer cells. *J Ethnopharmacol* (2017) 208:72–83. doi: 10.1016/j.jep.2017.07.001
27. Farahmand L, Darvishi B, Majidzadeh AK, Madjid Ansari A. Naturally occurring compounds acting as potent anti-metastatic agents and their suppressing effects on hedgehog and WNT/ β -catenin signalling pathways. *Cell Prolif* (2017) 50(1):e12299. doi: 10.1111/cpr.12299
28. Chen Y, Chen JQ, Ge MM, Zhang Q, Wang XQ, Zhu JY, et al. Sulforaphane inhibits epithelial-mesenchymal transition by activating extracellular signal-regulated kinase 5 in lung cancer cells. *J Nutr Biochem* (2019) 72:108219. doi: 10.1016/j.jnutbio.2019.108219
29. Jia M, Wang Y, Guo Y, Yu P, Sun Y, Song Y, et al. Nitidine chloride suppresses epithelial-mesenchymal transition and stem cell-like properties in glioblastoma by regulating JAK2/STAT3 signaling. *Cancer Med* (2021) 10:3113–28. doi: 10.1002/cam4.3869
30. Kaul R, Risinger AL, Mooberry SL. Eribulin rapidly inhibits TGF- β -induced snail expression and can induce slug expression in a Smad4-dependent manner. *Br J Cancer* (2019) 121:611–21. doi: 10.1038/s41416-019-0556-9
31. Hsieh YS, Chu SC, Huang SC, Kao SH, Lin MS, Chen PN. Gossypol reduces metastasis and epithelial-mesenchymal transition by targeting protease in human cervical cancer. *Am J Chin Med* (2021) 49:181–98. doi: 10.1142/S0192415X21500105
32. Kong D, Zhang Z, Chen L, Huang W, Zhang F, Wang L, et al. Curcumin blunts epithelial-mesenchymal transition of hepatocytes to alleviate hepatic fibrosis through regulating oxidative stress and autophagy. *Redox Biol* (2020) 36:101600. doi: 10.1016/j.redox.2020.101600
33. Divya T, Velavan B, Sudhandiran G. Regulation of transforming growth factor- β /Smad-mediated epithelial-mesenchymal transition by celastrol provides protection against bleomycin-induced pulmonary fibrosis. *Basic Clin Pharmacol Toxicol* (2018) 123:122–9. doi: 10.1111/bcpt.12975
34. Chen K, Zhang S, Ji Y, Li J, An P, Ren H, et al. Baicalein inhibits the invasion and metastatic capabilities of hepatocellular carcinoma cells via down-regulation of the ERK pathway. *PLoS One* (2013) 8:e72927. doi: 10.1371/journal.pone.0072927
35. Xing S, Yu W, Zhang X, Luo Y, Lei Z, Huang D, et al. Isoviolanthin extracted from *Dendrobium officinale* reverses TGF- β 1-Mediated Epithelial-Mesenchymal transition in hepatocellular carcinoma cells via deactivating the TGF- β /Smad and PI3K/Akt/mTOR signaling pathways. *Int J Mol Sci* (2018) 19(6):1556. doi: 10.3390/ijms19061556
36. Qin CD, Ma DN, Ren ZG, Zhu XD, Wang CH, Wang YC, et al. Astragaloside IV inhibits metastasis in hepatoma cells through the suppression of epithelial-mesenchymal transition via the Akt/GSK-3 β / β -catenin pathway. *Oncol Rep* (2017) 37:1725–35. doi: 10.3892/or.2017.5389
37. Xiao M, Hasmim M, Lequeux A, Moer KV, Tan TZ, Gilles C, et al. Epithelial to mesenchymal transition regulates surface PD-L1 via CMTM6 and CMTM7 induction in breast cancer. *Cancers (Basel)* (2021) 13(5):1165. doi: 10.3390/cancers13051165
38. Ge L, Li DS, Chen F, Feng JD, Li B, Wang TJ. TAZ overexpression is associated with epithelial-mesenchymal transition in cisplatin-resistant gastric cancer cells. *Int J Oncol* (2017) 51:307–15. doi: 10.3892/ijo.2017.3998
39. Ghosh S, Singh R, Vanwinkle ZM, Guo H, Vemula PK, Goel A, et al. Microbial metabolite restricts 5-fluorouracil-resistant colonic tumor progression by sensitizing drug transporters via regulation of FOXO3-FOXM1 axis. *Theranostics* (2022) 12:5574–95. doi: 10.7150/thno.70754
40. Ka C, Gautam S, Marshall SR, Tice LP, Martinez-Bartolome M, Fenner JL, et al. Receptor tyrosine kinases *ror1/2* and *ryk* are Co-expressed with multiple wnt signaling components during early development of Sea urchin embryos. *Biol Bull* (2021) 241:140–57. doi: 10.1086/715237
41. Zhang XY, Li YF, Ma H, Gao YH. Regulation of MYB mediated cisplatin resistance of ovarian cancer cells involves miR-21-wnt signaling axis. *Sci Rep* (2020) 10:6893. doi: 10.1038/s41598-020-63396-8
42. Zhang Y, Zhang Q, Chen H, Wang C. BCL9 promotes epithelial mesenchymal transition and invasion in cisplatin resistant NSCLC cells via β -catenin pathway. *Life Sci* (2018) 208:284–94. doi: 10.1016/j.lfs.2018.07.023
43. Zhang Y, Devocelle A, Desterke C, de Souza LEB, Hadadi É., Acloque H, et al. BMAL1 knockdown leans epithelial-mesenchymal balance toward epithelial properties and decreases the chemoresistance of colon carcinoma cells. *Int J Mol Sci* (2021) 22(10):5247. doi: 10.3390/ijms22105247
44. Yang Y, Jiang H, Li W, Chen L, Zhu W, Xian Y, et al. FOXM1/DVL2/Snail axis drives metastasis and chemoresistance of colorectal cancer. *Aging (Albany NY)* (2020) 12:24424–40. doi: 10.18632/aging.202300
45. Wu W, He K, Guo Q, Chen J, Zhang M, Huang K, et al. SSRP1 promotes colorectal cancer progression and is negatively regulated by miR-28-5p. *J Cell Mol Med* (2019) 23:3118–29. doi: 10.1111/jcmm.14134
46. Ge T, Liu T, Guo L, Chen Z, Lou G. MicroRNA-302 represses epithelial-mesenchymal transition and cisplatin resistance by regulating ATAD2 in ovarian carcinoma. *Exp Cell Res* (2020) 396:112241. doi: 10.1016/j.yexcr.2020.112241
47. Filipka M, Skrzypski M, Czetyrbok K, Stokowy T, Sasiłojć G, Supernat A, et al. MiR-192 and miR-662 enhance chemoresistance and invasiveness of squamous cell lung carcinoma. *Lung Cancer* (2018) 118:111–8. doi: 10.1016/j.lungcan.2018.02.002
48. Ashrafzadeh M, Zarrabi A, Hushmandi K, Kalantari M, Mohammadinejad R, Javaheri T, et al. Association of the epithelial-mesenchymal transition (EMT) with cisplatin resistance. *Int J Mol Sci* (2020) 21(11):4002. doi: 10.3390/ijms21114002
49. Wang H, Guo S, Kim SJ, Shao F, Ho JWK, Wong KU, et al. Cisplatin prevents breast cancer metastasis through blocking early EMT and retards cancer growth together with paclitaxel. *Theranostics* (2021) 11:2442–59. doi: 10.7150/thno.46460
50. Poh ME, Liam CK, Mun KS, Chai CS, Wong CK, Tan JL, et al. Epithelial-to-mesenchymal transition (EMT) to sarcoma in recurrent lung adenocarcinoma following adjuvant chemotherapy. *Thorac Cancer* (2019) 10:1841–5. doi: 10.1111/1759-7714.13156
51. Cheng Y, Mo F, Li Q, Han X, Shi H, Chen S, et al. Targeting CXCR2 inhibits the progression of lung cancer and promotes therapeutic effect of cisplatin. *Mol Cancer* (2021) 20:62. doi: 10.1186/s12943-021-01355-1
52. Zheng M, Niu Y, Bu J, Liang S, Zhang Z, Liu J, et al. ESRP1 regulates alternative splicing of CARM1 to sensitize small cell lung cancer cells to chemotherapy by inhibiting TGF- β /Smad signaling. *Aging (Albany NY)* (2021) 13:3554–72. doi: 10.18632/aging.202295
53. Nakamura K, Terai Y, Tanabe A, Ono YJ, Hayashi M, Maeda K, et al. CD24 expression is a marker for predicting clinical outcome and regulates the epithelial-mesenchymal transition in ovarian cancer via both the akt and ERK pathways. *Oncol Rep* (2017) 37:3189–200. doi: 10.3892/or.2017.5583
54. Bissey PA, Teng M, Law JH, Shi W, Bruce JP, Petit V, et al. MiR-34c downregulation leads to SOX4 overexpression and cisplatin resistance in nasopharyngeal carcinoma. *BMC Cancer* (2020) 20:597. doi: 10.1186/s12885-020-07081-z
55. Ma J, Zeng S, Zhang Y, Deng G, Qu Y, Guo C, et al. BMP4 promotes oxaliplatin resistance by an induction of epithelial-mesenchymal transition via MEK1/ERK/ELK1 signaling in hepatocellular carcinoma. *Cancer Lett* (2017) 411:117–29. doi: 10.1016/j.canlet.2017.09.041

56. Yan J, Xu H. Regulation of transforming growth factor-beta1 by circANKS1B/miR-515-5p affects the metastatic potential and cisplatin resistance in oral squamous cell carcinoma. *Bioengineered* (2021) 12:12420–30. doi: 10.1080/21655979.2021.2005221
57. Steins A, Ebbing EA, Creemers A, van der Zalm AP, Jibodh RA, Waasdorp C, et al. Chemoradiation induces epithelial-to-mesenchymal transition in esophageal adenocarcinoma. *Int J Cancer* (2019) 145:2792–803. doi: 10.1002/ijc.32364
58. Ni MM, Wang YR, Wu WW, Xia CC, Zhang YH, Xu J, et al. Novel insights on notch signaling pathways in liver fibrosis. *Eur J Pharmacol* (2018) 826:66–74. doi: 10.1016/j.ejphar.2018.02.051
59. Pagie S, Gérard N, Charreau B. Notch signaling triggered via the ligand DLL4 impedes M2 macrophage differentiation and promotes their apoptosis. *Cell Communication Signaling CCS* (2018) 16:4. doi: 10.1186/s12964-017-0214-x
60. Fukusumi T, Guo TW, Sakai A, Ando M, Ren S, Haft S, et al. The NOTCH4-HEY1 pathway induces epithelial-mesenchymal transition in head and neck squamous cell carcinoma. *Clin Cancer Res* (2018) 24:619–33. doi: 10.1158/1078-0432.CCR-17-1366
61. Zeng D, Liang YK, Xiao YS, Wei XL, Lin HY, Wu Y, et al. Inhibition of Notch1 reverses EMT and chemoresistance to cisplatin via direct downregulation of MCAM in triple-negative breast cancer cells. *Int J Cancer* (2020) 147:490–504. doi: 10.1002/ijc.32911
62. Xiao YS, Zeng D, Liang YK, Wu Y, Li MF, Qi YZ, et al. Major vault protein is a direct target of Notch1 signaling and contributes to chemoresistance in triple-negative breast cancer cells. *Cancer Lett* (2019) 440-441:156–67. doi: 10.1016/j.canlet.2018.09.031
63. Xu Y, Miao C, Jin C, Qiu C, Li Y, Sun X, et al. SUSD2 promotes cancer metastasis and confers cisplatin resistance in high grade serous ovarian cancer. *Exp Cell Res* (2018) 363:160–70. doi: 10.1016/j.yexcr.2017.12.029
64. Dai G, Liu G, Zheng D, Song Q. Inhibition of the notch signaling pathway attenuates progression of cell motility, metastasis, and epithelial-to-mesenchymal transition-like phenomena induced by low concentrations of cisplatin in osteosarcoma. *Eur J Pharmacol* (2021) 899:174058. doi: 10.1016/j.ejphar.2021.174058
65. Palagano E, Menale C, Sobacchi C, Villa A. Genetics of osteopetrosis. *Curr Osteoporosis Rep* (2018) 16:13–25. doi: 10.1007/s11914-018-0415-2
66. Mussbacher M, Salzmann M, Brostjan C, Hoesel B, Schoergenhofer C, Datler H, et al. Cell type-specific roles of NF- κ B linking inflammation and thrombosis. *Front Immunol* (2019) 10:85. doi: 10.3389/fimmu.2019.00085
67. Liao J, Yang Z, Carter-Cooper B, Chang ET, Choi EY, Kallakury B, et al. Suppression of migration, invasion, and metastasis of cisplatin-resistant head and neck squamous cell carcinoma through IKK β inhibition. *Clin Exp Metastasis* (2020) 37:283–92. doi: 10.1007/s10585-020-10021-7
68. Jin Y, Huynh DTN, Kang KW, Myung CS, Heo KS. Inhibition of p90RSK activation sensitizes triple-negative breast cancer cells to cisplatin by inhibiting proliferation, migration and EMT. *BMB Rep* (2019) 52:706–11. doi: 10.5483/BMBRep.2019.52.12.234
69. Zhao Y, He M, Cui L, Gao M, Zhang M, Yue F, et al. Chemotherapy exacerbates ovarian cancer cell migration and cancer stem cell-like characteristics through GLI1. *Br J Cancer* (2020) 122:1638–48. doi: 10.1038/s41416-020-0825-7
70. Qin T, Li B, Feng X, Fan S, Liu L, Liu D, et al. Abnormally elevated USP37 expression in breast cancer stem cells regulates stemness, epithelial-mesenchymal transition and cisplatin sensitivity. *J Exp Clin Cancer Res* (2018) 37:287. doi: 10.1186/s13046-018-0934-9
71. Wang Y, Li Q, Xu L, Chen J, Pu Y, Wang L, et al. Cancer stemness of CD10-positive cells regulated by hedgehog pathway promotes the resistance to cisplatin in oral squamous cell carcinoma. *Oral Dis* (2021) 27:1403–11. doi: 10.1111/odi.13673
72. He Y, Xie H, Yu P, Jiang S, Wei L. FOXC2 promotes epithelial-mesenchymal transition and cisplatin resistance of non-small cell lung cancer cells. *Cancer Chemotherapy Pharmacol* (2018) 82:1049–59. doi: 10.1007/s00280-018-3697-2
73. Yin C, Lin X, Wang Y, Liu X, Xiao Y, Liu J, et al. FAM83D promotes epithelial-mesenchymal transition, invasion and cisplatin resistance through regulating the AKT/mTOR pathway in non-small-cell lung cancer. *Cell Oncol (Dordrecht)* (2020) 43:395–407. doi: 10.1007/s13402-020-00494-9
74. Wu DM, Zhang T, Liu YB, Deng SH, Han R, Liu T, et al. The PAX6-ZEB2 axis promotes metastasis and cisplatin resistance in non-small cell lung cancer through PI3K/AKT signaling. *Cell Death Dis* (2019) 10:349. doi: 10.1038/s41419-019-1591-4
75. Liu X, Huang Z, Qian W, Zhang Q, Sun J. Silence of lncRNA UCA1 rescues drug resistance of cisplatin to non-small-cell lung cancer cells. *J Cell Biochem* (2019) 120:9243–9. doi: 10.1002/jcb.28200
76. Zhang P, Lu X, Shi Z, Li X, Zhang Y, Zhao S, et al. miR-205-5p regulates epithelial-mesenchymal transition by targeting PTEN via PI3K/AKT signaling pathway in cisplatin-resistant nasopharyngeal carcinoma cells. *Gene* (2019) 710:103–13. doi: 10.1016/j.gene.2019.05.058
77. Yoon C, Cho SJ, Chang KK, Park DJ, Ryeom SW, Yoon SS. Role of Rac1 pathway in epithelial-to-Mesenchymal transition and cancer stem-like cell phenotypes in gastric adenocarcinoma. *Mol Cancer Res MCR* (2017) 15:1106–16. doi: 10.1158/1541-7786.MCR-17-0053
78. Hu X, Feng Y, Zhang D, Zhao SD, Hu Z, Greshock J, et al. A functional genomic approach identifies FAL1 as an oncogenic long noncoding RNA that associates with BMI1 and represses p21 expression in cancer. *Cancer Cell* (2014) 26:344–57. doi: 10.1016/j.ccr.2014.07.009
79. Slack FJ, Chinnaiyan AM. The role of non-coding RNAs in oncology. *Cell* (2019) 179:1033–55. doi: 10.1016/j.cell.2019.10.017
80. Wang M, Zhang R, Zhang S, Xu R, Yang Q. MicroRNA-574-3p regulates epithelial mesenchymal transition and cisplatin resistance via targeting ZEB1 in human gastric carcinoma cells. *Gene* (2019) 700:110–9. doi: 10.1016/j.gene.2019.03.043
81. Wu S, Xie J, Shi H, Wang ZW. miR-492 promotes chemoresistance to CDDP and metastasis by targeting inhibiting DNMT3B and induces stemness in gastric cancer. *Biosci Rep* (2020) 40(3):BSR20194342. doi: 10.1042/BSR20194342
82. Wu DM, Hong XW, Wang LL, Cui XF, Lu J, Chen GQ, et al. MicroRNA-17 inhibition overcomes chemoresistance and suppresses epithelial-mesenchymal transition through a DEDD-dependent mechanism in gastric cancer. *Int J Biochem Cell Biol* (2018) 102:59–70. doi: 10.1016/j.biocel.2018.06.007
83. Cao L, Wan Q, Li F, Tang CE. MiR-363 inhibits cisplatin chemoresistance of epithelial ovarian cancer by regulating snail-induced epithelial-mesenchymal transition. *BMB Rep* (2018) 51:456–61. doi: 10.5483/BMBRep.2018.51.9.104
84. Kielbik M, Szulc-Kielbik I, Klink M. Impact of selected signaling proteins on SNAIL 1 and SNAIL 2 expression in ovarian cancer cell lines in relation to cells' cisplatin resistance and EMT markers level. *Int J Mol Sci* (2021) 22(2):980. doi: 10.3390/ijms22020980
85. Wang J, Liu L. MiR-149-3p promotes the cisplatin resistance and EMT in ovarian cancer through downregulating TIMP2 and CDKN1A. *J Ovarian Res* (2021) 14:165. doi: 10.1186/s13048-021-00919-5
86. Li H, Liang J, Qin F, Zhai Y. MiR-374b-5p-FOXP1 feedback loop regulates cell migration, epithelial-mesenchymal transition and chemosensitivity in ovarian cancer. *Biochem Biophys Res Commun* (2018) 505:554–60. doi: 10.1016/j.bbrc.2018.09.161
87. Li T, Xie J, Shen C, Cheng D, Shi Y, Wu Z, et al. Amplification of long noncoding RNA ZFAS1 promotes metastasis in hepatocellular carcinoma. *Cancer Res* (2015) 75:3181–91. doi: 10.1158/0008-5472.CAN-14-3721
88. Wang H, Huo X, Yang XR, He J, Cheng L, Wang N, et al. STAT3-mediated upregulation of lncRNA HOXD-AS1 as a ceRNA facilitates liver cancer metastasis by regulating SOX4. *Mol Cancer* (2017) 16:136. doi: 10.1186/s12943-017-0680-1
89. Farmer P, Bonnefoi H, Anderle P, Cameron D, Wirapati P, Becette V, et al. A stroma-related gene signature predicts resistance to neoadjuvant chemotherapy in breast cancer. *Nat Med* (2009) 15:68–74. doi: 10.1038/nm.1908
90. Byers LA, Diao L, Wang J, Saintigny P, Girard L, Peyton M, et al. An epithelial-mesenchymal transition gene signature predicts resistance to EGFR and PI3K inhibitors and identifies axl as a therapeutic target for overcoming EGFR inhibitor resistance. *Clin Cancer Res* (2013) 19:279–90. doi: 10.1158/1078-0432.CCR-12-1558
91. Bai X, Ni J, Beretov J, Graham P, Li Y. Cancer stem cell in breast cancer therapeutic resistance. *Cancer Treat Rev* (2018) 69:152–63. doi: 10.1016/j.ctrv.2018.07.004
92. Ji C, Yang L, Yi W, Xiang D, Wang Y, Zhou Z, et al. Capillary morphogenesis gene 2 maintains gastric cancer stem-like cell phenotype by activating a wnt/ β -catenin pathway. *Oncogene* (2018) 37:3953–66. doi: 10.1038/s41388-018-0226-z
93. Sui X, Cai J, Li H, He C, Zhou C, Dong Y, et al. p53-dependent CD51 expression contributes to characteristics of cancer stem cells in prostate cancer. *Cell Death Dis* (2018) 9:523. doi: 10.1038/s41419-018-0541-x
94. Cao S, Wang Z, Gao X, He W, Cai Y, Chen H, et al. FOXC1 induces cancer stem cell-like properties through upregulation of beta-catenin in NSCLC. *J Exp Clin Cancer Res CR* (2018) 37:220. doi: 10.1186/s13046-018-0894-0
95. Haspels HN, Rahman MA, Joseph JV, Gras Navarro A, Chekenya M. Glioblastoma stem-like cells are more susceptible than differentiated cells to natural killer cell lysis mediated through killer immunoglobulin-like receptors-human leukocyte antigen ligand mismatch and activation receptor-ligand interactions. *Front Immunol* (2018) 9:1345. doi: 10.3389/fimmu.2018.01345
96. Nguyen LV, Vanner R, Dirks P, Eaves CJ. Cancer stem cells: an evolving concept, nature reviews. *Cancer* (2012) 12:133–43. doi: 10.1038/nrc3184

97. Schatton T, Murphy GF, Frank NY, Yamaura K, Waaga-Gasser AM, Gasser M, et al. Identification of cells initiating human melanomas. *Nature* (2008) 451:345–9. doi: 10.1038/nature06489
98. Simbulan-Rosenthal CM, Gaur A, Zhou H, Abdusamad M, Qin Q, Dougherty R, et al. CD133 is associated with increased melanoma cell survival after multikinase inhibition. *J Oncol* (2019) 2019:6486173. doi: 10.1155/2019/6486173
99. Lu R, Zhao G, Yang Y, Jiang Z, Cai J, Hu H. Inhibition of CD133 overcomes cisplatin resistance through inhibiting PI3K/AKT/mTOR signaling pathway and autophagy in CD133-positive gastric cancer cells. *Technol Cancer Res Treat* (2019) 18:1533033819864311. doi: 10.1177/1533033819864311
100. Tian J, Li X, Si M, Liu T, Li J. CD271+ osteosarcoma cells display stem-like properties. *PLoS One* (2014) 9:e98549. doi: 10.1371/journal.pone.0098549
101. Jelski W, Laniewska-Dunaj M, Orywal K, Kochanowicz J, Rutkowski R, Szmikowski M. The diagnostic value of alcohol dehydrogenase (ADH) isoenzymes and aldehyde dehydrogenase (ALDH) measurement in the sera of patients with brain tumor. *Arch Med Sci AMS* (2017) 13:346–52. doi: 10.5114/aoms.2017.65366
102. Chaturvedi V, Fournier-Level A, Cooper HM, Murray MJ. Loss of Neogenin1 in human colorectal carcinoma cells causes a partial EMT and wound-healing response. *Sci Rep* (2019) 9:4110. doi: 10.1038/s41598-019-40886-y
103. Liao C, Wang Q, An J, Long Q, Wang H, Xiang M, et al. Partial EMT in squamous cell carcinoma: A snapshot. *Int J Biol Sci* (2021) 17:3036–47. doi: 10.7150/ijbs.61566
104. Sadeghi M, Ordway B, Rafiei I, Borad P, Fang B, Koomen JL, et al. Integrative analysis of breast cancer cells reveals an epithelial-mesenchymal transition role in adaptation to acidic microenvironment. *Front Oncol* (2020) 10:304. doi: 10.3389/fonc.2020.00304
105. Bocci F, Kumar Jolly M, Onuchic JN. A biophysical model uncovers the size distribution of migrating cell clusters across cancer types. *Cancer Res* (2019) 79:5527–35. doi: 10.1158/0008-5472.CAN-19-1726
106. Li Y, Fu L, Li JB, Qin Y, Zeng TT, Zhou J, et al. Increased expression of EIF5A2, via hypoxia or gene amplification, contributes to metastasis and angiogenesis of esophageal squamous cell carcinoma. *Gastroenterology* (2014) 146:1701–1713.e1709. doi: 10.1053/j.gastro.2014.02.029
107. Wu JS, Li ZF, Wang HF, Yu XH, Pang X, Wu JB, et al. Cathepsin b defines leader cells during the collective invasion of salivary adenoid cystic carcinoma. *Int J Oncol* (2019) 54:1233–44. doi: 10.3892/ijco.2019.4722
108. Zhu X, Chen L, Liu L, Niu X. EMT-mediated acquired EGFR-TKI resistance in NSCLC: Mechanisms and strategies. *Front Oncol* (2019) 9:1044. doi: 10.3389/fonc.2019.01044
109. Dongre A, Weinberg RA. New insights into the mechanisms of epithelial-mesenchymal transition and implications for cancer, nature reviews. *Mol Cell Biol* (2019) 20:69–84. doi: 10.1038/s41580-018-0080-4
110. Saxena K, Jolly MK, Balamurugan K. Hypoxia partial EMT collective migration: Emerging culprits metastasis. *Trans Oncol* (2020) 13:100845. doi: 10.1016/j.tranon.2020.100845
111. Dossus L, Benusiglio PR. Lobular breast cancer: incidence and genetic and non-genetic risk factors. *Breast Cancer Res BCR* (2015) 17:37. doi: 10.1186/s13058-015-0546-7
112. Li L, Han R, Xiao H, Lin C, Wang Y, Liu H, et al. Metformin sensitizes EGFR-TKI-resistant human lung cancer cells *in vitro* and *in vivo* through inhibition of IL-6 signaling and EMT reversal. *Clin Cancer Res an Off J Am Assoc Cancer Res* (2014) 20:2714–26. doi: 10.1158/1078-0432.CCR-13-2613
113. Biddle A, Gammon L, Liang X, Costea DE, Mackenzie IC. Phenotypic plasticity determines cancer stem cell therapeutic resistance in oral squamous cell carcinoma. *EBioMedicine* (2016) 4:138–45. doi: 10.1016/j.ebiom.2016.01.007
114. Hanrahan K, O'Neill A, Principe M, Bugler J, Murphy L, Fabre A, et al. The role of epithelial-mesenchymal transition drivers ZEB1 and ZEB2 in mediating docetaxel-resistant prostate cancer. *Mol Oncol* (2017) 11:251–65. doi: 10.1002/1878-0261.12030
115. Bosc C, Broin N, Fanjul M, Saland E, Farge T, Courdy C, et al. Autophagy regulates fatty acid availability for oxidative phosphorylation through mitochondria-endoplasmic reticulum contact sites. *Nat Commun* (2020) 11:4056. doi: 10.1038/s41467-020-17882-2
116. Eichner LJ, Brun SN, Herzog S, Young NP, Curtis SD, Shackelford DB, et al. Genetic analysis reveals AMPK is required to support tumor growth in murine kras-dependent lung cancer models. *Cell Metab* (2019) 29:285–302.e287. doi: 10.1016/j.cmet.2018.10.005
117. Poillet-Perez L, Xie X, Zhan L, Yang Y, Sharp DW, Hu ZS, et al. Autophagy maintains tumour growth through circulating arginine. *Nature* (2018) 563:569–73. doi: 10.1038/s41586-018-0697-7
118. Yang A, Herter-Sprig G, Zhang H, Lin EY, Biancur D, Wang X, et al. Autophagy sustains pancreatic cancer growth through both cell-autonomous and nonautonomous mechanisms. *Cancer Discovery* (2018) 8:276–87. doi: 10.1158/2159-8290.CD-17-0952
119. Shi RZ, He YF, Wen J, Niu YN, Gao Y, Liu LH, et al. Epithelial cell adhesion molecule promotes breast cancer resistance protein-mediated multidrug resistance in breast cancer by inducing partial epithelial-mesenchymal transition. *Cell Biol Int* (2021) 45:1644–53. doi: 10.1002/cbin.11598
120. Pastushenko I, Brisebarre A, Sifrim A, Fioramonti M, Revenco T, Boumahdi S, et al. Identification of the tumour transition states occurring during EMT. *Nature* (2018) 556:463–8. doi: 10.1038/s41586-018-0040-3
121. Liu D, Hong Y, Li Y, Hu C, Yip TC, Yu WK, et al. Targeted destruction of cancer stem cells using multifunctional magnetic nanoparticles that enable combined hyperthermia and chemotherapy. *Theranostics* (2020) 10:1181–96. doi: 10.7150/thno.38989
122. Scheel C, Eaton EN, Li SH, Chaffer CL, Reinhardt F, Kah KJ, et al. Paracrine and autocrine signals induce and maintain mesenchymal and stem cell states in the breast. *Cell* (2011) 145:926–40. doi: 10.1016/j.cell.2011.04.029
123. da Silva-Diz V, Lorenzo-Sanz L, Bernat-Peguera A, Lopez-Cerda M, Muñoz P. Cancer cell plasticity: Impact on tumor progression and therapy response. *Semin Cancer Biol* (2018) 53:48–58. doi: 10.1016/j.semcancer.2018.08.009
124. Hernández-Camarero P, Jiménez G, López-Ruiz E, Barungi S, Marchal JA, Perán E. Revisiting the dynamic cancer stem cell model: Importance of tumour edges. *Crit Rev Oncol Hematol* (2018) 131:35–45. doi: 10.1016/j.critrevonc.2018.08.004
125. Ni T, Li XY, Lu N, An T, Liu ZP, Fu R, et al. Snail1-dependent p53 repression regulates expansion and activity of tumour-initiating cells in breast cancer. *Nat Cell Biol* (2016) 18:1221–32. doi: 10.1038/ncb3425
126. Sorre B, Warmflash A, Brivanlou AH, Siggia ED. Encoding of temporal signals by the TGF- β pathway and implications for embryonic patterning. *Dev Cell* (2014) 30:334–42. doi: 10.1016/j.devcel.2014.05.022
127. Peitzsch C, Tyutyunnykova A, Pantel K, Dubrovskaya A. Cancer stem cells: The root of tumor recurrence and metastases. *Semin Cancer Biol* (2017) 44:10–24. doi: 10.1016/j.semcancer.2017.02.011
128. Malfettone A, Soukupova J, Bertran E, Crosas-Molist E, Lastra R, Fernando J, et al. Transforming growth factor- β -induced plasticity causes a migratory stemness phenotype in hepatocellular carcinoma. *Cancer Lett* (2017) 392:39–50. doi: 10.1016/j.canlet.2017.01.037
129. Kröger C, Afeyan A, Mraz J, Eaton EN, Reinhardt F, Khodor YL, et al. Acquisition of a hybrid E/M state is essential for tumorigenicity of basal breast cancer cells. *Proc Natl Acad Sci U.S.A.* (2019) 116:7353–62. doi: 10.1073/pnas.1812876116
130. Kim J, Villadsen R, Sørlie T, Fogh L, Grønlund SZ, Fridriksdottir AJ, et al. Tumor initiating but differentiated luminal-like breast cancer cells are highly invasive in the absence of basal-like activity. *Proc Natl Acad Sci U.S.A.* (2012) 109:6124–9. doi: 10.1073/pnas.1203203109
131. Liu S, Cong Y, Wang D, Sun Y, Deng L, Liu Y, et al. Breast cancer stem cells transition between epithelial and mesenchymal states reflective of their normal counterparts. *Stem Cell Rep* (2014) 2:78–91. doi: 10.1016/j.stemcr.2013.11.009
132. Bierie B, Pierce SE, Kroeger C, Stover DG, Pattabiraman DR, Thiru P, et al. Integrin- $\beta 4$ identifies cancer stem cell-enriched populations of partially mesenchymal carcinoma cells. *Proc Natl Acad Sci U.S.A.* (2017) 114:E2337–e2346. doi: 10.1073/pnas.1618298114
133. Pei YY, Li GC, Ran J, Wan XH, Wei FX, Wang L. Kinesin family member 11 enhances the self-renewal ability of breast cancer cells by participating in the wnt/ β -catenin pathway. *J Breast Cancer* (2019) 22:522–32. doi: 10.4048/jbc.2019.22.e51
134. Wang Y, Zhong Y, Hou T, Liao J, Zhang C, Sun C, et al. PM2.5 induces EMT and promotes CSC properties by activating notch pathway *in vivo* and *in vitro*. *Ecotoxicol Environ Saf* (2019) 178:159–67. doi: 10.1016/j.ecoenv.2019.03.086
135. Engler A, Rolando C, Giachino C, Saotome I, Erni A, Brien C, et al. Notch2 signaling maintains NSC quiescence in the murine ventricular-subventricular zone. *Cell Rep* (2018) 22:992–1002. doi: 10.1016/j.celrep.2017.12.094
136. Zhang Y, Ma Y, Zhao C, Zhang H, Pu Y, Yin L. Synergistic carcinogenesis of HPV18 and MNNG in het-1A cells through p62-KEAP1-NRF2 and PI3K/AKT/mTOR pathway. *Oxid Med Cell Longev* (2020) 2020:6352876. doi: 10.1155/2020/6352876
137. Bao S, Wu Q, McLendon RE, Hao Y, Shi Q, Hjelmeland AB, et al. Glioma stem cells promote radioresistance by preferential activation of the DNA damage response. *Nature* (2006) 444:756–60. doi: 10.1038/nature05236
138. Levina V, Marrangoni AM, DeMarco R, Gorelik E, Lokshin AE. Drug-selected human lung cancer stem cells: cytokine network, tumorigenic and metastatic properties. *PLoS One* (2008) 3:e3077. doi: 10.1371/journal.pone.0003077
139. Dallas NA, Xia L, Fan F, Gray MJ, Gaur P, Buren Gv2, et al. Chemoresistant colorectal cancer cells, the cancer stem cell phenotype, and

increased sensitivity to insulin-like growth factor-I receptor inhibition. *Cancer Res* (2009) 69:1951–7. doi: 10.1158/0008-5472.CAN-08-2023

140. Liu Y, Yang M, Luo J, Zhou H. Radiotherapy targeting cancer stem cells "awakens" them to induce tumour relapse and metastasis in oral cancer. *Int J Oral Sci* (2020) 12:19. doi: 10.1038/s41368-020-00087-0
141. Zhang Z, Qin S, Chen Y, Zhou L, Yang M, Tang Y, et al. Inhibition of NPC1L1 disrupts adaptive responses of drug-tolerant persister cells to chemotherapy. *EMBO Mol Med* (2022) 14:e14903. doi: 10.15252/emmm.202114903
142. Guler GD, Tindell CA, Pitti R, Wilson C, Nichols K, KaiWai Cheung T, et al. Repression of stress-induced LINE-1 expression protects cancer cell subpopulations from lethal drug exposure. *Cancer Cell* (2017) 32:221–237.e213. doi: 10.1016/j.ccell.2017.07.002
143. Rehman SK, Haynes J, Collignon E, Brown KR, Wang Y, Nixon AML, et al. Colorectal cancer cells enter a diapause-like DTP state to survive chemotherapy. *Cell* (2021) 184:226–242.e221. doi: 10.1016/j.cell.2020.11.018
144. Aldonza MBD, Ku J, Hong JY, Kim D, Yu SJ, Lee MS, et al. Prior acquired resistance to paclitaxel relays diverse EGFR-targeted therapy persistence mechanisms. *Sci Adv* (2020) 6(6):eaa7416. doi: 10.1126/sciadv.aav7416
145. Sharma SV, Lee DY, Li B, Quinlan MP, Takahashi F, Maheswaran S, et al. A chromatin-mediated reversible drug-tolerant state in cancer cell subpopulations. *Cell* (2010) 141:69–80. doi: 10.1016/j.cell.2010.02.027
146. Echeverria GV, Ge Z, Seth S, Zhang X, Jeter-Jones S, Zhou X, et al. Resistance to neoadjuvant chemotherapy in triple-negative breast cancer mediated by a reversible drug-tolerant state. *Sci Transl Med* (2019) 11(488):eaav0936. doi: 10.1126/scitranslmed.aav0936
147. Roesch A, Vultur A, Bogeski I, Wang H, Zimmermann KM, Speicher D, et al. Overcoming intrinsic multidrug resistance in melanoma by blocking the mitochondrial respiratory chain of slow-cycling JARID1B(high) cells. *Cancer Cell* (2013) 23:811–25. doi: 10.1016/j.ccr.2013.05.003
148. Visvader JE, Lindeman GJ. Cancer stem cells in solid tumours: accumulating evidence and unresolved questions. *Nat Rev Cancer* (2008) 8:755–68. doi: 10.1038/nrc2499
149. Chen J, Li Y, Yu TS, McKay RM, Burns DK, Kernie SG, et al. A restricted cell population propagates glioblastoma growth after chemotherapy. *Nature* (2012) 488:522–6. doi: 10.1038/nature11287
150. Maynard A, McCoach CE, Rotow JK, Harris L, Haderk F, Kerr DL, et al. Therapy-induced evolution of human lung cancer revealed by single-cell RNA sequencing. *Cell* (2020) 182:1232–1251.e1222. doi: 10.1016/j.cell.2020.07.017
151. Shiozawa Y, Pedersen EA, Patel LR, Ziegler AM, Havens AM, Jung Y, et al. GAS6/AXL axis regulates prostate cancer invasion, proliferation, and survival in the bone marrow niche. *Neoplasia* (2010) 12:116–27. doi: 10.1593/neo.91384
152. Flem-Karlsen K, McFadden E, Omar N, Haugen MH, Øy GF, Ryder T, et al. Targeting AXL and the DNA damage response pathway as a novel therapeutic strategy in melanoma. *Mol Cancer Ther* (2020) 19:895–905. doi: 10.1158/1535-7163.MCT-19-0290
153. Raouf S, Mulford IJ, Frisco-Cabanos H, Nangia V, Timonina D, Labrot E, et al. Targeting FGFR overcomes EMT-mediated resistance in EGFR mutant non-small cell lung cancer. *Oncogene* (2019) 38:6399–413. doi: 10.1038/s41388-019-0887-2
154. Du X, Qi F, Lu S, Li Y, Han W. Nicotine upregulates FGFR3 and RB1 expression and promotes non-small cell lung cancer cell proliferation and epithelial-to-mesenchymal transition via downregulation of miR-99b and miR-192. *BioMed Pharmacother* (2018) 101:656–62. doi: 10.1016/j.biopha.2018.02.113
155. Corbet C, Bastien E, Santiago de Jesus JP, Dierge E, Martherus R, Vander Linden C, et al. TGFβ2-induced formation of lipid droplets supports acidosis-driven EMT and the metastatic spreading of cancer cells. *Nat Commun* (2020) 11:454. doi: 10.1038/s41467-019-14262-3
156. Oren Y, Tsabar M, Cuoco MS, Amir-Zilberstein L, Cabanos HF, Hütter JC, et al. Cycling cancer persister cells arise from lineages with distinct programs. *Nature* (2021) 596:576–82. doi: 10.1038/s41586-021-03796-6
157. Russi S, Verma HK, Laurino S, Mazzone P, Storto G, Nardelli A, et al. Adapting and surviving: Intra and extra-cellular remodeling in drug-resistant gastric cancer cells. *Int J Mol Sci* (2019) 20(15):3736. doi: 10.3390/ijms20153736
158. Bugde S, Gonugunta VK, Penugurti V, Malisetty VL, Vadlamudi RK, Manavathi B. HPIP promotes epithelial-mesenchymal transition and cisplatin resistance in ovarian cancer cells through PI3K/AKT pathway activation. *Cell Oncol (Dordr)* (2017) 40:133–44. doi: 10.1007/s13402-016-0308-2
159. Yao Y, Pang T, Cheng Y, Yong W, Kang H, Zhao Y, et al. Positive correlative over-expression between eIF4E and snail in nasopharyngeal carcinoma promotes its metastasis and resistance to cisplatin. *Pathol Oncol Res* (2020) 26:1639–49. doi: 10.1007/s12253-019-00733-x
160. Han X, Zhou Y, You Y, Lu J, Wang L, Hou H, et al. TET1 promotes cisplatin-resistance via demethylating the vimentin promoter in ovarian cancer. *Cell Biol Int* (2017) 41:405–14. doi: 10.1002/cbin.10734
161. Mrkvicova A, Chmelarova M, Peterova E, Havelek R, Baranova I, Kazimirova P, et al. The effect of sodium butyrate and cisplatin on expression of EMT markers. *PLoS One* (2019) 14:e0210889. doi: 10.1371/journal.pone.0210889
162. Shi Q, Chen Q, Zhou Z, Zheng X, Huang X, Fang M, et al. Hypoxia-induced antizyme inhibitors 2 regulates cisplatin resistance through epithelial-mesenchymal transition pathway in non-small cell lung cancer. *Pulm Pharmacol Ther* (2021) 69:102048. doi: 10.1016/j.pupt.2021.102048
163. Kim TW, Lee SY, Kim M, Cheon C, Jang BH, Shin YC, et al. DSGOST regulates resistance via activation of autophagy in gastric cancer. *Cell Death Dis* (2018) 9:649. doi: 10.1038/s41419-018-0658-y
164. Hu C, Li M, Guo T, Wang S, Huang W, Yang K, et al. Anti-metastasis activity of curcumin against breast cancer via the inhibition of stem cell-like properties and EMT. *Phytomedicine Int J Phytotherapy Phytopharmacology* (2019) 58:152740. doi: 10.1016/j.phymed.2018.11.001
165. Li R, Wang Y, Liu Y, Chen Q, Fu W, Wang H, et al. Curcumin inhibits transforming growth factor-β1-induced EMT via PPARγ pathway, not smad pathway in renal tubular epithelial cells. *PLoS One* (2013) 8:e58848. doi: 10.1371/journal.pone.0058848
166. Gallardo M, Calaf GM. Curcumin and epithelial-mesenchymal transition in breast cancer cells transformed by low doses of radiation and estrogen. *Int J Oncol* (2016) 48:2534–42. doi: 10.3892/ijo.2016.3477
167. Zhao H, Yang Z, Wang X, Zhang X, Wang M, Wang Y, et al. Triptolide inhibits ovarian cancer cell invasion by repression of matrix metalloproteinase 7 and 19 and upregulation of e-cadherin. *Exp Mol Med* (2012) 44:633–41. doi: 10.3858/emmm.2012.44.11.072
168. Wang H, Zhang H, Tang L, Chen H, Wu C, Zhao M, et al. Resveratrol inhibits TGF-β1-induced epithelial-to-mesenchymal transition and suppresses lung cancer invasion and metastasis. *Toxicology* (2013) 303:139–46. doi: 10.1016/j.tox.2012.09.017
169. Yamashita N, Tokunaga E, Iimori M, Inoue Y, Tanaka K, Kitao H, et al. Epithelial paradox: Clinical significance of coexpression of e-cadherin and vimentin with regard to invasion and metastasis of breast cancer. *Clin Breast Cancer* (2018) 18:e1003–9. doi: 10.1016/j.clbc.2018.02.002
170. Ribeiro AS, Paredes J. P-cadherin linking breast cancer stem cells and invasion: A promising marker to identify an "Intermediate/Metastable" EMT state. *Front Oncol* (2014) 4:371. doi: 10.3389/fonc.2014.00371
171. Meredith SL, Bryant JL, Babur M, Riddell PW, Behrouzi R, Williams KJ, et al. Irradiation decreases the neuroendocrine biomarker pro-opiomelanocortin in small cell lung cancer cells *In vitro* and *In vivo*. *PLoS One* (2016) 11:e0148404. doi: 10.1371/journal.pone.0148404
172. Blondy S, David V, Verdier M, Mathonnet M, Perraud A, Christou N. 5-fluorouracil resistance mechanisms in colorectal cancer: From classical pathways to promising processes. *Cancer Sci* (2020) 111:3142–54. doi: 10.1111/cas.14532
173. Dong TT, Zhou HM, Wang LL, Feng B, Lv B, Zheng MH. Salinomycin selectively targets 'CD133+' cell subpopulations and decreases malignant traits in colorectal cancer lines. *Ann Surg Oncol* (2011) 18:1797–804. doi: 10.1245/s10434-011-1561-2
174. Qian W, Cai X, Qian Q, Zhang W, Wang D. Astragaloside IV modulates TGF-β1-dependent epithelial-mesenchymal transition in bleomycin-induced pulmonary fibrosis. *J Cell Mol Med* (2018) 22:4354–65. doi: 10.1111/jcmm.13725
175. Deng J, Bai X, Feng X, Ni J, Beretov J, Graham P, et al. Inhibition of PI3K/Akt/mTOR signaling pathway alleviates ovarian cancer chemoresistance through reversing epithelial-mesenchymal transition and decreasing cancer stem cell marker expression. *BMC Cancer* (2019) 19:618. doi: 10.1186/s12885-019-5824-9
176. Yao Y, Chen R, Wang G, Zhang Y, Liu F. Exosomes derived from mesenchymal stem cells reverse EMT via TGF-β1/Smad pathway and promote repair of damaged endometrium. *Stem Cell Res Ther* (2019) 10:225. doi: 10.1186/s13287-019-1332-8
177. Xiao K, He W, Guan W, Hou F, Yan P, Xu J, et al. Mesenchymal stem cells reverse EMT process through blocking the activation of NF-κB and hedgehog pathways in LPS-induced acute lung injury. *Cell Death Dis* (2020) 11:863. doi: 10.1038/s41419-020-03034-3