

Genome Sequence of *Vibrio campbellii* Strain UMTGB204, a Marine Bacterium Isolated from a Green Barrel Tunicate

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***Vibrio campbellii* strain UMTGB204 was isolated from a green barrel tunicate. The genome of this strain comprises 5,652,224 bp with 5,014 open reading frames, 9 rRNAs, and 116 tRNAs. It contains genes related to virulence and environmental tolerance. Gene clusters for the biosynthesis of nonribosomal peptides and bacteriocin were also identified.**

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Vibrio campbellii is widely distributed in marine environments (1). This gammaproteobacteria is an important pathogen to many aquatic organisms from both wild and cultured systems, most notably penaeid shrimp, several fish species, and mollusks (2, 3). *Vibrio campbellii* strain UMTGB204 was isolated from a green barrel tunicate in Bidong Island, a coral reef island in the South China Sea. The genome of this strain was sequenced in order to gain insights into its relationships with tunicates.

V. campbellii strain UMTGB204 was cultured in Marine Broth 2216 (Difco). Genomic DNA was then extracted using the GF-1 nucleic acid extraction kit (Vivantis, Malaysia). Sequencing was performed on the Illumina HiSeq2000 platform, generating 33,271,099 raw FASTQ paired-end reads. Two million reads were subsampled for error correction and *de novo* assembled using SPAdes version 3.1.0 (4). The resulting contigs were used for scaffolding and then gap-closed using SSPACE version 2.0 and Gap-Filler version 1.11 (5, 6). Sixty gap-filled contigs >3 kb with an N_{50} of 947,033 bp were produced, and the total sequence length was 5,652,224 bp with 70× coverage.

The Prokka version 1.8 annotation pipeline, comprising Prodigal version 2.60, RNAmmer version 1.2, and Aragorn version 1.2.36, was used to annotate the genome, predicting 5,014 open reading frames, 9 rRNAs, and 116 tRNAs (7–10). The predicted 16S rRNA was queried with BLASTn (11) against the nucleotide collection database, confirming that the strain was *Vibrio campbellii*. Further validation of the species was performed using *in silico* genome-to-genome comparison of the UMTGB204 strain to *Vibrio campbellii* 200612B (GenBank accession no. BANY00000000.1), showing a DNA-DNA hybridization probability of 90.03% (12). InterProScan5 was used to provide additional annotation to the predicted protein sequences (13). Furthermore, antiSMASH was used to identify the presence of secondary metabolite biosynthesis gene clusters in the genome (14).

Similar to the genomes of other *Vibrio campbellii* strains, strain

UMTGB204 carries genes that are responsible for virulence—such as the hemolysin, ToxR, and secretion systems (15, 16)—but unlike genomes of other reported strains, the proteorhodopsin-related gene was not identified in the genome of strain UMTGB204. Analysis of the genome also revealed genes associated with environmental adaptation, including osmotic and oxidative stress, thermal shock, and siderophores. Interestingly, several gene clusters for the biosynthesis of nonribosomal peptides and bacteriocin have been identified (contigs 1, 2, 10, and 19). This suggests that, as a survival mechanism in the marine environment, strain UMTGB204 possesses a better ability to compete with other closely related vibrios to colonize the host tunicate.

Nucleotide sequence accession number. This whole-genome shotgun project has been deposited in DDBJ/EMBL/GenBank under the accession number [JSE000000000](https://www.ncbi.nlm.nih.gov/nuccore/JSE000000000).

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REFERENCES

1. Thompson FL, Iida T, Swings J. 2004. Biodiversity of vibrios. *Microbiol Mol Biol Rev* 68:403–431. <http://dx.doi.org/10.1128/MMBR.68.3.403-431.2004>.
2. Gomez-Gil B, Soto-Rodríguez S, García-Gasca A, Roque A, Vazquez-Juarez R, Thompson FL, Swings J. 2004. Molecular identification of *Vibrio harveyi*-related isolates associated with diseased aquatic organisms. *Microbiology* 150:1769–1777. <http://dx.doi.org/10.1099/mic.0.26797-0>.
3. Haldar S, Chatterjee S, Sugimoto N, Das S, Chowdhury N, Hinenoya A, Asakura M, Yamasaki S. 2011. Identification of *Vibrio campbellii* isolated from diseased farm-shrimps from south India and establishment of its pathogenic potential in an artemia model. *Microbiology* 157:179–188. <http://dx.doi.org/10.1099/mic.0.041475-0>.
4. Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV,

- Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J Comput Biol* 19:455–477. <http://dx.doi.org/10.1089/cmb.2012.0021>.
5. Boetzer M, Henkel CV, Jansen HJ, Butler D, Pirovano W. 2011. Scaffolding pre-assembled contigs using SSPACE. *Bioinformatics* 27: 578–579. <http://dx.doi.org/10.1093/bioinformatics/btq683>.
 6. Boetzer M, Pirovano W. 2012. Toward almost closed genomes with GapFiller. *Genome Biol* 13:R56. <http://dx.doi.org/10.1186/gb-2012-13-6-r56>.
 7. Seemann T. 2014. Prokka: rapid prokaryotic genome annotation. *Bioinformatics* 30:2068–2069. <http://dx.doi.org/10.1093/bioinformatics/btu153>.
 8. Hyatt D, Chen GL, Locascio PF, Land ML, Larimer FW, Hauser LJ. 2010. Prodigal: prokaryotic gene recognition and translation initiation site identification. *BMC Bioinformatics* 11:119. <http://dx.doi.org/10.1186/1471-2105-11-119>.
 9. Lagesen K, Hallin P, Rødland EA, Staerfeldt HH, Rognes T, Ussery DW. 2007. RNAmmer: consistent and rapid annotation of ribosomal RNA genes. *Nucleic Acids Res* 35:3100–3108. <http://dx.doi.org/10.1093/nar/gkm160>.
 10. Laslett D, Canback B. 2004. ARAGORN, a program to detect tRNA genes and tmRNA genes in nucleotide sequences. *Nucleic Acids Res* 32:11–16. <http://dx.doi.org/10.1093/nar/gkh152>.
 11. Altschul SF, Gish W, Miller W, Myers EW, Lipman DJ. 1990. Basic local alignment search tool. *J Mol Biol* 215:403–410. [http://dx.doi.org/10.1016/S0022-2836\(05\)80360-2](http://dx.doi.org/10.1016/S0022-2836(05)80360-2).
 12. Meier-Kolthoff JP, Auch AF, Klenk HP, Göker M. 2013. Genome sequence-based species delimitation with confidence intervals and improved distance functions. *BMC Bioinformatics* 14:60. <http://dx.doi.org/10.1186/1471-2105-14-60>.
 13. Jones P, Binns D, Chang HY, Fraser M, Li W, Mcanulla C, McWilliam H, Maslen J, Mitchell A, Nuka G, Pesseat S, Quinn AF, Sangrador-Vegas A, Scheremetjew M, Yong SY, Lopez R, Hunter S. 2014. InterProScan 5: genome-scale protein function classification. *Bioinformatics* 30:1236–1240. <http://dx.doi.org/10.1093/bioinformatics/btu031>.
 14. Medema MH, Blin K, Cimermanic P, de Jager V, Zakrzewski P, Fischbach MA, Weber T, Takano E, Breitling R. 2011. antiSMASH: rapid identification, annotation and analysis of secondary metabolite biosynthesis gene clusters in bacterial and fungal genome sequences. *Nucleic Acids Res* 39(suppl 2):W339–W346. <http://dx.doi.org/10.1093/nar/gkr466>.
 15. Dias GM, Thompson CC, Fishman B, Naka H, Haygood MG, Crosa JH, Thompson FL. 2012. Genome sequence of the marine bacterium *Vibrio campbellii* DS40M4, isolated from open ocean water. *J Bacteriol* 194:904. <http://dx.doi.org/10.1128/JB.06583-11>.
 16. Amaral GR, Silva BS, Santos EO, Dias GM, Lopes RM, Edwards RA, Thompson CC, Thompson FL. 2012. Genome sequence of the bacterioplanktonic, mixotrophic *Vibrio campbellii* strain pel22a, isolated in the Abrolhos Bank. *J Bacteriol* 194:2759–2760. <http://dx.doi.org/10.1128/JB.00377-12>.