

# The frequency of *POLE*-mutation in endometrial carcinoma and prognostic implications: a systemic review and meta-analysis

Alaa Salah Jumaah<sup>1</sup>, Mais Muhammed Salim<sup>1</sup>, Hawraa Sahib Al-Haddad<sup>2</sup>, Katherine Ann McAllister<sup>3</sup>, Akeel Abed Yasseen<sup>1</sup>

<sup>1</sup>Department of Pathology and Forensic Medicine, Faculty of Medicine, University of Kufa, Kufa;

<sup>2</sup>Al-Furat Al-Awsat Hospital, Kufa, Iraq;

<sup>3</sup>School of Biomedical Science, University of Ulster, Northern Ireland, UK

**Background:** Endometrial carcinoma (EC) is classified into four distinct molecular subgroups including ultramutated DNA polymerase epsilon (*POLE*). *POLE*-mutated tumors have the best prognosis and are a promising target for immunotherapy. This meta-analysis consolidated the reported variation of *POLE*-mutant frequency and assessed prognostic value in EC. **Methods:** Internet searches explored scientific data bases: EMBASE, PubMed, and the Cochrane Central Register of Controlled Trials databases. Data was extracted from eligible studies including: sample size, number of positive *POLE*-mutant cases, sequencing information, clinicopathologic data, and survival data. Meta-analysis and a random-effects model produced pooled estimates of *POLE* frequency and prognostic parameters using 95% confidence intervals (CI), hazard ratios (HR), and odd ratios (OR). **Results:** Six thousand three hundred and forty-six EC patient cases were pooled from 25 studies. The pooled proportion of *POLE* gene mutation in EC was 8.59% (95% CI, 7.01 to 10.32), of which 8.22% (95% CI, 6.27 to 10.42) were type I and 0.93% (95% CI, 0.34 to 1.81) type 2. Clinicopathologic data showed that *POLE*-mutated tumors are mostly endometrioid. They present at higher levels in earlier stages (I–II) of EC (89.51%; 95% CI, 81.11 to 95.66) at the highest grade III (51.53%; 95% CI, 36.08 to 66.84) with reduced myometrial invasion (OR, 1.48, 95% CI, 0.99 to 2.20). Survival analysis indicated favorable overall survival (HR, 0.90), disease-specific survival (HR, 0.41), and progression-free survival (HR, 0.23) for *POLE* mutant EC. **Conclusions:** Almost one-tenth of EC patients have *POLE*-mutated tumors. Given their improved prognostic potential, identifying the *POLE* mutation status is key for the management of EC patients.

**Key Words:** Endometrial carcinoma; *POLE* gene

Received: February 27, 2020 Revised: July 18, 2020 Accepted: July 23, 2020

**Corresponding Author:** Akeel Abed Yasseen, PhD, Department of Pathology and Forensic Medicine, Faculty of Medicine, University of Kufa, Kufa, P.O. Box 21, Najaf Governorate, Iraq

Tel: +96-47811131586, E-mail: akeelyasseen@uokufa.edu.iq

Endometrial carcinoma (EC) is the most common cancer of the female reproductive system. In the United States, EC was the sixth-cause of cancer-related death in 2018, accounting for 11,350 deaths. The disease affects an estimated 1 out of every 37 and 49 women respectively during their lifetime in the United States and Australia [1,2]. The outlook for 5-year survival after treatment ranges from 74% to 91% [3,4]. EC has been historically classified into estrogen dependent (type I) and estrogen-independent (type II) cancers [4]. Type II tumors are less common, more clinically aggressive, and may have serous, clear cell or undifferentiated histology. In contrast, type I tumors present in 70%–80% of cases, with endometrioid histology and a more favorable outcome. The updated classification of EC identifies

four molecular subtypes according to The Cancer Genomic Atlas (TCGA): “*POLE*-mutated (ultramutated), microsatellite unstable (hypermutated), copy number low (endometrioid), and copy number high (serous-like)” [1]. The *POLE*-ultramutated subgroup holds great promise for the outlook of EC patients. The tumors have a more favorable outcome, and are usually noted to be of endometrioid type and associated with lymphoid infiltrates [5].

The *POLE* gene encodes the catalytic subunit of DNA polymerase epsilon, which synthesizes the leading strand of replicating DNA. The epsilon polymerase recognizes and removes mispaired nucleotides using exonuclease activity. This proofreading capacity of epsilon enables high fidelity DNA replication [6]. Mutations can occur in the exonuclease domain of the polymerase,

within hotspot regions [7]. These genetic alterations inactivate or suppress the proofreading abilities of the polymerase, causing increased replicative error rates and the ultra-mutated phenotype. Studies show that ultra-mutated *POLE* EC harbors up to 10-fold more mutations than the microsatellite instability subgroup [6,7]. In the TCGA study [8], whole genome and exon sequencing of EC tumors uncovered the *POLE* hotspot mutations of v411L and P286R. Follow-up studies mainly used the Sanger method [9-23], next generation sequencing [13,24] and unspecified sequencing approaches [10,11,25-29] to target the exonuclease domain (exons 9–14) of *POLE*. The reported proportion of mutated *POLE* is highly variable [8-31]: ranging from absent in studies of clear cell carcinoma [15,18] to levels as high as 43% in one study of rare undifferentiated/dedifferentiated EC histotypes [28]. In order to consolidate the studies, we conducted a meta-analysis of reported *POLE* gene mutation in EC to confirm its overall frequency. We also extracted clinicopathologic and survival data, to evaluate how mutated *POLE* can affect prognosis of EC patients.

## MATERIALS AND METHODS

This study was conducted according to the guidelines of Preferred Reporting for Systemic Review and Meta-analysis (PRISMA) statement [32].

### Literature search strategy

Searches were conducted according to the guidelines of PRISMA statement 2009 [32]. Two authors (A.S.J. and H.S.A.) searched independently the following data bases from inception to October 2019: Embase, PubMed, Cochrane central Register of Controlled trials, and Ovid. The reference lists were also scanned within the articles. There were no language limits and international papers were translated. All pathology and oncology journals indexed in the Scimago directory were reviewed and relevant papers scanned (A.S.J. and M.M.S.). The following search terms were used:

- 1) 'Endometrial cancer' or 'uterine cancer.'
- 2) '*POLE* gene' or 'ultramutated endometrial carcinoma.'

### Inclusion criteria

The following patient inclusion criteria were used:

- 1) EC or one of its histological variants was present in patients.
- 2) The expression of *POLE* gene was reported using genetic testing (e.g. sequencing, Sanger sequencing, next generation sequencing, polymerase chain reaction).

3) A full paper was published and studies published in abstract format only were excluded.

4) When similar studies were generated from the same patient, only the most recent investigation was included.

### Study selection

Studies were identified using different data bases. The title of the paper and abstract were assessed by two independent authors (A.S.J. and H.S.A.). The full texts were also reviewed independently by two authors (M.M.S. and K.A.M.). Any disagreement was resolved under guidance of the senior author (A.A.Y.).

### Data collection

The following data was extracted from eligible studies by two authors (A.S.J. and H.S.A.): study information (first author and year of publication), patient characteristics (sample size and gender), site of the study, and test method for *POLE* gene and proportion detected. Clinicopathologic data extracted included tumor stage and grade, presence of lymphovascular and myometrial invasion, and patient survival. If the relevant data was not available, it was recorded as NR (not reported). All datasets were checked independently (M.M.S.). Any disagreements were resolved by discussion and consultation with the senior author (A.A.Y.).

### Meta-analysis and statistical methods

The proportion of endometrial cancers that harbor *POLE* gene mutations was calculated using medcalc software [33]. The pooled proportion of *POLE* was calculated using the random effect model [32] for meta-analysis. For clinicopathologic meta-analysis, proportions of tumour stage, grade, lymphovascular invasion, myometrial invasion (MI), and survival analysis (overall survival, disease-free survival, and progression-free survival) were pooled from each study. The variation between datasets was assessed using the heterogeneity test with inconsistency index ( $I^2$ ) and Q statistic. The level of study heterogeneity was considered low at 25% ( $I^2 = 25%$ ), medium at 50% ( $I^2 = 50%$ ) or high at 75% ( $I^2 = 75%$ ). In regard to the Q statistic, a p-value of less than 0.1 was considered to represent significant heterogeneity. The possibility of publication bias was assessed by visual method using a funnel plot. This determined funnel plot asymmetry resulting from factors such as non-publication of studies with negative results.

### Sensitivity and subgroup analysis

Lastly, sensitivity analysis was conducted by omitting each study one-by-one to discover its contribution on the pooled meta-analysis results. Subgroup analysis was performed according to

geographical area (Asia, West-Europe, and America) and to different histological types to discover sources of heterogeneity. The subgroup analysis was further extended by using a meta regression model.

## RESULTS

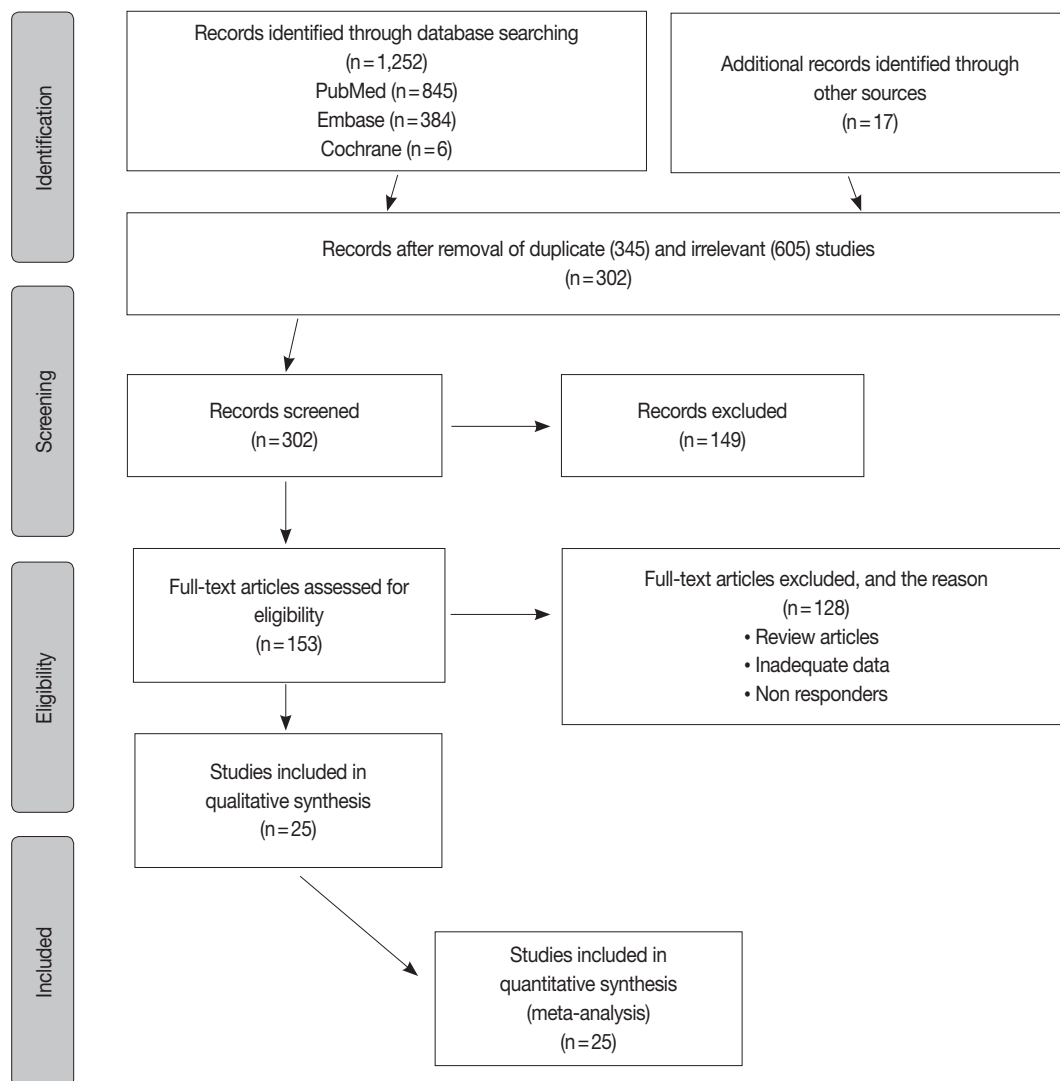
### Study selection for meta-analysis

The inclusion criteria were met by 1,252 studies, of which 345 studies were excluded for duplication, and 605 studies were excluded as irrelevant by reviewing the title and abstract. The full texts of the remaining 302 studies were considered for review. Approximately 25 studies were left for the final meta-analysis as illustrated in the flowchart diagram (Fig. 1). All of the studies

were published from 2013–2019, and there were no unpublished studies of relevance. The study characteristics and *POLE* gene sequencing methods are listed in (Table 1). The majority used Sanger sequencing to detect tumor mutations in exons 9–14 of the *POLE* exonuclease domain (Table 1) [8-29].

### Meta-analysis of EC

There were 6,346 EC patient cases pooled from the 25 studies and investigated for *POLE* gene tumor alterations. All studies (except one) were performed in western countries. The studies were published in English and full texts obtained. There was significant heterogeneity between the studies ( $Q = 109.57$ ,  $I^2 = 78.10\%$ ; 95% confidence interval [CI] for  $I^2$ , 68.15 to 84.94;  $p < .001$ ). The random effect model was used for final meta-analysis because



**Fig. 1.** Flow chart of the studies identified, screened and included for final meta-analysis. A total of 1,252 studies met the eligibility criteria for inclusion, of which 26 were selected for final meta-analysis.

**Table 1.** Study characteristics and *POLE* sequencing methodology

Study	EC cohort size	Proportion <i>POLE</i> -mutant	Country	Sequencing method	Location of exonuclease mutations
The Cancer Genome Atlas Research Network (2013) [8]	248	17	USA	Exome sequencing	Hotspots: Pro286Arg and Val411Leu
Auguste et al. (2018) [9]	102	9	Canada and Europe	Sanger sequencing	Exons 9 and 13
Talhok et al. (2015) [10]	143	12	Canada	Fluidigm-MiSeq and sanger sequencing	Exons 9–14
Stelloo et al. (2015) [11]	116	14	Europe and Australia	Sanger sequencing	Exons 9 and 13
Eggink et al. (2017) [12]	116	15	Europe and Australia	Sanger sequencing	Exons 9, 13 and 14
Wortman et al. (2018) [25]	344	16	Netherlands	Sequencing	Not reported
Kommoss et al. (2018) [26]	452	42	Germany	Sequencing	Exons 9–14
Bosse et al. (2018) [13]	376	48	USA, Canada, and Europe	Sanger or next-generation approaches	Hotspots in the exonuclease domain (exons 9–14)
Billingsley et al. (2015) [14]	535	30	USA	Sanger sequencing	Residues 268–471
Le Gallo et al. (2017) [15]	63	0	USA and Europe	Sanger sequencing	Not reported
Karnezis et al. (2017) [27]	460	42	Canada	Sequencing	Not reported
Talhok et al. (2017) [16]	319	30	Canada	Sanger sequencing	Exons 9–14
Rosa-Rosa et al. (2016) [17]	18	2	USA and Europe	Sanger sequencing	Exons 9 and 13
Wortman et al. (2018) [25]	416	16	Netherlands	Sequencing	Not reported
Espinosa et al. (2017) [28]	21	9	Spain	Sequencing	Exons 9 to 14
Stelloo et al. (2015) [11]	116	14	Europe	Sanger sequencing	Exons 9 and 13
Hoang et al. (2015) [18]	14	0	Canada	Sanger sequencing	Exons 9–14
DeLair et al. (2017) [29]	30	2	USA	Sequencing	Exons 9–14
Abdulfatah et al. (2019) [19]	60	2	USA	Sanger sequencing	Exons 9 and 13
Wong et al. (2016) [24]	47	14	Singapore	Next generation sequencing	Exons 9–14
Stelloo et al. (2016) [20]	834	49	Netherlands	Sanger sequencing	Exons 9 and 13
Imboden et al. (2019) [21]	599	38	Sweden	Sanger sequencing	Exons 9–14
Church et al. (2015) [7]	788	48	Europe	Sanger sequencing	Exons 9 and 13
Billingsley et al. (2016) [22]	72	7	USA	Sanger sequencing	Residues 268–471
Talhok et al. (2016) [23]	57	10	USA and Canada	Ultra-deep MiSeq or sanger sequencing	Exons 9–14

*POLE*, DNA polymerase epsilon; EC, endometrial carcinoma.

**Table 2.** The association between *POLE* mutated EC and clinicopathology characteristics

Clinicopathological characteristics in EC	Pooled % portion (95% CI, %)	No. of studies	I <sup>2</sup> (95% CI)	p-value	Model
Overall <i>POLE</i> mutation	8.59 (7.01–10.32)	25	78.10 (68.15–84.94)	<.001	Random effect
<i>POLE</i> mutation in type I	8.22 (6.27–10.42)	9	74.88 (51.43–87.00)	<.001	Random effect
<i>POLE</i> mutation in type II	0.93 (0.34–1.81)	10	75.32 (54.08–86.74)	<.001	Random effect
Stage I–II	89.51 (81.11–95.66)	10	69.09 (40.43–83.96)	<.001	Random effect
Stage III–IV	14.77 (5.99–26.59)	7	65.96 (23.79–84.79)	<.001	Random effect
Grade I–II	46.36 (30.66–62.43)	7	82.15 (64.34–91.06)	<.001	Random effect
Grade III	51.53 (36.08–66.84)	8	81.79 (65.23–90.46)	<.001	Random effect
Lymphovascular invasion	31.11 (10.44–56.86)	8	93.34 (89.15–95.91)	<.001	Random effect
Myometrial invasion less than 50%	49.90 (43.71–56.21)	7	22.10 (0.00–65.16)	.260	Fixed effect

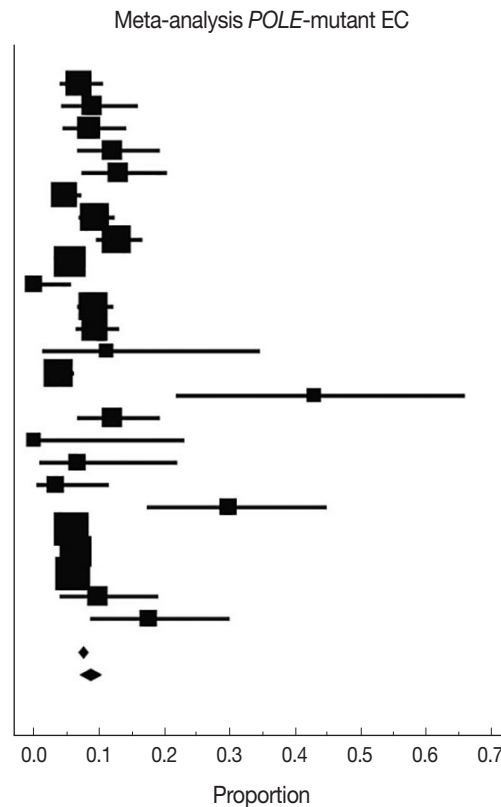
*POLE*, DNA polymerase epsilon; EC, endometrial carcinoma; CI, confidence interval.

of the significant heterogeneity. The pooled proportion of *POLE* gene mutated in EC was determined at 8.59% (95% CI, 7.01 to 10.32) as shown in Table 2. A forest plot representation of the EC patient cases comprising each study included for meta-analysis is shown in Fig. 2. Publication bias was assessed by funnel plot (Fig. 3) and the visual assessment was symmetrical.

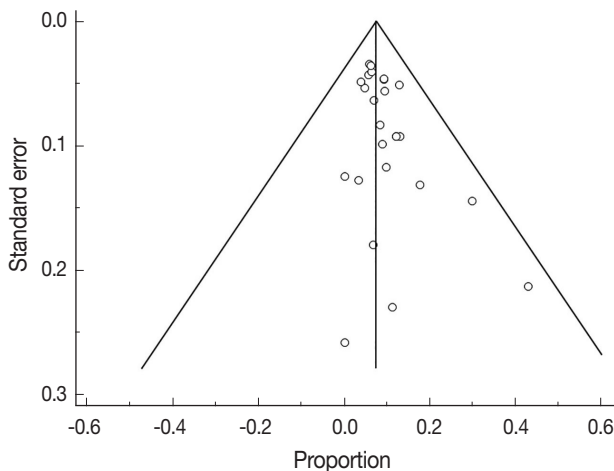
#### Analysis of EC histologic variants type I and type II

There were 3,363 patients included for the type 1 meta-analysis using a total of 9 studies (Supplementary Fig. S1). The mutated *POLE* gene proportion in type I EC was 8.22% (95% CI, 7.01 to 10.32). There was significant heterogeneity between studies (I<sup>2</sup> inconsistency = 74.88%; 95% CI, 51.43 to 87.00; p < .001) as shown in Table 2. A total of 3,423 patients with EC type II

The Cancer Genome Atlas Research Network (2013) [8]  
 Auguste et al. (2018) [9]  
 Talhouk et al. (2015) [10]  
 Stelloo et al. (2015) [11]  
 Eggink et al. (2017) [12]  
 Wortman et al. (2018) [25]  
 Kommos et al. (2018) [26]  
 Bosse et al. (2018) [13]  
 Billingsley et al. (2015) [14]  
 Le Gallo et al. (2017) [15]  
 Karnezis et al. (2017) [27]  
 Talhouk et al. (2017) [16]  
 Rosa-Rosa et al. (2016) [17]  
 Wortman et al. (2018) [25]  
 Espinosa et al. (2017) [28]  
 Stelloo et al. (2015) [11]  
 Hoang et al. (2015) [18]  
 DeLair et al. (2017) [29]  
 Abdulfatah et al. (2019) [19]  
 Wong et al. (2016) [24]  
 Stelloo et al. (2016) [20]  
 Imboden et al. (2019) [21]  
 Church et al. (2015) [7]  
 Billingsley et al. (2016) [22]  
 Talhouk et al. (2016) [23]  
 Total (fixed effects)  
 Total (random effects)



**Fig. 2.** Forest plot representation of the endometrial carcinoma (EC) patient cases for each study included for meta-analysis. *POLE*, DNA polymerase epsilon.



**Fig. 3.** Funnel plot for cases included for analysis. The visual assessment was symmetrical indicating no publication bias.

were obtained from 10 studies for the final meta-analysis (Supplementary Fig. S2). The histological types in EC type II are shown in (Supplementary Table S1). The pooled proportion of mutated *POLE* gene in type II was 0.93% (95% CI, 0.34 to 1.81) with a high  $I^2$  value (75.32%; 95% CI, 54.08 to 86.74;  $p < .001$ ).

### Subgroup analysis (country of origin)

In order to explore the role of heterogeneity, a subgroup analysis was performed according to the study site (Supplementary Fig. S3, S4). The studies were separated into two groups according to geographical area (USA and Canada vs. European countries). The heterogeneity was higher ( $I^2$ , inconsistency) for *POLE* genetic analysis in the European countries (78.41%; 95% CI, 59.27 to 88.55) compared to the USA and Canada (37.41%; 95% CI, 0.00 to 69.22).

### Sensitivity analysis

There were 2 identified study outliers [24,28]. Meta-analysis was performed after outlier exclusion. There was still significant heterogeneity ( $I^2 = 70.39%$  [95% CI for  $I^2$ , 54.78 to 80.61;  $p < .001$ ]) and exclusion did not significantly affect the final pooled proportion (pooled proportion = 7.76 [95% CI, 6.45 to 9.18]) (Supplementary Fig. S5). There were also two studies without *POLE* gene mutated EC [15,18]. Here, the pooled proportion was re-calculated using the random effect model (Supplementary Fig. S6). Significant heterogeneity was still present ( $I^2 = 76.31%$ ; 95% CI for  $I^2$ , 64.98 to 83.97;  $p < .001$ ). There were

10 studies [9,13,15,17,19–21,23,24,30] identified with a low sample size (< 100). These cases were excluded and the pooled proportions re-estimated (Supplementary Fig. S7). There was still significant heterogeneity ( $I^2 = 74.41\%$ ; 95% CI for  $I^2$ , 53.56 to 85.90;  $p < .001$ ).

### Meta-regression

Meta-regression was performed to identify the source of heterogeneity using the study country of origin and the *POLE* gene detection method (Supplementary Table S2). There was increased heterogeneity when considering the site of study performance in the European countries ( $I^2 = 78.41\%$ ) compared to United States and Canadian studies ( $I^2 = 37.41\%$ ).

### Clinicopathological characteristics

Clinicopathologic data was extracted from eligible studies of *POLE* mutant EC for meta-analysis (Supplementary Fig. S8–S14). The pooled proportions for stage, grade, lymphovascular invasion (LVI) and MI parameter are reported in Table 2. High heterogeneity was noted for pooled stage, grade, and LVI data. The pooled odd ratios were also calculated for *POLE*-mutant versus wild type *POLE* according to each clinicopathologic variable (Table 3).

### Tumor stage and grade in *POLE* mutant EC

The pooled proportion of mutant *POLE* presented at high levels of 89.51% (95% CI, 81.11 to 95.66) at the earliest EC stages of I–II (Table 2). This reduced to 14.77% by stages III–IV (95% CI, 5.99 to 26.59). The pooled odd ratio of stage I–II *POLE* mutant EC versus wild type was 3.72 (2.06 to 6.73), while stage III–IV was 0.26 (95% CI, 0.14 to 0.49) (Table 2, Supplementary Fig. S15, S16). The pooled collective proportions of grade I–II *POLE* mutant tumors was lower at 46.36% (95% CI, 30.66 to 62.43) compared to 51.53 of grade III (95% CI, 36.08 to 66.844) as shown in Table 2. The pooled odd ratio of grade I–II

*POLE* mutant EC versus wild type tumors was 0.40 (95% CI, 0.29 to 0.54) (Supplementary Fig. S17) with low heterogeneity ( $I^2 = 3.95\%$ , 95% CI, 0.00 to 69.18). Therefore, *POLE* mutated tumors present with a higher grade but at lower stage than wild type *POLE* mutant EC.

### LVI and MI in *POLE*-mutant EC

The pooled proportion of LVI was 31.11% (95% CI, 10.44 to 56.86). The pooled odd ratio of LVI positive in *POLE* mutant EC vs. wild type EC was 0.92 (0.643 to 1.34) (Table 3). The pooled proportion of MI either less or greater than 50% of myometrium in *POLE*-mutant tumor was equal at 49.90% (95% CI, 43.71 to 56.21) and 49.05% (95% CI, 39.17 to 58.98), respectively. Heterogeneity was low for pooled MI data (< 50% myometrium) at 22.10%; 95% CI, 0.00 to 65.16 (Table 2, Supplementary Figs. S13, S14). The pooled odd ratio of MI < 50% in *POLE* mutant EC versus MI < 50% in wild type *POLE* EC was 1.481 (95% CI, 0.99 to 2.20) with 47.63% heterogeneity (95% CI, 0.00 to 79.24) (Table 3). Overall, these findings imply that *POLE* mutant EC tumors have reduced ability to progress to myometrial invasion, which is an important prognostic finding.

### Endometrioid and non-endometrioid histologic types in *POLE* mutant EC

*POLE* mutant tumors were found to mainly present with endometrioid histologic type. The pooled proportions of type I and type II EC are shown in Table 2. The pooled odd ratio of endometrioid (type I) *POLE* mutant EC vs. endometrioid (type I) in wild type *POLE* EC was 1.72 (95% CI: 1.11 to 2.66), with very low heterogeneity ( $I^2 = 0.00\%$ ; 95% CI: 0.00 to 68.45) (Table 3, Supplementary Fig. S18).

### Survival analysis

The studies used for survival meta-analysis are listed in Table 4. Survival analysis was expressed using overall survival (OS),

**Table 3.** Pooled odd ratio of clinicopathology variables in *POLE*-mutant EC vs. wild type

Clinicopathology: <i>POLE</i> -mutant vs. wild type	Pooled odd ratio (95% CI)	No. of studies	$I^2$ (95% CI, %)	p-value for $I^2$	Model
Stage I–II EC	3.727 (2.063–6.732)	8	0.00 (0.00–25.07)	.890	Fixed effect
Stage III–IV EC	0.269 (0.147–0.494)	7	0.00 (0.00–53.51)	.716	Fixed effect
Grade I–II EC	0.400 (0.295–0.542)	8	3.95 (0.00–69.18)	.399	Fixed effect
Grade III EC	2.246 (1.655–3.048)	8	0.00 (0.00–29.91)	.865	Fixed effect
LVI	0.929 (0.643–1.341)	8	6.95 (0.00–70.15)	.376	Fixed effect
MI less than 50%	1.481 (0.996–2.202)	6	47.63 (0.00–79.24)	.089	Random effect
Type I endometrioid histology	1.721 (1.113–2.662)	9	0.00 (0.00–68.45)	.486	Fixed effect

*POLE*, DNA polymerase epsilon; EC, endometrial carcinoma; CI, confidence interval; LVI, lymphovascular invasion; MI, myometrial invasion.

**Table 4.** Survival analysis in *POLE* mutated EC

Study	OS estimated HR (95% CI)	DSS estimated HR (95% CI)	PFS estimated HR (95% CI)	Survival analysis test	Method
Talhok et al. (2017) [16]	1.01 (0.29–3.42)	0.42 (0.30–0.57)	-	Multivariable analysis	Kaplan-Meier survival analysis
Talhok et al. (2015) [10]	0.17 (0.01–1.98)	0.170 (0.01–1.99)	-	Multivariable analysis	Kaplan-Meier with log-rank significance testing and Cox proportional hazard regression models
Church et al. (2015) [7]	1.06 (0.58–1.91)	0.19 (0.02–1.31)	-	Multivariable analysis	Kaplan-Meier method and compared by the log-rank test
Karnezis et al. (2017) [27]	0.59 (0.21–1.60)	0.49 (0.12–1.90)	0.26 (0.04–1.49)	Univariable survival analysis	Kaplan-Meier survival curve
Stelloo et al. (2016) [20]	1.10 (0.39–3.10)			Multivariable analysis	Kaplan-Meier survival analysis
Bosse et al. (2018) [13]			0.23 (0.06–0.76)	Multivariable analysis	Kaplan-Meier survival curve
Pooled HR (95% CI)	0.90 (0.59–1.38)	0.41 (0.30–0.55)	0.23 (0.08–0.64)		
I <sup>2</sup> (95% CI, %)	0.00 (0.00–73.28)	0.00 (0.00–67.34)	0.00 (0.00–0.00)		

*POLE*, DNA polymerase epsilon; EC, endometrial carcinoma; OS, overall survival; DSS, disease-specific survival; PFS, progression-free survival; HR, hazard ratio; CI, confidence interval.

disease-specific survival (DSS), and progression-free survival (Supplementary Fig. S19–S21). All the survival parameters had a hazard ratio (HR) of less than 1. The estimated HR for OS was 0.90 (95% CI, 0.59 to 1.38) with low heterogeneity 0.00% (95% CI, 0.00 to 73.28). On the other hand, estimated HR for DSS was 0.41 (95% CI, 0.30 to 0.55), also with low heterogeneity (0.00%; 95% CI, 0.00 to 67.34). Likewise, the estimated HR of progression-free survival in *POLE*-mutant EC was 0.23 (95% CI, 0.08 to 0.64) with a low level of heterogeneity (I<sup>2</sup> = 0.00%, 95% CI, 0.00 to 0.00). These findings indicate better survival and favorable prognosis in *POLE* mutant EC patients.

## DISCUSSION

EC is the most common gynecologic malignancy in the western world [2,3,34], and survival rates are not improving. There is urgent need for strategies to improve outlook for patients with aggressive subtypes and advanced disease. TCGA first identified the very interesting molecular subset of *POLE*-ultramutated EC [1] that features a favorable prognostic potential, despite high tumor grading. Many follow-up studies investigated *POLE* mutant EC tumors, however the frequency reported is variable [8-31]. Some studies show ultra-high levels of mutation at 42.9% [28] compared to zero in others [15,18]. This meta-analysis aimed to resolve these datasets to estimate *POLE* gene mutational frequency in EC and the overall effect on patient prognosis.

Our meta-analysis determined that 8.595% of endometrial tumors harbor *POLE* gene mutations. The majority have endometrioid histology and present at the earlier stage of disease progression. Paradoxically the *POLE* mutant tumors present at

the highest grade (grade III, 51.5%) and yet have a better outcome with survival analysis. They also have reduced ability to progress to myometrial invasion which is an important prognostic marker. Many studies confirm that *POLE* mutant tumors have a better prognosis [6-8,13,22,28]. *POLE* mutations in high grade (grade III) endometrioid EC are shown to be associated with a lower risk of recurrence and death [22]. The presence of *POLE* mutations even offers a favorable prognosis for rare and aggressive undifferentiated EC [28]. *POLE* mutation could potentially act as a prognostic biomarker to guide treatment of women with grade III, early-stage disease, with either the common endometrioid or more rare histological types. The lower risk of occurrence means that administration of adjuvant therapy for these patient subsets could be inappropriate. *POLE* proofreading mutations have also been shown to elicit an anti-tumor response [35]. There is now an emerging link between high mutation burden in tumors, the immune response and improved prognosis in cancer patients. Indeed, ultramutated *POLE* tumors have been shown to feature higher immune infiltrations and programmed death-1 and programmed death-ligand 1 expression [36]. These immune cells may offset the survival risk caused by higher tumor grades in ultramutated *POLE*. Overall, mutant *POLE* is a key proportion of EC patients to target therapeutically for maximizing clinical impact, and a future target for immunotherapy [9].

However, it is important to note that significant heterogeneity was present in this meta-analysis. Indeed, heterogeneity is a key problem for meta-analysis studies. We tried to resolve the heterogeneity sources. Initially, all studies were re-checked with respect to data extraction and entry. The sensitivity analysis was conducted by re-estimating the pooled proportion of *POLE* gene after exclu-

sion of outlier studies and studies with low sample size. We also adopted a random effect model for final pooled estimation as this model assumed that effects estimated in different studies are not identical. We also tried to perform subgroup analysis with respect to patient age but unfortunately this parameter was not recorded in the majority of studies. Publication bias was investigated; in our study the plot was relatively symmetrical, indicating that publication bias is unlikely [37]. The presence of high heterogeneity was reduced by subgroup and meta-regression according to geographical distribution. European studies were found to be the main contributor to the heterogeneity. Therefore, data obtained from different countries can cause confounding effects and is a potential study limitation. The issue of heterogeneity in genetic studies is also further compounded with recent advances in sequencing technology. The improved yield of genetic testing can increase detection of variants of unknown significance. However in our *POLE* gene study the lack of standardized clarification of variants of unknown significance may have contributed to heterogeneity. This will be a key area to investigate during future studies.

## Conclusion

Our meta-analysis consolidates previous study estimates of *POLE*-mutated EC frequency and confirms its prognostic benefit for patients. The status and frequencies of the *POLE* gene mutation in EC has implications for medical management and future administration of immunotherapy. The *POLE* mutational status serves as an important prognostic marker, and grade III, early-stage disease patients with endometrioid histology could favor a change in medical management.

## Supplementary Information

The Data Supplement is available with this article at <https://doi.org/10.4132/jptm.2020.07.23>.

## Ethics Statement

The present study was approved by the Institutional Review Board of the University of Kufa (IRB approval No. UK-2018-0456) in accordance with the 1964 Helsinki declaration and its later amendments. Formal written informed consent was not required with a waiver issued by the Institutional Review Board of the University of Kufa. All the authors will be held responsible for any false statements or failure to follow the ethical guidelines.

## ORCID

Alaa Salah Jumaah	<a href="https://orcid.org/0000-0001-9709-1460">https://orcid.org/0000-0001-9709-1460</a>
Mais Muhammed Salim	<a href="http://orcid.org/0000-0001-8014-1863">http://orcid.org/0000-0001-8014-1863</a>
Hawraa Sahib Al-Haddad	<a href="http://orcid.org/0000-0002-8564-3202">http://orcid.org/0000-0002-8564-3202</a>
Katherine Ann McAllister	<a href="https://orcid.org/0000-0002-2893-7706">https://orcid.org/0000-0002-2893-7706</a>
Akeel Abed Yasseen	<a href="https://orcid.org/0000-0001-5050-4408">https://orcid.org/0000-0001-5050-4408</a>

## Author Contributions

Conceptualization: ASJ, MMS. Data curation: ASJ. Formal analysis: ASJ,

MMS. Methodology: ASJ, HSA, MMS. Project administration: HSA, MMS. Resources: HSA. Software: ASJ, MMS. Supervision: ASJ, AAY. Validation: MMS, HSA. Visualization: HSA, MMS. Writing—original draft: ASJ, AAY, KAM. Writing—review & editing: AAY, ASJ, KAM. Approval of final manuscript: all authors.

## Conflicts of Interest

The authors declare that they have no potential conflicts of interest.

## Funding Statement

No funding to declare.

## References

- Bell DW, Ellenson LH. Molecular genetics of endometrial carcinoma. *Annu Rev Pathol* 2019; 14: 339-67.
- Howlander N, Noone AM, Krapcho M, et al. SEER cancer statistics review, 1975-2011. Bethesda: National Cancer Institute, 2014.
- Siegel RL, Miller KD, Jemal A. Cancer statistics, 2015. *CA Cancer J Clin* 2015; 65: 5-29.
- Spurdle AB, Bowman MA, Shamsani J, Kirk J. Endometrial cancer gene panels: clinical diagnostic vs research germline DNA testing. *Mod Pathol* 2017; 30: 1048-68.
- Stelloo E, Nout RA, Naves LC, et al. High concordance of molecular tumor alterations between pre-operative curettage and hysterectomy specimens in patients with endometrial carcinoma. *Gynecol Oncol* 2014; 133: 197-204.
- Hussein YR, Weigelt B, Levine DA, et al. Clinicopathological analysis of endometrial carcinomas harboring somatic *POLE* exonuclease domain mutations. *Mod Pathol* 2015; 28: 505-14.
- Church DN, Stelloo E, Nout RA, et al. Prognostic significance of *POLE* proofreading mutations in endometrial cancer. *J Natl Cancer Inst* 2015; 107: 402.
- Cancer Genome Atlas Research Network, Kandoth C, Schultz N, et al. Integrated genomic characterization of endometrial carcinoma. *Nature* 2013; 497: 67-73.
- Auguste A, Genestie C, De Bruyn M, et al. Refinement of high-risk endometrial cancer classification using DNA damage response biomarkers: a TransPORTEC initiative. *Mod Pathol* 2018; 31: 1851-61.
- Talhok A, McConechy MK, Leung S, et al. A clinically applicable molecular-based classification for endometrial cancers. *Br J Cancer* 2015; 113: 299-310.
- Stelloo E, Bosse T, Nout RA, et al. Refining prognosis and identifying targetable pathways for high-risk endometrial cancer: a TransPORTEC initiative. *Mod Pathol* 2015; 28: 836-44.
- Eggink FA, Van Gool IC, Leary A, et al. Immunological profiling of molecularly classified high-risk endometrial cancers identifies *POLE*-mutant and microsatellite unstable carcinomas as candidates for checkpoint inhibition. *Oncoimmunology* 2017; 6: e1264565.
- Bosse T, Nout RA, McAlpine JN, et al. Molecular classification of grade 3 endometrioid endometrial cancers identifies distinct prognostic subgroups. *Am J Surg Pathol* 2018; 42: 561-8.
- Billingsley CC, Cohn DE, Mutch DG, Stephens JA, Suarez AA, Goodfellow PJ. Polymerase varepsilon (*POLE*) mutations in endometrial cancer: clinical outcomes and implications for Lynch syndrome testing. *Cancer* 2015; 121: 386-94.
- Le Gallo M, Rudd ML, Urlick ME, et al. Somatic mutation profiles of clear cell endometrial tumors revealed by whole exome and targeted gene sequencing. *Cancer* 2017; 123: 3261-8.



16. Talhouk A, McConechy MK, Leung S, et al. Confirmation of ProMiseE: a simple, genomics-based clinical classifier for endometrial cancer. *Cancer* 2017; 123: 802-13.
17. Rosa-Rosa JM, Leskela S, Cristobal-Lana E, et al. Molecular genetic heterogeneity in undifferentiated endometrial carcinomas. *Mod Pathol* 2016; 29: 1390-8.
18. Hoang LN, McConechy MK, Meng B, et al. Targeted mutation analysis of endometrial clear cell carcinoma. *Histopathology* 2015; 66: 664-74.
19. Abdulfatah E, Wakeling E, Sakr S, et al. Molecular classification of endometrial carcinoma applied to endometrial biopsy specimens: towards early personalized patient management. *Gynecol Oncol* 2019; 154: 467-74.
20. Stelloo E, Nout RA, Osse EM, et al. Improved risk assessment by integrating molecular and clinicopathological factors in early-stage endometrial cancer-combined analysis of the PORTEC cohorts. *Clin Cancer Res* 2016; 22: 4215-24.
21. Imboden S, Nastic D, Ghaderi M, et al. Phenotype of *POLE*-mutated endometrial cancer. *PLoS One* 2019; 14: e0214318.
22. Billingsley CC, Cohn DE, Mutch DG, Hade EM, Goodfellow PJ. Prognostic significance of *POLE* exonuclease domain mutations in high-grade endometrioid endometrial cancer on survival and recurrence: a subanalysis. *Int J Gynecol Cancer* 2016; 26: 933-8.
23. Talhouk A, Hoang LN, McConechy MK, et al. Molecular classification of endometrial carcinoma on diagnostic specimens is highly concordant with final hysterectomy: earlier prognostic information to guide treatment. *Gynecol Oncol* 2016; 143: 46-53.
24. Wong A, Kuick CH, Wong WL, et al. Mutation spectrum of *POLE* and *POLD1* mutations in South East Asian women presenting with grade 3 endometrioid endometrial carcinomas. *Gynecol Oncol* 2016; 141: 113-20.
25. Wortman BG, Creutzberg CL, Putter H, et al. Ten-year results of the PORTEC-2 trial for high-intermediate risk endometrial carcinoma: improving patient selection for adjuvant therapy. *Br J Cancer* 2018; 119: 1067-74.
26. Kommoss FK, Karnezis AN, Kommoss F, et al. L1CAM further stratifies endometrial carcinoma patients with no specific molecular risk profile. *Br J Cancer* 2018; 119: 480-6.
27. Karnezis AN, Leung S, Magrill J, et al. Evaluation of endometrial carcinoma prognostic immunohistochemistry markers in the context of molecular classification. *J Pathol Clin Res* 2017; 3: 279-93.
28. Espinosa I, Lee CH, D'Angelo E, Palacios J, Prat J. Undifferentiated and dedifferentiated endometrial carcinomas with *POLE* exonuclease domain mutations have a favorable prognosis. *Am J Surg Pathol* 2017; 41: 1121-8.
29. DeLair DF, Burke KA, Selenica P, et al. The genetic landscape of endometrial clear cell carcinomas. *J Pathol* 2017; 243: 230-41.
30. Meng B, Hoang LN, McIntyre JB, et al. *POLE* exonuclease domain mutation predicts long progression-free survival in grade 3 endometrioid carcinoma of the endometrium. *Gynecol Oncol* 2014; 134: 15-9.
31. Bellone S, Bignotti E, Lonardi S, et al. Polymerase epsilon (*POLE*) ultra-mutation in uterine tumors correlates with T lymphocyte infiltration and increased resistance to platinum-based chemotherapy in vitro. *Gynecol Oncol* 2017; 144: 146-52.
32. Moher D, Shamseer L, Clarke M, et al. Preferred reporting items for systematic review and meta-analysis protocols (PRISMA-P) 2015 statement. *Syst Rev* 2015; 4: 1.
33. MedCalc statistical software version 13.0.6 [Internet]. Ostend: MedCalc Software, 2014 [cited 2020 Feb 27]. Available from: <http://www.medcalc.org>.
34. Jumaah AS, Al-Haddad HS, Mahdi LH, et al. Increased *PTEN* gene expression in patients with endometrial carcinoma from areas of high risk depleted uranium exposure. *BMC Res Notes* 2019; 12: 708.
35. van Gool IC, Eggink FA, Freeman-Mills L, et al. *POLE* proofreading mutations elicit an antitumor immune response in endometrial cancer. *Clin Cancer Res* 2015; 21: 3347-55.
36. Bakhsh S, Kinloch M, Hoang LN, et al. Histopathological features of endometrial carcinomas associated with *POLE* mutations: implications for decisions about adjuvant therapy. *Histopathology* 2016; 68: 916-24.
37. Sterne JA, Sutton AJ, Ioannidis JB, et al. Recommendations for examining and interpreting funnel plot asymmetry in meta-analyses of randomised controlled trials. *BMJ* 2011; 343: d4002.