

Article

Estimating the Risk of Human Herpesvirus 6 and Cytomegalovirus Transmission to Ugandan Infants from Viral Shedding in Saliva by Household Contacts

Bryan T. Mayer ^{1,*}, Elizabeth M. Krantz ¹, Anna Wald ^{1,2,3,4}, Lawrence Corey ¹, Corey Casper ⁵, Soren Gantt ^{6,†} and Joshua T. Schiffer ^{1,†}

- ¹ Vaccine and Infectious Diseases Division, Fred Hutchinson Cancer Research Center, Seattle, WA 98109, USA; ekrantz@fredhutch.org (E.M.K.); nnawald@u.washington.edu (A.W.); lcorey@fredhutch.org (L.C.); jschiffe@fredhutch.org (J.T.S.)
- ² Department of Medicine, University of Washington, Seattle, WA 98195, USA
- ³ Department of Laboratory Medicine and Microbiology, University of Washington, Seattle, WA 98195, USA
- ⁴ Department of Epidemiology, University of Washington, Seattle, WA 98195, USA
- ⁵ Infectious Disease Research Institute, University of Washington, Seattle, WA 98102, USA; Corey.Casper@idri.org
- ⁶ BC Children's Hospital Research Institute, Vancouver, BC V5Z 4H4, Canada; sgantt@bcchr.ubc.ca
- * Correspondence: bmayer@fredhutch.org
- + These authors contributed equally to this work.

Received: 7 January 2020; Accepted: 29 January 2020; Published: 3 February 2020



Abstract: Human herpesvirus 6 (HHV-6) and cytomegalovirus (CMV) infections are common in early childhood. In a prospective Ugandan birth cohort study, most infants acquired HHV-6 (24/31; 77%) and CMV (20/30; 67%) during follow-up. To assess the transmission risk, we modeled a dose–response relationship between infant HHV-6 and CMV infections and weekly oral viral shedding by mothers and all other ("secondary") children in the home. Oral viral loads that were shed by mothers and secondary children were significantly associated with HHV-6 but not CMV transmission. While secondary children had higher and more frequent HHV-6 shedding than their mothers, they had a lower per-exposure transmission risk, suggesting that transmission to maternal contacts may be more efficient. HHV-6 transmission was relatively inefficient, occurring after <25% of all weekly exposures. Although HHV-6 transmission often occurs following repeated, low dose exposures, we found a non-linear dose–response relationship in which infection risk markedly increases when exposures reached a threshold of > 5 log₁₀ DNA copies/mL. The lack of association between oral CMV shedding and transmission is consistent with breastfeeding being the dominant route of infant infection for that virus. These affirm saliva as the route of HHV-6 transmission and provide benchmarks for developing strategies to reduce the risk of infection and its related morbidity.

Keywords: HHV-6; CMV; virology; transmission; epidemiology; biostatistics; viral excretion

1. Introduction

Human herpesvirus-6 (HHV-6) infects the vast majority of humans within the first few years of life, and febrile illness (roseola) associated with primary infection is a major cause of health care visits for young children [1–3]. Furthermore, HHV-6 leads to encephalitis in immunocompromised hosts and has been implicated as a possible contributor to other central nervous system diseases, including epilepsy and Alzheimer's disease [4–6]. Like other HHVs, HHV-6 establishes latency following primary infection and is persists within the host for life, with intermittent reactivation resulting in the shedding of infectious virus in saliva [7].



Cytomegalovirus (CMV) is also widespread globally and commonly infects young children, particularly in the developing world [8–10]. CMV is a major cause of illness following organ and stem cell transplantation and during untreated HIV infection [11–14]. CMV appears to be associated with immune senescence in the elderly, though the long-term consequences of infection are still being determined [15–19].

Because of the public health importance of HHV-6 and CMV infections and the interest in preventing HHV-6-related diseases, understanding the determinants of transmission is of great interest. CMV transmission to infants has been shown to occur most commonly via breast milk, though saliva and urine are other routes of infection during early childhood [9,20–23]. HHV-6 is likely transmitted through saliva, based on the frequency of the viral detection of the virus in the saliva of infected individuals [3]; in contrast, HHV-6 infection is not associated with breastfeeding and is rarely found in breast milk [9,24]. Thus, HHV-6 transmission is likely determined by the level of exposure to oral, viral shedding by close contacts. Exposure to high quantities of the virus at mucosal sites predicts the transmission of herpes simplex virus type 2 (HSV-2) [25,26] and HIV [27,28]. Estimating the risk of acquisition from the level of exposure to a virus at a mucosal surface has important ramifications for strategies designed to prevent virus transmission [26]. Effective treatment or vaccination strategies might have specific measurable effects on the transmission dose-response curve. Predicting transmission risk is challenging, however, because concurrent data from transmitters and susceptible individuals are required to adequately link exposure and infection. Here, we use longitudinal quantitative viral shedding data within households to estimate the transmission risk of HHV-6 and CMV in a cohort of infants on a weekly basis.

2. Materials and Methods

2.1. Study Cohort and Data

Study data were derived from a previously described household-based birth cohort study in Uganda [9]. Pregnant women attending prenatal care at Mulago Hospital in Kampala were eligible if they had at least one other ("secondary") child <7 years old living in the home and with documented HIV infection status. During the study, home visits were conducted within the first week after birth (median 2 days of age; range 0–9) and each week thereafter. At home visits, oropharyngeal swabs were collected in a standardized manner [29] from the mother and all children. The swabs were collected weekly from infants for up 119 weeks following birth, and from mothers and secondary children in the households for the first 52 weeks. An analysis of the clinical outcomes in this cohort by Gantt et al. [9] found a trend toward fever and increased paracetamol use associated with primary HHV-6 infection and no symptoms associated with CMV infection.

All oral swabs were tested for HHV-6 and CMV by quantitative (q)PCR, and HHV-6 typing was performed using the previously described methods [30–32]. Briefly, DNA extraction was performed using QIAamp DNA Mini Kits (Qiagen, Germantown, MD, USA) according to manufacturer's instructions. The 5R set of primers and probes were used to amplify HHV-6 [31], and discrimination between HHV-6A and 6B was performed by a separate PCR targeting the U94 gene using species-specific probes [32]. The amplification of CMV was performed using primers and probes that simultaneously target the glycoprotein B (gB, UL55) and immediate early viral protein 1 (IE1, UL123) coding regions, as described [30]. The cutoff for a positive HHV qPCR was 3 viral genome copies/reaction, or ~150 copies/mL of swab buffer [9]. HIV testing was performed according to national guidelines and was negative for all infants [9].

2.2. Definitions of Exposure and Transmission Events

We defined a potential salivary shedding exposure as a virus detected in a swab collected from mothers or secondary children when it met the following criteria: 1) it occurred before acquisition in the infant, or 2) it occurred at any time in a household in which no infant acquisition was observed. When there were multiple secondary children in one household measured on the same week, the aggregate sum was used to create a single measurement. In addition to considering the exposure from mother and secondary children independently, we constructed a household measure calculated as the sum of exposures from mothers and the summed secondary children measurements from contemporaneous samples.

While the study visit schedule was designed to be weekly, the actual sampling schedule often deviated from weekly visits. To standardize exposure intervals, the study time was considered relative to infants' date of birth and one exposure measurement was used per week. If there were multiple measurements per week, we aggregated these values by taking the maximum. If weekly measurements were missing, they were imputed using linear interpolation between the previous and subsequent observed concentrations. In 5 households, exposure data was missing for the first week following infant birth, so linear interpolation could not be performed. For 4 of those households, the first exposure observation occurred during the second week after the infant was born, and this concentration was used to impute the first week exposure. In the fifth household, the first exposure observation was at 7 weeks. This household was included in the descriptive exposure analysis but was excluded from the risk analysis. The interpolation never incorporated concentrations measured after transmission was observed.

The criteria for determining transmission to infants and corresponding infection times were based on the onset of the repeated detection of viral DNA at high levels in oral swabs or in plasma from infants and mothers confirmed with serology for CMV [9]. To estimate the weekly transmission risk, an exposure was considered infectious if it occurred approximately one week prior to detected infection. Because of the limited sample size and variable sampling schedule, we allowed an infectious exposure window around one week, between 4 and 14 days, following an exposure. Infected infants were considered right-censored if there were no infectious exposures measured during this window. For uninfected infants, the infant was considered right-censored after their final observed exposure. The timing of transmission events was displayed using cumulative incidence curves with transmission risk beginning at the date of infant birth and the time to transmission being calculated relative to that date. The cumulative incidence was estimated using the Kaplan–Meier survival estimator assuming viral infections were not competing risks.

2.3. Survival Analysis and Dose-Response Modeling

To estimate the weekly risk of transmission by household members, we constructed a time-dependent dose-response model using a survival model. In contrast to a Cox proportional hazards model, where the hazard is not generally estimated, we constructed models by directly estimating the hazard to determine the dose-response relationship between viral load exposure and risk.

As sampling is generally weekly, we assumed that the true transmission time was interval censored and occurred during the week between the last observation prior to infection and the infection time. Therefore, our model was constructed to estimate the weekly transmission risk given weekly measurements of exposures. The resulting weekly infection probability for a given prior weekly exposure (E) was calculated as follows:

$$Pr(infection) = 1 - e^{-(b_0 + b_E E)}, \tag{1}$$

where b_E is the risk parameter associated with the exposure and the b0 parameter is a constant weekly risk parameter capturing the remaining risk not associated with exposure (i.e., when exposure was 0 DNA copies/mL). For the combined model, the weekly infection probability was calculated as follows:

$$\Pr(infection) = 1 - e^{-(b_0 + b_s S + b_M M)},$$
(2)

where S and M indices correspond to secondary children and mother exposures, respectively.

The parameters in these models were estimated from the data by maximizing a survival likelihood. Details on the model and model optimization procedures are available in Appendix A. To determine whether household exposures significantly contributed to infection risk, the model incorporating exposure in the household was statistically compared to a null model including just the constant weekly risk parameter, b_0 . This was done using a likelihood ratio test, which is described in more detail in Appendix A. The predictions of the weekly risk were calculated from the individual (Equation (1)) and combined (Equation (2)) model using the fitted parameters using the range of exposures observed in the data.

An infectious dose of viral load associated with 50% risk (ID_{50}) was calculated by solving for the exposure that gives a 50% infection risk given the fitted parameters using Equations (1) and (2). ID_{25} and ID_{75} were also calculated. For the combined model (Equation (2)), ID calculations were done for one exposure source at a time (the other was assumed to be 0 DNA copies/mL).

2.4. Sensitivity Analysis

We performed a series of sensitivity analyses. First, we iteratively refit the model, leaving out one household to evaluate individual household leverage on the parameter estimates. Next, to evaluate the sensitivity to exposure interpolation, we iteratively refit the model, allowing an increasing maximum amount of exposure interpolation by households. In other words, we fit the model including households with 20% interpolation or less, then 25% interpolation or less, etc., until all households were included. Lastly, in combination with the interpolation analysis, we also fit the model using only data from households with transmission to assess whether there was a categorical difference that determined whether infants were not infected that could modulate exposure risk.

2.5. Software, Data, and Code Availability

Programming and analysis was conducted using the R programming language (CRAN) [33]. Data processing and visualizations were conducted using the tidyverse [34]. Kaplan–Meier estimates were generated using the survival package [35]. Exact Wilcox rank tests were performed using the coin package [36,37]. Optimization was done using the optim function with the BLGS [38] and Nelder–Mead [39] algorithms. Latin hypercube samples were implemented using the lhs package [40]. A research compendium for this analysis was created using the workflowr package [41]. The workflowr package generated the following research website containing all data and analysis code used to generate results and figures: http://bryanmayer.github.io/HHVtransmission.

3. Results

3.1. HHV-6 and CMV Transmission Occurred at High Rate Early During Infancy

Thirty-two newborn infants, 32 mothers (17 HIV-infected and 15 HIV-uninfected), and 49 secondary children (4 HIV-infected and 7 unknown) were followed for a median of 57 weeks. Primary HHV-6 infections were observed in 24 of the 31 at-risk infants during the study (one household had no detectable virus during the study). All infections were typed as HHV-6B and 1 was a co-infection with HHV-6B and HHV-6A. CMV was acquired at a similar rate, with 20 infections observed among 30 infants at risk of infection during the study (two cases of congenital CMV transmission were excluded). The majority of transmissions occurred prior to one year with no difference in cumulative incidence observed between infants with HIV+ and HIV- mothers for either virus (Figure 1). As exposure information was only collected through the first year, the four CMV transmissions and one HHV-6 transmission that occurred after one year were right-censored at one year for further analysis.



Figure 1. Time taken for the transmission of human herpesvirus 6 (HHV-6) and cytomegalovirus (CMV). Cumulative incidence curves for all transmission events to infants following birth stratified by maternal HIV status with censoring denoted by a plus. The P-values from log-rank tests comparing survival curves (one minus proportion infected) between infants with HIV+ and HIV- mothers are displayed in the bottom-right of each panel. The household exposure information was collected through one year (52 weeks; vertical, dashed black line). All events occurring after one year were right-censored for an estimation of transmission risk.

3.2. Secondary Children Shed at Higher Viral Loads than Mothers

Overall, during the at-risk period for the infants, there were 684 exposure weeks for HHV-6 and 819 exposure weeks for CMV to evaluate for transmission. Not every week had a measured exposure; secondary children were more likely to have missing data with 21% observations missing compared to 6% in mothers across the viruses (Table S1). Observed (non-missing) exposures were summarized for each household contact and virus by the frequency of positive measurements (percent of weeks with positive oral swabs) and then the mean and maximum oral viral loads (log₁₀ DNA copies/mL) over the exposure period. For both viruses, secondary children uniformly had higher shedding frequencies and viral loads compared to mothers in the same household (Figure 2a–c). Summarized exposure measurements were not significantly correlated between mothers and secondary children within the same household, although there were modest positive correlations estimated for HHV-6 (Spearman correlation *p*-values > 0.05 for all, Table S2). Overall household exposure was assessed by taking the sum of maternal and secondary children viral loads at each week. In the majority of households, viral loads shed by secondary children contributed to the majority of the household exposure sum measure, comprising a median of 99.7% (IQR: 94.6–99.9%) of the CMV household sum and 91.2% (IQR: 80.5–98.8%) of the HHV-6 household sum (Figure S1).



Figure 2. Comparisons of exposure patterns across households and among household members. Comparing viral exposure patterns for HHV-6 and CMV between mother and secondary children within a household by (**a**) percent positive weekly swab measurements; (**b**) mean log viral load (VL; DNA copies/mL); and (**c**) maximum log viral load. The diagonal line in the scatterplot denotes equivalent summary measures between household mother and secondary children (i.e., the points below the line indicate lower measurements from the household mother). Shedding patterns by exposure source compared between households without and with transmission by (**d**) the percent of positive weekly measurements; (**e**) the mean log viral load; and (**f**) the maximum log viral load. Boxes represent the interquartile range, stems represent values within 1.5 of the IQR, and points represent the raw data. All observed exposures measured in the infant's first year of life and prior to transmission times are shown.

3.3. Correlation between household shedding and transmission to infants

We next compared shedding frequency, median viral loads, and maximum viral loads between households with (prior to) and without a transmission event for each virus (Figure 2d–f, Table S3). Generally, for both HHV-6 and CMV, households with transmission events had higher oral shedding frequencies and magnitudes than households without observed transmission in the first year. For HHV-6, this trend was uniformed and more evident. For all three outcomes, secondary children and overall household exposures were significantly higher in the households with HHV-6 transmissions (exact Wilcoxon test *p*-values < 0.05, Table S3). There were no significant differences between shedding patterns from any exposure source between households with and without infant CMV transmission (Table S3).

3.4. HHV-6, but not CMV, Acquisition in Infants was Associated with Viral Loads of Oral Shedding Exposures

We next estimated the weekly risk of transmission by household members using the longitudinal, weekly exposure data. Based on a sensitivity analysis, one household was removed from the HHV-6 transmission model, see Appendix B. All exposure sources were significantly associated with infant HHV-6 infection but not CMV (Table 1). The weekly risk was calculated based on the model estimates and the mean viral load (among positive swabs), using secondary children and mothers by virus as inputs. Without including exposure, the average (constant) weekly risk for HHV-6 was 3.60% and it was 1.96% for CMV. In the HHV-6 models including exposure sources, the constant weekly risk was lower because the exposure sources explain additional variation in weekly risk (Table 1). For secondary children, the estimated weekly risk absent exposure was 2.07% compared to 2.92% given the exposure of the mean positive oral viral load. For mothers, the estimated weekly risk absent exposure was 2.88% compared to 3.39% given as a mean exposure. For either household contact, risk was substantial (> 90%) given the exposure of the maximum observed viral load. As secondary children viral loads comprise a large proportion of household sum exposure, risk due to household sum was similar to secondary children. For CMV, the estimated constant weekly risk in models including exposure sources was 1.92% and did not differ based on the exposure source. With the exception of the household removed from the HHV-6 model, model results were generally robust to sensitivity analysis; for further details and estimated model parameters (Table A1), see Appendix B.

Table 1. Estimates of the weekly risk for each virus with no exposure (constant weekly risk) and by exposure source. Models of risk were fit without exposure (null model) and independently for each exposure source. Weekly risk with exposure includes constant risk from the model. The mean exposure was calculated among positive swabs. The P-values were calculated from the model to test if risk attributable to exposure is significantly different from null model without exposure risk (log-likelihood ratio test).

		No Exposure	Mean Exp	osure	Maximum E	xposure	
Virus	Exposure Source	Weekly Risk (%)	Log ₁₀ DNA copies/mL	Risk (%)	Log ₁₀ DNA Copies/mL	Risk (%)	<i>p</i> -Value
	Null	3.60					
HHV-6	Secondary children	2.07	3.98	2.92	6.50	94.66	< 0.001
	Mother	2.88	2.82	3.39	5.68	97.75	0.007
	Household sum	2.01	3.94	2.79	6.50	94.63	< 0.001
	Null	1.96					
CMV	Secondary children	1.92	3.72	1.92	6.72	1.92	0.939
	Mother	1.92	2.56	1.92	3.88	1.92	0.939
	Household sum	1.92	3.77	1.92	6.72	1.92	0.939

3.5. A Combined Exposure Model for HHV-6 Highlights Differential Risk between Mother and Secondary Children Exposures

While the household sum model is a model of combined household risk, it assumes that risk exposure is equivalent between secondary children and mothers. We next estimated a combined exposure transmission risk that includes and adjusts for separate risk estimates by household secondary children and mothers. The combined model for HHV-6, but not CMV, was statistically significant (Table 2). In the combined model, the estimated constant weekly risk was 1.80%, which was lower than any estimate from the individual models (Table 1), indicating that additional risk is explained by including both exposures simultaneously with a different risk. Per mean exposure, absent exposure from another contact source, and secondary children exposure had a higher weekly risk compared with maternal exposure: 2.55% vs. 2.22% a result following from higher viral loads in secondary children (Figure 2).

Table 2. Estimates of weekly risk for each virus with no exposure (constant weekly risk) and by exposure source. Models of risk were fitted to adjust for both exposure sources and risks, which were calculated assuming that other exposure sources were 0 DNA copies/mL. The weekly risk with exposure includes constant risk from the model. P-values calculated from the model to test if risk attributable to exposure is significantly different from the null model without exposure risk (log-likelihood ratio test).

	No Exposure	Secondary Chi	Secondary Children Exposure ¹		Maternal Exposure ¹		
Virus	Weekly Risk (%)	Mean Exposure Risk (%)	Maximum Exposure Risk (%)	Mean Exposure Risk (%)	Maximum Exposure Risk (%)	<i>p</i> -Value	
HHV-6	1.80	2.55	92.39	2.22	95.58	< 0.001	
CMV	1.92	1.92	1.92	1.92	1.92	0.997	

¹ Mean and maximum exposures (DNA copies/mL) shown in Table 1.

For equivalent viral loads, the risk of HHV-6 infection was higher from maternal exposures. This is evident by higher maximum maternal exposure risk despite lower maximum viral load (Tables 1 and 2) and differences in infectious doses resulting in a given risk of infection (Table 3). From the combined exposure model, the estimated infectious oral exposure, resulting in 50% probability of infection (ID_{50}), was 5.01 log₁₀ DNA copies/mL from maternal exposures compared to 5.92 log₁₀ DNA copies/mL from secondary children, which is almost a 10-fold difference. Using the single exposure models, ID_{50} estimates showed a similar difference between sources: 4.92 and 5.87 log₁₀ DNA copies/mL from mother and secondary children, respectively. As exposure to CMV was not associated with infection risk, ID calculations were not performed for CMV.

Table 3. HHV-6 weekly infectious exposure (ID, log_{10} DNA copies/mL) associated with 25%, 50%, and 75% probability of infection (ID₂₅, ID₅₀, and ID₇₅) estimated from the HHV-6 risk models by exposure source. For the exposure combined model, calculations assume a single exposure source. Exposure was not associated with CMV transmission and so IDs were not calculated.

		HHV-6 Infectiou	HHV-6 Infectious Dose (ID, log ₁₀ DNA copies/mL)			
Model	Exposure Source	ID ₂₅	ID ₅₀	ID ₇₅		
Individual	Secondary Children	5.47	5.87	6.17		
	Mother	4.51	4.92	5.23		
Combined	Secondary Children	5.53	5.92	6.23		
	Mother	4.61	5.01	5.32		

3.6. The Dose–Response Relationship between the Viral Load of HHV-6 oral Shedding Exposure and Weekly Infant Acquisition

Using the estimated transmission models, we can describe the dose–response relationship by calculating the probability of transmission per weekly exposure across the range of potential exposures. Comparing the estimated dose-response curve from the combined exposure model to the corresponding observed data, a clear dose–response relationship was evident between weekly oral HHV-6 shedding and weekly transmission (Figure 3a). In general, the probability of transmission on any given week was relatively low, with the majority of individual exposures having a < 25% probability and only high exposures (> 5 \log_{10} DNA copies/mL for mothers or > 6 \log_{10} DNA copies/mL for secondary children) result in higher predicted risk. For CMV, as there was no risk attributable to exposure, the dose–response curves were flat, always predicting the 2% constant risk estimated from the model (Figure S2a).



Figure 3. Estimation of HHV-6 acquisition risk from exposures by week using the combined exposure model (Tables 2 and A1). (a) Individual HHV-6 dose–response relationship for mother and secondary children oral viral load (VL, DNA copies/mL) exposures where bars depict the percentage of infections observed among total exposures in the data and lines depict estimated risk from the model. (b) A heatmap depicting the distribution of total combined exposures in a given viral load bin from mothers and secondary children in a household. The text depicts the fraction of total infections over total exposures. (c) A heatmap depicting the risk of HHV-6 infection from combined household mother and secondary children exposures estimated from the model. The text depicts the proportion of infections observed in the data using binned viral loads from (b) (equivalent to fraction displayed).

We next looked at the weekly risk resulting from combined exposures from both secondary children and mother within the household. Most weekly exposures were either low without observed transmission (less than the lower limit of detection) or in the 3-5 log₁₀ DNA copies/mL range where the majority of transmissions were observed (Figure 3b). Of note, there were 107 exposure weeks with no measured household HHV-6 shedding measured and no instances of HHV-6 infections. In contrast, for CMV, there were 4 infections recorded among the 217 weeks with no household CMV shedding (Figure S2b). Using the model, the majority of combined exposures were associated with a low weekly probability of HHV-6 transmission (Figure 3c). The predicted risk increases dramatically for HHV-6 exposures higher than 5.5 log₁₀ DNA copies/mL from secondary infants and 4.5 log₁₀ DNA copies/mL from the mother. Though these high exposures were generally rare in the data, they were associated with an increased observed transmission risk, particularly when the mother had a detectable viral load. Together, the model and the data suggest that the majority of HHV-6 infections arise from repeated exposures to oral viral load shedding by all members in the household, but with varying degrees of contact efficiency. Episodes of high viral shedding by any household member substantially increases risk.

4. Discussion

Here, we estimated risk of HHV-6 and CMV transmission to infants using weekly measurements of oral shedding exposures by household contacts. Oral viral shedding by contacts was strongly associated with incident HHV-6 infection in infants, which affirms suggestions that saliva is a major transmission route [3,24] and is consistent with the epidemiologic data showing that parental saliva-sharing behavior and having older siblings are both associated with HHV-6 acquisition risk [42]. HHV-6 transmission showed a clear dose–response relationship with weekly exposure to oral viral loads from both mothers and secondary children, with different associated risks. Infection generally resulted from repeated, low infectivity exposures. However, the relationship between viral load and transmission appears non-linear and the data suggest that exposure to bursts of oral shedding with viral loads >1 million copies dramatically increases the risk of transmission.

We did not find that oral exposure was a significant risk for infant CMV acquisition, which is consistent with breast milk being the dominant route in this age group or this infant cohort specifically [9]. Although oral shedding by household contacts did not appear to be relevant for infant acquisition of CMV in this setting, we posit that there is a relationship between the CMV viral load of an exposure and transmission. We did not collect breast milk from mothers in this cohort, but others have reported a positive correlation between the viral load in breast milk and the risk of infant CMV acquisition [22,43,44], and we have shown a dose–response between oral CMV viral load shedding by young children and transmission to their mothers [45]. Breast milk does not appear to be a route of HHV-6 transmission [24]; rather, saliva has been thought to be the major source of HHV-6 transmission based on the frequency of viral detection in saliva [3], which our data strongly support.

While our model estimated the weekly risk from a single oral viral load measurement, there are likely many exposures in a given week that vary in magnitude. In contrast to our approach, a mechanistic dose–response relationship would describe the probability of infection at each exposure. Data informing such a model would be extremely difficult, if not impossible, to collect. If weekly measurements are a good estimate of the average daily exposures within a week, then we expect the true risk per contact to be lower than our weekly risk estimate because there are likely many exposures per week. Specifically, while mothers have lower viral loads, they likely expose their infants more than siblings, and therefore may have similar per exposure risks. In this case, infants become high-risk when their mothers have a high shedding event due to exposure frequency, which could exponentially increase transmission risk even amongst low risk, individual exposures. Regardless, this is consistent with our conclusion that HHV-6 transmission may result from repeated, inefficient exposures. Ultimately, with increased exposure sampling granularity, a true per-contact risk could be

estimated with this model approach and has been previously shown in HSV-2, a system with discrete, sexual contacts [26].

Time-dependent exposure and transmission data are challenging to collect, and our analysis was therefore limited by a small sample size and censoring. To accommodate, we used interpolation for missing exposures, interval censoring approaches to model survival risk, an independent weekly risk assumption, and a linear risk model structure within the survival function. In addition, all HHV-6 positive samples could not be typed and so the exact proportion that were HHV-6B is unknown. These approaches result in assumptions that add uncertainty to our model results that are difficult to quantify. Reassuringly, we found that the oral shedding of HHV-6 by household members is a dose-dependent risk factor for transmission with or without data interpolation. While our estimated dose–response relationship for HHV-6 is consistent with these data and the known transmission routes of HHV-6, the validation of the relationship requires additional study.

Primary HHV-6 infection accounts for 10%–20% of febrile illness in the first 2-3 years of life in high-income countries and is associated with a high rate of physician visits [3,7]. Thus, the prevention of HHV-6 infection is of significant public health interest. Linking HHV-6 exposure to infection risk provides insight into the viral determinants of transmission. In general, the classification of a subpopulation as high- or low-risk depends on both the frequency of exposure and the susceptibility to infection. There is no known immunologic correlate of protection against primary infection with HHV-6, which infects nearly 100% of people worldwide. Using the dose–response relationships and the subsequent infectious doses (e.g., ID₅₀) uncovered in this study, we established a baseline link between oral shedding and the weekly risk of transmission. Interventions that affect the susceptibility of infants (i.e., vaccination) would theoretically increase necessary quantities of exposure. The study of dose–response relationships before and after specific interventions could be used as a precise metric to develop and evaluate the effectiveness of interventions to prevent HHV-6 infection and its related morbidity.

Supplementary Materials: The following are available online at http://www.mdpi.com/1999-4915/12/2/171/s1. Figure S1: Composition of household sum viral load; Figure S2: Estimation of CMV acquisition risk from exposures by week using the combined exposure model; Table S1: Total observed and interpolated exposures by virus and exposure source; Table S2: Spearmen correlations for exposure outcomes between secondary children and mothers within a household; Table S3: Comparison of exposure outcomes by exposure source between households by infant transmission status.

Author Contributions: Conceptualization, B.T.M., S.G. and J.T.S.; methodology, B.T.M. and E.M.K.; software, B.T.M.; formal analysis, B.T.M.; investigation, B.T.M., A.W., L.C., C.C., S.G. and J.T.S.; data curation, B.T.M. and E.M.K.; writing—original draft preparation, B.T.M., S.G. and J.T.S.; writing—review and editing, all authors; visualization, B.T.M.; supervision, J.T.S. and S.G.; funding acquisition, A.W., L.C., C.C., S.G. and J.T.S. All authors have read and agreed to the published version of the manuscript.

Funding: This work was supported by the National Institutes of Health Roadmap KL2 Clinical Scholar Training Program (grant KL2-RR025015-01 to S.G.), the University of Washington Center for AIDS Research (new investigator awards P30-AI027757 [to S.G.], P01-AI 030731 [to L.C. and A.W.], R01-CA138165 [to C.C.], and P30-CA015704 [to C.C. and L.C.], and a BC Children's Hospital Investigator Salary Award [to S.G.]).

Conflicts of Interest: The authors declare no conflict of interest related to this research. The funders had no role in the design of the study; in the collection, analyses, or interpretation of data; in the writing of the manuscript, or in the decision to publish the results.

Appendix A

This appendix contains additional, detailed information on the survival model, optimization, and statistical testing. Noting that the survival probability is one minus the infection probability (equation (1)), the survival function for any given week, j, with exposure, E_j, is

$$S(j) = e^{-(b_0 + b_E E_j)}.$$
 (A1)

From there, we considered five risk models,

$$S_0(j) = e^{-b_0},$$
 (A2a)

$$S_m(i) = e^{-(b_0 + b_m m_j)},$$
 (A2b)

$$S_s(j) = e^{-(b_0 + b_s s_j)},$$
 (A2c)

$$S_{hh}(j) = e^{-(b_0 + b_{hh}(s_j + m_j))}$$
, and (A2d)

$$S_{full}(j) = e^{-(b_0 + b_m m_j + b_s s_j)}.$$
 (A2e)

Each equation describes each potential exposure source: 1) exposure-independent or null model (equation A2a), 2) mother exposure only (equation A2b; m_j denotes maternal exposure in the jth week), 3) secondary children exposure only (equation A2c; s_j denotes secondary children exposure in the jth week), 4) total household exposure (sum in the jth week, $m_j + s_j$) with common risk (equation A2d), and 5) full risk model with differential risk by mother or secondary children exposure (equation A2e).

For each risk model, S_k (equation A2a – A2e), we assumed that at-risk infants were independent, and the risk associated with weekly exposure was unique from other exposures (i.e., non-infectious exposure weeks are exchangeable). That is, the accumulation of risk or survival probability does not depend on the order of exposures [46]. The overall negative log-likelihood function for all n infants was then defined as follows

$$\sum_{i=1}^{n} \log L_k(T_i) = \sum_{i=1}^{n} \sum_{j=1}^{T_i} \log S_k(i,j) + \sum_{i=1}^{n} \Delta_i * \log(1 - S_k(T_i + 1)).$$
(A3)

Here, the likelihood of the ith infant was calculated across their unique exposure series (S(i,j)) for each jth week for all T_i weeks without infection, and their infection week, T_i + 1, when infection was observed ($\Delta_i = 1$). From this formulation, the maximum likelihood estimates for the parameters were estimated by minimizing the negative log-likelihood.

With the exception of the null model (equation A2a), parameters were estimated numerically. Numerical estimation was performed as follows for each model: 1) for each parameter, 25 initial values were drawn randomly using Latin hypercube sampling [47] (range: -20 to 0 on the log-scale), 2) the likelihood was minimized separately using both the Nelder-Mead and BLGS algorithms; and then 3) the best fitted parameters were determined from the overall minimum optimization. For the null model, the maximum likelihood estimate of the constant risk parameter is

$$\hat{b}_{0,null} = -\log\left(1 - \frac{\sum_{i=1}^{n} \Delta_i}{\sum_{i=1}^{n} T_i}\right).$$
(A4)

Lastly, exposure-specific risk was tested for statistical significance by comparing the exposure risk models to the null model and performing a log-likelihood ratio test. The test statistic was calculated as 2 times the difference in the negative log-likelihoods and the p-value was then calculated from the chi-square distribution with degrees of freedom matching the total parameters tested: one for the individual models (equations A2b-A2d) and two for the combined (equation A2e).

Appendix B

Sensitivity analysis and final model estimates. In our sensitivity analysis, the HHV-6 risk estimation for secondary children exposure was extremely sensitive to one household (Family AB, Figure A1a). Inclusion of that single household reduced the risk estimate by approximately 10-fold. Upon further inspection, this household had two secondary children, one of which was only present for half the study and had some of the highest exposure levels in the study (Figure A1b). Given the stability of estimation excluding this household (Figure A1a), they were removed from the final, presented

model results. It is plausible that while that second secondary child represents high exposure risk, they never contacted the infant. We next evaluated sensitivity to interpolation. While interpolation rates ranged >50% for some households and was occasionally applied for more than 2 consecutive weeks, estimates were stable to inclusion of those households (Figure A2). No substantial differences were seen refitting the model using only infected households (Figure A2b). For CMV, all estimates across sensitivity analysis resulted in insignificant risk attributable to exposure (Figures A1 and A2). For HHV-6, across all sensitivity analysis, risk estimates were different between mother and secondary children exposure.



Figure A1. Transmission model sensitivity analysis to individual households. (a) Exposure risk coefficient estimate by exposure source and model re-estimated iteratively excluding one household at a time for both viruses; and HHV-6 was additionally evaluated completely excluding household AB. Estimates shown for both individual (single exposure) and combined exposure models. (b) HHV-6 viral load (VL, DNA copies/mL) for household AB by exposure sources. Infant AB was infected following the final exposure.



Exposure Source

Secondary children

Mother

Figure A2. Transmission model sensitivity analysis to exposure interpolation and infection status. (a) Summary of interpolation by household (points) by maximum consecutive weeks interpolated and percentage of exposures weeks interpolation by exposure source for each virus. (b) Estimated exposure risk coefficients for each virus allowing for a maximum total interpolation by household. For example, the risk estimate at 20% maximum interpolation includes only households with 20% interpolation or less. Estimates shown for both individual (single exposure) and combined exposure models. All households are included at 60%.

Virus	Model	Parameter	Estimate	-Log likelihood	
	Constant risk (null)	b ₀	3.67×10^{-2}	91.2	
	Cocon dowy shildron	b ₀	2.10×10^{-2}	24.2	
	Secondary children	b _S	9.13×10^{-7}	84.8	
		b ₀	2.93×10^{-2}	07 5	
ННИ-6	Mother	b _M	$7.96 imes 10^{-6}$	87.5	
11110-0		b ₀	2.03×10^{-2}	01.6	
	Household sum	$b_{\rm HH}$	9.12×10^{-7}	84.6	
		b ₀	1.82×10^{-2}		
	Combined model	b _M	6.56×10^{-6}	84.2	
	-	b _S	8.03×10^{-7}		

Table A1. Coefficient estimates and log likelihood from the survival analysis. For CMV, the log likelihood value was equivalent for all models (see equations (1 - 2) and Appendix A).

15 of 17

Virus	Model	Parameter	Estimate	-Log likelihood
	Constant risk (null)	b ₀	1.98×10^{-2}	
	Cocon dowy shildron	b ₀	1.94×10^{-2}	
	Secondary children	b _S	0	
		b0	1.94×10^{-2}	74.1
CMV	Mother	b _M	0	
		b ₀	1.94×10^{-2}	
	Household sum	b _{HH}	0	
		b ₀	1.94×10^{-2}	
	Combined model	b _M	0	
	-	b _S	0	

 Table A1. Cont.

Abbreviations: HH: household sum; M: mother; S: secondary children

References

- Condon, L.M.; Cederberg, L.E.; Rabinovitch, M.D.; Liebo, R.V.; Go, J.C.; Delaney, A.S.; Schmeling, D.O.; Thomas, W.; Balfour, H.H., Jr. Age-specific prevalence of Epstein-Barr virus infection among Minnesota children: Effects of race/ethnicity and family environment. *Clin Infect Dis.* 2014, *59*, 501–508. [CrossRef] [PubMed]
- Kusuhara, K.; Takabayashi, A.; Ueda, K.; Hidaka, Y.; Minamishima, I.; Take, H.; Fujioka, K.; Imai, S.; Osato, T. Breast milk is not a significant source for early Epstein-Barr virus or human herpesvirus 6 infection in infants: A seroepidemiologic study in 2 endemic areas of human T-cell lymphotropic virus type I in Japan. *Microbiol. Immunol.* 1997, 41, 309–312. [CrossRef] [PubMed]
- 3. Zerr, D.M.; Meier, A.S.; Selke, S.S.; Frenkel, L.M.; Huang, M.L.; Wald, A.; Rhoads, M.P.; Nguy, L.; Bornemann, R.; Morrow, R.A.; et al. A population-based study of primary human herpesvirus 6 infection. *N. Engl. J. Med.* **2005**, *352*, 768–776. [CrossRef] [PubMed]
- Eimer, W.A.; Vijaya Kumar, D.K.; Navalpur Shanmugam, N.K.; Rodriguez, A.S.; Mitchell, T.; Washicosky, K.J.; Gyorgy, B.; Breakefield, X.O.; Tanzi, R.E.; Moir, R.D. Alzheimer's Disease-Associated beta-Amyloid Is Rapidly Seeded by Herpesviridae to Protect against Brain Infection. *Neuron* 2018, 100, 1527–1532. [CrossRef] [PubMed]
- Readhead, B.; Haure-Mirande, J.V.; Funk, C.C.; Richards, M.A.; Shannon, P.; Haroutunian, V.; Sano, M.; Liang, W.S.; Beckmann, N.D.; Price, N.D.; et al. Multiscale Analysis of Independent Alzheimer's Cohorts Finds Disruption of Molecular, Genetic, and Clinical Networks by Human Herpesvirus. *Neuron* 2018, 99, 64–82 e67. [CrossRef] [PubMed]
- 6. Zerr, D.M. Human herpesvirus 6 and central nervous system disease in hematopoietic cell transplantation. *J. Clin. Virol.* **2006**, *37*, S52–S56. [CrossRef]
- 7. Agut, H.; Bonnafous, P.; Gautheret-Dejean, A. Laboratory and clinical aspects of human herpesvirus 6 infections. *Clin. Microbiol. Rev.* **2015**, *28*, 313–335. [CrossRef]
- 8. Kenneson, A.; Cannon, M.J. Review and meta-analysis of the epidemiology of congenital cytomegalovirus (CMV) infection. *Rev. Med. Virol.* **2007**, 17, 253–276. [CrossRef]
- Gantt, S.; Orem, J.; Krantz, E.M.; Morrow, R.A.; Selke, S.; Huang, M.L.; Schiffer, J.T.; Jerome, K.R.; Nakaganda, A.; Wald, A.; et al. Prospective Characterization of the Risk Factors for Transmission and Symptoms of Primary Human Herpesvirus Infections Among Ugandan Infants. *J. Infect. Dis* 2016, 214, 36–44. [CrossRef]
- 10. Pass, R.F. Epidemiology and transmission of cytomegalovirus. J. Infect. Dis 1985, 152, 243–248. [CrossRef]
- 11. Boeckh, M.; Geballe, A.P. Cytomegalovirus: Pathogen, paradigm, and puzzle. *J. Clin. Invest.* **2011**, 121, 1673–1680. [CrossRef] [PubMed]
- 12. Mentec, H.; Leport, C.; Leport, J.; Marche, C.; Harzic, M.; Vilde, J.L. Cytomegalovirus colitis in HIV-1-infected patients: A prospective research in 55 patients. *AIDS* **1994**, *8*, 461–467. [CrossRef]
- 13. Maclean, H.; Dhillon, B. Cytomegalovirus retinitis: Diagnosis and treatment. *Int J. STD AIDS* **1993**, *4*, 322–325. [CrossRef]

- Adachi, K.; Xu, J.; Ank, B.; Watts, D.H.; Camarca, M.; Mofenson, L.M.; Pilotto, J.H.; Joao, E.; Gray, G.; Theron, G.; et al. Congenital Cytomegalovirus and HIV Perinatal Transmission. *Pediatr Infect. Dis. J.* 2018, 37, 1016–1021. [CrossRef] [PubMed]
- 15. Looney, R.J.; Falsey, A.; Campbell, D.; Torres, A.; Kolassa, J.; Brower, C.; McCann, R.; Menegus, M.; McCormick, K.; Frampton, M.; et al. Role of cytomegalovirus in the T cell changes seen in elderly individuals. *Clin. Immunol.* **1999**, *90*, 213–219. [CrossRef]
- Karrer, U.; Sierro, S.; Wagner, M.; Oxenius, A.; Hengel, H.; Koszinowski, U.H.; Phillips, R.E.; Klenerman, P. Memory inflation: Continuous accumulation of antiviral CD8+ T cells over time. *J. Immunol.* 2003, 170, 2022–2029. [CrossRef] [PubMed]
- Weltevrede, M.; Eilers, R.; de Melker, H.E.; van Baarle, D. Cytomegalovirus persistence and T-cell immunosenescence in people aged fifty and older: A systematic review. *Exp. Gerontol.* 2016, 77, 87–95. [CrossRef] [PubMed]
- Simanek, A.M.; Dowd, J.B.; Pawelec, G.; Melzer, D.; Dutta, A.; Aiello, A.E. Seropositivity to cytomegalovirus, inflammation, all-cause and cardiovascular disease-related mortality in the United States. *PLoS ONE* 2011, 6, e16103. [CrossRef] [PubMed]
- Brodin, P.; Jojic, V.; Gao, T.; Bhattacharya, S.; Angel, C.J.; Furman, D.; Shen-Orr, S.; Dekker, C.L.; Swan, G.E.; Butte, A.J.; et al. Variation in the human immune system is largely driven by non-heritable influences. *Cell* 2015, 160, 37–47. [CrossRef] [PubMed]
- 20. Amin, M.M.; Bialek, S.R.; Dollard, S.C.; Wang, C. Urinary Cytomegalovirus Shedding in the United States: The National Health and Nutrition Examination Surveys, 1999-2004. *Clin. Infect. Dis.* **2018**, *67*, 587–592. [CrossRef] [PubMed]
- 21. Prendergast, A.J.; Goga, A.E.; Waitt, C.; Gessain, A.; Taylor, G.P.; Rollins, N.; Abrams, E.J.; Lyall, E.H.; de Perre, P.V. Transmission of CMV, HTLV-1, and HIV through breastmilk. *Lancet Child. Adolesc Health* **2019**, *3*, 264–273. [CrossRef]
- 22. Hamprecht, K.; Maschmann, J.; Vochem, M.; Dietz, K.; Speer, C.P.; Jahn, G. Epidemiology of transmission of cytomegalovirus from mother to preterm infant by breastfeeding. *Lancet* **2001**, *357*, 513–518. [CrossRef]
- 23. Stagno, S.; Reynolds, D.W.; Pass, R.F.; Alford, C.A. Breast milk and the risk of cytomegalovirus infection. *N. Engl J. Med.* **1980**, 302, 1073–1076. [CrossRef] [PubMed]
- Emery, V.C.; Clark, D.A. HHV-6A, 6B, and 7: Persistence in the population, epidemiology and transmission. In *Human Herpesviruses: Biology, Therapy, and Immunoprophylaxis*; Arvin, A., Campadelli-Fiume, G., Mocarski, E., Moore, P.S., Roizman, B., Whitley, R., Yamanishi, K., Eds.; Cambridge University Press: Cambridge, UK, 2007.
- Corey, L.; Wald, A.; Patel, R.; Sacks, S.L.; Tyring, S.K.; Warren, T.; Douglas, J.M., Jr.; Paavonen, J.; Morrow, R.A.; Beutner, K.R.; et al. Once-daily valacyclovir to reduce the risk of transmission of genital herpes. *N. Engl. J. Med.* 2004, 350, 11–20. [CrossRef]
- 26. Schiffer, J.T.; Mayer, B.T.; Fong, Y.; Swan, D.A.; Wald, A. Herpes simplex virus-2 transmission probability estimates based on quantity of viral shedding. *J. R Soc. Interface* **2014**, *11*, 20140160. [CrossRef]
- 27. Quinn, T.C.; Wawer, M.J.; Sewankambo, N.; Serwadda, D.; Li, C.; Wabwire-Mangen, F.; Meehan, M.O.; Lutalo, T.; Gray, R.H. Viral load and heterosexual transmission of human immunodeficiency virus type 1. Rakai Project Study Group. *N. Engl. J. Med.* **2000**, *342*, 921–929. [CrossRef]
- Baeten, J.M.; Kahle, E.; Lingappa, J.R.; Coombs, R.W.; Delany-Moretlwe, S.; Nakku-Joloba, E.; Mugo, N.R.; Wald, A.; Corey, L.; Donnell, D.; et al. Genital HIV-1 RNA predicts risk of heterosexual HIV-1 transmission. *Sci. Transl. Med.* 2011, *3*, 77ra29. [CrossRef]
- 29. Johnston, C.; Orem, J.; Okuku, F.; Kalinaki, M.; Saracino, M.; Katongole-Mbidde, E.; Sande, M.; Ronald, A.; McAdam, K.; Huang, M.L.; et al. Impact of HIV infection and Kaposi sarcoma on human herpesvirus-8 mucosal replication and dissemination in Uganda. *PLoS ONE* **2009**, *4*, e4222. [CrossRef]
- 30. Boeckh, M.; Huang, M.; Ferrenberg, J.; Stevens-Ayers, T.; Stensland, L.; Nichols, W.G.; Corey, L. Optimization of quantitative detection of cytomegalovirus DNA in plasma by real-time PCR. *J. Clin. Microbiol.* **2004**, *42*, 1142–1148. [CrossRef]
- 31. Cone, R.W.; Huang, M.L.; Ashley, R.; Corey, L. Human herpesvirus 6 DNA in peripheral blood cells and saliva from immunocompetent individuals. *J. Clin. Microbiol* **1993**, *31*, 1262–1267. [CrossRef]

- Zerr, D.M.; Gupta, D.; Huang, M.L.; Carter, R.; Corey, L. Effect of antivirals on human herpesvirus 6 replication in hematopoietic stem cell transplant recipients. *Clin. Infect. Dis* 2002, 34, 309–317. [CrossRef] [PubMed]
- 33. R Core Team. *R: A Language and Environment for Statistical Computing;* R Foundation for Statistical Computing: Vienna, Austria, 2019.
- 34. Wickham, H.; Averick, M.; Bryan, J.; Chang, W.; McGowan, L.D.A.; François, R.; Grolemund, G.; Hayes, A.; Henry, L.; Hester, J.; et al. Welcome to the tidyverse. *J. Open Source Softw.* **2019**, *4*, 1686. [CrossRef]
- 35. Therneau, T.M. *A Package for Survival Analysis in S*, version 2.38; 2015; Available online: http://cran.r-project. org/package=survival (accessed on 31 January 2020).
- 36. Hothorn, T.; Hornik, K.; van de Wiel, M.A.; Zeileis, A. Implementing a Class of Permutation Tests: The coin Package. *J. Stat. Softw.* **2008**, *28*. [CrossRef]
- Hothorn, T.; Hornik, K.; van de Wiel, M.A.; Zeileis, A. A Lego System for Conditional Inference. *Am. Stat.* 2006, *60*, 257–263. [CrossRef]
- 38. Nocedal, J.; Wright, S.J. Numerical Optimization, 2nd ed.; Springer: New York, NY, USA, 2006.
- 39. Nelder, J.A.; Mead, R. A Simplex Method for Function Minimization. Comput. J. 1965, 7, 308–313. [CrossRef]
- 40. Carnell, R. *lhs: Latin Hypercube Samples*, version 1.0.1; 2019; Available online: https://github.com/bertcarnell/lhs (accessed on 6 January 2020).
- 41. Blischak, J.; Carbonetto, P.; Stephens, M. Creating and sharing reproducible research code the workflowr way [version 1; peer review: 3 approved]. *F1000Research* **2019**, *8*. [CrossRef]
- 42. Rhoads, M.P.; Magaret, A.S.; Zerr, D.M. Family saliva sharing behaviors and age of human herpesvirus-6B infection. *J. Infect.* **2007**, *54*, 623–626. [CrossRef]
- 43. Murata, H.; Nii, R.; Ito, M.; Ihara, T.; Komada, Y. Quantitative detection of HCMV-DNA in saliva from infants and breast milk on real-time polymerase chain reaction. *Pediatr. Int.* **2009**, *51*, 530–534. [CrossRef]
- 44. Slyker, J.; Farquhar, C.; Atkinson, C.; Asbjornsdottir, K.; Roxby, A.; Drake, A.; Kiarie, J.; Wald, A.; Boeckh, M.; Richardson, B.; et al. Compartmentalized cytomegalovirus replication and transmission in the setting of maternal HIV-1 infection. *Clin. Infect. Dis.* **2014**, *58*, 564–572. [CrossRef]
- 45. Boucoiran, I.; Mayer, B.T.; Krantz, E.M.; Marchant, A.; Pati, S.; Boppana, S.; Wald, A.; Corey, L.; Casper, C.; Schiffer, J.T.; et al. Nonprimary Maternal Cytomegalovirus Infection After Viral Shedding in Infants. *Pediatr Infect. Dis. J.* **2018**, *37*, 627–631. [CrossRef]
- Mayer, B.T.; Koopman, J.S.; Ionides, E.L.; Pujol, J.M.; Eisenberg, J.N. A dynamic dose-response model to account for exposure patterns in risk assessment: A case study in inhalation anthrax. *J. R Soc. Interface* 2011, *8*, 506–517. [CrossRef] [PubMed]
- 47. Stocki, R. A method to improve design reliability using optimal Latin hypercube sampling. *Comput. Assist. Mech. Eng. Sci.* **2005**, *12*, 393–411.



© 2020 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (http://creativecommons.org/licenses/by/4.0/).