No Association between PAWR Gene Polymorphisms and Tardive Dyskinesia in Schizophrenia Patients

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Tardive dyskinesia (TD) is a hyperkinetic movement disorder associated with the prolonged use of antipsychotic drugs. Since prostate apoptosis response 4 (Par-4) is a key ligand of the dopamine D2 receptor, the Par-4 gene (PAWR) is a good candidate gene to study in the context of TD susceptibility. We examined the association between PAWR gene polymorphisms and TD. Three single nucleotide polymorphisms of PAWR were selected for the analysis: rs7979987, rs4842318, and rs17005769. Two hundred and eighty unrelated Korean schizophrenic patients participated in this study (105 TD and 175 non-TD patients). Genotype/allele-wise and haplotype-wise analyses were performed. There were no significant differences in genotype and allele frequencies between the two groups. Haplotype analysis also did not reveal a difference between the two groups. Within the limitations imposed by the size of the clinical sample, these findings suggest that PAWR gene variants do not significantly contribute to an increased risk of TD. **Psychiatry Investig 2012;9:191-194**

Key Words Schizophrenia, Tardive dyskinesia, PAWR, Polymorphism.

INTRODUCTION

Tardive dyskinesia (TD) is a hyperkinetic movement disorder associated with the prolonged use of antipsychotic drugs.¹ The annual incidence was reported to be 3.9% for secondgeneration antipsychotics and 5.5% for first-generation antipsychotics, while the prevalence is 13.1% for second-generation antipsychotics, 15.6% for antipsychotic-free patients, and 32.4% for first-generation antipsychotics.² Although hypotheses such as dopamine receptor supersensitivity, imbalance between D1 and D2 receptor-mediated effects and the GABA hypothesis have been suggested, the pathophysiology of TD is not well defined.^{1,3-9} Among the risk factors known for TD, age is the most robust.¹⁰⁻¹³ Other risk factors include nonwhite ethnicity, extrapyramidal symptoms, lack of response to

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treatment, worse premorbid functioning, negative symptoms, long treatment duration, worsening of psychosis and prolactine-related sexual dysfunction.¹⁴

Genetic susceptibility has been also examined in many studies. There have been substantial case-control association studies on TD, and a number of studies have been conducted on the genes involved in drug metabolism such as cytochrome P450 (CYP), monoamine receptor genes, and oxidative stress related genes. As a result, relatively consistent findings have been reported for certain polymorphisms such as CYP2D6*10,¹⁵ dopamine D2 receptor (DRD2) Taq1A,¹⁶ dopamine D3 receptor (DRD3) Ser9Gly,¹⁷ serotonin 2A receptor (HTR2A) T102C,¹⁸ and manganese superoxide dismutase (MnSOD) Ala9Val,^{19,20} though they are still controversial.

Recently, Park et al.²¹ identified prostate apoptosis response 4 (Par-4) as a regulatory component in dopamine signaling. In their model of the involvement of Par-4 in the regulation of DRD2 signaling, Par-4/DRD2 complex formation via the calmodulin binding motif in the third cytoplasmic loop was suggested to inhibit cAMP signaling from DRD2. Moreover, they also observed that a mutant mouse lacking the DRD2 interaction domain of Par-4 displayed depression-like behaviors.

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Since TD mainly develops in patients treated with a DRD2 blocker, the Par-4 gene (PAWR) could be associated with TD. In this study, we selected three SNPs of PAWR and examined their association with TD susceptibility in Korean schizophrenia patients.

METHODS

Subjects and assessment

All subjects were ethnically homogeneous Korean and were examined by trained psychiatrists using the Korean version of the Structured Clinical Interview for DSM-IV, leading to a diagnosis based on DSM-IV criteria.²² Written informed consent was obtained from each patient, and the study protocol was approved by the Ethics Committee of the Korea University Medical Center. None of the patients had a significant comorbid neurological illness, mental retardation, or history of substance abuse. All subjects were inpatients at collaborating hospitals of Korea University. Other findings from these subjects have been reported previously.^{20,23-28}

Assessment with the Abnormal Involuntary Movement Scale (AIMS)²⁹ was used to recruit 105 schizophrenic patients with TD and 175 schizophrenic patients without TD. Our test sample included TD patients who had received atypical antipsychotic treatment (25 patients), which reportedly has a lower risk of TD than typical antipsychotic treatment, and who had been treated for less than ten years (18 patients). All of the non-TD patients in the test sample had received typical antipsychotic treatment for at least ten years. We applied these different inclusion criteria since the occurrence of TD in patients treated for less than ten years is indicative of a higher genetic susceptibility. Their current antipsychotic dosages were converted into chlorpromazine equivalents. All patients were maintained on stable dosages of antipsychotics for at least three months before the assessment of TD. The TD diagnosis was performed according to the Research Diagnostic Criteria for TD (RDC-TD) on the basis of AIMS.³⁰ We rated the first seven items of the AIMS to determine the total AIMS score of abnormal movements in the patients. Subjects with two or more two-point ratings or one or more threepoint ratings on the first seven items of the AIMS were diagnosed with TD according to RDC-TD. Clinical symptoms were measured by the Positive and Negative Syndrome Scale (PANSS).31

Genetic markers

Three single nucleotide polymorphisms (SNPs) were selected for the analysis: rs7979987, rs4842318, and rs17005769. To evaluate the coverage rate of the three SNPs for the total PAWR gene variation we consulted the HapMap database (release #27, population: CHB, http://hapmap.ncbi.nlm.nih. gov). There were 41 SNPs reported to have a minimal allele frequency of more than 0.1. We performed 2- and 3- marker aggressive tagging using the Tagger program implemented in Haploview 4.2 (http://www.broadinstitute.org/haploview). We found that 35 of 41 (85%) SNPs were captured by the three SNPs.

Genotyping

Blood samples (5-10 mL) from subjects were collected in EDTA, and genomic DNA was isolated using a NucleoSpin® Blood DNA Extraction Kit (Macherev-Nagel, Germany) according to the manual. Genotyping was performed by highresolution melting (HRM) curve analysis. PCR was performed in a volume of 20 µL per reaction with a 96-well Bio-Rad CFX96 Real Time PCR System (Bio-Rad, Hercules, CA, USA). Reaction mixtures included 1.5 µL of genomic DNA as template, 200 mM of each primer [rs7979987-F: GGA CAT TAT CTC TGG GTT AG, rs7979987-R: ACA TAA GCA CAT ACA CCA AC, rs4842318-F: GAA GGC TAC ACG CTA CAT T, rs4842318-R: AGA TGA GTC TGA AGG TCC AAT A, rs17005769-F: TCT ACA ATC TGT CCT TAG AAA CTT AA, rs17005769-R: CTT TCA AGG TTG TTA ATG GCA TAA] (BMS, Daejeon, Korea), 1×Sso Fast EvaGreen SuperMix (Bio-Rad), and sterile H2O. The amplification protocol started with 98°C/3 min followed by 39 cycles of 98°C/10 s and 58°C/20 s. After an initial step of 95°C/10 s and 65°C/10 s, melting curves were generated from 65°C to 95°C in increments of 0.3°C/cycle. Melting profiles were analyzed with the Bio-Rad Precision Melt Software.

Statistical analysis

Differences between categorical and continuous variables were evaluated by the chi-square test and Student's t-test, respectively. The total PANSS and AIMS scores were evaluated by ANCOVA with sex, age, duration of antipsychotics and smoking status as covariates. Genotype deviation from Hardy-Weinberg equilibrium (HWE) was evaluated by the chi-square test for goodness of fit. Haplotype analysis was performed using the UNPHASED 3.1.4 software.³² A power calculation was performed using G*Power3.1.3.³³ The cutoff p-value was set at 0.05.

RESULTS

There were significant differences in age (non-TD: $43.47\pm$ 9.174 years; TD: 46.29 ± 9.722 years, p=0.016) and smoking status of female subjects (non-TD: smoker/nonsmoker= 18/64, TD: smoker/nonsmoker=21/27, p=0.009), while no significant differences were observed between groups with re-

gard to sex, duration of illness, duration of antipsychotic treatment, current antipsychotic dosage and smoking status. Total AIMS scores (mean \pm SD) were higher in TD subjects (non-TD: 0.74 \pm 1.062 vs. TD: 7.66 \pm 4.287, p=0.010). Total PANSS scores were not significantly different (non-TD: 87.87 \pm 18.77 vs. TD: 92.43 \pm 19.49, p=0.095) although the general PANSS subscores were higher in TD patients than in non-TD patients (non-TD: 42.87 \pm 9.80 vs. TD: 46.29 \pm 11.07, p=0.010). Genotype frequencies of all SNPs were in Hardy-Weinberg equilibrium. There were no significant differences in genotype or allele frequencies between the two groups (Table 1). Haplotype analysis also detected no significant difference between the two groups in either two- (data not shown) or three-marker haplotypes (Table 2).

DISCUSSION

Prostate apoptosis response-4 (Par-4) is a key ligand of DRD2. Par-4 is expressed in the striatal neurons along with DRD2 and interacts with DRD2 in neural cells.³⁴ Par-4 normally enhances DRD2 signaling and thereby inhibits dopamine/DRD2-mediated neurotransmission.²¹ Par-4 specifically interacts with DRD2 via its leucine domain, and it can be coimmunoprecipitated with DRD2 in mouse brain lysate. Once DRD2 is activated, Par-4/DRD2 complex formation is necessary for the maintenance of inhibitory tone in dopamine-mediated cAMP signaling. The binding of Par-4 to DRD2 regulates cAMP signaling, and subsequently affects the function of the DA-cAMP-CREB pathway.²¹ Therefore, Par-4 (PAWR) is a good candidate gene to study in the context of TD.

However, in this study, the three PAWR SNPs we analyzed were not associated with TD in terms of genotype, allele, or haplotype. These findings indicate that PAWR may not related to TD susceptibility. However, it is still possible that other regions of the PAWR gene are associated with TD. The studied SNPs only captured approximately 85% of the genetic variations in the PAWR gene according to the HapMap database. On the other hand, it is also possible that the regulatory regions outside of the PAWR gene are associated with TD. The negative association of the analyzed SNPs could be due to type II error. The power was 0.86 and 0.92 to detect an effect size of 0.20 in the analysis of genotype and allele frequencies, respectively. There are additional limitations to generalization of the results of this study. Although anticholinergics and benzodiazepines had been reported to be beneficial in some cases of TD, we could not control these drugs. There was significant age difference between the TD and non-TD group, although we tried to control the age as a covariate in some analyses. Recently, Liou et al.35 tested whether variation in the PAWR gene might be related to TD susceptibility with five SNPs as genetic markers: rs1705769, rs7305141, rs8176874, rs7955388 and rs2307220. However, their results did not indicate any association between PAWR genetic polymorphisms and TD. Our finding supports their results. In conclusion, we

Table 1. PAWR Genotype and allele distributions between TD and non-TD subjects

SNP ID	Position	Allele	Phenotype	Genotype			Allele		p value		
		1/2		1/1	1/2	2/2	1	2	Genotype	Allele	HWE
rs7979987	78522768	G/C	Non-TD	103	64	8	0.7714	0.2286	0.469	0.354	0.953
			TD	66	37	2	0.8048	0.1952			0.630
rs4842318	78568298	C/T	Non-TD	70	92	13	0.6629	0.3371	0.660	0.713	0.226
			TD	42	52	11	0.6476	0.3524			0.830
rs17005769	78579258	T/C	Non-TD	94	75	6	0.7514	0.2486	0.651	0.726	0.314
			TD	56	43	6	0.7381	0.2619			0.935

TD: tardive dskinesia, HWE: Hardy-Weinberg Equilibrium, SNP: single nuclotide polymorphism

Table 2. Haplotype analysis for PAWR between TD and non-TD subjects

Haplotype	Overeall a visible o	Haplotype fr	- p value	
(rs7979987, rs4842318, rs17005769)	Overall p value	Non-TD	Non-TD TD	
C-C-T		0.229	0.189	0.300
C-T-T		0.000	0.006	0.378
G-C-C	0.80	0.241	0.256	0.718
G-C-T	0.80	0.194	0.203	0.815
G-T-C		0.008	0.006	0.968
G-T-T		0.329	0.340	0.771

TD: tardive dyskinesia

did not identify any association between three PAWR SNPs and TD in Korean schizophrenia patients. However, further studies with a larger sample size and more genetic markers will be necessary to confirm these results.

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