



Article Effect of Allopregnanolone on Spatial Memory and Synaptic Proteins in Animal Model of Metabolic Syndrome

Shaimaa Nasr Amin ^{1,2,*}, Shaimaa Abdalaleem Abdalgeleel ^{3,*}, Mubarak Ali Algahtany ⁴, Sherif Ahmed Shaltout ^{1,5}, Walaa Bayoumie El Gazzar ^{1,6} and Dalia Azmy Elberry ²

- ¹ Department of Basic Medical Sciences, Faculty of Medicine, Hashemite University, Zarqa 13133, Jordan; Sherif@hu.edu.jo (S.A.S.); wallagazzar@hu.edu.jo (W.B.E.G.)
- ² Department of Medical Physiology, Faculty of Medicine, Cairo University, Cairo 11451, Egypt; dalia_azmy@cu.edu.eg
- ³ Department of Biostatistics and Epidemiology, National Cancer Institute, Cairo University, Cairo 11451, Egypt
 ⁴ Division of Neurosurgery, Department of Surgery, College of Medicine, King Khalid University,
- Abha 61421, Saudi Arabia; Mbalgahtany@kku.edu.sa
- ⁵ Department of Pharmacology, Faculty of Medicine, Benha University, Benha 13511, Egypt
- ⁶ Department of Biochemistry, Faculty of Medicine, Benha University, Benha 13511, Egypt
- * Correspondence: shaimaa@hu.edu.jo (S.N.A.); shaimaa.abdalgeleel@nci.cu.edu.eg (S.A.A.); Tel.: +962-770507906 (S.N.A.)

Abstract: Metabolic Syndrome (MetS) is considered a common disorder, especially with a sedentary lifestyle and unhealthy food consumption. Cognitive impairment is one of the MetS consequences that worsens the quality of life of the patients. The study aimed to assess the therapeutic effect of the neurosteroid Allopregnalonone on spatial memory and, therefore, the expression of two synaptic plasticity markers in the hippocampus. Thirty-two male rats were divided into four groups: control groups, MetS, and MetS + Allopregnalone. Spatial memory has been evaluated by the Y-maze task and blood pressure measured by the rat tail method. Biochemical evaluation of serum glucose, insulin, lipid profile, and hippocampal expression of Synaptophysin and Associated Protein 43 (GAP-43) were performed for assessing Allopregnanolone on serum and hippocampal markers. Allopregnanolone therapy improved working spatial memory, hypertension, and biochemical markers measured in the serum and hippocampus.

Keywords: metabolic syndrome; spatial memory; synaptic plasticity; synaptophysin; GAP-43; allopregnanolone

1. Introduction

Metabolic syndrome (MetS) is a common metabolic disorder correlated to a sedentary lifestyle and junk food consumption. The syndrome manifestations are abnormal lipid profile, hypertension, hyperglycemia, insulin resistance (IR), and abdominal obesity [1].

Cognitive dysfunction and dementia are known comorbidities associated with MetS. The suggested mechanisms for impaired cognition in MetS may be related to oxidative stress, abnormal metabolism, inflammation, and vascular reactivity abnormalities. These abnormalities, in turn, disturb the homeostasis of the internal environment around the neurons under basal conditions and more impairment during activity [2]. As encountered in the metabolic syndrome, insulin resistance is found to reduce neurosteroids [3], and neurosteroids administration has been found to improve hypertension in obesity [4].

Neurosteroids are steroid hormones synthesized de novo in the brain and peripheral nervous tissues [5]. Allopregnanolone is an essential neurosteroid in the brain, synthesized locally or in the periphery, and reaches the brain by crossing the blood-brain barrier [6]. Allopregnanolone levels were reported to be downregulated in neurodegenerative diseases [7].

The current work aimed to evaluate the impact of the neurosteroid Allopregnanolone administration as a modulator for Gamma-Amino-Butyric Acid (GABA)-A receptors on



Citation: Amin, S.N.; Abdalgeleel, S.A.; Algahtany, M.A.; Shaltout, S.A.; El Gazzar, W.B.; Elberry, D.A. Effect of Allopregnanolone on Spatial Memory and Synaptic Proteins in Animal Model of Metabolic Syndrome. *Brain Sci.* 2021, *11*, 644. https://doi.org/ 10.3390/brainsci11050644

Academic Editor: Rita Barone

Received: 29 March 2021 Accepted: 11 May 2021 Published: 15 May 2021

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). working spatial memory and hippocampal expression of Synaptophysin and Associated Protein 43 (GAP 43) expression in an animal model of metabolic syndrome (induced by high fructose diet for eight weeks).

2. Materials and Methods

2.1. Experimental Animals

Thirty-two male Sprague–Dawley rats weighing 100–150 g, eight weeks old, were used in this study. Rats were obtained and housed in the ophthalmology research institute, Giza, Egypt. The animals were housed in stainless steel cages (4 rats/cage to avoid isolation stress) at the temperature range of 22 ± 2 °C, under a 12:12 light/dark cycle. The bedding in the cages was wood chips. Rats were allowed to acclimatize for ten days before experimentation; adequate water and chow were available ad libitum.

2.2. Induction of Metabolic Syndrome

Induction of metabolic syndrome in the albino rat model will be achieved by administration of a high-fructose diet (HFD) for eight weeks. HFD is found to induce insulin resistance in rats 3–4 weeks after consuming 60% HFD daily [8]. High-fructose diet (HFD) composed of: starch 2%, sucrose 2%, fructose 60.0%, protein 20.0%, fat, 5.0%, vitamins 1.0%, minerals 5.0% [9]. This formula was mixed with a normal diet in a ratio of 6:4; thus, rats received 60% HFD daily.

2.3. Experimental Design

Rats were divided into the following groups (8 rats/group):

Group I- Negative control: consumed ad libitum water and standard chow diet (7% simple sugars, 3% fat, 50% polysaccharide, 15% protein), energy 3.5 kcal/g [10], and received the vehicle for Allopregnanolone.

Group II: Allopregnanolone group (Positive control): Consumed ad libitum water and standard chow diet (7% simple sugars, 3% fat, 50% polysaccharide, 15% protein), energy 3.5 kcal/g [11] and received Allopregnanolone (Sigma Aldrich, St. Louis, MO, USA) (20 mg/kg, orally) for eight weeks. Allopregnanolone initially dissolved in absolute ethanol then diluted with saline, yielding a final concentration of 20 mg/mL of administration solution. The percentage of ethanol and saline in the final volume is 20% and 80%, respectively [12].

Group III: Metabolic Syndrome (MetS): consumed ad libitum water and HFD for 8 weeks (starch 2%, sucrose 2%, fructose 60.0%, protein 20.0%, fat, 5.0%, vitamins 1.0%, minerals 5.0%) [9]. This formula was mixed with a normal diet in a ratio of 6:4; thus, rats received 60% HFD daily.

Group IV: MetS + *Allopregnanolone group:* received diet as described in group III and starting from week three till the end of week 8; Allopregnanolone administered as described in group II.

Randomization was performed while selecting the rats for each group. For groups, no randomization was done; all rats within the same group received the same protocol for therapy and measurements. At the time of testing, rats were seventeen weeks old. The Y-maze test evaluated rats' spatial working memory, and on the second-day rats, blood pressure was measured (the same day of euthanasia and sampling).

Beginning \rightarrow 10 days acclimatization \rightarrow 8 weeks of modeling \rightarrow Y-maze test \rightarrow after one day of Y-maze testing, blood pressure evaluated followed by euthanasia and sampling.

2.4. Assessment of Spatial Working Memory

2.4.1. Y-Maze Task

Spontaneous alternation in rats refers to rodents' natural tendency to spontaneously choose alternate arms in a Y-maze, and it is considered a quick and relatively simple test of spatial working memory [7].

2.4.2. Apparatus

A wooden Y-maze has three equal size arms (60 cm long, 11.5 cm wide, and 25 cm high). Rats placed in the center of the apparatus to explore the maze for 8 min.

Parameters measured are the number of arm entries, the number of alternations (manually recorded), and the alternation score. Any three consecutive choices of three different arms were counted as a correct choice or alternation. The alternation score is a calculation that reflects the spatial working memory using the number of correct choices or alternation and the total number of arm entries as follow:

Alternation score (%) = (number of alternations)/(total arm entries -2) × 100 [7].

Tests were performed during the light phase of the light/dark cycle after 30 min of acclimatization. Blinding was used during the Y-maze test and analysis of the videos as the investigator performed the tests and the analysis provided with the animals as serial numbers without declaring the groups or therapies. The maze was cleaned with 20% ethanol, and we waited till its odor evaporated before placing another animal to avoid the effect of any olfactory cues.

2.5. In Vivo Measurement of the Arterial Blood Pressure

The blood pressure of animals was indirectly measured by a non-invasive blood pressure monitor (LE 5001, LETICA scientific Instruments, Espania) from conscious rats' tail by the tail-cuff technique. In the tail-cuff technique, animals are warmed for 30 min at 28 °C in a thermostatically controlled heating cabinet (Ugo Basille, Italy) to detect tail artery pulse better. The tail will be passed through a miniaturized cuff, and a tail-cuff sensor connected to an amplifier (LE 5001, LETICA Scientific Instruments, Espania) and BP will be recorded on a chart [13].

At the end of the work, blood samples were collected from retroorbital venous sinuses to measure serum glucose, insulin, and lipid profile. Animals were euthanized, then brains were extracted and dissected at the level of the hippocampus for assessment of the expression of Synaptophysin and Growth Associated Protein 43 (GAP-43):

2.6. Biochemical Measurements

Serum glucose was performed according to the method of Passing and Bablok [14], 1983. The kits were supplied by Diamond Diagnostics Inc., Holliston, MA, USA.

Serum insulin measured by enzyme-linked immunosorbent assay (ELISA) kits (DRG, USA) according to manufacturer's instructions.

Homeostasis model assessment of insulin resistance (HOMA-IR):

HOMA-IR was calculated as fasting insulin $(U/L) \times$ fasting glucose (mg/dL)/405, as described by Matthews et al. [15].

HOMA value of more than 4.0 is an index for insulin resistance [16].

Serum cholesterol measured according to the method of Allain et al. [17]. Serum triglycerides were performed according to the method of Glick et al. [18]. Serum HDL-cholesterol was performed according to the method of Lopez-Virella et al. [19].

2.7. Gene Expression by Real-Time PCR of Synaptophysin and Growth Associated Protein 43 (GAP-43)

Total RNA extraction:

According to manufacturer instructions, total RNA was extracted from hippocampal homogenate using RNA Isolation System (Qiagen, Germantown, MD, USA). The RNA concentrations and purity were measured with an ultraviolet spectrophotometer.

Complementary DNA (cDNA) synthesis: The cDNA was synthesized from 1 µg RNA using SuperScript III First-Strand Synthesis System described in the manufacturer's protocol (#K1621, Fermentas, Waltham, MA, USA).

Real-time quantitative PCR: Real-time PCR amplification and analysis were performed using an Applied Biosystem with software version 3.1 (StepOne[™], Foster City, CA, USA). The reaction contained SYBR Green Master Mix (Applied Biosystems), gene-specific primer pairs, which were shown in Table 1 and designed with Gene Runner Software (Hasting Software, Inc., Hasting, NY, USA) from RNA sequences from the gene bank. All primer sets had a calculated annealing temperature of 60° . Quantitative RT-PCR was performed in a 25-µL reaction volume consisting of 2X SYBR Green PCR Master Mix (Applied Biosystems), 900 nM of each primer, and 2 µL of cDNA. Amplification conditions were: 2 min at 50°, 10 min at 95°, and 40 cycles of denaturation for 15 s, and annealing/extension at 60° for 10 min. Data from real-time assays were calculated using the v1·7 sequence detection software from PE Biosystems (Foster City, CA, USA). Relative expression of studied gene mRNA was calculated using the comparative Ct method. All values were normalized to glyceraldehyde-3-phosphate dehydrogenase, used as the control housekeeping gene, and reported as fold change over background levels detected in the diseased groups.

Table 1. The primer sequence of the studied genes.

Primer Sequence	Accession Number
Forward primer: 5'-TCCAATCAGATGTAGTCTGGTCAG-3' Reverse primer: 5'-AGGCCTTCTCCTGAGCTCTT-3'	NM_009305.2
Forward primer: 5'-TTTCCTCTCTGTCCTGCTC-3' Reverse primer: 5'-TGGACTTGGGATCTTTCCTG-'3	NM_008083.2
Forward primer: 5'-GGTCGGTGTGAACGGATTTGG-3' Reverse primer: 5'-ATGTAGGCCATGAGGTCCACC-3'	NM_001289726.1
	Forward primer: 5'-TCCAATCAGATGTAGTCTGGTCAG-3' Reverse primer: 5'-AGGCCTTCTCCTGAGCTCTT-3' Forward primer: 5'-TTTCCTCTCTGTCCTGCTC-3' Reverse primer: 5'-TGGACTTGGGATCTTTCCTG-'3 Forward primer: 5'-GGTCGGTGTGAACGGATTTGG-3'

GAP-43: Growth Associated Protein 43.

2.8. Statistical Methods

The Statistical Package of Social Science (SPSS) (version 26) was used to generate results. The normality of the data was tested using the Kolmogorov–Smirnov single-sample test. As the data were normally distributed, they were presented as mean and standard deviation (SD). For comparison, an ANOVA test was used to compare groups. Post hoc multiple comparisons were made using the Tukey test. Pearson correlation was used to correlate continuous data. A ($p \le 0.05$) was considered significant.

3. Results

As shown in Table 2:

Mean serum glucoses: it was significantly higher in MetS than in negative control for Allopregnanolone, positive control for Allopregnanolone and MetS + Allopregnanolone (*p*-value ≤ 0.001 , < 0.001 and 0.005 respectively).

In addition, it is significantly lower in MetS+ Allopregnanolone than in negative control for Allopregnanolone and positive control for Allopregnanolone (*p*-value < 0.001).

Mean serum insulin: it was significantly lower in negative control for Allopregnanolone than other groups (*p*-value < 0.001), while it was significantly higher in the MetS group than positive control for Allopregnanolone and MetS+ Allopregnanolone (*p*-value < 0.001 and 0.002 respectively).

Mean HOMA-IR: it was significantly higher in MetS than in other groups ((p-value < 0.001), and it was significantly higher in MetS + Allopregnanolone than negative control for Allopregnanolone and positive control for Allopregnanolone (p-value < 0.001).

Mean serum Triglycerides: it was significantly higher in MetS than negative control for Allopregnanolone and positive control for Allopregnanolone (p-value < 0.001), and it was significantly higher in MetS + Allopregnanolone than negative control for Allopregnanolone and positive control for Allopregnanolone (p-value < 0.001; 0.001 respectively)

Mean serum Cholesterol: it was significantly higher in MetS than in negative control for Allopregnanolone, positive control for Allopregnanolone and MetS + Allopregnanolone (p-value = < 0.001, < 0.001 and 0.005 respectively).

Measured Parameters	Negative Control for Allopregnanolone	Positive Control for Allopregnanolone	MetS	MetS+ Allo- pregnanolone	F-Values	<i>p</i> -Value
Serum Glucose (mg/dL)	$97.3\pm12.9~^{\rm a}$	$107.8\pm8.2~^{\rm a}$	$250.8\pm53.8~^{\rm b}$	189.0 ± 36.6 ^c	37.6	< 0.001
Serum Insulin (µIU/L)	9.2 ± 0.9 ^a	$14.4\pm1.1~^{\rm b}$	$21.3\pm3.9~^{\rm c}$	$16.7\pm2.0~^{\rm b}$	40.1	< 0.001
HOMA-IR	$2.20\pm0.34~^{a}$	$3.72\pm0.44~^{a}$	12.97 ± 2.8 $^{\rm b}$	$7.85\pm2.03~^{c}$	33.5	< 0.001
Serum Triglycerides (mg/dL)	$75.5\pm14.9~^{\rm a}$	$91.2\pm22.9~^{\rm a}$	$143.4\pm17.5^{\text{ b}}$	$126.7\pm5.6^{\text{ b}}$	28.7	<0.001
Serum Cholesterol (mg/dL)	130.9 ± 3.9 a	$146.2\pm25.4~^{\rm a}$	204.6 ± 16.3 ^b	$153.9\pm45.1~^{\rm a}$	11.1	<0.001
Serum HDL (mg/dL)	$58.3\pm3.4~^{\rm a}$	$56.9\pm9.1~^{\rm a}$	$28.6\pm3.2^{\text{ b}}$	$43.1\pm5.1~^{\rm c}$	47.1	< 0.001
Hippocampal Synaptophysin (relative expression)	1.8 ± 0.2 a	$1.3\pm0.1~^{\rm b}$	0.1 ± 0.03 c	$0.9\pm0.1~^{\rm d}$	155.1	<0.001
Hippocampal GAP43 (relative expression)	$1.1\pm0.03~^{\rm a}$	1.2 ± 0.3 $^{\rm a}$	$4.5\pm0.6^{\text{ b}}$	$2.7\pm0.5~^{\rm c}$	112.9	<0.001

Table 2. Biochemical serum and hippocampal markers.

There was a statistically significant difference between a variable with a different letter, no significant difference between variables with the same letter. Degree of freedom between groups equal 3 and within groups equal to 28. MetS: Metabolic syndrome; HOMA-IR: Homeostasis model assessment of insulin resistance; HDL: high-density lipoproteins; GAP 43: Growth Associated Protein 43.

Mean Serum HDL: it was significantly lower in MetS than in other groups (p-value < 0.001), and it significantly lower in MetS + Allopregnanolone than in other groups (p-value < 0.001).

Mean Hippocampal Synaptophysin: All groups were significant from each other, with the lowest mean in the MetS group (p-value < 0.001).

Mean Hippocampal Growth Associated Protein 43 (GAP43) (relative expression): it was significantly higher MetS than other groups (*p*-value < 0.001), the mean higher in MetS + Allopregnanolone than in positive control for Allopregnanolone (*p*-value < 0.001). Table 3 shows:

Table 3. Arterial blood pressure measurements in the studied groups.

Measured Parameters	Negative Control for Allopregnanolone	Positive Control for Allopregnanolone	MetS	MetS + Allopregnanolone	F-Value	<i>p-</i> Value
Systolic BP (mmHg)	$116.8\pm4.1~^{\rm a}$	$104.9\pm4.3~^{\rm b}$	$151.5\pm11.0~^{\rm c}$	$118.5\pm5.1~^{\rm a}$	70.3	< 0.001
Diastolic BP (mmHg)	$84.5\pm6.6~^{\rm a}$	$86.4\pm3.6~^{\rm a}$	$113.6\pm6.3^{\text{ b}}$	90.9 ± 2.3 ^a	56.6	<0.001
MAP (mmHg)	96.6 ± 4.2 a	91.5 ± 3.7 a	$128.3\pm8.7^{\text{ b}}$	100.4 ± 3.4 a	72.6	< 0.001

There was a statistically significant difference between a variable with a different letter, no significant difference between variables with the same letter. Degree of freedom between groups equal 3 and within groups equal to 28. MAP: Mean arterial blood pressure.

Mean systolic blood pressure: it was significantly higher in MetS than in other groups (p-value < 0.001), and it was significantly higher in MetS + Allopregnanolonen than in positive control for Allopregnanolone (p value = 0.002)

Mean diastolic blood pressure was significantly higher in the MetS group than in other groups (p-value < 0.001).



Mean arterial blood pressure: significantly higher in the MetS group than in other groups (*p*-value < 0.001). As shown in Figure 1:

Figure 1. Alteration score in the studied groups in the Y-maze test. Alternation score (%) = (number of alternations)/(total arm entries -2) × 100. Data are expressed as mean \pm SD; n = 8 in each group.

Alternation score in Y-Maze (%), negative control for Allopregnanolone (33.2 \pm 5.4), positive control for Allopregnanolone 37.1 \pm 4.7, MetS 22.6 \pm 11.6 and for MetS+ Allopregnanolone (39.3 \pm 11.0) with a significant lower mean in MetS group than positive control for Allopregnanolone and for MetS + Allopregnanolone (*p*-value = 0.031 and 0.009 respectively).

Table 4 and Figures 2 and 3 show:

Table 4. Correlation between alternation score in Y-maze (%) and biochemical markers.

Biochemical Parameters	Alternation Score in Y-Maze (%)			
	Correlation Coefficient (r)	p Value		
Serum Glucose (mg/dL)	-0.086	0.640		
Serum Insulin ((µIU/L)	-0.334	0.062		
Serum Triglycerides (mg/dL)	-0.301	0.094		
Serum Cholesterol (mg/dL)	-0.204	0.262		
Serum HDL (mg/dL)	0.317	0.077		
Hippocampal Synaptophysin (relative expression)	0.366	0.039		
Hippocampal Growth Associated Protein 43 (GAP43) (relative expression)	-0.414	0.019		



Figure 2. Significant negative correlation between the alternation score and Hippocampal Growth Associated Protein 43 expression.



Figure 3. Significant positive correlation between alternation score and hippocampal synaptophysin expression.

Significant positive correlation between alternation score and hippocampal synaptophysin expression (*p*-value = 0.039), Significant negative correlation between the alternation score and Hippocampal GAP 43 expression (*p*-value = 0.019), however no significant correlation was found between serum glucose and serum insulin, serum triglycerides, serum cholesterol, serum HDL and alternation score (*p*-value = 0.640, 0.062, 0.094, 0.262 and 0.077 respectively).

4. Discussion

MetS is a critical universal disorder that affects multiple brain regions, especially the hippocampus, concerned with memory and learning processes [20,21]. A commonly used

maze test is the Y-maze task [22], used in our study to assess working spatial memory via calculating the alternation score. Our results revealed marked deterioration of spatial working memory in the form of a significant ($p \le 0.05$) decrease in the alternation score of the MetS group compared to the negative control group (Figure 1). These results concord with previous studies that revealed a significant defect in spatial working memory processes in rodents with diet-induced MetS, using the Barnes maze [23], the Morris water maze [24], and novel objects test (NORT) [20].

The present study showed a significant decrease in the relative expression of hippocampal Synaptophysin of MetS (Table 2). This effect most probably was a reason for the apparent spatial working memory changes in the MetS group, evidenced by the significant positive correlation between the alternation score and relative expression of Synaptophysin (Figure 3). This outcome was consistent with the work done by Trevino et al. [20], who claimed that immunoreactivity to Synaptophysin in the hippocampus of high-calorie dietinduced MetS animals (with memory impairments) compared to control. Synaptophysin is an essential marker of neuronal plasticity [25]. It is an abundant integral membrane protein of presynaptic vesicles involved in regulating neurotransmitter release and synaptic plasticity [26]. Synaptophysin plays a crucial role in neurogenesis and neuronal interaction, particularly in degenerative diseases such as Alzheimer's disease [27].

Astrocytes play a vital role in maintaining neurotransmission and maintaining synapses, reflecting cognitive function [28]. Among various messengers released by astrocytes, Thrombospondin-1 (TSP-1) [26]. TSP-1 deficiency impairs the presynaptic proteins, such as Synaptophysin [29]. Moreover, activation of Toll-like receptor 9 (TLR9) by hyperglycemia mediates oxidative stress and astrocytic dysfunction, leading to reduced TSP-1 secretion and synaptophysin expression [30]. Since the metabolic disorders in MetS (dyslipidemia, hyperglycemia, and IR); were all demonstrated in our results; this may suggest excessive oxidative stress and inflammation, increasing the susceptibility of the CNS to injury [31].

Furthermore, fructose may directly affect brain functions and increase hippocampal and cortical glucose transporter 5 (GLUT5) [32]. Fructose has higher reactivity than glucose, producing several alterations and increasing oxidative stress events [33].

GAP-43, as with Synaptophysin, is expressed in the terminals of presynaptic vesicles [34]. It worth noting that GAP-43 is involved in synaptic plasticity processes such as long-term potentiation (LTP) and is correlated to anatomical arborization of new synapses [35]. Unexpectedly and contradictory to many previous studies, our results revealed a significant increase in the relative expression of hippocampal GAP 43 in the MetS group compared to the control group (Table 2).

Additionally, there is a significant negative correlation between alternation score and relative expression of GAP 43 (Table 4, Figure 2). These findings are not concomitant with Gu et al., [36] research which demonstrated that streptozotocin (STZ)-induced diabetes in rat models produced marked deterioration of spatial learning and memory with downregulation of GAP-43 expression in the hippocampus. This discrepancy in results might be due to the diversity of the animal models employed [36]. However, our results agree with the work done by Sandelius et al. [37].

Besides, increased GAP-43 in the area of infarction in experimental brain ischemia has been reported indicating neuronal injury and possible regeneration [38,39]. These previous studies may partly explain our results, as it was documented that fructose administration leads to a reduction in cerebral blood flow (CBF) in the hippocampus and other parts of the brain [40,41]. Moreover, Hung et al. [42] reported that lipopolysaccharide (LPS) induced GAP-43 through ROS and nuclear factor-B (NF-B) [42]. This fact may be linked to ROS accumulation with high fructose intake [43].

Our results demonstrated the prominent therapeutic effect of Allopregnanolone on working spatial memory, evidenced by the significant improvement of alternation score in the Allopregnanolone group compared to the MetS group (Figure 1). These results are concomitant with the study done by Park et al., [6], who assessed the effect of Allopregnanolone on spatial learning and memory in aged mice using the radial arm water maze. Additionally, Frye et al. [44] demonstrated that Allopregnanolone administration improved T-maze and water maze task performance.

It worth noting that Allopregnanolone was approved in the united states of America for the treatment of postpartum depression [45]. Allopregnanolone is a positive modulator of GABAAR activity [5]. GABA is the primary inhibitory neurotransmitter in the brain and plays a vital role in learning and memory [46] and synaptic plasticity [47]. The hippocampus has numerous GABAA receptors that participate in molecular mechanisms of memory formation [48].

In a pilot study performed by Napoli et al. [49], allopregnanolone treatment significantly impacted GABA metabolism, oxidative stress, and other mitochondria-related outcomes. Allopregnanolone effectively lowered the content of the three markers of oxidative stress, including 2-hydroxybutyric acid [49], which is an early marker for both insulin resistance and impaired glucose regulation from increased lipid peroxidation and oxidative stress [50]. This effect may propose a primitive explanation for improving spatial memory (increased alternation score) after Allopregnanolone administration in our study.

Furthermore, Frye et al. [51] showed cognitive enhancement of progesterone (induce the formation of Allopregnanolone) that was exerted through increasing GABAA receptors activity of the hippocampus and hippocampal brain-derived neurotrophic factor (BDNF) level. BDNF the neurotrophic factor that controls synaptic plasticity [52,53]. Several studies reported increased BDNF levels following Allopregnanolone administration [54–56].

In Xu et al., [57] study, infusion of BDNF into the hippocampus reversed that cognitive impairment due to OA (okadaic acid) injection and upregulated Synaptophysin expression. Zhong et al. [58] also claimed the protective effect of BDNF on rat hippocampal neurons exposed to high glucose in vitro by increasing Synaptophysin levels. These studies propose a logical cause for the marked increase of relative expression of Synaptophysin in the allopregnanolone group compared to the MetS group (Table 2).

Regarding GAP-43, our study showed a prominent decrease in its relative expression in the allopregnanolone group compared to the MetS group (Table 3). However, there is still a significant difference between the relative expression of GAP-43 in the allopregnanolone group compared to the negative control group (Table 3). This result can also be explained by the ability of Allopregnanolone to increase hippocampal BDNF level [51]. It is worth noting that BDNF increases antioxidant enzymes' activity in cells and fights against free radicals [59].

Besides, previous studies have shown that Allopregnanolone also can increase hippocampal neurogenesis [42]. Allopregnanolone increases the expression of genes that promote mitosis and reduces antimitotic gene expression [60,61]. In the present study, allopregnanolone administration, a ligand to GABAA receptors [62], caused a decremental decrease in serum glucose, serum insulin, and HOMA-IR in group IV in comparison to group III. These findings run parallel to what was previously reported by Tian et al. [63]. It has been reported that GABAARs are expressed in the primary islet β -cells [64] that regulate insulin secretion together with changes in glucose levels [65].

Another finding of our study is that the role of Allopregnanolone in improving the lipid profile indicated by the decreased levels of serum triglycerides and cholesterol. Previous studies have reported that the mechanism of Allopregnanolone action is thought to involve pregnane X receptor (PXR) activation [66]. PXR transcriptionally activates Insig-1, a protein with antilipogenic properties, which accordingly reduces the nuclear protein level of the active sterol regulatory element-binding protein 1 (SREB*P*-1) with subsequent reduction of transcription of specific lipogenic genes, causing a decrease in fatty acid, triglyceride and cholesterol synthesis [67,68].

Our findings indicate that AlloP has antihypertensive effects in experimentally induced metabolic syndrome in rats. It produced a marked decrease in systolic blood pressure (SBP), diastolic blood pressure (DBP), and mean blood pressure (MBP). These findings are in line with a previous study that found that the neurosteroid allopregnanolone effectively reduces BP and also the pressor response to stress in a mouse model of neurogenic hypertension [69]. Additionally, other clinical studies showed that high levels of Allopregnanolone were correlated with a blunted SBP response to cocaine cues in cocaine-dependent individuals [70]. The previous studies support these findings reported the antihypertensive effect of GABA-rich Chlorella in spontaneously hypertensive rats (SHR) and borderline hypertension subjects [71]. High blood pressure could be a classical feature of metabolic syndrome, and it has been reported that metabolic syndrome is present in up to 30% of hypertensive patients [72]. The sympathetic system activation is characteristic of several metabolic disorders., many of which occur more frequently in obese individuals. Obesity potentiates sympathetic system activation in patients with hypertension [73]. Multiple studies have demonstrated that metabolic syndrome is accompanied by a hyperadrenergic state independent of the presence of hypertension [74].

The elevated plasma insulin levels may account for multiple metabolic syndrome features since insulin has been demonstrated to change glucose and lipid metabolism, enhance atherogenesis, and increase blood pressure [75]. Human studies revealed a heterogenous sympathetic response with increased muscle sympathetic activity induced by insulin [76,77].

The arterial baroreceptor control of sympathetic nerve traffic undergoes a clear-cut impairment in patients with metabolic syndrome, in which both the sympathoinhibitory and the sympathoexcitatory baroreflex components are involved [74]. The BP lowering actions of Allopregnanolone involve sympatho-inhibition via upregulation of GABAA receptors that specifically contain δ subunits within the hypothalamus and amygdala [69]. These δ subunits containing GABAA receptors are presumably extrasynaptic and very sensitive to GABA and Allopregnanolone but relatively insensitive to the action of benzo-diazepines [11,78].

5. Conclusions

The present study revealed a positive impact of allopregnanolone therapy in animal models of metabolic syndrome. Allopregnanolone therapy improved working spatial memory and biochemical markers of metabolic syndrome (serum glucose, insulin, insulin resistance, lipid profile). Moreover, it improved the expression of hippocampal Synaptophysin that may explain the improved performance in Y-maze. Further studies are recommended to evaluate other aspects of cognition with a molecular evaluation of other synaptic plasticity markers, especially BDNF and signaling pathways, to understand better the mechanism involved in allopregnanolone effects. It is advised to separate the drug's effect from the alcohol (vehicle), so further investigations are advised with comparing MetS treated with Allopregnanolone dissolved in the vehicle with MetS treated with vehicle alone. Besides, more investigations are required to evaluate the effect of Allopregnanolone in females with metabolic syndrome considering fluctuations in female sex hormones in estrus or menstrual cycle and their effects on the brain.

Author Contributions: S.N.A.: Conceptualization, methodology, investigation validation, supervision, project administration, writing—original draft preparation, writing—review and editing. S.A.A.: methodology data curation; writing. M.A.A.: investigation, writing—original draft preparation. S.A.S.: investigation, writing—original draft preparation. W.B.E.G.: investigation, writing—original draft preparation. D.A.E.: investigation, writing—original draft preparation. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Institutional Review Board Statement: The study was conducted according to the Declaration of Helsinki's guidelines and approved by the Institutional Review Board of Hashemite University (NO.13/6/2020/2021).

Informed Consent Statement: Not applicable.

Data Availability Statement: The data that support the findings of this study are available from the corresponding author S.N.A., upon reasonable request.

Conflicts of Interest: The authors declare no conflict of interest.

References

- 1. Kassi, E.; Pervanidou, P.; Kaltsas, G.; Chrousos, G. Metabolic syndrome: Definitions and controversies. *BMC Med.* **2011**, *9*, 48. [CrossRef]
- Yates, K.F.; Sweat, V.; Yau, P.L.; Turchiano, M.M.; Convit, A. Impact of metabolic syndrome on cognition and brain: A selected review of the literature. *Arter. Thromb. Vasc. Biol.* 2012, *32*, 2060–2067. [CrossRef] [PubMed]
- Genazzani, A.D.; Strucchi, C.; Luisi, M.; Casarosa, E.; Lanzoni, C.; Baraldi, E.; Ricchieri, F.; Mehmeti, H.; Genazzani, A.R. Metformin administration modulates neurosteroids secretion in non-obese amenorrhoic patients with polycystic ovary syndrome. *Gynecol. Endocrinol.* 2006, 22, 36–43. [CrossRef] [PubMed]
- 4. Head, G.A.; Jackson, K.L.; Gueguen, C. Potential Therapeutic Use of Neurosteroids for Hypertension. *Front. Physiol.* **2019**, *10*, 1477. [CrossRef]
- 5. Lloyd-Evans, E.; Waller-Evans, H. Biosynthesis and signalling functions of central and peripheral nervous system neurosteroids in health and disease. *Essays Biochem.* **2020**, *64*, 591–606. [CrossRef] [PubMed]
- Parks, E.E.; Logan, S.; Yeganeh, A.; Farley, J.A.; Owen, D.B.; Sonntag, W.E. Interleukin 6 reduces allopregnanolone synthesis in the brain and contributes to age-related cognitive decline in mice. *J. Lipid Res.* 2020, *61*, 1308–1319. [CrossRef] [PubMed]
- 7. Noorbakhsh, F.; Baker, G.B.; Power, C. Allopregnanolone and neuroinflammation: A focus on multiple sclerosis. *Front. Cell. Neurosci.* **2014**, *8*, 134. [CrossRef]
- Gajda, A.; Pellizzon, M.; Ricci, M.; Ulman, E. Diet-Induced Metabolic Syndrome in Rodent Models. Animal Lab News. 2007. Available online: http://www.eps-cjgroup.com/lsg/service/researchdiets/pdf/Diet-Induced%20Metabolic%20Syndrome% 20models.pdf (accessed on 1 September 2020).
- Nagai, Y.; Yonemitsu, S.; Erion, D.M.; Iwasaki, T.; Stark, R.; Weismann, D.; Dong, J.; Zhang, D.; Jurczak, M.J.; Löffler, M.G.; et al. The role of peroxisome proliferator-activated receptor gamma coactivator-1 beta in the pathogenesis of fructose-induced insulin resistance. *Cell Metab.* 2009, *9*, 252–264. [CrossRef]
- Samuelsson, A.M.; Matthews, P.A.; Argenton, M.; Christie, M.R.; McConnell, J.M.; Jansen, E.H. Diet-induced obesity in female mice leads to offspring hyperphagia, adiposity, hypertension, and insulin resistance: A novel murine model of developmental programming. *Hypertension* 2008, *51*, 383–392. [CrossRef]
- 11. Maguire, J.; Mody, I. GABAAR plasticity during pregnancy: Relevance to postpartum depression. *Neuron* **2008**, *59*, 207–213. [CrossRef] [PubMed]
- 12. Afrazi, S.; Esmaeili-Mahani, S.; Sheibani, V.; Abbasnejad, M. Neurosteroid allopregnanolone attenuates high glucose-induced apoptosis and prevents experimental diabetic neuropathic pain: In vitro and in vivo studies. *J. Steroid. Biochem. Mol. Biol.* 2014, 139, 98–103. [CrossRef]
- 13. Irvine, R.J.; White, J.; Chan, R. The influence of restraint on blood pressure in the rat. *J. Pharmacol. Toxicol.* **1997**, *38*, 157–162. [CrossRef]
- 14. Passing, H.; Bablok, W. A new biometrical procedure for testing the equality of measurements from two different analytical methods. Application of linear regression procedures for method comparison studies in clinical chemistry, Part I. J. Clin. Chem. Clin. Biochem. **1983**, *21*, 709–720. [CrossRef] [PubMed]
- Matthews, D.R.; Hosker, J.P.; Rudenski, A.S.; Naylor, B.A.; Treacher, D.F.; Turner, R.C. Homeostasis model assessment: Insulin resistance and beta-cell function from fasting plasma glucose and insulin concentrations in man. *Diabetologia* 1985, 28, 412–419. [CrossRef]
- 16. Becker, R.H.A.; Frick, A.D.; Burger, F.; Potgieter, J.H.; Scholtz, H. Insulin glulisine, a new rapid-acting insulin analogue, displays a rapid time-action profile in obese non-diabetic subjects. *Exp. Clin. Endocrinol. Diabetes* **2005**, *113*, 435–443. [CrossRef]
- 17. Allain, C.C.; Poon, L.S.; Chan, C.S.; Richmond, W.; Fu, P.C. Enzymatic determination of total serum cholesterol. *Clin. Chem.* **1974**, 20, 470–475. [CrossRef]
- Glick, M.R.; Ryder, K.W.; Jackson, S.A. Graphical comparisons of interferences in clinical chemistry instrumentation. *Clin. Chem.* 1986, 32, 470–474. [CrossRef] [PubMed]
- 19. Lopez-virella, M.F.; Stone, P.; Ellis, S.; Collwel, J. Cholesterol determination in high density lipoprotein by three different methods. *Clin. Chem.* **1977**, *23*, 882. [CrossRef]
- Treviño, S.; Vázquez-Roque, R.A.; López-López, G.; Perez-Cruz, C.; Moran, C.; Handal-Silva, A.; González-Vergara, E.; Flores, G.; Guevara, J.; Díaz, A. Metabolic syndrome causes recognition impairments and reduced hippocampal neuronal plasticity in rats. *J. Chem. Neuroanat.* 2017, *82*, 65–75. [CrossRef] [PubMed]
- Haberman, R.P.; Colantuoni, C.; Koh, M.T.; Gallagher, M. Behaviorally activated mRNA expression profiles produce signatures of learning and enhanced inhibition in aged rats with preserved memory. *PLoS ONE* 2013, *8*, e83674. [CrossRef]
- 22. Arai, K.; Matsuki, N.; Ikegaya, Y.; Nishiyama, N. Deterioration of spatial learning performances in lipopolysaccharide-treated mice. *Jpn. J. Pharmacol.* 2001, *87*, 195–201. [CrossRef]
- 23. Dallagi, Y.; Rahali, D.; Perrotte, M.; Dkhili, H.; Korsan, A.; El May, M.V.; El Fazaa, S.; Ramassamy, C.; El Golli, N. Date seeds alleviate behavioural and neuronal complications of metabolic syndrome in rats. *Arch. Physiol. Biochem.* **2020**, *8*, 1–15. [CrossRef]

- Treviño, S.; Aguilar-Alonso, P.; Flores Hernandez, J.A.; Brambila, E.; Guevara, J.; Flores, G.; Lopez-Lopez, G.; Muñoz-Arenas, G.; Morales-Medina, J.C.; Toxqui, V. A high calorie diet causes memory loss, metabolic syndrome and oxidative stress into hippocampus and temporal cortex of rats. *Synapse* 2015, *69*, 421–433. [CrossRef]
- Muñoz-Arenas, G.; Pulido, G.; Treviño, S.; Vázquez-Roque, R.; Flores, G.; Moran, C.; Handal-Silva, A.; Guevara, J.; Venegas, B.; Díaz, A. Effects of metformin on recognition memory and hippocampal neuroplasticity in rats with metabolic syndrome. *Synapse* 2020, 74, e22153. [CrossRef] [PubMed]
- 26. Jayakumar, A.R.; Tong, X.Y.; Shamaladevi, N.; Barcelona, S.; Gaidosh, G.; Agarwal, A.; Norenberg, M.D. Defective synthesis and release of astrocytic thrombospondin-1 mediates the neuronal TDP-43 proteinopathy, resulting in defects in neuronal integrity associated with chronic traumatic encephalopathy: In vitro studies. *J. Neurochem.* **2017**, *140*, 645–661. [CrossRef] [PubMed]
- Choi, J.-K.; Carreras, I.; Aytan, N.; Jenkins-Sahlin, E.; Dedeoglu, A.; Jenkins, B.G. The effects of aging, housing and ibuprofen treatment on brain neurochemistry in a triple transgene Alzheimer's disease mouse model using magnetic resonance spectroscopy and imaging. *Brain Res.* 2014, 1590, 85–96. [CrossRef] [PubMed]
- Dallérac, G.; Rouach, N. Astrocytes as new targets to improve cognitive functions. *Prog. Neurobiol.* 2016, 144, 48–67. [CrossRef]
 Elberry, D.A.; Amin, S.N.; Esmail, R.S.E.N.; Rashed, L.A.; Gamal, M.M. Effect of undifferentiated versus hepatogenic partially
- differentiated mesenchymal stem cells on hepatic and cognitive functions in liver cirrhosis. *Excli J.* 2016, *15*, 652–670. [CrossRef]
 30. Zhao, Y.; Pu, D.; Sun, Y.; Chen, J.; Luo, C.; Wang, M.; Zhou, J.; Lv, A.; Zhu, S.; Liao, Z. High glucose-induced defective thrombospondin-1 release from astrocytes via TLR9 activation contributes to the synaptic protein loss. *Exp. Cell Res.* 2018, *363*, 171–178. [CrossRef] [PubMed]
- 31. Stranahan, A.M.; Cutler, R.G.; Button, C.; Telljohann, R.; Mattson, M.P. Diet-induced elevations in serum cholesterol are associated with alterations in hippocampal lipid metabolism and increased oxidative stress. *J. Neurochem.* **2011**, *118*, 611–615. [CrossRef]
- 32. Funari, V.A.; Crandall, J.E.; Tolan, D.R. Fructose metabolism in the cerebellum. *Cerebellum* 2007, 6, 130. [CrossRef]
- Rivera, D.S.; Lindsay, C.B.; Codocedo, J.F.; Carreño, L.E.; Cabrera, D.; Arrese, M.A.; Vio, C.P.; Bozinovic, F.; Inestrosa, N.C. Long-Term, Fructose-Induced Metabolic Syndrome-Like Condition Is Associated with Higher Metabolism, Reduced Synaptic Plasticity and Cognitive Impairment in Octodon degus. *Mol. Neurobiol.* 2018, 55, 9169–9187. [CrossRef] [PubMed]
- 34. Nemes, A.D.; Ayasoufi, K.; Ying, Z.; Zhou, Q.-G.; Suh, H.; Najm, I.M. Growth Associated Protein 43 (GAP-43) as a Novel Target for the Diagnosis, Treatment and Prevention of Epileptogenesis. *Sci. Rep.* **2017**, *7*, 17702. [CrossRef] [PubMed]
- 35. Holahan, M.R. A Shift from a Pivotal to Supporting Role for the Growth-Associated Protein (GAP-43) in the Coordination of Axonal Structural and Functional Plasticity. *Front. Cell. Neurosci.* **2017**, *11*, 266. [CrossRef] [PubMed]
- Gu, J.; Cheng, X.; Luo, X.; Yang, X.; Pang, Y.; Zhang, X.; Zhang, Y.; Liu, Y. Luteolin Ameliorates Cognitive Impairments by Suppressing the Expression of Inflammatory Cytokines and Enhancing Synapse-Associated Proteins GAP-43 and SYN Levels in Streptozotocin-Induced Diabetic Rats. *Neurochem. Res.* 2018, 43, 1905–1913. [CrossRef] [PubMed]
- Sandelius, Å.; Portelius, E.; Källén, Å.; Zetterberg, H.; Rot, U.; Olsson, B.; Toledo, J.B.; Shaw, L.M.; Lee, V.M.Y.; Irwin, D.J.; et al. Elevated CSF GAP-43 is Alzheimer's disease specific and associated with tau and amyloid pathology. *Alzheimers Dement.* 2019, 15, 55–64. [CrossRef] [PubMed]
- 38. Gorup, D.; Bohaček, I.; Miličević, T.; Pochet, R.; Mitrečić, D.; Križ, J.; Gajović, S. Increased expression and colocalization of GAP43 and CASP3 after brain ischemic lesion in mouse. *Neurosci. Lett.* **2015**, *15*, 176–182. [CrossRef]
- Sandelius, Å.; Cullen, N.C.; Källén, Å.; Rosengren, L.; Jensen, C.; Kostanjevecki, V.; Vandijck, M.; Zetterberg, H.; Blennow, K. Transient increase in CSF GAP-43 concentration after ischemic stroke. *BMC Neurol.* 2018, 18, 202. [CrossRef]
- 40. Page, K.A.; Chan, O.; Arora, J.; Belfort-DeAguiar, R.; Dzuira, J.; Roehmholdt, B.; Cline, G.W.; Naik, S.; Sinha, R.; Constable, R.T.; et al. Effects of Fructose vs Glucose on Regional Cerebral Blood Flow in Brain Regions Involved with Appetite and Reward Pathways. *JAMA* 2013, *309*, 63–70. [CrossRef] [PubMed]
- Calvo-Ochoa, E.; Hernández-Ortega, K.; Ferrera, P.; Morimoto, S.; Arias, C. Short-Term High-Fat-and-Fructose Feeding Produces Insulin Signaling Alterations Accompanied by Neurite and Synaptic Reduction and Astroglial Activation in the Rat Hippocampus. *J. Cereb. Blood Flow Metab.* 2014, 34, 1001–1008. [CrossRef]
- Hung, C.-C.; Lin, C.-H.; Chang, H.; Wang, C.-Y.; Lin, S.-H.; Hsu, P.-C.; Sun, Y.-Y.; Lin, T.-N.; Shie, F.-S.; Kao, L.-S.; et al. Astrocytic GAP43 Induced by the TLR4/NF-κB/STAT3 Axis Attenuates Astrogliosis-Mediated Microglial Activation and Neurotoxicity. J. Neurosci. 2016, 36, 2027–2043. [CrossRef]
- Mastrocola, R.; Nigro, D.; Cento, A.S.; Chiazza, F.; Collino, M.; Aragno, M. High-fructose intake as risk factor for neurodegeneration: Key role for carboxy methyllysine accumulation in mice hippocampal neurons. *Neurobiol. Dis.* 2016, 89, 65–75. [CrossRef]
- 44. Frye, C.A.; Koonce, C.J.; Walf, A.A. Progesterone, compared to medroxyprogesterone acetate, to C57BL/6, but not 5α-reductase mutant, mice enhances object recognition and placement memory and is associated with higher BDNF levels in the hippocampus and cortex. *Neurosci. Lett.* 2013, 551, 53–57. [CrossRef] [PubMed]
- 45. Meltzer-Brody, S.; Kanes, S.J. Allopregnanolone in postpartum depression: Role in pathophysiology and treatment. *Neurobiol Stress.* **2020**, *12*, 100212. [CrossRef] [PubMed]
- 46. Bonansco, C.; Fuenzalida, M. Plasticity of Hippocampal Excitatory-Inhibitory Balance: Missing the Synaptic Control in the Epileptic Brain. *Neural. Plast.* **2016**, e8607038. [CrossRef] [PubMed]

- Kim, J.; Son, Y.; Kim, J.; Lee, S.; Kang, S.; Park, K.; Kim, S.H.; Kim, J.C.; Kim, J.; Takayama, C.; et al. Developmental and degenerative modulation of GABAergic transmission in the mouse hippocampus. *Int. J. Dev. Neurosci.* 2015, 47, 320–332. [CrossRef]
- 48. Whissell, P.D.; Rosenzweig, S.; Lecker, I.; Wang, D.-S.; Wojtowicz, J.M.; Orser, B.A. γ-aminobutyric acid type A receptors that contain the δ subunit promote memory and neurogenesis in the dentate gyrus. *Ann. Neurol.* **2013**, *74*, 611–621. [CrossRef]
- 49. Napoli, E.; Schneider, A.; Wang, J.Y.; Trivedi, A.; Carrillo, N.R.; Tassone, F.; Rogawski, M.; Hagerman, R.J.; Giulivi, C. Allopregnanolone Treatment Improves Plasma Metabolomic Profile Associated with GABA Metabolism in Fragile X-Associated Tremor/Ataxia Syndrome: A Pilot Study. *Mol. Neurobiol.* **2019**, *56*, 3702–3713. [CrossRef]
- 50. Gall, W.E.; Beebe, K.; Lawton, K.A.; Adam, K.-P.; Mitchell, M.W.; Nakhle, P.J.; Ryals, J.A.; Milburn, M.V.; Nannipieri, M.; Camastra, S.; et al. Group, for the the RS α-Hydroxybutyrate Is an Early Biomarker of Insulin Resistance and Glucose Intolerance in a Nondiabetic Population. *PLoS ONE* 2010, *5*, e10883. [CrossRef]
- Frye, C.A.; Lembo, V.F.; Walf, A.A. Progesterone's Effects on Cognitive Performance of Male Mice Are Independent of Progestin Receptors but Relate to Increases in GABAA Activity in the Hippocampus and Cortex. *Front. Endocrinol.* 2021, *11*, 552805. [CrossRef] [PubMed]
- 52. Kowiański, P.; Lietzau, G.; Czuba, E.; Waśkow, M.; Steliga, A.; Moryś, J. BDNF: A Key Factor with Multipotent Impact on Brain Signaling and Synaptic Plasticity. *Cell. Mol. Neurobiol.* **2018**, *38*, 579–593. [CrossRef] [PubMed]
- 53. Pistell, P.J.; Morrison, C.D.; Gupta, S.; Knight, A.G.; Keller, J.N.; Ingram, D.K.; Bruce-Keller, A.J. Cognitive impairment following high fat diet consumption is associated with brain inflammation. *J. Neuroimmunol.* **2010**, *219*, 25–32. [CrossRef]
- Wang, T.; Ye, X.; Bian, W.; Chen, Z.; Du, J.; Li, M.; Zhou, P.; Cui, H.; Ding, Y.Q.; Qi, S.; et al. Allopregnanolone Modulates GABAAR-Dependent CaMKIIδ3 and BDNF to Protect SH-SY5Y Cells Against 6-OHDA-Induced Damage. *Front Cell Neurosci.* 2020, 13, 569. [CrossRef]
- 55. Almeida, F.B.; Nin, M.S.; Barros, H.M.T. The role of Allopregnanolone in depressive-like behaviors: Focus on neurotrophic proteins. *Neurobiol. Stress.* 2020, 12, 100218. [CrossRef]
- Almeida, F.B.; Barros, H.M.T.; Pinna, G. Neurosteroids and Neurotrophic Factors: What Is Their Promise as Biomarkers for Major Depression and PTSD? *Int. J. Mol. Sci.* 2021, 22, 1758. [CrossRef]
- 57. Xu, A.-H.; Yang, Y.; Sun, Y.-X.; Zhang, C.-D. Exogenous brain-derived neurotrophic factor attenuates cognitive impairment induced by okadaic acid in a rat model of Alzheimer's disease. *Neural Regen. Res.* **2018**, *13*, 2173–2181. [CrossRef]
- Zhong, Y.; Zhu, Y.; He, T.; Li, W.; Li, Q.; Miao, Y. Brain-derived neurotrophic factor inhibits hyperglycemia-induced apoptosis and downregulation of synaptic plasticity-related proteins in hippocampal neurons via the PI3K/Akt pathway. *Int. J. Mol. Med.* 2019, 43, 294–304. [CrossRef] [PubMed]
- Sampaio, T.B.; Savall, A.S.; Gutierrez, M.E.Z.; Pinton, S. Neurotrophic factors in Alzheimer's and Parkinson's diseases: Implications for pathogenesis and therapy. *Neural Regen. Res.* 2017, 12, 549–557. [CrossRef] [PubMed]
- Wang, J.M.; Johnston, P.B.; Ball, B.G.; Brinton, R.D. The Neurosteroid Allopregnanolone Promotes Proliferation of Rodent and Human Neural Progenitor Cells and Regulates Cell-Cycle Gene and Protein Expression. J. Neurosci. 2005, 25, 4706–4718. [CrossRef]
- 61. Singh, C.; Liu, L.; Wang, J.M.; Irwin, R.W.; Yao, J.; Chen, S.; Henry, S.; Thompson, R.F.; Brinton, R.D. Allopregnanolone restores hippocampal-dependent learning and memory and neural progenitor survival in aging 3xTgAD and nonTg mice. *Neurobiol. Aging* **2012**, *33*, 1493–1506. [CrossRef] [PubMed]
- 62. Haage, D.; Johansson, S. Neurosteroid modulation of synaptic and GABAevoked currents in neurons from the rat medial preoptic nucleus. *J. Neurophysiol.* **1999**, *82*, 143–151. [CrossRef]
- 63. Tian, J.; Dang, H.N.; Yong, J.; Chui, W.S.; Dizon MP, G.; Yaw CK, Y.; Kaufman, D.L. Oral treatment with gamma-aminobutyric acid improves glucose tolerance and insulin sensitivity by inhibiting inflammation in high fat diet-fed mice. *PLoS ONE* **2011**, *6*, e25338. [CrossRef]
- 64. Xu, E.; Kumar, M.; Zhang, Y.; Ju, W.; Obata, T.; Zhang, N.; Liu, S.; Wendt, A.; Deng, S.; Ebina, Y.; et al. Intra-islet insulin suppresses glucagon release via GABA-GABAA receptor system. *Cell Metab.* **2006**, *3*, 47–58. [CrossRef]
- 65. Braun, M.; Ramracheya, R.; Bengtsson, M.; Clark, A.; Walker, J.N.; Johnson, P.R.; Rorsman, P. Gamma-Aminobutyric Acid (GABA) Is an Autocrine Excitatory Transmitter in Human Pancreatic Beta-Cells. *Diabetes* **2010**, *59*, 1694–1701. [CrossRef] [PubMed]
- 66. Langmade, S.J.; Gale, S.E.; Frolov, A.; Mohri, I.; Suzuki, K.; Mellon, S.H.; Walkley, S.U.; Covey, D.F.; Schaffer, J.E.; Ory, D.S. Pregnane X receptor (PXR) activation: A mechanism for neuroprotection in a mouse model of Niemann-Pick C disease. *Proc. Natl. Acad. Sci. USA* 2006, 103, 13807–13812. [CrossRef]
- 67. Roth, A.; Looser, R.; Kaufmann, M.; Blättler, S.M.; Rencurel, F.; Huang, W.; Moore, D.D.; Meyer, U.A. Regulatory cross-talk between drug metabolism and lipid homeostasis: Constitutive androstane receptor and pregnane X receptor increase Insig-1 expression. *Mol. Pharmacol.* 2008, *73*, 1282–1289. [CrossRef] [PubMed]
- Monnerie, H.; Romer, M.; Jensen, B.K.; Millar, J.S.; Jordan-Sciutto, K.L.; Kim, S.F.; Grinspan, J.B. Reduced sterol regulatory element-binding protein (SREBP) processing through site-1 protease (S1P) inhibition alters oligodendrocyte differentiation in vitro. J. Neurochem. 2017, 140, 53–67. [CrossRef] [PubMed]
- 69. Stevenson, E.; Johns, E.; Marques, F.; Jackson, K.; Davern, P.; Evansc, R.; Head, G. Positive allosteric modulation of GABA-A receptors attenuates high blood pressure in Schlager hypertensive mice. *J. Hypertens.* **2017**, *35*, 546–557. [CrossRef] [PubMed]

- 70. Milivojevic, V.; Fox, H.C.; Sofuoglu, M.; Covault, J.; Sinha, R. Effects of progesterone stimulated Allopregnanolone on craving and stress response in cocaine dependent men and women. *Psychoneuroendocrinology* **2016**, *65*, 44–53. [CrossRef]
- Shimada, M.; Hasegawa, T.; Nishimura, C.; Kan, H.; Kanno, T.; Nakamura, T.; Matsubayashi, T. Anti-Hypertensive Effect of g-Aminobutyric Acid (GABA)-Rich Chlorella on High-Normal Blood Pressure and Borderline Hypertension in Placebo-Controlled Double Blind Study. *Clin. Exp. Hypertens.* 2009, *31*, 342–354. [CrossRef] [PubMed]
- 72. Schillaci, G.; Pirro, M.; Vaudo, G.; Gemelli, F.; Marchesi, S.; Porcellati, C.; Mannarino, E. Prognostic value of the metabolic syndrome in essential hypertension. *J. Am. Coll. Cardiol.* **2004**, *43*, 1817–1822. [CrossRef]
- 73. Grassi, G.; Seravalle, G.; Dell'Oro, R.; Quarti Trevano, F.; Bombelli, M.; Scopelliti, F.; Facchini, A.; Mancia, G. Comparative effects of candesartan and hydrochlorothiazide on blood pressure, insulin sensitivity, and sympathetic drive in obese hypertensive individuals: Results of the CROSS study. *J. Hypertens.* 2003, *21*, 1761–1769. [CrossRef] [PubMed]
- 74. Grassi, G.; Dell'Oro, R.; Quarti Trevano, F.; Scopelliti, F.; Seravalle, G.; Paleari, F.; Gamba, P.L.; Mancia, G. Neuroadrenergic and reflex abnormalities in patients with metabolic syndrome. *Diabetologia* **2005**, *48*, 1359–1365. [CrossRef]
- 75. Reaven, G.M.; Lithell, H.; Landsberg, L. Hypertension and associated metabolic abnormalities—the role of insulin resistance and the sympathoadrenal system. *N. Engl. J. Med.* **1996**, *334*, 374–381. [CrossRef] [PubMed]
- 76. Vollenweider, P.; Tappy, L.; Randin, D.; Schneiter, P.; Jequier, E.; Nicod, P.; Scherrer, U. Differential effects of hyperinsulinemia and carbohydrate metabolism on sympathetic nerve activity and muscle blood flow in humans. *J. Clin. Investig.* 1993, 92, 147–154. [CrossRef] [PubMed]
- Gudbjornsdottir, S.; Friberg, P.; Elam, M.; Attvall, S.; Lonnroth, P.; Wallin, B.G. The effects of metformin and insulin on sympathetic nerve activity, norepinehrine spillover and blood pressure in obese, insulin resistant, normoglycemic, hypertensive men. *Blood Press* 1994, 3, 394–403. [CrossRef] [PubMed]
- Maguire, J.; Stell, B.; Rafizadeh, M.; Mody, I. Ovarian cycle-linked changes in GABAA receptors mediating tonic inhibition alter seizure susceptibility and anxiety. *Nat. Neurosci.* 2005, *8*, 797–804. [CrossRef] [PubMed]