A longitudinal VBM study monitoring treatment with erythropoietin in patients with Friedreich ataxia

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Abstract

Background: Recombinant human erythropoietin (rhuEPO) has received considerable attention because of its neuroprotective properties. It has recently been reported that rhuEPO increases frataxin levels in combination with clinical improvement in rhuEPO treated patients with Friedreich ataxia (FRDA).

Purpose: To determine possible therapy dependent intracranial volume changes after treatment with rhuEPO using voxel-based morphometry (VBM).

Material and Methods: Nine FRDA patients were scanned on the same 1.5-Tesla MRI scanner before and after treatment with rhuEPO. FRDA patients received 5000 IU rhuEPO thrice weekly subcutaneously for a time period of 8 weeks followed by 2000 IU thrice weekly over 6 months. To test for re-test reliability a control group of 12 healthy volunteers were scanned twice on the same scanner without rhuEPO treatment. Neurological state was defined by the Friedreich Ataxia Rating Scale (FARS) and the Scale for the Assessment and Rating of Ataxia (SARA). Statistical parametric mapping software was used for image processing and statistical analysis.

Results: When comparing follow-up scans after rhuEPO treatment with baseline scans (P < 0.001 uncorrected) an increase of gray matter volume was observed bilaterally in the Pulvinar and the posterior parietal cortex. Moreover, clinical improvement detected using specific Ataxia scores correlated with VBM results in the pulvinar.

Conclusion: Given the limitation of a small sample size, our study confirms previous findings that MRI may serve as reliable biomarker in neurodegenerative diseases as well as in monitoring of microstructural changes representing disease progression and/or therapy effects.

Keywords

CNS, thalamus, parietal cortex, Friedreich ataxia, voxel-based morphometry, erythropoietin

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Introduction

Friedreich ataxia (FRDA) is the most common inherited ataxia and in most cases due to a gene repeat expansion which provides instructions for producing an enzyme called "glucosidase alpha acid" (GAA) on chromosome 9q13 coding for a mitochondrial protein named Frataxin (1).

Neuropathological studies have shown that FRDA is characterized by neuronal loss and white matter (WM) tract degeneration in the spinal cord, brainstem and cerebellum (2). A magnetic resonance imaging (MRI) study using voxel-based morphometry (VBM) in FRDA patients revealed symmetrical volume loss in dorsal medulla and cerebellum as compared to

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controls. Both atrophy of the cerebellum and medulla correlated with clinical severity and disease duration (3).

Recombinant human erythropoietin (rhuEPO) has received considerable attention because of its neuroprotective and cardioprotective properties (4–8). An *in vitro* study found that rhuEPO increases frataxin levels in isolated lymphocytes from FRDA patients. Subsequently, we were able to show significant increase in frataxin-levels in FRDA patients treated with rhuEPO in a two months "proof-of-concept" study and clinical improvement in a 6-month follow-up study (9–11). Furthermore a recent diffusion tensor imaging (DTI) study (12) revealed rather unspecific increase in fractional anisotropy (FA) in the cerebral hemispheres bilateraly in FRDA patients after rhuEPO treatment.

VBM is designed to detect significant regional MRI signal differences *in vivo* by applying voxel-wise statistics in the context of Gaussian random fields (13–15). The aim of this study was to detect therapy dependent intracranial volume changes using VBM in FRDA patients after treatment with rhuEPO.

Material and Methods

Subjects

Nine adult FRDA patients (6 men, 3 women, mean age at onset, 21.5 years; mean duration of disease, 14.5 years; GAA repeat length range between 240 and 1000) (Table 1) and a control group of 12 healthy volunteers (6 men, 6 women; mean age at examination, 25.4 years) were examined twice on the same MR scanner within this VBM study. Neurological state defined by the Friedreich Ataxia Rating Scale (FARS) in FRDA patients was 59.83 points (range, 36–86 points) (16). The Scale for the Assessment and Rating of Ataxia (SARA) revealed mean 20 points (range, 13–27 points) (17).

Study design

In an open-label, phase II, proof-of-concept study FRDA patients received 5000 IU rhuEPO thrice weekly subcutaneously for a time period of 8 weeks followed by 2000 IU rhuEPO thrice weekly over 6 months (9). The healthy control group received no rhuEPO.

MR assessment

MR assessment was performed according a predefined protocol before and after treatment with rhuEPO. Additional exclusion criteria for MR assessment were

 Table 1. Demographic and clinical parameters of study participants.

Age* (years)/Sex	GAA repeats	DD	SARA	FARS
26/M	650	7	18	56
26/M	1000	12	26.5	86
46/M	800	14	23.5	68
51/F	190	15	20.5	44.5
29/F	320	9	21	52
32/M	370	10	14.5	36
29/F 1000		13	25.5	83
18/M	500		13	49
41/M	420	21	19	64

Patient ID in accordance with prior publication on this sample (11). *Age at study entry.

DD, disease duration (years); FARS, Friedreich Ataxia Rating Scale; GAA, number of trinucleotide repeats on the shorter allele; SARA, Scale for the Assessment and Rating of Ataxia.

metal in the body that precludes MRI and agoraphobia. The MR assessment of the healthy control group was performed in an interval of 2 months (baseline scan, follow-up scan 2 months later). All MRI scans were proofed by an experienced radiologist (MS) to avoid structural abnormalities or artifacts.

Data acquisition

All participants were scanned on the same 1.5 Tesla MRI scanner (Magnetom Avanto, Siemens, Erlangen, Germany) using a T1-weighted 3D sequence with a repetition time (TR) of 9.7 ms, an echo time (TE) of 4 ms, a matrix size of 256×256 , and a field of view of 230 mm. The resulting sagittal slices had a thickness of 1.5 mm and an in-plane resolution of 0.98×0.98 mm.

Voxel based morphometry (VBM)

Statistical parametric mapping software (SPM2; The Wellcome Department of Cognitive Neurology, London, UK) was used for image processing and statistical analysis. We applied the optimized VBM protocol to the image data as reported by Good et al. (15). This protocol includes a study specific template, extraction of miss-segmented areas and the modulation of the data with the Jacobian determinants.

The creation of the study group-specific template was performed to minimize the scanner specific bias by averaging all images from the study specific subject group, after being normalized using linear 12-parameter affine-only transformation. Custom tissue probability maps were obtained by segmenting the individual normalized images into gray matter (GM), white matter (WM), and cerebrospinal fluid (CSF), averaging and smoothing with an isotropic Gaussian kernel of 8 mm full-width at half maximum (FWHM).

The optimized VBM protocol includes two segmentation steps. First segmentation was performed in native space and non-brain tissue was removed automatically by modulation with an individually derived brain-tissue mask. Second segmentation was performed after applying the normalization parameters to the original whole brain images (as described below) including once again removing of non-brain tissue followed by reslicing onto a voxel size of $1 \times 1 \times 1$ mm.

The spatial normalization parameters were estimated by matching the native spaced individual GM image with the study-specific GM template using combined 12 parameter linear and non-linear $(7 \times 9 \times 7)$ basic functions) transformation. The parameters resulting from this spatial normalization step were then reapplied to the original structural images.

Multiplying voxel values of the segmented images with the Jacobian determinants was implemented to convert the GM segments into measures of absolute gray matter volume (GMV), as opposed to relative GMV, following spatial normalization.

Finally, all modulated images were smoothed with a 10 mm FWHM Gaussian kernel to reduce confounding by individual variation in gyral anatomy. This also has the effect of rendering the data more normally distributed (by the central limit theorem) (13,14).

Smoothed and segmented gray matter partitions have been subjected for statistical analysis choosing the factorial design "paired t-test".

The statistical parametric maps were thresholded at P uncorrected <0.001 with an extent threshold of 20 contiguous voxels. Finally, smoothed GM segments were entered into a voxel-based multiple regression analysis to investigate linear correlations between GM volume and primary clinical outcome measures (FARS and SARA) (10).

Results

When comparing the follow-up scans after treatment with recombinant human erythropoietin (rhuEPO) with the baseline scans an increase of GMV (Table 2) was observed bilaterally in the posterior part of the thalamus (pulvinar) (Fig. 1) and in the posterior parietal lobe (Brodmann area 7) (Fig. 2) (P < 0.001uncorrected).

There were no regions showing significant decrease in GMV, and there was no significant group difference in white matter volume (WMV).

Correlation analysis (Spearman rank correlation analysis performed with SPSS, Version 15; SPSS Inc., Chicago, IL, USA) for FRDA patients revealed an **Table 2.** Relative increase of gray matter volume in patients with Friedreich ataxia after treatment with recombinant human erythropoietin (rHuEPO) relative to baseline examination before treatment (P < 0.001 uncorrected).

			Significant differences in gray matter			
			MNI coordinates*		tes*	Peak gray matter change
Region		BA	x	у	z	Z-score
Parietal lobe						
	Right	7	21	-69	45	3.50
	Left	7	-9	-76	50	4.16
Thalamus/						
Pulvinar	Right		18	-28	3	3.41
	Left		-16	-3 I	3	3.61

*Coordinates are given in MNI space (Montreal Neurological Institute).

association between age at onset of disease and GAA repeat length (R = -0.927, P = 0.003). Voxel-based multiple regression analysis showed that there were significant correlations between GMV in rather unspecific cortical and subcortical structures and individual clinical ataxia scores such as SARA and FARS. Additionally, scores in SARA and FARS showed a significant negative correlation (P < 0.001 uncorrected) with increased GMV in the pulvinar and posterior parietal cortex bilaterally. The healthy control group showed no significant change of GMV (P < 0.001 uncorrected).

Discussion

In the last years, the non-erythroid, especially neuroprotective functions of EPO have been increasingly recognized. These functions probably base on suppression of apoptosis and promotion of neurogenesis, as suggested by several experimental animal studies (18–26).

In the present study, we report on a volume increase in the dorsal thalamus (Pulvinar) measured with the help of VBM in a patient collective with FRDA, receiving EPO over a period of several months. As our initial VBM analysis had shown no changes in primary disease-specific areas (e.g. cerebellum), we also looked for supratentorial changes, in accordance with previously published DTI results (12) within the same study collective. This pulvinar volume increase correlated with an improvement in ataxia rating scales. Additional significant volume increase was found in the posterior parietal cortex, where higher association areas are in permanent interaction with the pulvinar.



Fig. 1. Areas of subcortical GMV increase when baseline MR scans of patients with Friedreich ataxia are compared to the follow-up scans after treatment with recombinant human erythropoietin (rHuEPO). Results are superimposed on the slices of a TI-weighted mean-picture (a) axial, (b) coronal, and on the slices of a modulated mean picture (c) sagital, (d) coronal, (e) axial of all study participants. The color bars give the corresponding Z-scores. Threshold was set at T = 3.4 (corresponding to a *P* value of <0.001 uncorrected).



Fig. 2. Results in SPM render-style show areas of cortical GMV increase when baseline MR scans of patients with Friedreich ataxia are compared to the follow-up scans after treatment with recombinant human erythropoietin (rHuEPO). Threshold was set at T = 3.4 (corresponding to a *P* value of <0.001 uncorrected).

Parietal association areas play a major role in linking incoming sensory information with the primary motor area. This is of major interest, as FRDA is an autosomalrecessive neurodegenerative disorder mainly affecting sensory neurons in the dorsal root ganglia as well as the spinocerebellar tract, pyramidal tract and the dorsal columns (27). Underlying mechanisms of dynamic structural alterations detected by VBM are not entirely clear. Macroscopic alterations may be based on changes at the level of synaptic bulk, or they might include increased cell genesis of glial and/or neuronal cells. EPO has been shown to enhance the levels of BDNF (brain derived neurotropic factor) (28) which results in an improvement of the mTOR (mammalian target of rapamycin) pathway, representing an integral cell growth regulator (29), which might result in changes of cellular volume.

Another study administering large systemic doses of rhEPO in mice revealed transient enhancement of hippocampal neurogenesis (30).

PET studies investigating glucose metabolism in FRDA patients have shown that glucose metabolism is initially increased in several brain regions like brain stem, cerebellum, and thalamus and declines with disease severity (31). Alternatively, altered regional volume as measured in our VBM study might therefore be related to an improvement of energy supply in the dorsal thalamus.

Since our healthy control group showed no significant changes over time, it can be assumed that VBM with longer intra-individual scan intervals is a stable method. This finding is in accordance with a recent study showing that regional GMV was relatively stable within the same scanner, but can significantly change when comparing images acquired on different scanners (32).

Given the limitation of small sample size in this pilot study further confirmation of our findings are of major interest. Future interventional animal studies combining structural and functional MRI with histological verification are needed to enlighten the impact of MRI in monitoring of microstructural changes representing disease progression and/or therapy effects.

In conclusion, our study confirms previous findings that MRI may serve as reliable biomarker in neurodegenerative diseases as well as in monitoring of microstructural changes representing disease progression and/or therapy effects.

References

- 1. Klockgether T. *Handbook of ataxia disorders*. New York: Marcel Dekker, 2000.
- Lowe J, Lennox G, Leigh PN. Disorders of movement and system degeneration. Greenfield's Neuropathology, 6th edn. New York: Oxford University Press, 1997.
- Della Nave R, Ginestroni A, Giannelli M, et al. Brain structural damage in Friedreich's ataxia. J Neurol Neurosurg Psychiatry 2008;79:82–85.
- 4. Bogoyevitch MA. An update on the cardiac effects of erythropoietin cardioprotection by erythropoietin and the lessons learnt from studies in neuroprotection. Cardiovasc Res 2004;63:208–216.
- Juul S. Erythropoietin in the central nervous system, and its use to prevent hypoxic-ischemic brain damage. Acta Paediatr Suppl 2002;91:36–42.
- 6. Li W, Maeda Y, Yuan RR, Elkabes S, et al. Beneficial effect of erythropoietin on experimental allergic encephalomyelitis. Ann Neurol 2004;56:767–777.

- Siren AL, Ehrenreich H. Erythropoietin–a novel concept for neuroprotection. Eur Arch Psychiatry Clin Neurosci 2001;251:179–184.
- 8. Smith KJ, Bleyer AJ, Little WC, et al. The cardiovascular effects of erythropoietin. Cardiovasc Res 2003;59: 538–548.
- Boesch S, Sturm B, Hering S, Goldenberg H, et al. Friedreich's ataxia: clinical pilot trial with recombinant human erythropoietin. Ann Neurol 2007;62:521–524.
- Boesch S, Sturm B, Hering S, et al. Neurological effects of recombinant human erythropoietin in Friedreich's ataxia: a clinical pilot trial. Mov Disord 2008;23: 1940–1944.
- 11. Sturm B, Stupphann D, Kaun C, et al. Recombinant human erythropoietin: effects on frataxin expression in vitro. Eur J Clin Invest 2005;35:711–717.
- Egger K, Clemm von Hohenberg C, Schocke MF, et al. White Matter Changes in Patients with Friedreich Ataxia after Treatment with Erythropoietin. J Neuroimaging Epub ahead of print 9 September 2013. doi: 10.1111/ jon.12050.
- 13. Ashburner J, Andersson JL, Friston KJ. Image registration using a symmetric prior-in three dimensions. Hum Brain Mapp 2000;9:212-225.
- Ashburner J, Friston KJ. Voxel-based morphometry-the methods. Neuroimage 2000;11:805–821.
- 15. Good CD, Johnsrude IS, Ashburner J, et al. A voxelbased morphometric study of ageing in 465 normal adult human brains. Neuroimage 2001;14:21–36.
- Schmitz-Hubsch T, du Montcel ST, Baliko L, et al. Scale for the assessment and rating of ataxia: development of a new clinical scale. Neurology 2006;66:1717–1720.
- Subramony SH. SARA–a new clinical scale for the assessment and rating of ataxia. Nat Clin Pract Neurol 2007;3:136–137.
- Liu XB, Wang JA, Yu SP, et al. Therapeutic strategy of erythropoietin in neurological disorders. CNS Neurol Disord Drug Targets 2008;7:227–234.
- Grasso G, Sfacteria A, Meli F, et al. The role of erythropoietin in neuroprotection: therapeutic perspectives. Drug News Perspect 2007;20:315–320.
- Iwai M, Cao G, Yin W, et al. Erythropoietin promotes neuronal replacement through revascularization and neurogenesis after neonatal hypoxia/ischemia in rats. Stroke 2007;38:2795–2803.
- Liu XF, Fawcett JR, Thorne RG, et al. Non-invasive intranasal insulin-like growth factor-I reduces infarct volume and improves neurologic function in rats following middle cerebral artery occlusion. Neurosci Lett 2001; 308:91–94.
- Xiong Y, Lu D, Qu C, et al. Effects of erythropoietin on reducing brain damage and improving functional outcome after traumatic brain injury in mice. J Neurosurg 2008;109:510–521.
- Yu YP, Xu QQ, Zhang Q, et al. Intranasal recombinant human erythropoietin protects rats against focal cerebral ischemia. Neurosci Lett 2005;387:5–10.
- 24. Siren AL, Radyushkin K, Boretius S, et al. Global brain atrophy after unilateral parietal lesion and its prevention by erythropoietin. Brain 2006;129:480–489.

- 25. Sargin D, Hassouna I, Sperling S, et al. Uncoupling of neurodegeneration and gliosis in a murine model of juvenile cortical lesion. Glia 2009;57:693–702.
- 26. Taoufik E, Petit E, Divoux D, et al. TNF receptor I sensitizes neurons to erythropoietin- and VEGF-mediated neuroprotection after ischemic and excitotoxic injury. Proc Natl Acad Sci U S A 2008;105:6185–6190.
- Fogel BL, Perlman S. Clinical features and molecular genetics of autosomal recessive cerebellar ataxias. Lancet Neurol 2007;6:245–257.
- Viviani B, Bartesaghi S, Corsini E, et al. Erythropoietin protects primary hippocampal neurons increasing the expression of brain-derived neurotrophic factor. J Neurochem 2005;93:412–421.
- Lee CH, Inoki K, Guan KL. mTOR pathway as a target in tissue hypertrophy. Annu Rev Pharmacol Toxicol 2007;47:443–467.

- Ransome MI, Turnley AM. Systemically delivered Erythropoietin transiently enhances adult hippocampal neurogenesis. J Neurochem 2007;102:1953–1965.
- Gilman S, Junck L, Markel DS, et al. Cerebral glucose hypermetabolism in Friedreich's ataxia detected with positron emission tomography. Ann Neurol 1990;28: 750–757.
- Takao H, Hayashi N, Ohtomo K. Effects of the use of multiple scanners and of scanner upgrade in longitudinal voxel-based morphometry studies. J Magn Reson Imaging 2013;38:1283–1291.