## nature portfolio

Jennifer Zagelbaum, Allana Schooley, Junfei Zhao, Benjamin R. Schrank, Elsa Callen, Shan Zha, Max E. Gottesman, André Nussenzweig,

Corresponding author(s):

Raul Rabadan, Job Dekker, and Jean Gautier

Last updated by author(s): Sep 14, 2022

## **Reporting Summary**

Nature Portfolio wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Portfolio policies, see our <u>Editorial Policies</u> and the <u>Editorial Policy Checklist</u>.

For all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section

٠.	ナつ	+	-	ics
_ ``	_			

n/a	Confirmed
	$\square$ The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement
	A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
	The statistical test(s) used AND whether they are one- or two-sided  Only common tests should be described solely by name; describe more complex techniques in the Methods section.
$\boxtimes$	A description of all covariates tested
	A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
	A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
	For null hypothesis testing, the test statistic (e.g. <i>F</i> , <i>t</i> , <i>r</i> ) with confidence intervals, effect sizes, degrees of freedom and <i>P</i> value noted <i>Give P values as exact values whenever suitable.</i>
$\boxtimes$	For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings
$\boxtimes$	For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes
	Estimates of effect sizes (e.g. Cohen's <i>d</i> , Pearson's <i>r</i> ), indicating how they were calculated
	Our was collection on statistics for high gives contains articles on many of the points above

## Software and code

Policy information about availability of computer code

Data collection

Illumina HiSeq 4000 machine (Paired End 50 bp module), Illumina MiSeq sequencer, NIS-elements, MetaCyte software (Metasystems, version 3.10.6)

Data analysis

Data was mapped to mm10 reference genome using BWA-MEM (BWA-0.7.17 (r1188)). For Hi-C analyses, Python 3.7.10, matplotlib, MACS3 (https://github.com/macs3-project/MACS), and HOMER were used. For HTGTS analyses, HTGTS pipeline (https://github.com/robinmeyers/transloc\_pipeline), R package ChIPseeker, and GSEAPreranked were used.

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Portfolio guidelines for submitting code & software for further information.

## Data

Policy information about availability of data

All manuscripts must include a data availability statement. This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A description of any restrictions on data availability
- For clinical datasets or third party data, please ensure that the statement adheres to our policy

High throughput sequencing data have been deposited to Gene Expression Omnibus under accession number GSE183059

Human resea	arch parti	cipants				
Policy information a	about <u>studies i</u>	nvolving human research participants and Sex and Gender in Research.				
Reporting on sex	and gender	N/A				
Population characteristics		N/A				
Recruitment		N/A				
Ethics oversight		N/A				
Note that full informa	tion on the appı	roval of the study protocol must also be provided in the manuscript.				
	. 6.					
Field-spe	citic re	porting				
Please select the or	ne below that i	s the best fit for your research. If you are not sure, read the appropriate sections before making your selection.				
X Life sciences		Behavioural & social sciences Ecological, evolutionary & environmental sciences				
For a reference copy of th	he document with	all sections, see <u>nature.com/documents/nr-reporting-summary-flat.pdf</u>				
Life scien	ices sti	udy design				
All studies must disc	close on these	points even when the disclosure is negative.				
Sample size	For Hi-C experiments, 2 biological replicates were performed. For all other data, experiments were performed in at least triplicate					
Data exclusions	No data was ex	xcluded from analysis				
Replication	All experiments have at least two independent biological replicates. All findings described in the manuscript were confirmed in all individual replicates.					
Randomization	Randomization of this study was not necessary as we did not allocate datasets into experimental groups.					
Blinding	Blinding is not required . Results were directly linked with the data and this is not a clinical or genetic study with large numbers of samples.					
Doportin	a for s	accific materials systems and mathods				
		pecific materials, systems and methods				
		about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.				
Natariala Carre		N/ath ada				
Materials & experimental systems  n/a   Involved in the study  Methods  n/a   Involved in the study						
n/a   Involved in the study						
Eukaryotic o	cell lines	Flow cytometry				
Palaeontolo	ogy and archaed					
Animals and	d other organisr					
☐ Clinical data						
Dual use re	search of conce	rn				
Eukaryotic ce	ell lines					
		and Sex and Gender in Research				
Cell line source(s)	icy information about <u>cell lines and Sex and Gender in Research</u> Well line source(s)  Mouse embryonic fibroblasts					
Authentication						
Authentication Centines were authenticated by A. Nussenzweig (MEF)						

Mycoplasma contamination

Cells tested negative for mycoplasma